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(19) **United States**(12) **Patent Application Publication****Van Der Werf et al.**(10) **Pub. No.: US 2007/0128224 A1**(43) **Pub. Date: Jun. 7, 2007**(54) **NOVEL STRAIN OF SARS-ASSOCIATED
CORONAVIRUS AND APPLICATIONS
THEREOF**

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(52) **U.S. Cl.** **424/221.1**; 435/5; 435/69.3;
435/326; 435/456; 530/350;
530/388.3; 536/23.72; 977/802

(57)

ABSTRACT

The invention relates to a novel strain of severe acute respiratory syndrome (SARS)-associated coronavirus, resulting from a sample collected in Hanoi (Vietnam), reference number 031589, nucleic acid molecules originating from the genome of same, proteins and peptides coded by said nucleic acid molecules and, more specifically, protein N and the applications thereof, for example, as diagnostic reagents and/or as a vaccine.

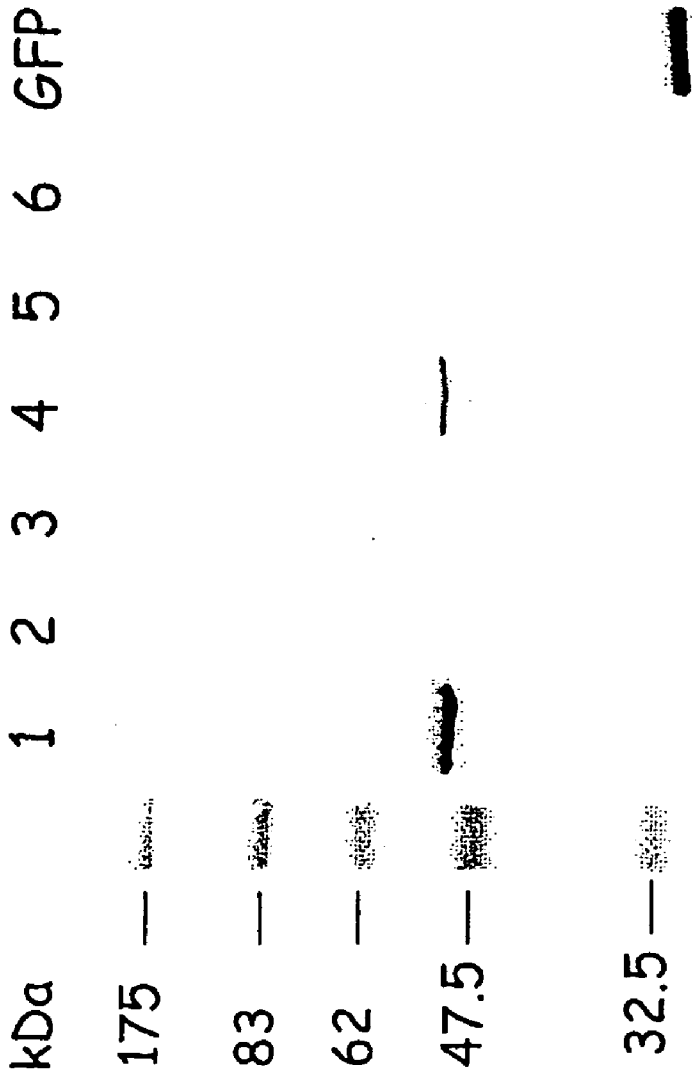


FIGURE 1

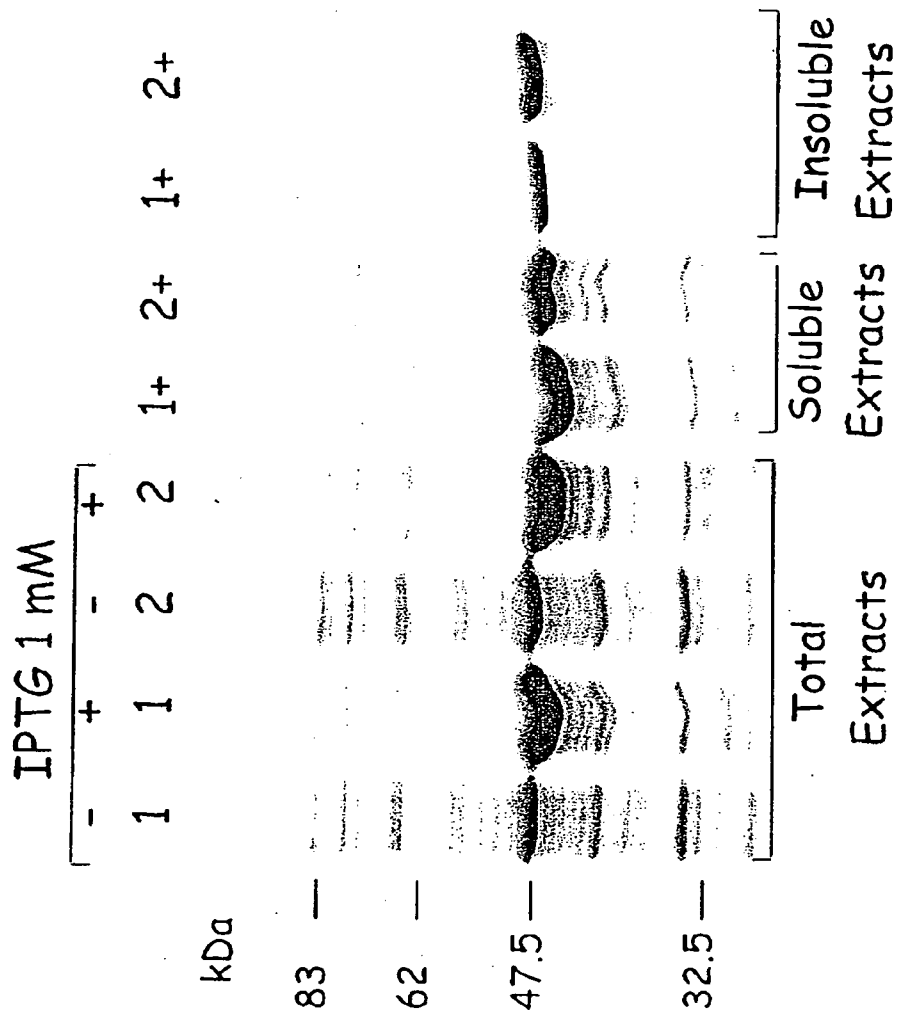


FIGURE 2

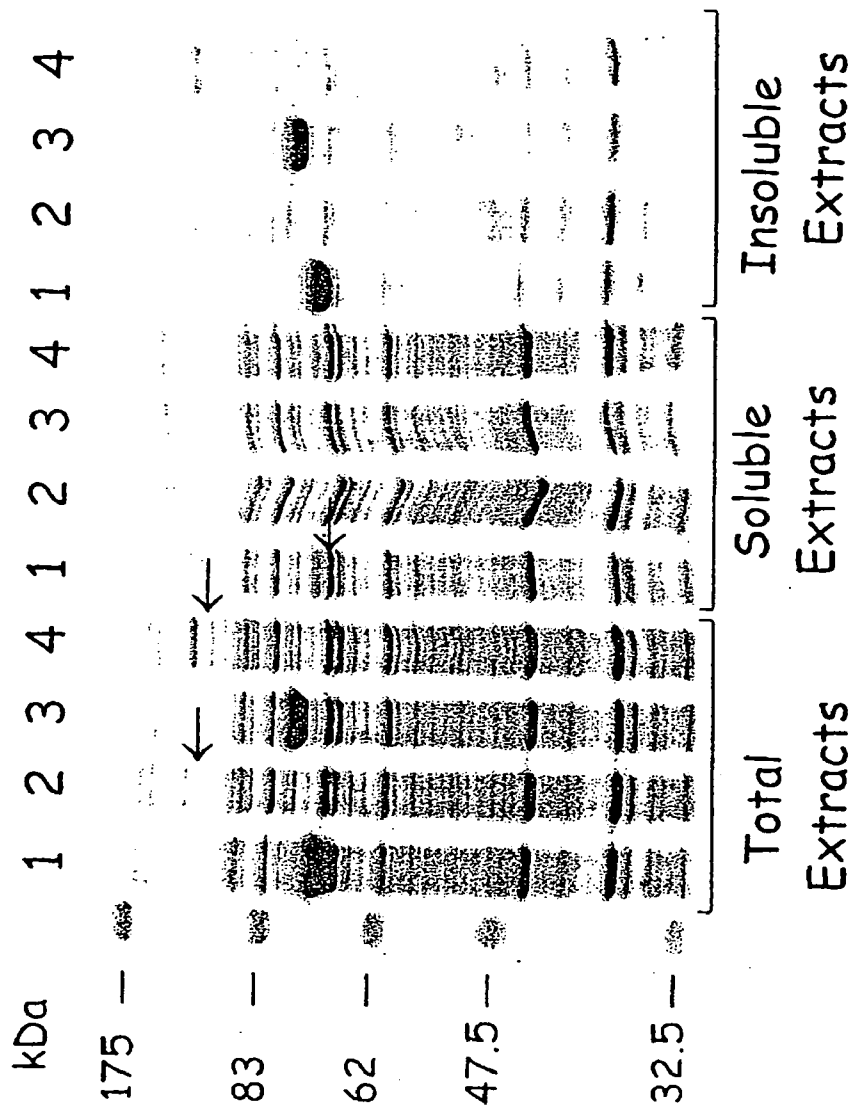


FIGURE 3

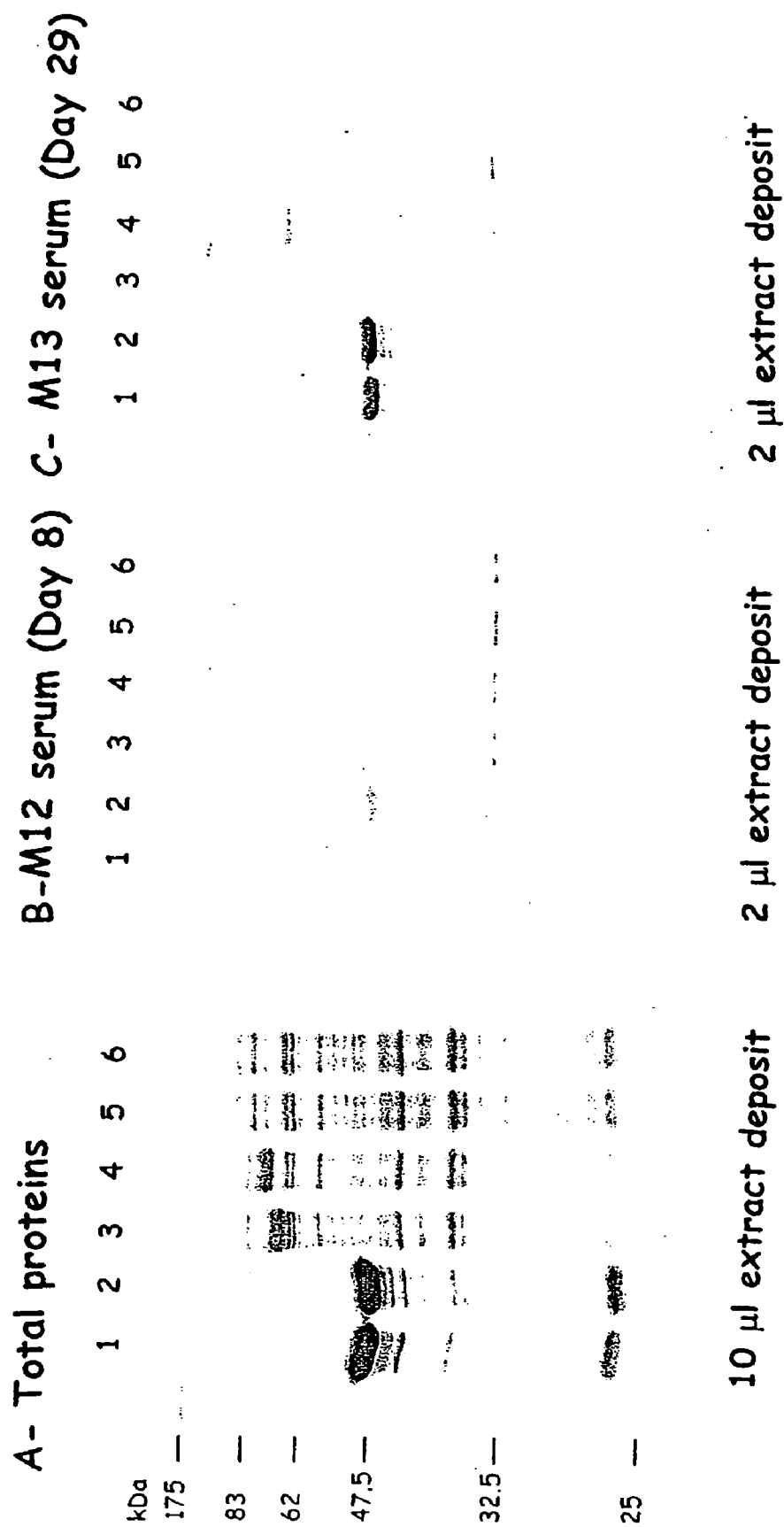


FIGURE 4

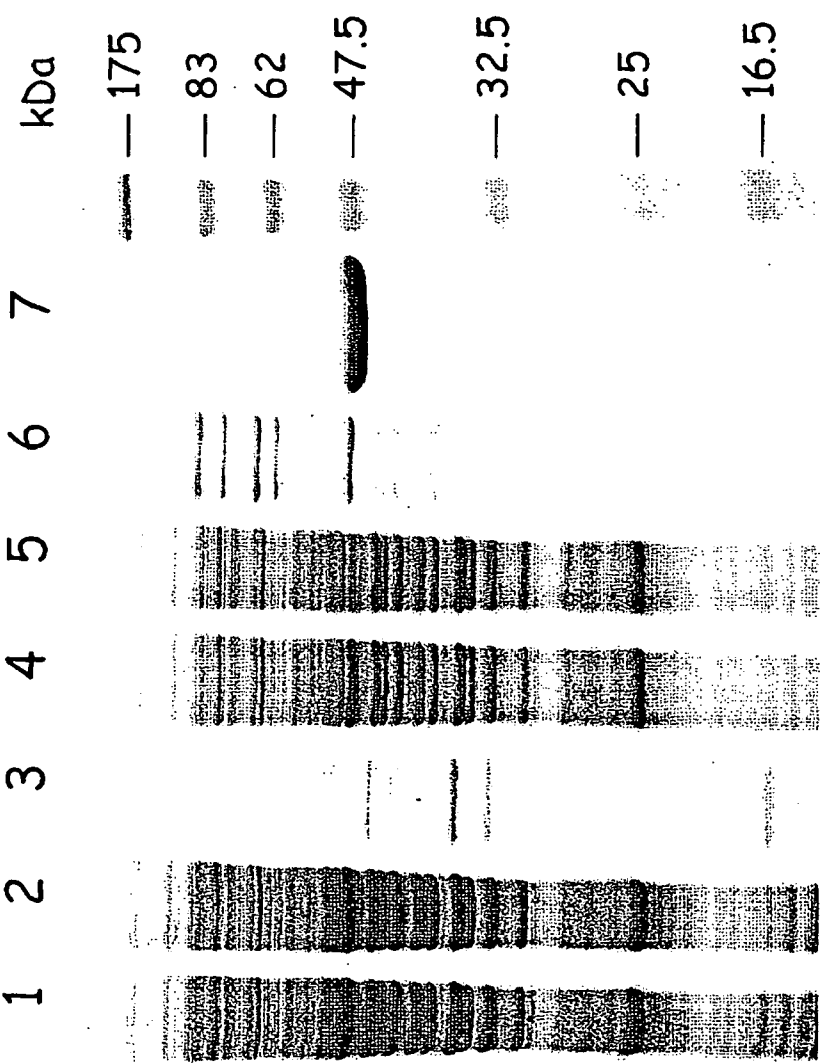


FIGURE 5

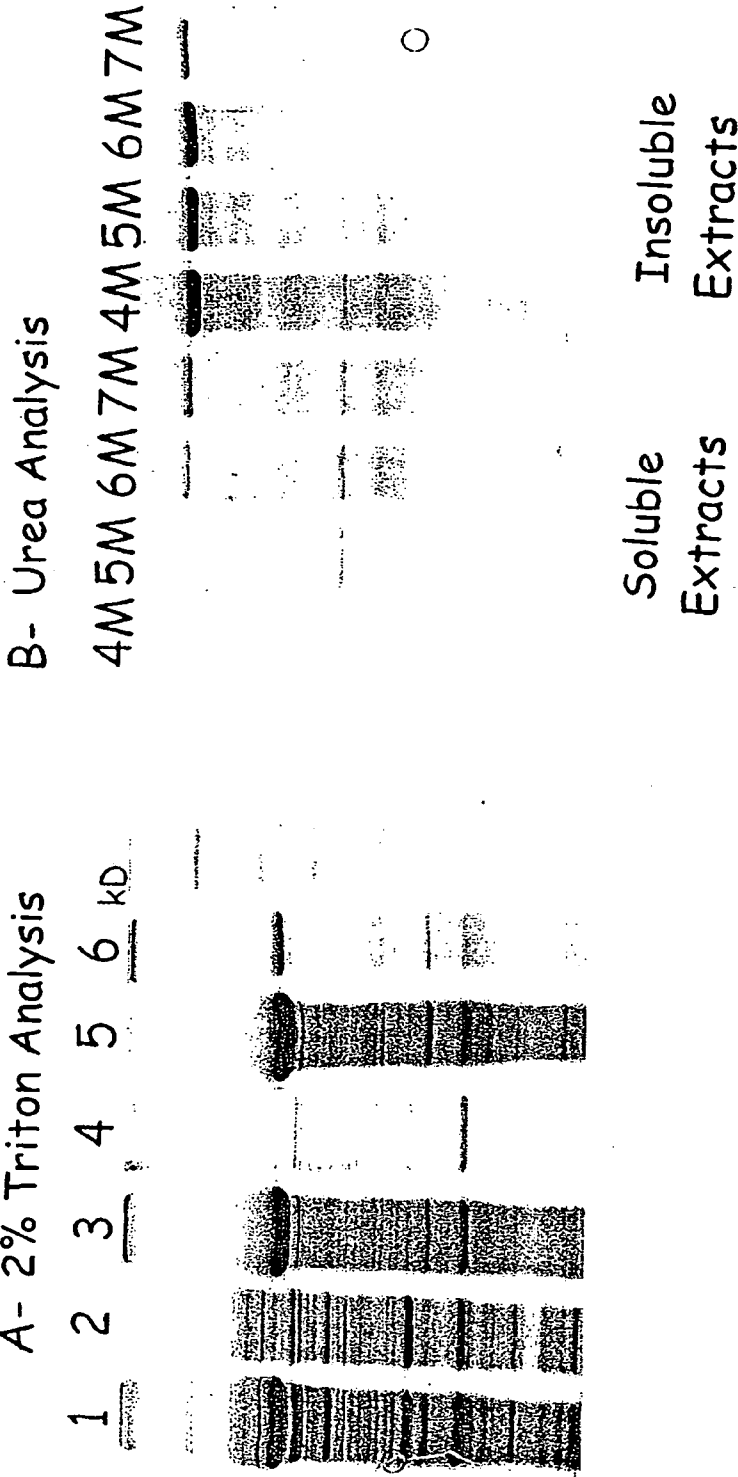


FIGURE 6

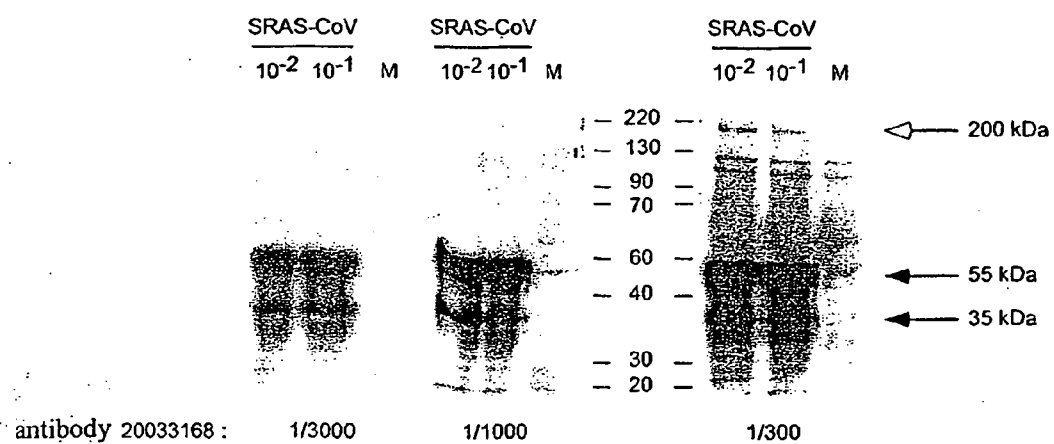


FIGURE 7

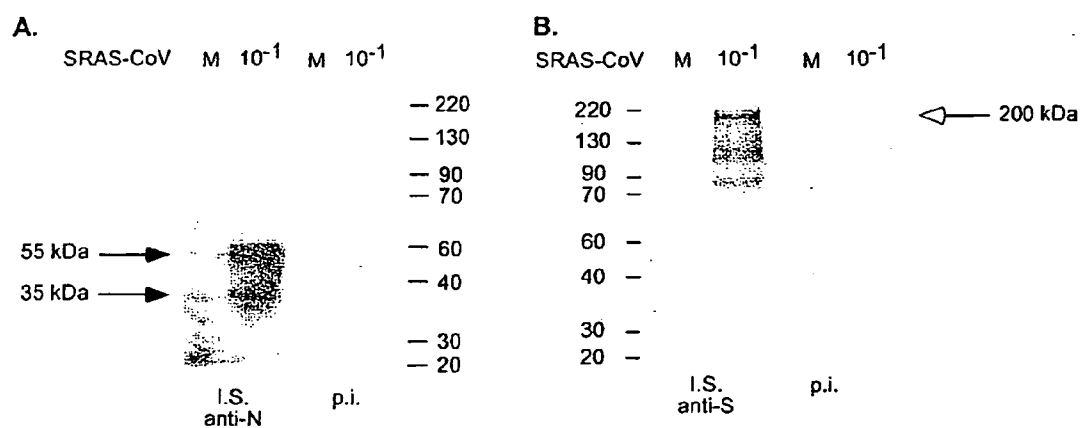
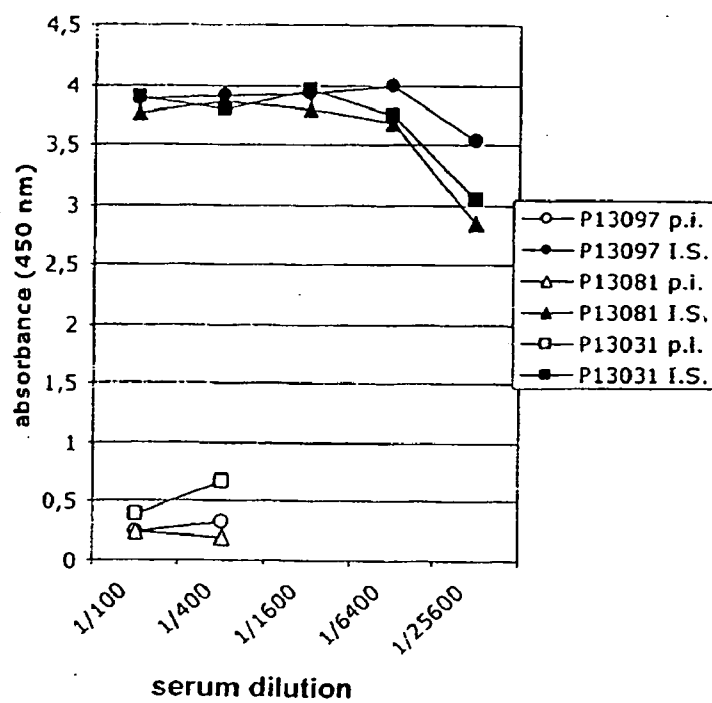


FIGURE 8

A



B

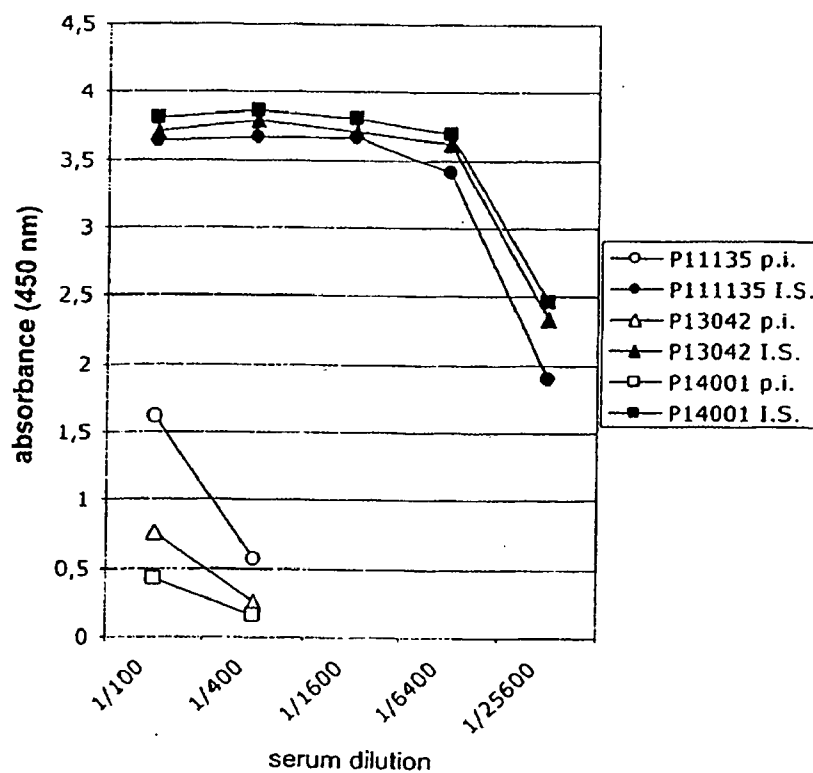


FIGURE 9

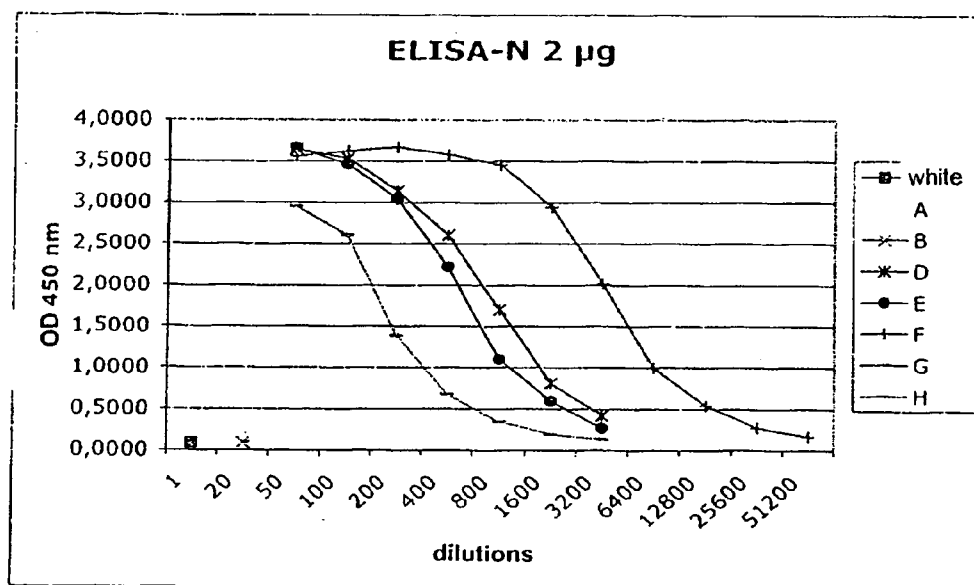
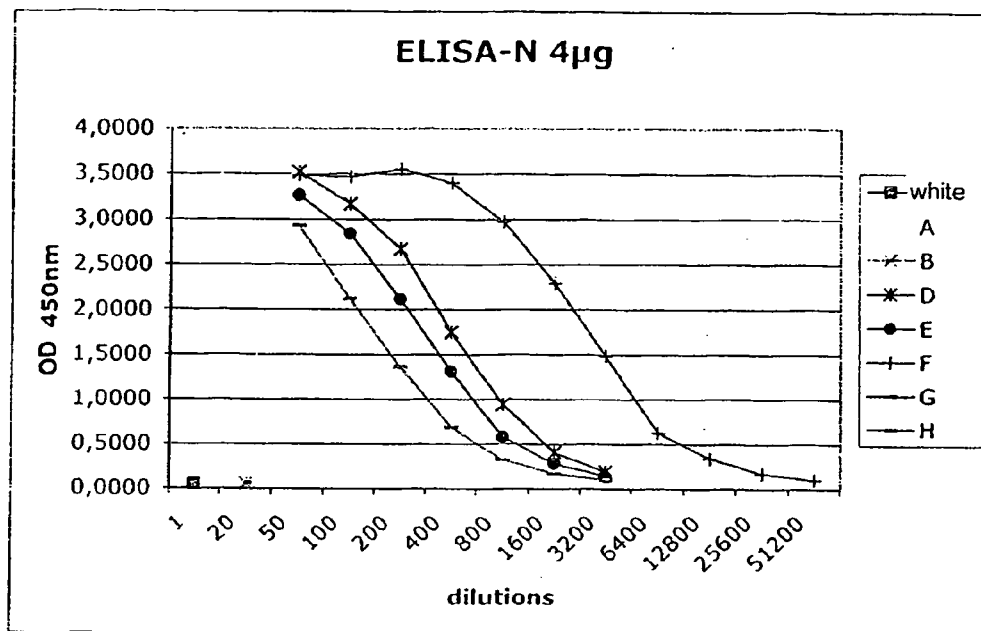


FIGURE 10a

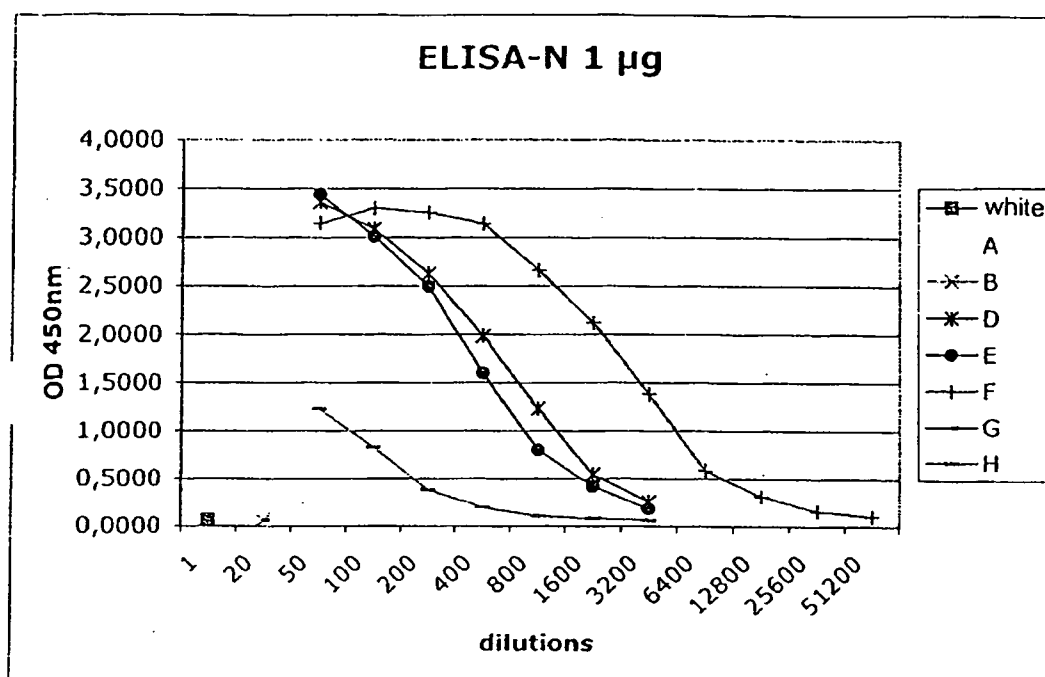


FIGURE 10b

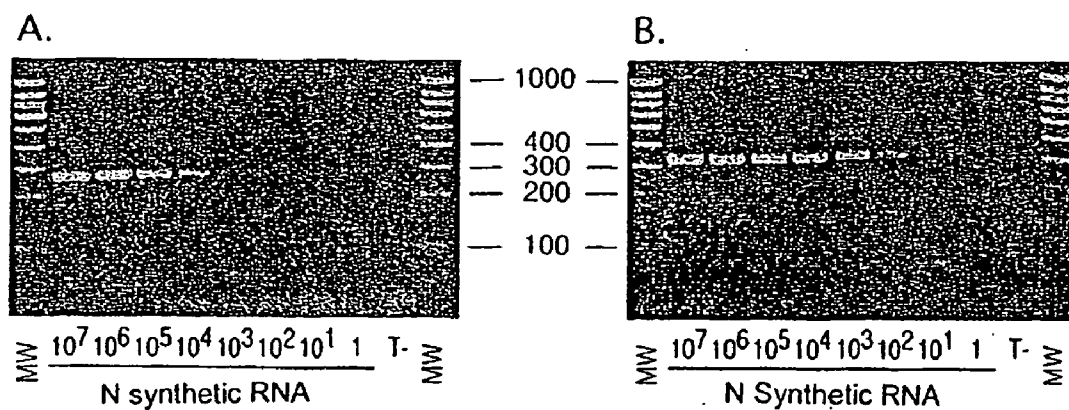


FIGURE 11

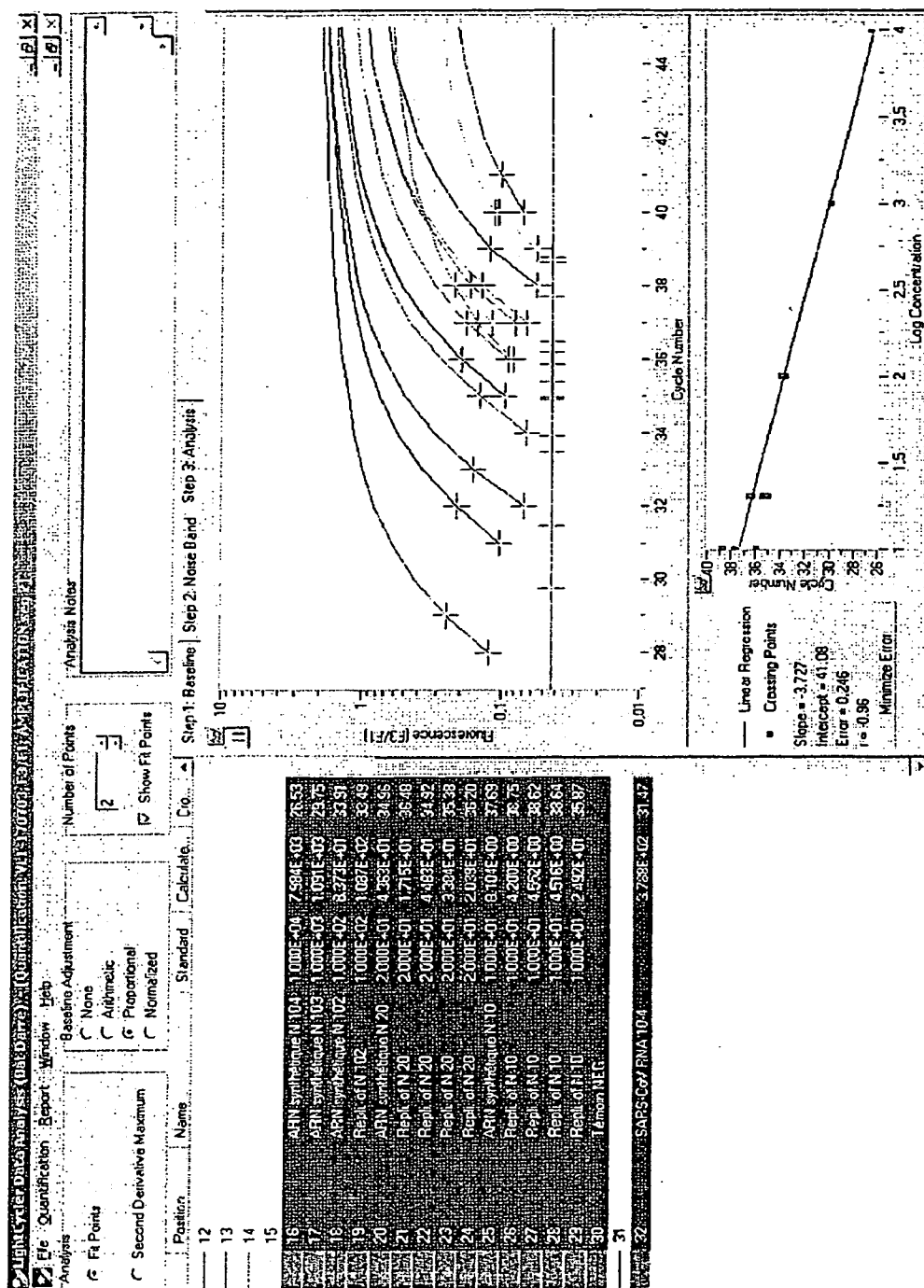


FIGURE 12

							>> RmaI
	>> HindII	>> MaeII>	< Eco57I		>> Esp3I	>> MaeII	
	>> HincII	>	< AflIII>	< DdeI	>> BsmAI		>> MaeI
TTGGTGTCAA	CGAGAAACA	CACGTCCAAC	TCAGTTTGCC	TGTCCTTCAG	GTTAGAGACG	TGCTAGTGC	
290	300	310	320	330	340	350	

FIGURE 13.1

```

    >< Sau96I
    >< PssI
    >< Pali
    >< NspIV
    >< MnlI
    >< HaeIII
    >< EcoO109I
    >< DraII>< MboII >< PmlI
    >< MnlI >< CfrI3I >< PmaCI
    >< Ksp632I >< BsuRI >< MaeII
    >< HinfI >< BsiZI>< EcoNI >< Eco72I
    >< EarI >< BshI >< BslI >< BsaAI
    >< PleI >< Eam1104I>< AsuI >< BsiYI>< BbrPI >< MnlI
TGGCTTCGGG GACTCTGTGG AAGAGGCCCT ATCGGAGGCA CGTGAACACC TCAAAAATGG CACTTGTGGT
    360      370      380      390      400      410      420

    >< Tru9I
    >< RsaI >< SfaNI
    >< RnaI >< Csp6I >< BspWI >< MseI
    >< MaeI >< AluI >< AfaI >< AluI >< MaeII
CTAGTAGAGC TGGAAAAAGG CGTACTGCCC CAGCTTGAAC AGCCCTATGT GTTCATTAAA CGTTCTGATG
    430      440      450      460      470      480      490

    >< Pali
    >< HaeIII
    >< Tru9I >< GdiII >< RsaI
    >< MseI >< EaeI McrI ><
    >< Esp4I >< BsuRI >< BsmI BsiEI ><
    >< AflII >< BshI >< AluI >< BscCI >< AfaI
CCTTAAGCAC CAATCACGGC CACAAGGTCG TTGAGCTGGT TGCAGAAATG GACGGCATTG AGTACGGTCG
    500      510      520      530      540      550      560

    >< NspI
    >< ScaI >< NspHI
    >< RsaI >< NlaIII
    >< Csp6I >< BslI
    >< BsrI >< BsiYI >< MboII
    >< AciI >< AfaI >< AflIII >< MunI >< AciI
TAGCGGTATA ACATGGGAG TACTCGTGCC ACATGTGGGC GAAACCCCAA TTGCATACCG CAATGTTCTT
    570      580      590      600      610      620      630

    >< TthHB8I
    >< TaqI
    >< Sau3AI
    >< NdeII
    >< MboI
    >< DpnII
    >< DpnI
    >< ClaI
    >< Bsu15I
    >< BspOI
    >< NlaIV
    >< BspAI
    >< MspI >< Bsp143I
    >< HpaII >< Bsp106I
    >< HapII >< BsiXI MaeIII >
    >< Cfr10I >< BscI>< SfaNI DdeI ><
    >< BscBI >< AluI >< BanIII BfrI ><
CTTCGTAAGA ACGGTAATAA GGGAGCCGGT GGTCATAGCT ATGGCATCGA TCTAAAGTCT TATGACTTAG
    640      650      660      670      680      690      700

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FIGURE 13.2

```

>< Sau3AI
>< NdeII
>< MboI
>< HphI
>< DpnII
>< BspAI
>< AlwI>< DpnI
>< AluI >< Bsp143I
>< MboII >< BsrI
>< DdeI
>< SstI
>< SduI
>< SacI
>< NspII
>< MnlI
>< HgiAI
>< Eco24I
>< TthHB8I
>< TaqI
>< SalI
>< RtrI
>< HindII
>< HincII
>< BsgI
>< AluI
>< MaeIII
>< AccI
>< ThaI
>< ThaI
>< MvnI
>< MvnI
>< HinPII
>< Hin6I
>< HhaI
>< CfoI
>< BstUI
>< BstUI
>< Bsp50I
>< Bsp50I
>< AciI
>< AccII
>< MnlI
>< SfaNI
>< AccII
>< TthHB8I
>< TthHB8I
>< TaqI
>< TaqI
>< MnlI
>< Ksp632I
>< HinfI>< PleI
>< Eam1104I
>< EarI >< BbvI>< AccI >< Fnu4HI
>< MboII >< MaeIII
>< NlaIII ><
>< NlaIII
>< EcoRII ><
>< DsaV ><
>< TthHB8I
>< TaqI
>< SfuI
>< NspV>< Tru9I
>< LspI>< MseI

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FIGURE 13.3

```

>< MvaI      >< Hin6I      >< SduI      >< Csp45I
>< Ecl136I   >< HhaI      >< NspII     >< BstBI
>< BstOI     >< MaeII     >< HgiAI     >< Bsp119I
>< BstNI     >< Eco47III   >< Bsp1286I  >< BsiCI
>< BsiLI     >< CfoI      >< BmyI      >< Bpu14I
>< ApyI >< DdeI >< Bsp143II >< AluI   >< Alw21I   >< AsuII
CTGGTTCAC T GAGCGCTCTG ATAAGAGCTA CGAGCACCAG ACACCCTTCG AAATTAAGAG TGCCAAGAAA
990      1000      1010      1020      1030      1040      1050

                                >< Tru9I
                                >< MseI
                                >< BsmI
                                >< BscCI
                                >< MnlI
TTTGACACTT TCAAAGGGGA ATGCCCAAAG TTTGTGTTTC CTCTTAACTC AAAAGTCAAA GTCATTCAAC
1060      1070      1080      1090      1100      1110      1120

>< PmlI
>< PmaCI
>< MaeII
>< Eco72I
>< BsaAI      >< NlaIII      >< RsaI
>< BbrPI      >< Bst1107I >< Csp6I
>< AflIII     >< MnlI>< DdeI     >< AccI      >< AfaI
CACGTGTTGA AAAGAAAAAG ACTGAGGGTT TCATGGGGCG TATACGCTCT GTGTACCCTG TTGCATCTCC
1130      1140      1150      1160      1170      1180      1190

>< SfaNI
>< MaeIII     >< AccI      NlaIII ><
ACAGGAGTGT AACAATATGC ACTTGTCTAC CTTGATGAAA TGTAATCATT GCGATGAAGT TTCATGGCAG
1200      1210      1220      1230      1240      1250      1260

                                >< SinI
                                >< Sau96I
                                PssI ><
                                >< Psp5II
                                >< PpuMI
                                >< NspIV
                                >< NspHII
                                >< Eco47I
                                >< DraII
                                >< Cfr13I
                                >< BsiZI
                                >< Bmel8I
                                >< AvaII
                                >< AsuI
>< MaeII     EcoO109I >< AflIII >
ACGTGCGACT TTCTGAAAGC CACTTGTGAA CATTGTGGCA CTGAAAATTT AGTTATTGAA GGACCTACTA
1270      1280      1290      1300      1310      1320      1330

                                Van91I ><
                                SinI ><
                                Sau96I ><
                                PflMI ><
                                NspIV ><
                                NspHII >
                                Eco47I ><
                                Cfr13I ><
                                BslI ><
                                BsiZI ><
                                BsiYI ><
                                Bmel8I ><
                                AvaII ><
                                AsuI ><

>< RsaI
>< NspI
>< NlaIV
>< NlaIII
>< NspHI>< KpnI
>< Eco64I
>< Csp6I
>< BscBI
>< BanI
>< Asp718
>< AfaI
>< AccBII

```

FIGURE 13. 4

```

    >< Acc65I          >< SfcI          >< NlaIII          AccB7I ><
CATGTGGGTA CCTACCTACT AATGCTGTAG TGAAAATGCC ATGTCCTGCC TGTCAAGACC CAGAGATTGG
    1340          1350          1360          1370          1380          1390          1400

                                >< TthHB8I
                                >< TaqI>< MnlI
                                >< HinfI

    >< DdeI          >< PleI          >< AciI
ACCTGAGCAT AGTGTTCAG ATTATCACAA CCACTCAAAC ATTGAAATC GACTCCGCAA GGGAGGTAGG
    1410          1420          1430          1440          1450          1460          1470

    >< RmaI          NlaIV ><
    >< MnlI          >< BsrI
    >< MaeI          >< BbvI          >< Fnu4HI          BscBI ><
ACTAGATGTT TTGGAGGCTG TGTGTTTGCC TATGTTGGCT GCTATAATAA GCGTGCCTAC TGGGTTCCTC
    1480          1490          1500          1510          1520          1530          1540

                                XhoII ><
                                Sau3AI ><
                                NdeII ><
                                MflI ><
                                MboI ><

                                >< MaeIII          >< Eco31I          DpnII ><
                                >< Pali          >< HaeIII          >< BsrI          >< MnlI DpnI >
    >< RmaI          >< BsuRI          >< BsrI          >< BsmAI          BstYI ><
    >< MnlI          >< DdeI          >< BspWI          >< BsaI>< HphI          BspAI ><
    >< MaeI          >< BshI>< BglI          >< Alw26I          BspI43I >
GTGCTAGTGC TGATATTGGC TCAGGCCATA CTGGCATTAC TGGTGACAAT GTGGAGACCT TGAATGAGGA
    1550          1560          1570          1580          1590          1600          1610

                                > < Tru9I
                                > < MseI
                                >< MaeII          >< Tru9I
                                >< HpaI          > < MnlI
                                >< HindII          > < Ksp632I
                                >< HinfI >< PleI >< HincII          > < EarI
    >< AlwI          >< DdeI          >< AflIII          >< MseI          > < Eam1104I
TCTCCTTGAG ATACTGAGTC GTGAACGTGT TAACATTAAC ATTGTTGGCG ATTTTCATTT GAATGAAGAG
    1620          1630          1640          1650          1660          1670          1680

    >< MboII          PleI ><
    >< BstXI          >< SfaNI          > < HinfI
GTTGCCATCA TTTTGGCATC TTTCTCTGCT TCTACAAGTG CCTTTATTGA CACTATAAAG AGTCTTGATT
    1690          1700          1710          1720          1730          1740          1750

                                >< StyI
                                >< MaeIII
                                >< EcoT14I
                                >< Eco130I
                                >< PleI          >< BssT1I          BslI ><
                                >< MaeIII          >< BsaJI          BsiYI ><
                                >< HinfI>< AciI          >< BsaJI          BsiYI ><
ACAAGTCTTT CAAAACCATT GTTGAGTCCT GCGGTAAC TAAGTTACC AAGGGAAAGC CCGTAAAAGG
    1760          1770          1780          1790          1800          1810          1820

                                >< Sau3AI          >< Van91I
                                >< NdeII          >< PflMI
                                >< MboI          >< DraIII
                                >< DpnII          >< BslI
                                >< DpnI >< Tru9I          >< EsiYI
                                >< BspAI >< MseI          >< BbvI          >< MnlI
                                >< BspI43I          >< AccB7I          Fnu4HI ><

```

FIGURE 135

```

TGCTTGAAC ATTGGACAAC AGAGATCACT TTTAACACCA CTGTGTGGTT TTCCCTCACA GGCTGCTGGT
1830      1840      1850      1860      1870      1880      1890

      >< ThaI
      >< SfaNI
      >< MvnI
      >< HinPII
    >< HinPII
    >< Hin6I
    >< Hin6I
      >< HhaI
    >< Sau3AI    >< HhaI
    >< NdeII      >< CfoI
    >< MboI        >< CfoI
    >< DpnII       >< BstUI
    >< DpnI    >< BssHII
    >< BspAI      >< Bsp50I
    >< Bsp143I    >< AccII      >< Fnu4HI    >< BbvI
GTTATCAGAT CAATTTTTCG GCGCACACTT GATGCAGCAA ACCACTCAAT TCCTGATTG CAAAGAGCAG
1900      1910      1920      1930      1940      1950      1960

      >< TthHB8I
      >< StyI
      >< NcoI
      >< HindII
      >< HincII
      >< HinfI
      >< EcoT14I
      >< Eco57I
      >< TaqI>< Eco130I
    >< Sali >< DsaI
    >< RtrI >< BssT1I
      >< BsaHI
      >< BbiII>< NlaIII
      >< AclI >< HgaI
    >< MaeIII
      >< BbvI
      >< MaeII >< AccI>< BsaJI    HphI ><
CTGTCACCAT ACTTGATGGT ATTTCTGAAC AGTCATTACG TCTGTGCGAC GCCATGGTTT ATACTTCAGA
1970      1980      1990      2000      2010      2020      2030

      >< RsaI
      >< NdeI      >< Csp6I
    >< BspMI
      >< MaeIII >< BsrI >< AfaI    >< DdeI
CCTGCTCACC AACAGTGTCA TTATTATGGC ATATGTAAC TGTGGTCTTG TACAACAGAC TTCTCAGTGG
2040      2050      2060      2070      2080      2090      2100

      >< StuI
      >< PstI
      >< HaeIII
      >< Eco147I
    >< SduI    >< DdeI
    >< NspII
    >< Bsp1286I
    >< BmyI
      >< BsuRI
      >< BshI
      >< AatI      >< MnlI    DdeI ><
      >< BfrI ><
TTGTCTAATC TTTTGGGCAC TACTGTTGAA AAACCTCAGG CTATCTTTGA ATGGATTGAG GCGAAACTTA
2110      2120      2130      2140      2150      2160      2170

      >< TfiI
      >< HinfI
      >< SfaNI >< BsgI    >< FokI    Tth111I ><
      >< AspI ><
GTGCAGGAGT TGAATTTCTC AAGGATGCTT GGGAGATTCT CAAATTTCTC ATTACAGGTG TTTTGGACAT
2180      2190      2200      2210      2220      2230      2240

```

FIGURE 13.6


```

Tru9I ><
MseI ><
HpaI >
HindII >
HincII >
>< Eco57I
CGTCAAGGGT CAAATACAGG TTGCTTCAGA TAACATCAAG GATTGTGTAA AATGCTTCAT TGATGTTGTT
2250      2260      2270      2280      2290      2300      2310

>< Sau3AI
>< NdeII
>< MboI
> < MaeIII
>< FbaI
>< DpnII
>< DpnI
>< BspAI
>< Bsp143I
>< TthHB8I
>< TaqI
AACAAGGCAC TCGAAATGTG CATTGATCAA GTCACATATCG CTGGCGCAAA GTTGCGATCA CTCAACTTAG
2320      2330      2340      2350      2360      2370      2380

>< PvuII
>< MaeII
>< Bst1107I
>< BsaAI
>< BbvI
>< HphI
>< DrdI
>< AccI
GTGAAGTCTT CATCGCTCAA AGCAAGGGAC TTTACCGTCA GTGTATACGT GGCAAGGAGC AGCTGCAACT
2390      2400      2410      2420      2430      2440      2450

>< Tru9I
>< NlaIV
>< MseI
>< MnlI
>< Esp4I
>< Eco64I
>< BscBI
>< NlaIII >< BanI
>< AflII
>< BbvI
>< AccBII
>< MaeIII
ACTCATGCCT CTTAAGGCAC CAAAAGAAGT AACCTTTCTT GAAGGTGATT CACATGACAC AGTACTTACC
2460      2470      2480      2490      2500      2510      2520

> < XhoI
>< TthHB8I
>< TthHB8I>< TaqI
> < SlaI
> < Paer7I
> < NspIII
>< HphI >< HinII
> < Eco88I
> < CcrI
>< Esp3I >< BsaHI
> < BcoI
>< BsmAI >< BbiII
> < AvaI >< HgaI
>< TaqI > < Ama87I>< BsmBI
>< DdeI>< MnlI
>< Alw26I >< AcyI
TCTGAGGAGG TTGTTCTCAA GAACGGTGAA CTCGAAGCAC TCGAGACGCC CGTTGATAGC TTCACAAATG
2530      2540      2550      2560      2570      2580      2590

```

FIGURE 13.7

```

                                >< PstI >< NlaIII
                                >< HaeIII >< MnlI
                                >< BsuRI >< DdeI >< Tru9I
                                >< BshI >< BfrI >< MseI
>< AluI >< BsrI
GAGCTATCGT TGGCACACCA GTCTGTGTAA ATGGCCTCAT GCTCTTAGAG ATTAAGGACA AAGAACAATA
2600 2610 2620 2630 2640 2650 2660

                                >< VneI
                                Tru9I ><
                                >< SnaI
                                >< SduI
                                >< NspII
                                MseI ><
                                >< HgiAI
                                Bsp1286I >< BslI ><
                                BsiYI ><
                                >< BmyI
                                >< ApaLI
                                >< Tru9I >< Alw44I
                                >< MseI >< Alw21I
CTGCGCATTG TCTCCTGGTT TACTGGCTAC AAACAATGTC TTTGCTTAA AAGGGGGTGC ACCAATTAAA
2670 2680 2690 2700 2710 2720 2730

                                >< TfiI
>< MaeIII >< MboII >< MaeIII >< HinfI AluI ><
GGTGTAACCT TTGGAGAAGA TACTGTTTGG GAAGTTCAAG GTTACAAGAA TGTGAGAATC ACATTTGAGC
2740 2750 2760 2770 2780 2790 2800

                                >< RsaI
                                >< NlaIV
                                MaeIII ><
                                >< MspI >< KpnI
                                >< HpaII
                                >< HapII
                                >< Eco64I
                                >< SduI >< Csp6I
                                >< NspII >< TfiI >< BscBI
                                >< HgiAI >< BanI
                                >< Bsp1286I >< Asp718
                                >< BmyI >< HinfI >< AfaI
                                >< Alw21I >< AccB1I
                                >< AflIII >< MseI >< AccI >< Acc65I
TTGATGAACG TGTTGACAAA GTGCTTAATG AAAAGTGCTC TGTCTACACT GTTGAATCCG GTACCGAAGT
2810 2820 2830 2840 2850 2860 2870

                                >< Sau3AI
                                >< NdeII
                                >< MboI
                                >< DpnII
                                >< DpnI
                                >< NspI
                                >< NspHI
                                >< NlaIII
                                >< MboII >< BspAI
                                >< DdeI >< MboI >< Bsp143I
                                >< Mali >< AlwNI
                                >< BbsI >< AlwNI
TACTGAGTTT GCATGTGTTG TAGCAGAGGC TGTGTGAAG ACTTTACAAC CAGTTTCTGA TCTCCTTACC
2880 2890 2900 2910 2920 2930 2940

                                >< Sau3AI
                                >< NdeII
                                >< MboI
                                >< DpnII
                                >< DpnI
                                >< BspAI

```

FIGURE 13.8

```

    >< NlaIII>< Bsp143I          >< AluI          >< SfaNI
AACATGGGTA TTGATCTTGA TGAGTGGAGT GTAGCTACAT TCTACTTATT TGATGATGCT GGTGAAGAAA
    2950          2960          2970          2980          2990          3000          3010

                                >< SfaNI
                                >< MnlI
                                >< Ksp632I          >< MnlI
                                >< EarI          >< MboII
    >< MboII          >< GsuI
                                >< MnlI          >< Eam1104I          >< MboII
    >< HphI          >< MaeII>< BpmI          >< MnlI          >< Eam1104I          >< MboII
ACTTTTCATC ACGTATGTAT TGTTCTTTT ACCCTCCAGA TGAGGAAGAA GAGGACGATG CAGAGTGTGA
    3020          3030          3040          3050          3060          3070          3080

                                >< RsaI
                                >< RsaI
                                >< NlaIII
                                >< MnlI          >< FokI
                                >< Csp6I          Eco31I ><
                                >< Csp6I          >< MamI BsmAI ><
                                >< MboII          >< AfaI          >< BsiBI BsaI ><
    >< MboII          >< AfaI          >< BsaB1Alw26I ><
GGAAGAAGAA ATTGATGAAA CCTGTGAACA TGAGTACGGT ACAGAGGATG ATTATCAAGG TCTCCCTCTG
    3090          3100          3110          3120          3130          3140          3150

    >< NlaIV>< PvuII>< XmnI
    >< Eco64I >< Psp5I          >< TthHB8I
    >< MnlI >< DdeI          >< TaqI          >< MnlI          >< MboII
    >< BscBI>< NspBII >< MnlI          >< Ksp632I          >< MboII >< MboII
    >< BanI          >< MnlI          >< EarI          >< BsrI
    >< AccBII >< AluI >< Asp700I          >< Eam1104I >< MboII>< BbsI
GAATTTGGTG CCTCAGCTGA AACAGTTCGA GTTGAGGAAG AAGAAGAGGA AGACTGGCTG GATGATACTA
    3160          3170          3180          3190          3200          3210          3220

                                >< Tru9I
    >< FokI          >< MseI          >< Eco57I
    >< DdeI          >< BsrI>< MboII BsrI ><
CTGAGCAATC AGAGATTGAG CCAGAACCAG AACCTACACC TGAAGAACCA GTTAATCAGT TTACTGGTTA
    3230          3240          3250          3260          3270          3280          3290

    >< Tru9I          >< MnlI
    >< MseI          >< Tru9I >< HindII>< Tru9I          >< DraIII
    >< DraI          >< MseI          >< HincII>< MseI          >< BspWI
TTTAAAACTT ACTGACAATG TTGCCATTAA ATGTGTTGAC ATCGTTAAGG AGGCACAAAG TGCTAATCCT
    3300          3310          3320          3330          3340          3350          3360

                                >< VneI
                                >< SnaI
                                >< SduI
                                >< NspII
                                >< HgiAI
                                >< Bsp1286I
                                >< BmyI
                                >< ApaLI
    >< HphI          >< NlaIII          >< Alw44I
    >< BbvI          >< Fnu4HI          >< BspMI          >< Alw21I
ATGGTGATTG TAAATGCTGC TAACATACAC CTGAAACATG GTGGTGGTGT AGCAGGTGCA CTCACAAGG
    3370          3380          3390          3400          3410          3420          3430

                                >< Sau96I
                                >< Pali
                                >< NspIV
                                >< HaeIII
    >< NlaIV          >< Cfr13I

```

FIGURE 13.9

```

>< Eco64I
>< BscBI
>< BanI
>< AccB1I>< NlaIII
CAACCAATGG TGCCATGCAA AAGGAGAGTG ATGATTACAT TAAGCTAAAT GGCCCTCTTA CAGTAGGAGG
3440 3450 3460 3470 3480 3490 3500

>< BsuRI
>< Tru9I
>< MseI
>< AluI
>< AsuI
>< MnlI
>< SinI
>< Sau96I
>< NspIV
>< NspHI>< NspHII
>< Eco47I
>< Cfr13I
>< NlaIII
>< BspMI
>< Bsi2I
>< Bme18I
>< AvaII MnlI ><
>< DdeI
>< NspI>< AsuI FokI ><
GTCTTGTTTG CTTTCTGGAC ATAATCTTGC TAAGAAGTGT CTGCATGTTG TTGGACCTAA CCTAAATGCA
3510 3520 3530 3540 3550 3560 3570

>< Tru9I
>< HphI> < MseI
>< Esp4I
>< AluI
>< NdeI
>< AflIII>< Fnu4HI
>< BbvI
GGTGAGGACA TCCAGCTTCT TAAGGCAGCA TATGAAAATT TCAATTCACA GGACATCTTA CTGCACCAT
3580 3590 3600 3610 3620 3630 3640

RsaI ><
Csp6I ><
AfaI ><
>< Eco57I
>< BcgI
TGTTGTCAGC AGGCATATTT GGTGCTAAAC CACTTCAGTC TTTACAAGTG TGCGTGCAGA CGGTTCGTAC
3650 3660 3670 3680 3690 3700 3710

>< BsgI
>< BcgI/a
>< BspMI
>< AluI
>< NlaIII
ACAGGTTTAT ATTGCAGTCA ATGACAAAGC TCTTTATGAG CAGGTTGTCA TGGATTATCT TGATAACCTG
3720 3730 3740 3750 3760 3770 3780

>< MnlI
>< RmaI
>< MaeI
>< Eco57I
>< BscBI
>< TfiI
>< MboII
>< HinfI
>< DdeI
AAGCCTAGAG TGGAAGCACC TAAACAAGAG GAGCCACCAA ACACAGAAGA TTCCAAAACCT GAGGAGAAAT
3790 3800 3810 3820 3830 3840 3850

>< Tru9I
>< StuI
>< Pali
>< MseI
>< MnlI
>< MaeIII
>< HaeIII
>< Eco65I
>< Eco147I
>< Eco91I
>< BsuRI
>< BstXI ><
>< BshI
>< BstPI
>< AatI
>< BstEII
CTGTCGTACA GAAGCCTGTC GATGTGAAGC CAAAAATTAA GGCCTGCATT GATGAGGTTA CCACAACACT
3860 3870 3880 3890 3900 3910 3920

TfiI ><
NlaIII ><
HinfI ><
>< DdeI
>< EcoRV
>< HindIII

```

FIGURE 13.10

```

>< BsrI      >< MboII      >< MaeIII      >< Eco32I      >< AluI
GGAAGAACT AAGTTTCTTA CCAATAAGTT ACTCTTGT TT GCTGATATCA ATGGTAAGCT TTACCATGAT
3930      3940      3950      3960      3970      3980      3990

      >< NspI
      >< NspHI
      >< NlaIII
      >< MnlI      >< SfaNI
      > < EcoNI
      >< DdeI      >< MboII >< BslI      > < NlaIII
>< DdeI      >< BfrI      >< HphI      >< BsiYI      >< FokI
TCTCAGAACA TGCTTAGAGG TGAAGATATG TCTTTCCTTG AGAAGGATGC ACCTTACATG GTAGGTGATG
4000      4010      4020      4030      4040      4050      4060

      >< SpeI
      >< RmaI
      >< MaeI      >< EcoRV>< HphI      >< SfaNI
      >< HphI      >< Eco32I      >< MnlI      >< DdeI
TTATCACTAG TGGTGATATC ACTTGTGTG TAATACCCTC CAAAAAGGCT GGTGGCACTA CTGAGATGCT
4070      4080      4090      4100      4110      4120      4130

      >< ScrFI
      >< RsaI
      >< MvaI
      >< EcoRII
      >< Ecl136I
      >< DsaV
      >< Csp6I >< EcoNI
      >< BstOI
      >< BstNI
      >< BsiLI
      >< BsaJI
      >< BsaAI      >< BslI
      >< MaeII>< ApyI
      >< AluI      >< BsrI      >< AfaI      >< BsiYI
CTCAAGAGCT TTGAAGAAAG TGCCAGTTGA TGAGTATATA ACCACGTACC CTGGACAAGG ATGTGCTGGT
4140      4150      4160      4170      4180      4190      4200

      >< Tru9I
      >< MseI
      >< DdeI      >< Esp4I      >< RsaI
      >< MnlI      >< BspWI      >< Csp6I
      >< FokI      >< AluI      >< AflIII      >< Eco57I >< AfaI
TATACACTTG AGGAAGCTAA GACTGCTCTT AAGAAATGCA AATCTGCATT TTATGTACTA CCTTCAGAAG
4210      4220      4230      4240      4250      4260      4270

      >< ScrFI
      >< MvaI
      >< EcoRII
      >< XmnI      >< Ecl136I      NlaIII ><
      > < Ksp632I      >< RmaI      >< DsaV      Ksp632I ><
      > < EarI      > < TfiI>< MboII      >< BstOI      >< EarI
      > < Eam1104I      >< MaeI      >< BstNI      Eam1104I ><
      > < DdeI      > < HinfI      >< BsiLI      BsmAI ><
      >< BspWI      >< Asp700I      >< ApyI      Alw26I ><
CACCTAATGC TAAGGAAGAG ATTCTAGGAA CTGTATCCTG GAATTTGAGA GAAATGCTTG CTCATGCTGA
4280      4290      4300      4310      4320      4330      4340

      >< VspI      >< Zsp2I
      >< Tru9I      >< Ppu10I
      >< MseI      >< NsiI
      >< MboII      >< NlaIII      >< FokI
      >< Eco57I      >< Mph1103I      >< FokI

```

FIGURE 13.11

```

                >< AsnI          >< EcoT22I          >< BspWI
                >< AseI          >< AvaIII          >< BglI          >< MaeII
AGAGACAAGA AAATTAATGC CTATATGCAT GGATGTTAGA GCCATAATGG CAACCATCCA ACGTAAGTAT
4350          4360          4370          4380          4390          4400          4410

                >< SfaNI
                >< Tru9I          >< HindII          >< TfiI          >< SpeI
                >< MseI          >< HincII>< MboII          >< RmaI
                >< MnlI          >< DrdI >< HinfI          >< MaeI
AAAGGAATTA AAATTCAAGA GGCATCGTT GACTATGGTG TCCGATTCTT CTTTATACT AGTAAAGAGC
4420          4430          4440          4450          4460          4470          4480

                >< MaeIII
                >< SfcI          >< Fnu4HI          >< MunI
                >< AluI          >< AluI          >< AciI          >< MaeIII ><
CTGTAGCTTC TATTATTACG AAGCTGAACT CTCTAAATGA GCCGCTTGTC ACAATGCCAA TTGGTTATGT
4490          4500          4510          4520          4530          4540          4550

                >< ThaI
                >< MvnI
                >< MboII
                >< HinPII
                >< HinPII
                >< Hin6I
                >< Hin6I
                >< HhaI
                >< Tru9I          >< HhaI
                >< NlaIII          >< Fnu4HI
                >< MseI          >< CfoI
                >< MnlI          >< CfoI
                >< Ksp632I          >< BstUI
                >< EarI          >< BssHII>< BspWI          >< Tru9I
                >< Eam1104I          >< Bsp50I          >< MseI
                >< BbvI          >< AccII          >< AluI          >< HphI ><
GACACATGGT TTTAATCTTG AAGAGGCTGC GCGCTGTATG CGTTCTCTTA AAGCTCCTGC CGTAGTGTC
4560          4570          4580          4590          4600          4610          4620

                >< MaeIII
                >< SfaNI          >< AlwNI          >< MnlI >< MnlI>< DdeI
GTATCATCAC CAGATGCTGT TACTACATAT AATGGATACC TCACCTTCGTC ATCAAAGACA TCTGAGGAGC
4630          4640          4650          4660          4670          4680          4690

                >< SinI
                >< Sau96I
                >< NspIV
                >< NspHII
                >< SduI          >< Eco47I
                >< NspII          >< Cfr13I
                >< HgiAI          >< BsiZI
                >< Bsp1286I          >< Bme18I          >< RsaI
                >< BmyI          >< AvaII          >< Csp6I
                >< Alw21I          >< AsuI          >< AfaI
ACTTTGTAGA AACAGTTTCT TTGGCTGGCT CTTACAGAGA TTGGTCCTAT TCAGGACAGC GTACAGAGTT
4700          4710          4720          4730          4740          4750          4760

                >< TthHB8I
                >< TaqI
                >< SduI
                >< Van9II          >< NspII
                >< Tru9I          >< RsaI          >< PflMI          >< Eco24I
                >< MseI          >< HphI          >< BslI          >< Bsp1286I
                >< Esp4I          >< Csp6I          >< BsiYI          >< BmyI          >< GsuI ><

```

FIGURE 13.12

```

    >< AflIII >< MaeIII >< AfaI >< AccB7I >< BanIIBpmI ><
AGGTGTTGAA TTTCTTAAGC GTGGTGACAA AATTGTGTAC CACACTCTGG AGAGCCCCGT CGAGTTTCAT
    4770      4780      4790      4800      4810      4820      4830

                                >< Tru9I
                                >< P1eI >< EcoNI
                                >< MnlI >< BslI
                                >< BsmAI >< BsiYI
    >< MnlI >< HphI >< HinfI >< Alw26I >< AciI >< MseI
CTTGACGGTG AGGTTCTTTC ACTTGACAAA CTAAAGAGTC TCTTATCCCT GCGGGAGGTT AAGACTATAA
    4840      4850      4860      4870      4880      4890      4900

                                >< AluI >< NdeI
AAGTGTTCAC AACTGTGGAC AACACTAATC TCCACACACA GCTTGTGGAT ATGTCTATGA CATATGGACA
    4910      4920      4930      4940      4950      4960      4970

    >< SinI
    >< Sau96I
    >< NspIV
    >< NspHII
    >< Eco47I
    >< Cfr13I
    >< Bsi2I
    >< Bme18I
    >< AvaII
    >< AsuI
                                >< MaeIII >< Tru9I
                                >< FokI >< MseI
                                >< BspHI
GCAGTTTGGT CCAACATACT TGGATGGTGC TGATGTTACA AAAATTAAAC CTCATGTAAA TCATGAGGGT
    4980      4990      5000      5010      5020      5030      5040

                                > < TthHB8I
                                > < TaqI
    >< RsaI
    > < RmaI
    > < MaeI
    >< Csp6I
    >< AfaI
                                >< SnaBI
                                >< MaeII >< HindIII
                                >< Eco105I
                                >< BsaAI >< AluI >< AfaI
                                >< ScaI
                                >< RsaI
                                >< Csp6I
                                >< AfaI
AAGACTTTCT TTGTACTACC TAGTGATGAC ACACTACGTA GTGAAGCTTT CGAGTACTAC CATACTCTTG
    5050      5060      5070      5080      5090      5100      5110

    >< RsaI
    >< NspI
    >< NspHI
    >< NlaIII
    > < Csp6I >< Tru9I
    >< AflIII >< MseI
    >< AfaI >< DraI
                                MnlI >
                                BslI ><
                                BsiYI ><
ATGAGAGTTT TCTTGGTAGG TACATGTCTG CTTTAAACCA CACAAAGAAA TGGAAATTC CTCAAGTTGG
    5120      5130      5140      5150      5160      5170      5180

    >< Tru9I >< Tru9I
    >< MseI >< MseI
    >< MunI >< RmaI
    >< MaeI >< AluI >
TGGTTTAACT TCAATTAAAT GGGCTGATAA CAATTGTTAT TTGTCTAGTG TTTTATTAGC ACTTCAACAG
    5190      5200      5210      5220      5230      5240      5250

                                >< SfaNI
                                >< SduI
                                >< NspII
                                >< Eco24I
                                >< Bsp1286I
                                >< BmyI
                                >< BbvI Fnu4HI ><
                                >< BanII >< BspWI
    >< MnlI

```

FIGURE 13.13

```

CTTGAAGTCA AATTCAATGC ACCAGCACTT CAAGAGGCTT ATTATAGAGC CCGTGCTGGT GATGCTGCTA
5260      5270      5280      5290      5300      5310      5320

>< VneI
>< SnoI
    >< SduI
    >< NspII
    >< HgiAI
    >< Bsp1286I
    >< BmyI
>< ApaLI
>< Alw44I
    >< Alw21I
    >< AluI
    >< HphI
ACTTTTGTGC ACTCATACTC GCTTACAGTA ATAAACTGT TGGCGAGCTT GGTGATGTCA GAGAACTAT
5330      5340      5350      5360      5370      5380      5390

    > < SphI
    > < PaeI
    > < NspI
    > < NspHI >< TfiI
    >< Tru9I
    >< SfcI > < NlaIII>< HinfI
    >< MseI
GACCCATCTT CTACAGCATG CTAATTTGGA ATCTGCAAAG CGAGTTCTTA ATGTGGTGTG TAAACATTGT
5400      5410      5420      5430      5440      5450      5460

    >< RsaI
    >< Tru9I
    >< Csp6I
    >< MseI
    >< AluI
    >< AfaI
    >< Esp4I >
    >< AfIII >
GGTCAGAAAA CTACTACCTT AACGGGTGTA GAAGCTGTGA TGTATATGGG TACTCTATCT TATGATAATC
5470      5480      5490      5500      5510      5520      5530

    >< RsaI
    >< MboII
    >< RmaIHinfI ><
    >< Csp6I
    >< MaeI >< BbsI
    >< AfaI
>< Tru9I
    >< SfaNI
>< MseI
    >< NlaIII
TTAAGACAGG TGTTCATT CCATGTGTGT GTGGTCGTGA TGCTACACAA TATCTAGTAC AACAGAGTC
5540      5550      5560      5570      5580      5590      5600

    >< RsaI
    >< PleI
    >< DdeI
    >< Csp6I
    >< BsgI
    >< BspWI >< BspMI
    >< AfaI
TTCTTTTGTG ATGATGTCTG CACCACCTGC TGAGTATAAA TTACAGCAAG GTACATTCTT ATGTGCGAAT
5610      5620      5630      5640      5650      5660      5670

    >< RsaI
    >< Eco31I
    >< DdeI
    >< MaeIII
    >< BsmAI
    >< Csp6I
    >< BsaI
    >< Alw26I
    >< MnlI ><
    >< AfaI >< BsrI
    >< HphI >
GAGTACACTG GTAACATCA GTGTGGTCAT TACACTCATA TAACTGCTAA GGAGACCCTC TATCGTATTG
5680      5690      5700      5710      5720      5730      5740

    >< SstI
    >< SinI
    >< SduI
    >< Sau96I
    >< SacI
    >< NspIV
    >< NspII
    >< NspHII
    >< HgiAI
    >< Eco24I
    >< Eco47I
    >< Ecl136II
    >< Cfr13I
    >< Bsp1286I
    >< BsiZI
    >< BmyI
    >< Bmel8I

```

FIGURE 13. 14


```

    >< BanII
    >< Alw21I
    >< AluI
    ACGGAGCTCA CCTTACAAAG ATGTCAGAGT ACAAAGGACC AGTGACTGAT GTTTTCTACA AGGAAACATC
    5750      5760      5770      5780      5790      5800      5810

    >< TthHB8I
    >< TaqI >< MaeIII
    TTACTACTACA ACCATCAAGC CTGTGTCGTA TAAACTCGAT GGAGTTACTT ACACAGAGAT TGAACCAAAA
    5820      5830      5840      5850      5860      5870      5880

    >< RsaI
    >< Csp6I
    >< SfcI >< BbvI
    >< Fnu4HI >< AfaI
    TTGGATGGGT ATTATAAAAA GGATAATGCT TACTATACAG AGCAGCCTAT AGACCTTGTA CCAACTCAAC
    5890      5900      5910      5920      5930      5940      5950

    Tru9I ><
    SwaI ><
    MseI ><
    > < NspI
    > < NspHI
    > < NlaIII
    >< AflIII
    CATTACCAAA TGCGAGTTTT GATAATTTC AACTCACATG TTCTAACACA AAATTGCTG ATGATTTAAA
    5960      5970      5980      5990      6000      6010      6020

    >< MboII
    >< AluI >< AluI >< MaeIII
    TCAAATGACA GGCTTCACAA AGCCAGCTTC ACGAGAGCTA TCTGTCACAT TCTTCCCAGA CTTGAATGGC
    6030      6040      6050      6060      6070      6080      6090

    >< SfcI
    GATGTAGTGG CTATTGACTA TAGACACTAT TCAGCGAGTT TCAAGAAAGG TGCTAAATTA CTGCATAAGC
    6100      6110      6120      6130      6140      6150      6160

    >< Tru9I
    >< ScrFI
    >< MvaI
    >< MseI
    >< EcoRII
    >< Ecl136I
    >< DsaV
    >< BstOI
    >< BstNI
    >< BsiLI
    >< MunI
    >< BstXI
    CAATTGTTTG GCACATTAAC CAGGCTACAA CCAAGACAAC GTTCAAACCA AACACTTGGT GTTTACGTTG
    6170      6180      6190      6200      6210      6220      6230

    > < RsaI
    >< Csp6I
    > < AfaI >< BsrI
    TCTTTGGAGT ACAAAGCCAG TAGATACTTC AAATTCATTT GAAGTTCTGG CAGTAGAAGA CACACAAGGA
    6240      6250      6260      6270      6280      6290      6300

    >< HindII
    >< HincII
    ATGGACAATC TTGCTTGTGA AAGTCAACAA CCCACCTCTG AAGAAGTAGT GGAAAATCCT ACCATACAGA
    6310      6320      6330      6340      6350      6360      6370

    >< MboII
    >< MnlI
    >< Eco57I

```

FIGURE 13.15

```

    >< MaeIII
    >< MaeII
AGGAAGTCAT AGAGTGTGAC GTGAAAACTA CCGAAGTTGT AGGCAATGTC ATACTTAAAC CATCAGATGA
    6380      6390      6400      6410      6420      6430      6440

    >< XhoII
    >< Sau3AI
    >< NlaIII
    >< NdeII
    >< MflI
    >< MboI
    >< DpnII
    >< DpnI
    >< BstYI
    >< BspAI
    >< Tru9I
    >< MseI
    >< BspHI >< Bsp143I>< Fnu4HI
    >< MaeIII >< MnlI >< BbvI >< AlwI
AGGTGTAAAA GTAACACAAG AGTTAGGTCA TGAGGATCTT ATGGCTGCTT ATGTGGAAAA CACAAGCATT
    6450      6460      6470      6480      6490      6500      6510

    >< SauI
    >< RmaI
    >< MstII
    >< MaeI
    >< Eco8II
    >< DdeI
    >< CvnI
    >< Bsu36I
    >< Bse2II
    >< BfrI> < Tru9I
    >< AxyI> < MseI>< MunI >< NlaIII
    >< Tru9I
    >< MseI >< AluJ >< AocI >< DraI >< BbvI Fnu4HI ><
ACCATTAAGA AACCTAATGA GCTTTCACCTA GCCTTAGGTT TAAAAACAAT TGCCACTCAT GGTATTGCTG
    6520      6530      6540      6550      6560      6570      6580

    >< VspI >< StyI
    >< Tru9I >< EcoT14I >< DdeI
    >< MseI >< Eco130I >< BslI
    >< AsnI >< BssTII >< BsiYI
    >< AseI >< BsaJI >< BfrI >< Fnu4HI
CAATTAATAG TGTTCCTTGG AGTAAAATTT TGGCTTATGT CAAACCATTC TTAGGACAAG CAGCAATTAC
    6590      6600      6610      6620      6630      6640      6650

    >< HinPII
    >< Hin6I
    >< HhaI >< Tru9I
    >< DdeI >< MaeII>< MseI
    >< BbvI >< CfoI >< DraIII
    >< AflIII
AACATCAAAT TGCCTAAGA GATTAGCACA ACGTGTGTTT AACAATTATA TGCCTTATGT GTTTACATTA
    6660      6670      6680      6690      6700      6710      6720

    >< RsaI >< RsaI>< XbaI
    >< Csp6I >< Csp6I >< RmaI
    >< MunI >< AfaI >< AfaI >< MaeI >< AluI
TTGTTCCAAT TGTGTACTTT TACTAAAAGT ACCAATTCTA GAATTAGAGC TTCACTACCT ACAACTATTG
    6730      6740      6750      6760      6770      6780      6790

    >< VspI
    >< Tru9I
    >< NaeI
    >< MspI
    >< MseI

```

FIGURE 13. 16

```

                                >< HpaII
                                >< HapII
                                >< Cfr10I >< FokI
                                >< AsnI
                                >< AseI>< HphI>< MaeIII
                                >< Tru9I
                                >< DdeI    MaeIII >
                                >< MseI    >< BfrI    >< BbvI
CTAAAAATAG TGTTAAGAGT GTTGCTAAAT TATGTTTGGG TGCCGGCATT AATTATGTGA AGTCACCCAA
6800      6810      6820      6830      6840      6850      6860

                                >< Tru9I    >< DdeI    MaeIII >
                                >< MseI    >< BfrI    >< BbvI
ATTTTCTAAA TTGTTACAA TCGCTATGTG GCTATTGTG TTAAGTATTT GCTTAGGTTC TCTAATCTGT
6870      6880      6890      6900      6910      6920      6930

                                >< SduI
                                >< NspII
                                >< HgiAI
                                >< Bsp1286I
                                >< BmyI
                                >< Alw21I
                                > < RsaI
                                >< Csp6I
                                > < AfaI
GTAAGTGTG CTTTGGTGT ACTCTTATCT AATTTTGGTG CTCCTTCTTA TTGTAATGGC GTTAGAGAAT
6940      6950      6960      6970      6980      6990      7000

                                Tru9I ><
                                MseI ><
                                >< Fnu4HI
                                >< MaeIII
                                >< MaeII
                                >< BbvI >
TGTATCTTAA TTCGTCTAAC GTTACTACTA TGGATTCTG TGAAGGTTCT TTTCCTTGCA GCATTTGTTT
7010      7020      7030      7040      7050      7060      7070

                                > < TfiI
                                >< MamI
                                > < HinfI
                                >< BsiBI
                                >< XmnI>< MaeIII
                                >< BsaBI >< AluI
                                >< Asp700I
                                >< AfaI ><
AAGTGGATTA GACTCCCTTG ATTCTTATCC AGCTCTTGAA ACCATTACAG TGACGATTTC ATCGTACAAG
7080      7090      7100      7110      7120      7130      7140

                                >< Pali
                                >< NspBII
                                >< HaeIII
                                >< GdiII
                                >< Fnu4HI
                                >< EaeI
                                >< DdeI
                                >< BsuRI
                                >< RmaI
                                >< BshI >< BslI
                                >< MaeI
                                >< AciI>< BsiYI
CTAGACTTGA CAATTTTAGG TCTGGCCGCT GAGTGGGTTT TGGCATATAT GTTGTTTACA AAATTCTTTT
7150      7160      7170      7180      7190      7200      7210

                                >< BspMI
                                >< RmaI
                                >< AluI
                                >< MaeI
ATTTATTAGG TCTTTCAGCT ATAATGCAGG TGTTCTTTGG CTATTTTGCT AGTCATTTCA TCAGCAATTC
7220      7230      7240      7250      7260      7270      7280

                                RsaI ><
                                >< MboII
                                >< NlaIV
                                >< Eco64I
                                > < RsaI >< BscBI
                                >< Csp6I >< BanI
                                > < AfaI>< AccBI
                                >< NlaIII
                                >< BmyI
                                >< BsaBI ><
                                >< AfaI ><

```

FIGURE 13.17

```

TTGGCTCATG TGGTTTATCA TTAGTATTGT ACAAATGGCA CCCGTTTCTG CAATGGTTAG GATGTACATC
7290      7300      7310      7320      7330      7340      7350

                                TthHB8I ><
                                >< TaqI
                                MnlI ><
                                >< NdeI
                                >< Ksp632I
                                >< EarI
                                >< MboII EarI ><
>< FokI
>< Eam1104I>< AluI>< MboII >< NlaIII Eam1104I ><
TTCTTTGCTT CTTTCTACTA CATATGGAAG AGCTATGTTC ATATCATGGA TGGTTGCACC TCTTCGACTT
7360      7370      7380      7390      7400      7410      7420

                                XhoII ><
                                Sau3AI ><
                                NlaIII ><
                                NdeII ><
                                MflI ><
                                MboI ><
                                >< ThaI
                                >< MvnI
                                >< EarI
>< HinPII
>< Hin6I
>< HhaI
>< Bsp50I >< RsaI
>< NlaIII >< CfoI
>< AflIII >< Csp6I
>< BspWI >< BspWI
>< AccII >< AfaI
GCATGATGTG CTATAAGCGC AATCGTGCCA CACGCGTTGA GTGTACAACT ATTGTTAATG GCATGAAGAG
7430      7440      7450      7460      7470      7480      7490

                                >< Pali
                                >< HaeIII
                                >< DsaI
                                >< MunI
>< MboII
>< BsuRI
>< DpnI
>< BspI43I
>< MnlI
>< BsaJI
>< PleI>< HinfI
>< MunI
>< BsmAI ><
>< Alw26I ><
ATCTTTCTAT GTCTATGCAA ATGGAGGCCG TGGCTTCTGC AAGACTCACA ATTGGAATTG TCTCAATTGT
7500      7510      7520      7530      7540      7550      7560

                                >< RsaI
                                >< Csp6I
                                >< BsrI
                                >< AfaI
                                >< GsuI
                                >< BpmI
                                >< MaeIIIDraI ><
                                >< BsrI
GACACATTTT GCACTGGTAG TACATTCATT AGTGATGAAG TTGCTCGTGA TTTGTCACCTC CAGTTTAAAA
7570      7580      7590      7600      7610      7620      7630

                                >< ThaI
                                >< MvnI
                                >< HphI
                                >< HinPII ><
                                >< HinPII
                                >< Hin6I
                                >< Hin6I
                                >< HhaI ><
                                >< HhaI
                                >< CfoI ><
                                >< CfoI
                                >< BstUI
                                >< BssHII
                                >< Bsp50I ><
                                >< BsrI
                                >< AccII
GACCAATCAA CCCTACTGAC CAGTCATCGT ATATTGTTGA TAGTGTTGCT GTGAAAAATG GCGCGCTTCA
7640      7650      7660      7670      7680      7690      7700

```

FIGURE 13. 18

```

                                >< FokI
                                >< BsmAI
                                >< Alw26I
                                >< MnlI
                                >< AciI
CCTCTACTTT GACAAGGCTG GTCAAAAGAC CTATGAGAGA CATCCGCTCT CCCATTTTGT CAATTTAGAC
7710      7720      7730      7740      7750      7760      7770

                                >< VspI
                                >< Tru9I
                                >< MseI
                                >< AsnI
                                >< AseI
                                >< BcgI/a
> < AluI
AATTGAGAG CTAACAACAC TAAAGGTTCA CTGCCTATTA ATGTCATAGT TTTTGATGGC AAGTCCAAAT
7780      7790      7800      7810      7820      7830      7840

                                >< SfcI
                                >< PvuII
                                >< RsaI
                                >< Psp5I
                                >< PleI
                                >< Csp6I
                                >< NspBII
                                >< HinfI
                                >< DdeI
                                >< BcgI
                                >< AfaI
                                >< AluI
GCGACGAGTC TGCTTCTAAG TCTGCTTCTG TGTACTACAG TCAGCTGATG TGCCAACCTA TTCTGTTGCT
7850      7860      7870      7880      7890      7900      7910

                                TthHB8I ><
                                TaqI ><
                                SalI ><
                                RtrI ><
                                HindII >
                                HincII >
                                >< ScaI
                                >< RsaI
                                >< Tru9I
                                >< Csp6I
                                >< SfaNI
                                >< Eco57I
                                >< AluI
                                >< MaeII
                                >< AfaI
                                >< MseI
                                >< AccI
TGACCAAGCT CTTGTATCAG AC GTTGGAGA TAGTACTGAA GTTCCGTTA AGATGTTTGA TGCTTATGTC
7920      7930      7940      7950      7960      7970      7980

                                >< Tru9I
                                >< MseI
                                > < Esp4I
                                >< SfcI
                                > < AflIII
                                >< BspWI
                                >< AluI
GACACCTTTT CAGCAACTTT TAGTGTTCTT ATGGAAAAAC TTAAGGCACT TGTGCTACA GCTCACAGCG
7990      8000      8010      8020      8030      8040      8050

                                >< PvuII
                                >< Psp5I
                                >< NspBII
                                >< Fnu4HI
                                >< AluI
                                >< BbvI
AGTTAGCAAA GGGTGTAGCT TTAGATGGTG TCCTTTCTAC ATTCGTGTCA GCTGCCCGAC AAGGTGTTGT
8060      8070      8080      8090      8100      8110      8120

                                >< HindII
                                >< BsmAI
                                >< DdeI
                                >< HincII
                                >< FokI
                                >< Alw26I
                                >< BfrI
TGATACCGAT GTTGACACAA AGGATGTTAT TGAATGTCTC AAACTTTCAC ATCACTCTGA CTTAGAAGTG
8130      8140      8150      8160      8170      8180      8190

                                >< XhoII
                                Sau3AI ><
                                >< NdeII
                                >< MflI
                                >< MboI
                                >< NlaIII
                                >< HgaI
                                >< HinfI
                                >< DpnII
                                DpnI ><

```

FIGURE 13.19

```

                                Bsp143I ><
                                >< BsaHI >< BstYI
                                >< BbiII >< BspAI
                                >< AcyI >< BglII
                                >< MaeIII>< HphI
                                >< MaeIII >< HphI >< NlaIII
ACAGGTGACA GTTGTAACAA TTTCATGCTC ACCTATAATA AGGTTGAAAA CATGACGCCC AGAGATCTTG
8200      8210      8220      8230      8240      8250      8260

                                >< NspI
                                >< NspHI
                                >< NlaIII
>< HinPII
>< Hin6I
>< HhaI
>< CfoI
                                >< BspWI >< MaeIII
GCGCATGTAT TGACTGTAAT GCAAGGCATA TCAATGCCCA AGTAGCAAAA AGTCACAATG TTTCATCAT
8270      8280      8290      8300      8310      8320      8330

                                >< NspI
                                >< NspHI >< PvuII
                                >< NlaIII >< Psp5I
                                >< Eam1105I >< NspBII
                                >< BbvI >< Fnu4HI
                                >< AflIII >< AluI >< BbvI >< Fnu4HI
CTGGAATGTA AAAGACTACA TGTCTTTATC TGAACAGCTG CGTAAACAAA TTCGTAGTGC TGCCAAGAAG
8340      8350      8360      8370      8380      8390      8400

                                >< RmaI
                                >< MboII >< MaeI >< Eam1105I
AACAAACATAC CTTTGTAGACT AACTTGTGCT ACAACTAGAC AGGTTGTCAA TGTCATAACT ACTAAAATCT
8410      8420      8430      8440      8450      8460      8470

                                >< Tru9I
                                >< Pali
                                >< MseI
                                >< HaeIII
                                >< ScaI >< Esp4I
                                >< RsaI >< Tru9I >< BsuRI
                                >< Csp6I >< MseI >< BshI
                                >< AfaI >< DraI >< AflII >< BbvI
CACTCAAGGG TGTAAGATT GTTAGTACTT GTTTTAACT TATGCTTAAG GCCACATTAT TGTGCGTTCT
8480      8490      8500      8510      8520      8530      8540

                                >< RsaI
                                >< Csp6I
                                >< BsrI >< NlaIII
                                >< Fnu4HI >< AfaI >< MaeIII
TGCTGCATTG GTTTGTTATA TCGTTATGCC AGTACATACA TTGTCAATCC ATGATGGTTA CACAAATGAA
8550      8560      8570      8580      8590      8600      8610

                                >< MaeIII
                                >< MaeIII
                                >< MaeIII >< FokI
ATCATTGGTT ACAAAGCCAT TCAGGATGGT GTCACCTCGTG ACATCATTTT TACTGATGAT TGTTTTGCAA
8620      8630      8640      8650      8660      8670      8680

                                SfcI >
                                >< NspI
                                >< NspHI >< NlaIII
                                >< NlaIII >< HgaI >< BstXI >< BbvI >< AluI
ATAAACATGC TGGTTTTGAC GCATGGTTTA GCCAGCGTGG TGGTTCATAC AAAAAAGACA AAAGCTGCCC
8690      8700      8710      8720      8730      8740      8750

```

FIGURE 13. 20

```

                                >< ScrFI
                                >< ScrFI >< RsaI
                                >< MvaI >< MspI
                                >< EcoRII >< HpaII
                                >< Ecl136I>< NciI
                                >< DsaV >< HapII
                                >< BstOI>< DsaV
                                >< BstNI >< Csp6I
                                >< BsiLI >< BcnIDdeI ><
                                >< ApyI >< AfaI
                                >< Fnu4HI
                                >< AluI
TGTTAGTAGCT GCTATCATT CAAGAGAGAT TGGTTTCATA GTGCCTGGCT TACCGGGTAC TGTGCTGAGA
8760 8770 8780 8790 8800 8810 8820

                                > < MaeIII >< HphI >< MnlI >< BspWI
GCAATCAATG GTGACTTCTT GCATTTTCTA CCTCGTGTTC TTAGTGCTGT TGGCAACATT TGCTACACAC
8830 8840 8850 8860 8870 8880 8890

                                Tru9I >
                                SfaNI ><
                                >< RsaI
                                MseI >
                                >< BspWI >< Fnu4HI >< Csp6I
                                >< BbvI>< MnlI >< DdeI >< AfaI
CTTCCAACT CATTGAGTAT AGTGATTTG CTACCTCTGC TTGCGTTCTT GCTGCTGAGT GTACAATTTT
8900 8910 8920 8930 8940 8950 8960

                                > < RmaI
                                >< MnlI
                                >< FokI
                                > < MaeI
TAAGGATGCT ATGGGCAAAC CTGTGCCATA TTGTTATGAC ACTAATTGCT TAGAGGGTTC TATTTCTTAT
8970 8980 8990 9000 9010 9020 9030

                                ScrFI >
                                MvaI >
                                MnlI ><
                                EcoRII ><
                                Ecl136I >
                                DsaV ><
                                BstOI >
                                BstNI >
                                BsiLI >
                                >< NlaIV
                                >< FokI
                                >< BscBI
                                >< AluI
                                >< BscBI
                                >< ApyI >
AGTGAGCTTC GTCCAGACAC TCGTTATGTG CTTATGGATG GTTCCATCAT ACAGTTTCCT AACACTTACC
9040 9050 9060 9070 9080 9090 9100

                                >< RsaI
                                >< SfcI >< NspI
                                >< ScaI >< NspHI
                                >< RsaI >< NlaIII
                                >< Csp6I >< NlaIII
                                >< AfaI >< Csp6I
                                >< BpmI >< DdeI >< AccI >< AfaI
TGGAGGGTTC TGTTAGAGTA GTAACAACTT TTGATGCTGA GTAGTGTAGA CATGGTACAT GCGAAAGGTC
9110 9120 9130 9140 9150 9160 9170

                                >< SstI
                                >< SduI
                                >< SacI
                                NspII ><
                                HgiAI ><
                                Eco24I ><
                                Bsp1286I ><

```

FIGURE 13.21

```

                                Ecl136II ><>< BmyI
                                BanII ><
                                >< Tru9I
                                Alw21I ><
                                >< BsrI
                                >< MseI
                                >< AluI
AGAAGTAGGT ATTTGCCTAT CTACCACTGG TAGATGGGTT CTTAATAATG AGCATTACAG AGCTCTATCA
  9180      9190      9200      9210      9220      9230      9240

                                >< TfiI
                                >< SfaNI
                                >< HinfI
                                >< AluI
                                >< MnlI
GGAGTTTTCT GTGGTGTGA TGCGATGAAT CTCATAGCTA ACATCTTTAC TCCTCTTGTTG CAACCTGTGG
  9250      9260      9270      9280      9290      9300      9310

                                >< MaeIII
                                HphI ><
                                >< Eco57I
                                >< BbvI Fnu4HI ><
GTGCTTTAGA TGTGTCTGCT TCACTAGTGG CTGGTGGTAT TATTGCCATA TTGGTGACTT GTGCTGCCTA
  9320      9330      9340      9350      9360      9370      9380

                                >< RsaI
                                >< Csp6I
                                >< NlaIII
                                >< MaeII
                                >< BbvI
                                >< Fnu4HI
                                >< AflIII
                                >< AfaI>< HphI
                                >< BspWI
CTACTTTATG AAATTCAGAC GTGTTTTTGG TGAGTACAAC CATGTTGTTG CTGCTAATGC ACTTTTGTTC
  9390      9400      9410      9420      9430      9440      9450

                                >< RsaI
                                >< NlaIV
                                >< KpnI
                                >< Eco64I
                                >< Csp6I
                                >< BscBI
                                >< Asp718
                                >< BanI >< AluI
                                >< AfaI
                                >< AccBII
                                >< Acc65I
                                >< ScrFI
                                >< NciI
                                >< MspI
                                >< HpaII
                                >< HinfI
                                >< HapII
                                >< PleI
                                >< BcnI
                                >< DdeI
                                >< AluI>< DsaV
                                >< AccI
TTGATGTCTT TCACTATACT CTGTCTGGTA CCAGCTTACA GCTTTCTGCC GGGAGTCTAC TCAGTCTTTT
  9460      9470      9480      9490      9500      9510      9520

                                >< RsaI
                                >< Csp6I
                                >< AfaI >< HphI
                                >< HphI
                                NlaIII ><
ACTTGTAATT GACATTCTAT TTCACCAATG ATGTTTCATT CTTGGCTCAC CTTCAATGGT TTGCCATGTT
  9530      9540      9550      9560      9570      9580      9590

TTCTCCTATT GTGCCTTTTT GGATAACAGC AATCTATGTA TTCTGTATTT CTCTGAAGCA CTGCCATTGG
  9600      9610      9620      9630      9640      9650      9660

                                >< TthHB8I
                                >< RsaI
                                >< MnlI
                                >< MnlI
                                >< Csp6I
                                >< Tru9I
                                >< PleI
                                >< BcgI/a >< TaqI
                                >< MseI
                                >< DdeI
                                >< NlaIII
                                >< BbvI
                                >< Eco57I
                                >< BfrI
                                >< HinfI
                                >< MseI
                                >< MaeIII
                                >< AfaI Fnu4HI ><
TTCTTTAACA ACTATCTTAG GAAAAGAGTC ATGTTTAATG GAGTTACATT TAGTACCTTC GAGGAGGCTG
  9670      9680      9690      9700      9710      9720      9730

                                >< RsaI
                                >< Csp6I
                                >< BcgI
                                >< RsaI
                                >< Csp6I
                                >< BsmAI

```

FIGURE 13.22


```

    >> AfaI          >> AfaI          >> Alw26I
CTTTGTGTAC CTTTTTGCTC AACAAGGAAA TGTACCTAAA ATTGCGTAGC GAGACACTGT TGCCACTTAC
  9740      9750      9760      9770      9780      9790      9800

                                >> NlaIV
                                >> DdeI
                                >> RsaI
                                >> Csp6I
                                >> BscBI
                                >> AfaI
                                >> BfrI   AluI >>
ACAGTATAAC AGGTATCTTG CTCTATATAA CAAGTACAAG TATTTCAGTG GAGCCTTAGA TACTACCAGC
  9810      9820      9830      9840      9850      9860      9870

    >> Fnu4HI
                                >> DdeI
                                >> BfrI
    >> BbvI   >> AluI   >> BbvI          >> DdeI >> AlwNI
TATCGTGAAG CAGCTTGCTG CCACTTAGCA AAGGCTCTAA ATGACTTTAG CAACTCAGGT GCTGATGTTT
  9880      9890      9900      9910      9920      9930      9940

                                >> SfcI          >> BsmI
                                >> PstI          >> BscCI
TCTACCAACC ACCACAGACA TCAATCACTT CTGCTGTCTC GCAGAGTGGT TTTAGGAAAA TGGCATTCCT
  9950      9960      9970      9980      9990      10000     10010

    >> RsaI
    >> NlaIII
    >> MaeIII
    >> Csp6I          >> Tru9I
    >> AfaI          >> MseI
GTCAGGCAAA GTTGAAGGGT GCATGGTACA AGTAACCTGT GGAAGTACAA CTCTTAATGG ATTGTGGTTG
  10020     10030     10040     10050     10060     10070     10080

                                XhoII >>
                                Sau3AI >>
                                >> Tru9I   NdeII >>
                                >> NspI     MflI >>
                                >> NspHI     MboI >>
                                >> NlaIII     DpnII >>
                                >> MseI     BstYI >>
                                >> MboII     BspAI >>
                                >> BbsI     BglII >>
    >> FokI          >> NspI
    >> Bst1107I     >> NspHI
    >> AccI          >> NlaIII
    >> AflIII
GATGACACAG TATACTGTCC AAGACATGTC ATTTGCACAG CAGAAGACAT GCTTAATCCT AACTATGAAG
  10090     10100     10110     10120     10130     10140     10150

                                PalI >
                                MscI >
                                HaeIII >
                                EaeI >>
                                BsuRI >
                                BshI >
                                BalI >
    >> DpnI >> MboII
    >> Bsp143I          >> AluI
ATCTGCTCAT TCGCAAATCC AACCATAGCT TTCTTGTTCA GGCTGGCAAT GTTCAACTTC GTGTTATTGG
  10160     10170     10180     10190     10200     10210     10220

                                >> DdeI > > Tru9I
                                >> BfrI > > MseI
                                >> DdeI
CCATTCTATG CAAAATTGTC TGCTTAGGCT TAAAGTTGAT ACTTCTAACC CTAAGACACC CAAGTATAAA
  10230     10240     10250     10260     10270     10280     10290

    >> ScrFI
    >> MvaI
    >> EcoRII
    >> Ecl136I          >> SphI

```

FIGURE 13.23

```

                >< DsaV                >< PaeI
                >< BstOI                >< NspI
                >< BstNI                >< NspHI
                >< BsiLI                >< RmaI >< NlaIII
                >< ApyI                >< MaeI >< HphI
TTTGTCCGTA TCCAACCTGG TCAAACATTT TCAGTTCTAG CATGCTACAA TGGTTCACCA TCTGGTGTTC
10300      10310      10320      10330      10340      10350      10360

                >< Sau3AI
                >< NdeII
                >< MboI>< NlaIII
                >< DpnII
                >< Eco3II
                >< BsmAI                >< Tru9I>< DpnI
                >< BsaI>< NlaIII        >< MseI >< Bsp143I
                >< Alw26I            >< MseI >< BspAI>< AlwI
ATCAGTGTGC CATGAGACCT AATCATACCA TTAAAGGTTT TTTCTTAAT GGATCATGTG GTAGTGTTCG
10370      10380      10390      10400      10410      10420      10430

                >< Zsp2I
                >< Ppu10I
                >< NsiI>< SfaNI
                >< NdeI
                >< Mph1103I            RsaI ><
                >< EcoT22I            Csp6I ><
                >< Tru9I                >< AvaIII >< AluI AfaI ><
                >< MseI                >< EcoT22I >< AluI AfaI ><
TTTTAACATT GATTATGATT GCGTGTCTTT CTGCTATATG CATCATATGG AGCTTCCAAC AGGAGTACAC
10440      10450      10460      10470      10480      10490      10500

                >< SinI
                >< Sau96I
                >< NspIV
                >< NspHII
                >< Eco47I            >< SfcI
                >< Cfr13I            RsaI ><
                >< BsiZI            PstI ><
                >< Bme18I >< HindII    >< Fnu4HI
                >< Csp6I>< DdeI        >< AvaII >< HincII    >< Csp6I ><
                >< AfaI>< BfrI        >< AsuI>< BsgI >< BbvI >< BspMI AfaI ><
                >< BspWI
GCTGGTACTG ACTTAGAAGG TAAATTCTAT GGTCCATTTG TTGACAGACA AACTGCACAG GCTGCAGGTA
10510      10520      10530      10540      10550      10560      10570

                >< Tru9I                >< NlaIII
                >< MseI >< BbvI            >< Fnu4HI            HphI ><
CAGACACAAC CATAACATTA AATGTTTGG CATGGCTGTA TGCTGCTGTT ATCAATGGTG ATAGGTGGTT
10580      10590      10600      10610      10620      10630      10640

                >< Tru9I
                >< TfiI
                >< MseI                >< RsaI
                >< HphI                >< Tru9I >< Csp6I
                >< HinfI            >< MseI >< AfaI
TCTTAATAGA TTCACCACTA CTTTGAATGA CTTTAACCTT GTGGCAATGA AGTACAACCTA TGAACCTTTG
10650      10660      10670      10680      10690      10700      10710

                >< SinI
                >< Sau96I
                >< PssI
                >< Psp5II
                >< PpuMI
                >< NspIV
                >< NspHII
                >< NlaIV

```

FIGURE 13. 24

```

                >< Eco0109I
                >< Eco47I
    >< Sau3AI
    >< NdeII
    >< MboI
    >< DpnII>< NlaIII
    >< DpnI >< HindII
    >< BspAI >< HincII
    >< Bsp143I
ACACAAGATC ATGTTGACAT ATTGGGACCT CTTTCTGCTC AAACAGGAAT TGCCGTCTTA GATATGTGTG
10720      10730      10740      10750      10760      10770      10780

                >< Eco147I
                >< DraII
                >< Cfr13I
                >< BsiZI
                >< BscBI
                >< Bme18I
                >< DdeI
                >< BfrI
                >< BbvI
                >< MnlI
                >< StyI
                >< RsaI
                >< EcoT14I
                >< Eco130I
                >< SfcI
                >< Fnu4HI
                >< BbvI
                >< BbvI
                >< AluI
                >< PstI
                >< AfaI
                >< Csp6I
                >< BssT1I
                >< BsaJI
CTGCTTTGAA AGAGCTGCTG CAGAATGGTA TGAATGGTCG TACTATCCTT GGTAGCACTA TTTTAGAAGA
10790      10800      10810      10820      10830      10840      10850

                >< StyI
                >< EcoT14I
                >< Eco130I
                >< BssT1I
                >< MboII
                >< MaeIII>< BsaJI
TGAGTTTACA CCATTTGATG TTGTTAGACA ATGCTCTGGT GTTACCTTCC AAGGTAAGTT CAAGAAAATT
10860      10870      10880      10890      10900      10910      10920

                >< SfaNI
                >< SduI
                >< NspII
                >< Tru9I
                >< MseI
                >< Tru9I>< Bsp1286I
                >< MseI>< BmyI
                >< FokI
                >< TfiI
                >< HinfI
                >< Csp6I><
                >< AfaI><
GTTAAGGGCA CTCATCATTG GATGCTTTTA ACTTTCTTGA CATCACTATT GATTCTTGTT CAAAGTACAC
10930      10940      10950      10960      10970      10980      10990

                >< XmnI
                >< BsmI
                >< BscCI
                >< Asp700I
                >< BbvI
                >< BbvI><
                >< MaeIII
AGTGGTCACT GTTTTCTTT GTTTACGAGA ATGCTTTCTT GCCATTTACT CTTGGTATTA TGGCAATTGC
11000      11010      11020      11030      11040      11050      11060

                >< NspI
                >< NspHI
                >< NlaIII
                >< BspWI
                >< Fnu4HI>< BspWI
                >< BscCI
                >< MaeIII
                >< Tru9I
                >< MseI
                >< AccI>< NspHI>< AluI
                >< BsaBI>< NlaIII
TGCATGTGCT ATGCTGCTTG TTAAGCATAA GCACGCATTC TTGTGCTTGT TTCTGTTACC TTCTCTTGCA
11070      11080      11090      11100      11110      11120      11130

                >< SfaNI
                >< RmaI
                >< NspI
                >< NlaIII
                >< NheI
                >< MaeI
                >< BsiBI
                >< BspHI
                >< MamI
                >< HphI
                >< NlaIII
                >< BsaBI>< NlaIII
ACAGTTGCTT ACTTTAATAT GGTCTACATG CCTGCTAGCT GGGTGATGCG TATCATGACA TGGCTTGAAT
11140      11150      11160      11170      11180      11190      11200

```

FIGURE 13.25

```

                                >< Tru9I
                                >< MseI
                                >< Esp4I
                                >< Eco57I
                                >< AluI
                                >< AflIII
                                >< AluI
TGGCTGACAC TAGCTTGTCT GGTATATAGGC TTAAGGATTG TGTATGTAT GCTTCAGCTT TAGTTTGTCT
11210      11220      11230      11240      11250      11260      11270

                                >< RmaI
                                >< MaeII
                                >< MaeI
                                >< NlaIII
                                >< SfaNI
                                >< Fnu4HI
                                >< BspHI
                                >< AluI
                                >< BbvI
                                >< AflIII
TATTCTCATG ACAGCTCGCA CTGTTTATGA TGATGCTGCT AGACGTGTTT GGACACTGAT GAATGTCATT
11280      11290      11300      11310      11320      11330      11340

                                >< Sau96I
                                >< Pali
                                >< NspIV
                                >< NlaIII
                                >< HaeIII
                                >< DdeI
                                >< Sau3AI
                                >< NdeII
                                >< MboI
                                >< DpnII
                                >< DpnI
                                >< Bsp143I
                                >< BspAI
                                >< AluI
                                >< AsuI
                                >< Cfr13I
                                >< BsuRI
                                >< BsiZI
                                >< BshI
                                >< BfrI
ACACTTGTTT ACAAAGTCTA CTATGGTAAT GCTTTAGATC AAGCTATTTC CATGTGGGCC TTAGTTATTT
11350      11360      11370      11380      11390      11400      11410

                                >< RmaI
                                >< NlaIII
                                >< MaeI
                                >< SfcI
                                >< AluI
                                >< AluI
                                >< MaeIII
                                >< MnlI
                                >< MaeIII
CTGTAACCTC TAACTATTCT GGTGTCGTTA CGACTATCAT GTTTTATAGCT AGAGCTATAG TGTTTGTGTG
11420      11430      11440      11450      11460      11470      11480

                                >< BsrI
                                >< NlaIII
                                >< BfrI
                                >< DdeI
TGTTGAGTAT TACCCATTGT TATTTATTAC TGGCAACACC TTACAGTGTA TCATGCTTGT TTATTGTTTC
11490      11500      11510      11520      11530      11540      11550

                                >< Pali
                                >< HaeIII
                                >< Fnu4HI
                                >< BsuRI
                                >< BbvI
                                >< Fnu4HI
                                >< BspWI
                                >< BbvI
                                >< BspWI
                                >< BshI
                                >< Eco57I
                                >< MaeIII
TTAGGCTATT GTTGCTGCTG CTACTTTGGC CTTTCTGTT TACTCAACCG TTAGTTCAGG CTTACTCTTG
11560      11570      11580      11590      11600      11610      11620

                                >< ScrFI
                                >< MvaI
                                >< EcoRII
                                >< Ecl136I
                                >< DsaV
                                >< BstOI
                                >< BstNI
                                >< BsiLI
                                >< BsaJI
                                >< BsaJI
                                >< Eco31I
                                >< BsmAI
                                >< BsaI

```

FIGURE 13. 26

```

>< DrdI >< Alw26I
GTGTTTATGA CTACTTGGTC TCTACACAAG AATTTAGGTA TATGAAGTCC CAGGGGCTTT TGCCTCCTAA
11630 11640 11650 11660 11670 11680 11690

>< Tru9I
>< MseI
>< SfaNI >< HindIII> < Tru9I
>< MnlI >< AluI >< MseI >< MnlI >< NlaIII
GAGTAGTATT GATGCTTCA AGCTTAACAT TAAGTTGTTG GGTATTGGAG GTAAACCATG TATCAAGGTT
11700 11710 11720 11730 11740 11750 11760

>< VneI
>< SnaI
>< SduI
>< NspII
>< HgiAI
>< Bsp1286I
>< BmyI >< RsaI
>< RsaI >< ApaLI >< MboII
>< Csp6I >< Alw44I >< Csp6I DdeI >
>< AfaI >< MaeII >< Alw21I >< AfaI BfrI >
GCTACTGTAC AGTCTAAAT GTCTGACGTA AAGTGCACAT CTGTGGTACT GCTCTCGGTT CTCAACAAC
11770 11780 11790 11800 11810 11820 11830

>< NspII> < RsaI
>< DraIII
>< SduI>< Csp6I
>< Bsp1286I
>< MboII >< HinfI >< PfuI >< BmyI >< AfaI >< MboII
TTAGAGTAGA GTCATCTTCT AAATTGTGGG CACAATGTGT ACAACTCCAC AATGATATTC TTCTTGCAAA
11840 11850 11860 11870 11880 11890 11900

>< TthHB8I
>< TaqI
>< HindIII >< MboII SfcI ><
>< AluI >< Eco57I >< NlaIII
AGACACAACT GAAGCTTTCG AGAAGATGGT TTCTCTTTTG TCTGTTTTGC TATCCATGCA GGGTGCTGTA
11910 11920 11930 11940 11950 11960 11970

>< VspI
>< Tru9I >< Ksp632I
>< MseI >< TthHB8I >< EarI
>< AsnI >< TaqI >< MboII >< Eam1104I
>< AseI>< MnlI >< BcgI/a >< Eco57I >< Eco57I >< BcgI
GACATTAATA GGTGTGCGA GGAAATGCTC GATAACCGTG CTACTCTTCA GGCTATTGCT TCAGAATTTA
11980 11990 12000 12010 12020 12030 12040

>< StuI
>< ScrFI
>< Pali
>< MvaI>< HaeIII
>< EcoRII>< Eco147I
>< Ecl136I
>< DsaV >< BsuRI
>< BstOI
>< BstNI
>< BspWI
>< EsiLI
>< Fnu4HI >< BsaJI >< BshI TfiI ><
>< NdeI >< BspWI>< MnlI >< BglI >< SfcI HinfI ><
>< AciI >< ApyI>< AatI >< AluI

```

FIGURE 13. 27

```

GTTCTTTACC ATCATATGCC GCTTATGCCA CTGCCCAGGA GGCCTATGAG CAGGCTGTAG CTAATGGTGA
12050      12060      12070      12080      12090      12100      12110

      >< XmnI      >< Tru9I      >< SfaNI
      >< HphI      >< MseI      >< DdeI
      >< Asp700I   >< Eco57I   >< BbvI Fnu4HI ><
TTCTGAAGTC GTTCTCAAAA AGTTAAAGAA ATCTTTGAAT GTGGCTAAAT CTGAGTTGA CCGTGATGCT
12120      12130      12140      12150      12160      12170      12180

                                      XhoII ><
                                      Sau3AI ><
                                      NdeII ><
                                      MnlI >
                                      >< MnlI
                                      >< MflI
                                      >< MboI
      > < Sau3AI
      > < NdeII
      > < MboI
      > < DpnII
      >< DpnI
      >< BspWI
      >< BspAI
      >< Bsp143I
      >< NlaIII
      >< Bsp6I
      >< AfaI BglIII ><
GCCATGCAAC GCAAGTTGGA AAAGATGGCA GATCAGGCTA TGACCCAAAT GTACAAACAG GCAAGATCTG
12190      12200      12210      12220      12230      12240      12250

      >< SpeI
      >< RmaI
      >< MaeIII
      >< MaeI
      >< MboII
      >< BspWI
      >< EarI>< BfrI >< AluI
AGGACAAGAG GGCAAAAGTA ACTAGTGCTA TGCAAACAAT GCTCTTCACT ATGCTTAGGA AGCTTGATAA
12260      12270      12280      12290      12300      12310      12320

      >< ThaI
      >< MvnI
      >< HinPII
      >< Hin6I
      >< HhaI
      >< CfoI
      >< BstUI
      >< Tru9I
      >< MseI
      >< Bsp50I
      >< AccII
      >< SfcI ><
TGATGCACTT AACAAACATTA TCAACAATGC GCGTGATGGT TGTGTTCCAC TCAACATCAT ACCATTGACT
12330      12340      12350      12360      12370      12380      12390

      >< RsaI
      >< NlaIV
      >< Eco64I
      >< Csp6I
      >< BslI
      >< BsiYI>< KpnI
      >< BscBI
      >< BanI
      >< Asp718
      >< AfaI
      >< NlaIII
      >< BstXI
      >< Fnu4HI >< BbvI
      >< AccB1I
      >< Acc65I
      >< MaeIII
      >< BsgI ><
ACAGCAGCCA AACTCATGGT TGTGTGCCCT GATTATGGTA CCTACAAGAA CACTTGTGAT -GGTAACACCT
12400      12410      12420      12430      12440      12450      12460

      >< Zsp2I
      >< Ppu10I

```

FIGURE 13.28

```

    >< NsiI
    >< Mph1103I
    >< NdeI>< EcoT22I
    >< AvaIII >< SfaNI
    >< SfaNI
    >< AciI
    DdeI ><
    BfrI ><
    TTACATATGC ATCTGCACTC TGGGAAATCC AGCAAGTTGT TGATGCGGAT AGCAAGATTG TTCAACTTAG
    12470 12480 12490 12500 12510 12520 12530

    >< Pali
    >< HaeIII >< MnlI >< DdeIDdeI ><
    >< BsuRI >< MaeIII >< BspWI
    >< Tru9I>< NlaIII
    >< MseI>< HphI >< XcmI>< BshI >< AluI BspWI ><
    TGAAATTAAC ATGGACAATT CACCAAATTT GGCTTGGCCT CTTATTGTTA CAGCTCTAAG AGCCAACTCA
    12540 12550 12560 12570 12580 12590 12600

    RsaI ><
    NlaIV ><
    KpnI ><
    >< Fnu4HI
    Eco64I ><
    Csp6I ><
    BscBI ><
    Asp718 ><
    AfaI ><
    >< AciI>< BanI
    AccB1I ><
    >< Tru9I
    >< PvuII
    >< Psp5I
    >< NspBII
    >< MseI
    >< AluI >< SfcI
    >< DdeI>< BsrI
    >< PshAI
    Acc65I ><
    GCTGTAAAC TACAGAATAA TGAAGTGTAGT CCAGTAGCAC TACGACAGAT GTCCTGTGCG GCTGGTACCA
    12610 12620 12630 12640 12650 12660 12670

    >< TthHB8I
    >< TaqI
    >< SfuI
    >< NspV
    >< MnlI
    >< LspI
    >< Csp45I
    >< BstBI
    >< Bsp119I
    >< BsiCI
    >< Bpu14I
    >< AsuII
    >< RsaI
    >< Csp6I
    >< AluI
    >< AfaI
    CACAAACAGC TTGTACTGAT GACAATGCAC TTGCCTACTA TAACAATTTCG AAGGGAGGTA GGTTCGTGCT
    12680 12690 12700 12710 12720 12730 12740

    >< XhoII
    >< Sau3AI
    >< NdeII
    >< MflI
    >< MboI
    >< DpnII
    >< DpnI
    >< BstYI
    >< BspAI
    >< Bsp143I
    >< BglII
    >< TfiI
    >< RmaI
    >< HinfI
    >< MaeI
    >< DdeI
    >< RsaI
    >< Csp6I
    >< Csp6I>< RsaI
    >< AfaI>< AfaI
    GGCATTACTA TCAGACCACC AAGATCTCAA ATGGGCTAGA TTCCCTAAGA GTGATGGTAC AGGTACAATT
    12750 12760 12770 12780 12790 12800 12810

    >< Sau96I
    >< PssI
    >< Pali
    >< NspIV

```

FIGURE 13.29

```

                                >< HaeIII
                                >< EcoO109I
                                >< DraII
                                >< CfrI3I
                                >< BsuRI
                                >< Bsi2I      RsaI >
                                >< BshI      Csp6I ><
                                >< AsuI      AfaI >
TACACAGAAC TGGAACCACC TTGTAGGTTT GTTACAGACA CACCAAAAGG GCCTAAAGTG AAATACTTGT
12820      12830      12840      12850      12860      12870      12880

                                >< SfcI
                                > < MboII
                                MaeII ><
                                >< Fnu4HI >< RsaI
                                >< Eco57I >< Csp6I
                                > < BbsI
                                >< Tru9I
                                >< MseI >< MnlI
                                >< BbvI      >< AluI      >< AfaI
ACTTCATCAA AGGCTTAAAC AACCTAAATA GAGGTATGGT GCTGGGCAGT TTAGCTGCTA CAGTACGTCT
12890      12900      12910      12920      12930      12940      12950

                                >< RsaI
                                >< SfcI >< Csp6I
                                >< BspWI      >< AfaI      >< BspMI
TCAGGCTGGA AATGCTACAG AAGTACCTGC CAATTCAACT GTGCTTTCCT TCTGTGCTTT TGCAGTAGAC
12960      12970      12980      12990      13000      13010      13020

                                >< RmaI
                                >< MnlI
                                >< MaeI      >< HphI
CCTGCTAAAG CATATAAGGA TTACCTAGCA AGTGAGGAGC AACCAATCAC CAACTGTGTG AAGATGTTGT
13030      13040      13050      13060      13070      13080      13090

                                >< SinI
                                >< Sau96I
                                >< NspIV
                                >< NspHII
                                >< NlaIII
                                >< Eco47I
                                >< Eam1105I
                                >< CfrI3I
                                >< Bsi2I
                                >< Bme18I >< XcmI
                                >< AvaII      PleI ><
                                >< AfaI      >< AfaI      >< MaeIII      >< AluI      >< AsuI > < HinfI
GTACACACAC TGGTACAGGA CAGGCAATTA CTGTACACC AGAAGCTAAC ATGGACCAAG AGTCCTTTGG
13100      13110      13120      13130      13140      13150      13160

                                >< TfiI
                                >< MaelIII
                                >< SfaNI
                                >< NlaIII      >< FokI
                                >< HinfI
TGGTGCTTCA TGTTGTCTGT ATTGTAGATG CCACATTGAC CATCCAAATC CTAAGGATT CTGTGACTTG
13170      13180      13190      13200      13210      13220      13230

                                > < RsaI
                                >< MaeII
                                >< Csp6I
                                > < AfaI
                                >< DdeI
                                >< BfrI
AAAGGTAAGT ACGTCCAAAT ACCTACCACT TGTGCTAATG ACCCAGTGGG TTTTACACTT AGAAACACAG
13240      13250      13260      13270      13280      13290      13300

                                >< ThaI

```

FIGURE 13.30


```

                                >< SfaNI
                                >< MvnI
                                >< BstUI
                                >< Bsp50I
                                >< AciI
                                >< SfaNI ><
                                >< MaeIII ><
                                >< AccIISfaNI ><
                                ><
>< RsaI
>< Csp6I
>< AfaI >< AciI
TCTGTACCGT CTGCGGAATG TGGAAAGGTT ATGGCTGTAG TTGTGACCAA CTCCGCGAAC CCTTGATGCA
13310 13320 13330 13340 13350 13360 13370

                                >< Zsp2I
                                >< SfaNI
                                >< Mph1103I>< Tru9I
                                >< Ppu10I>< MaeII
                                >< NsiI>< FokI
                                >< EcoT22I >< MseI
                                >< Fnu4HI ><
                                >< BsgI ><
                                >< BbvI
                                ><
>< AciI>< AvaIII >< DraI >< AciI >< Fnu4HI >< AciI ><
GTCTGCGGAT GCATCAACGT TTTTAAACGG GTTTGCGGTG TAAGTGACGC CCGTCTTACA CCGTGCGGCA
13380 13390 13400 13410 13420 13430 13440

>< SpeI
>< ScaI
>< RsaI
>< RmaI
>< MaeI
>< Csp6I >< SfcI >< BspWI
>< BspWI >< AfaI >< AccI >< BcgI/a >< BcgI ><
CAGGCACTAG TACTGATGTC GTCTACAGGG CTTTGTATAT TTACAACGAA AAAGTTGCTG GTTTTGCAAA
13450 13460 13470 13480 13490 13500 13510

                                >< ScrFI
                                >< MvaI
                                >< MnlI
                                >< EcoRII
                                >< Ecl136I
                                >< BstOI
                                >< BstNI
                                >< BslI
                                >< DsaV >< BsiYI
                                >< BsiLI
                                >< ApyI
                                >< PleI
                                >< FokI >< HinfI
GTTCCTAAAA ACTAATTGCT GTCGCTTCCA GGAGAAGGAT GAGGAAGGCA ATTTATTAGA CTCTTACTTT
13520 13530 13540 13550 13560 13570 13580

                                >< NlaIII
                                >< Ksp632I
                                >< EarI
                                >< Eam1104I
                                >< BsmAI
                                >< Tru9I
>< Tru9I
>< MseI
>< MnlI
>< Alw26I >< MboII >< MseI
GTAGTTAAGA GGCATACTAT GTCTAACTAC CAACATGAAG AGACTATTTA TAACTTGTTT AAAGATTGTC
13590 13600 13610 13620 13630 13640 13650

                                >< RsaI
                                >< NlaIV
                                >< NlaIII
                                >< KpnI
                                >< HphI
                                >< Eco64I
                                >< Csp6I
                                >< BscBI
                                >< BanI
                                >< Asp718

```

FIGURE 13.31

```

>< NspBII
>< AclI
CAGCGGTTGC TGTCCATGAC TTTTCAAGT TTAGAGTAGA TGGTGACATG GTACCACATA TATCACGTCA
13660 13670 13680 13690 13700 13710 13720

>< MaeIII >< AfaI
> < AccBII MaeII ><
> < Acc65I > < HgaI
GCGTCTAACT AAATACACAA TGGCTGATTT AGTCTATGCT CTACGTCATT TTGATGAGGG TAATTGTGAT
13730 13740 13750 13760 13770 13780 13790

>< MnlI
>< MaeII
GCGTCTAACT AAATACACAA TGGCTGATTT AGTCTATGCT CTACGTCATT TTGATGAGGG TAATTGTGAT
13730 13740 13750 13760 13770 13780 13790

>< Tru9I
>< MseI >< MaeIII >< MunI
ACATTAAGAG AAATACTCGT CACATACAAT TGCTGTGATG ATGATTATTT CAATAAGAAG GATTGGTATG
13800 13810 13820 13830 13840 13850 13860

>< Thai
>< MvnI
>< MluI
>< BstUI
>< Bsp50I
>< RsaI
>< HphI
>< TfiI >< AflIII >< DdeI >< Csp6I Tru9I ><
>< HinfI >< AccII >< BfrI >< AfaI MseI ><
ACTTCGTAGA GAATCCTGAC ATCTTACGCG TATATGCTAA CTTAGGTGAG CGTGACGCC AATCATTATT
13870 13880 13890 13900 13910 13920 13930

XhoII >
Sau3AI >
NdeII >
MflI >
MboI >
DpnII >
BstYI >
BspAI >
> < SfaNI
>< RsaI
>< Csp6I
>< AfaI >< SfaNI
AAAGACTGTA CAATTCTGCG ATGCTATGCG TGATGCAGGC ATTGTAGGCG TACTGACATT AGATAATCAG
13940 13950 13960 13970 13980 13990 14000

> < ScrFI
> < MvaI
>< Fnu4HI
>< EcoRII
> < Ecl136I
> < BstOI
> < BstNI
>< Tru9I
>< MseI >< RsaI >< BslI
>< DpnI >< Csp6I >< HphI >< BsiYI
>< Bsp143I >< BsrI >< Csp6I >< BsiLI
>< AlwI >< AfaI >< BbvI >< ApyI
GATCTTAATG GGAAGTGGTA CGATTTCGGT GATTTCGTAC AAGTAGCACC AGGCTGCGGA GTTCCTATTG
14010 14020 14030 14040 14050 14060 14070

>< SfaNI
>< HinfI
>< Fnu4HIpleI ><
>< DdeI
>< BspWI NdeI ><
TGGATTCATA TTAATCATTG CTGATGCCCA TCCTCACTTT GACTAGGGCA TTGGCTGCTG AGTCCCATAT
14080 14090 14100 14110 14120 14130 14140

>< Sau3AI
>< NdeII

```

FIGURE 13.32

```

>< MboI
>< MamI
>< DpnII
>< DpnI
>< BspWI
>< BspAI
>< BspI43I
>< BsiBI
>< BsaBI >< FokI
GGATGCTGAT CTCGCAAAAC CACTTATTAA GTGGGATTG CTGAAATATG ATTTTACGGA AGAGAGACTT
14150 14160 14170 14180 14190 14200 14210

>< Ksp632I
>< Eam1104I
>< BsmAI
>< EarI >< AspI ><
>< Alw26I

>< SinI
>< Sau96I
>< NspIV
>< NspHII
>< NlaIV
>< TthHB8I
>< TaqI
>< McrI
>< Ksp632I
>< EarI
>< Eam1104I
>< BsmAI
>< MboII
>< Alw26I
TGTCTCTTCG ACCGTTATTT TAAATATTGG GACCAGACAT ACCATCCCAA TTGTATTAAC TGTTTGGATG
14220 14230 14240 14250 14260 14270 14280

>< FokI
>< MseI
>< DraI
>< AsuI
>< MunI
>< MseI
>< Tru9I
>< BsiEI
>< MseI
>< AvaII
>< Bsi2I
>< Bmel8I
>< AvaII
>< AsuI
>< SinI
>< Sau96I
>< NspIV
>< NspHII
>< Eco47I
>< Cfr13I
>< Bsi2I
>< Bmel8I
>< AvaII
>< AsuI
ATAGGTGTAT CCTTCATTGT GCAAACTTTA ATGTGTTATT TTCTACTGTG TTCCACCTA CAAGTTTGG
14290 14300 14310 14320 14330 14340 14350

>< SpeI
>< RmaI
>< MaeI
>< SspI
>< BsrI
ACCACTAGTA AGAAAAATAT TTGTAGATGG TGTTCTTTT GTTGTITCAA CTGGATACCA TTTTCGTGAG
14360 14370 14380 14390 14400 14410 14420

>< RsaI
>< HinfI >< PfuI
>< Csp6I
>< AfaI
>< HgaI >< AluI
>< FokI
>< AccII
>< BbvI
TTAGGAGTCG TACATAATCA GGATGTAAAC TTACATAGCT CGCGTCTCAG TTTCAAGGAA CTTTGTAGTG
14430 14440 14450 14460 14470 14480 14490

>< 2sp2I
>< SphI
>< Ppu10I
>< PaeI
>< NspI

```

FIGURE 13.33

```

>< Sau3AI      >< NspHI
>< NdeII      >< NsiI
>< MboI      >< NlaIII
>< DpnII     >< Mph1103I
> < DpnI     >< Fnu4HI
>< Fnu4HI>< BspWI >< EcoT22I
>< BspAI      >< BspWI
> < Bsp143I> < AvalIII > < AlwNI
>< AlwI      >< AluI      >< AluI >< BbvI >< MaeI
ATGCTGCTGA TCCAGCTATG CATGCAGCTT CTGGCAATTT ATTGCTAGAT AAACGCACTA CATGCTTTTC
14500      14510      14520      14530      14540      14550      14560

>< ScrFI
>< NciI
>< MspI
>< HpaII
>< HapII
>< Fnu4HI
>< AlwNI
>< AluI
AGTAGCTGCA CTAACAAACA ATGTTGCTTT TCAAAGTGTG AAACCCGGTA ATTTTAATAA AGACTTTTAT
14570      14580      14590      14600      14610      14620      14630

>< Tru9I
>< MseI
GACTTTGCTG TGTCTAAAGG TTTCTTTAAG GAAGGAAGTT CTGTTGAACT AAAACACTTC TTCTTTGCTC
14640      14650      14660      14670      14680      14690      14700

>< FokI
>< Fnu4HI
AGGATGGCAA CGCTGCTATC AGTGATTATG ACTATTATCG TTATAATCTG CCAACAATGT GTGATATCAG
14710      14720      14730      14740      14750      14760      14770

>< VspI
>< Tru9I
>< MseI
>< AsnI
>< AseI
>< MaeIII
ACAACCTCTA TTCGTAGTTG AAGTTGTTGA TAAATACTTT GATTGTTACG ATGGTGGCTG TATTAATGCTC
14780      14790      14800      14810      14820      14830      14840

>< Tru9I
>< MseI
>< HpaI
>< HindII
>< HincII
AACCAAGTAA TCGTTAACAA TCTGGATAAA TCAGCTGGTT TCCCATTTAA TAAATGGGGT AAGGCTAGAC
14850      14860      14870      14880      14890      14900      14910

>< SfaNI
>< Sau3AI
>< NdeII
>< MboI
>< DpnII
>< DpnI
>< Bsp143I
>< PleI
>< HinfI>< MnlI
>< BspAI >< AlwI
TTTATTATGA CTCAATGAGT TATGAGGATC AAGATGCACT TTTCGCGTAT ACTAAGCGTA ATGTCATCCC
14920      14930      14940      14950      14960      14970      14980

>< SstI
>< SduI
>< SacI

```

FIGURE 13.34

```

                                >< NspII
                                >< HgiAI
                                >< Eco24I
                                >< Ecl136II
                                >< Bsp1286I
                                >< BmyI
                                >< BanII
                                >< Alw21I
                                >< AluI
                                >< AluI
                                >< BspWI
                                >< AflIII
                                >< Esp4I
                                >< HinfI
                                >< MseI
                                >< TfiI
                                >< Tru9I
TACTATAACT CAAATGAATC TTAAGTATGC CATTAGTGCA AAGAATAGAG CTCGCACCGT AGCTGGTGTG
14990      15000      15010      15020      15030      15040      15050

                                RmaI ><
                                >< MnlI
                                MaeI ><
                                >< Fnu4HI
                                >< AciI
TCTATCTGTA GTACTATGAC AAATAGACAG TTTCATCAGA AATTATTGAA GTCAATAGCC GCCACTAGAG
15060      15070      15080      15090      15100      15110      15120

                                >< Tru9I
                                >< MseI
GAGCTACTGT GGTAATTGGA ACAAGCAAAGT TTTACGGTGG CTGGCATAAT ATGTTAAAAA CTGTTTACAG
15130      15140      15150      15160      15170      15180      15190

                                NspI ><
                                NspHI ><
                                NlaIII ><
                                >< NlaIII
                                DdeI ><
                                BspWI ><
                                >< MaeIII
                                BfrI ><
TGATGTAGAA ACTCCACACC TTATGGGTTG GGATTATCCA AAATGTGACA GAGCCATGCC TAACATGCTT
15200      15210      15220      15230      15240      15250      15260

                                >< Pali
                                >< HaeIII
                                >< BsuRI
                                >< BshI
                                >< MnlI
                                >< MaeIII
                                SfcI ><
AGGATAATGG CCTCTCTGTG TCTTGCTCGC AAACATAACA CTTGCTGTAA CTTATCACAC CGTTTCTACA
15270      15280      15290      15300      15310      15320      15330

                                Tru9I ><
                                ScrFI >
                                MvaI >
                                >< MseI
                                FokI ><
                                EcoRII ><
                                Ecl136I >
                                DsaV ><
                                BstOI >
                                BstNI >
                                >< NlaIII
                                >< Fnu4HI
                                BsiLI >
                                >< AciI
                                ApyI >
GGTAGCTAA CGAGTGTGCG CAAGTATTAA GTGAGATGGT CATGTGTGGC GGCTCACTAT ATGTTAAACC
15340      15350      15360      15370      15380      15390      15400

                                >< SfaNI
                                >< MspI
                                >< HpaII
                                >< HapII
                                >< HphI
                                >< BspWI
                                >< Tru9I
                                MaeIII ><
                                >< MseI
                                AluI ><

```

FIGURE 13.35

```

AGGTGGAACA TCATCCGGTG ATGCTACAAC TGCTTATGCT AATAGTGTCT TTAACATTG TCAAGCTGTT
15410      15420      15430      15440      15450      15460      15470

>< BspWI                                     >< DrdI
ACAGCCAATG TAAATGCACT TCTTTCAACT GATGGTAATA AGATAGCTGA CAAGTATGTC CGCAATCTAC
15480      15490      15500      15510      15520      15530      15540

                                     >< AluI       > < AciI
                                     >< Sau3AI
                                     >< NdeII
                                     >< MboI
> < MamI
                                     >< FbaI
                                     >< DpnII
                                     >< DpnI
                                     >< BspHI
                                     >< BspAI
                                     >< Bsp143I
                                     >< BsiQI
                                     >< SfcI       > < BsiBI>< NlaIII
                                     >< BsmAI       > < BsaBI>< FokI
                                     >< Alw26I      >< BclI>< EcoRI       FokI ><
AACACAGGCT CTATGAGTGT CTCTATAGAA ATAGGGATGT TGATCATGAA TTCGTGGATG AGTTTTACGC
15550      15560      15570      15580      15590      15600      15610

                                     >< TfiI
                                     >< SfaNI
                                     >< NlaIII
                                     >< BspMI       >< HinfI       >< MaeIII
TTACCTGCGT AAACATTTCT CCATGATGAT TCTTTCTGAT GATGCCGTTG TGTGCTATAA CAGTAACTAT
15620      15630      15640      15650      15660      15670      15680

                                     > < RmaI
                                     >< NheI >< Tru9I
>< Fnu4HI       > < MaeI       >< Tru9I
>< AciI       >< AluI >< MseI >< MseI       MnlI ><
GCGGCTCAAG GTTTAGTAGC TAGCATTAAAG AACTTTAAGG CAGTTCTTTA TTATCAAAAT AATGTGTTCA
15690      15700      15710      15720      15730      15740      15750

                                     >< SinI
                                     >< Sau96I
                                     >< PssI
                                     >< Psp5II
                                     >< PpuMI
                                     >< NspIV
                                     >< NspHII
                                     >< Eco0109I
                                     >< Eco47I
                                     >< DraII
                                     >< Cfr13I
                                     >< BsiZI
                                     >< Bme18I
                                     >< AvaII
                                     >< AsuI       >< MnlI
>< NlaIII       >< BsmAI       >< Alw26I
TGCTGAGGC AAAATGTTGG ACTGAGACTG ACCTTACTAA AGGACCTCAC GAATTTTGCT CACAGCATAC
15760      15770      15780      15790      15800      15810      15820

                                     >< XhoII
                                     >< Sau3AI
                                     >< NdeII
                                     >< MflI
                                     >< MboI

```

FIGURE 13. 36

```

                >< RsaI          >< DpnII
                >< MaeII        >< DpnI   > < SspI
                >< Csp6I        >< BstYI   HinPII ><
    >< Tru9I
    >< RmaI          >< BsaAI        >< BspMI   Hin6I ><
    >< MaeI          >< AflIII       >< BspAI    HhaI ><
    >< BspWI>< MseI          >< AfaI    >< AlwI>< Bsp143I   CfoI ><
    AATGCTAGTT AAACAAGGAG ATGATTACGT GTACCTGCCT TACCCAGATC CATCAAGAAT ATTAGGCGCA
    15830      15840      15850      15860      15870      15880      15890

                >< RsaI          >< SfaNI
                >< TthHB8I        >< Csp6I        >< MaeIII
                >< TaqI          >< AfaI          BsrI ><
    GGCTGTTTTG TCGATGATAT TGTCAAAACA GATGGTACAC TTATGATTGA AAGGTTCTGT TCACTGGCTA
    15900      15910      15920      15930      15940      15950      15960

    > < FokI
    >< BspWI
    TTGATGCTTA CCCACTTACA AAACATCCTA ATCAGGAGTA TGCTGATGTC TTTCACCTGT ATTTACAATA
    15970      15980      15990      16000      16010      16020      16030

                >< Van91I
                >< PflMI
                >< NspI
                > < Pali>< NspHI
                > < MscI>< NlaIII
                > < HaeIII
                > < BsuRI
                >< BsrI
                >< EaeI   >< BslI   >< NspI
                > < BshI>< BsiYI >< NspHI
                >< NlaIII   >< AflIII >< AflIII
    >< MaeIII   >< AluI > < BalI>< AccB7I >< NlaIII
    CATTAGAAAG TTACATGATG AGCTTACTGG CCACATGTTG GACATGTATT CCGTAATGCT AACTAATGAT
    16040      16050      16060      16070      16080      16090      16100

                >< RsaI> < NlaIV
                >< MnlI
                >< Csp6I   >< DdeI          >< RsaI
                >< BsrI >< MnlI          >< Csp6I
                >< AfaI> < BscBI          >< AfaI          SfcI ><
    AACACCTCAC GGTACTGGGA ACCTGAGTTT TATGAGGCTA TGTACACACC ACATACAGTC TTGCAGGCTG
    16110      16120      16130      16140      16150      16160      16170

                >< NlaIV
                >< EcoNI
                >< Eco31I
                >< Eco64I>< BsmAI
                >< BscBI >< BslI
                >< BanI   >< BsiYI
                >< AclI   >< BsaI
                >< BspWI
                >< AccB1I>< Alw26I   BbvI ><
    TAGGTGCTTG TGTATTGTGC AATTCACAGA CTTCACTTCG TTGCGGTGCC TGTATTAGGA GACCATTCTT
    16180      16190      16200      16210      16220      16230      16240

                >< Tth111I
                >< Fnu4HI   >< NlaIII
                >< BspWI >< AspI          > < Tru9I
                >< BspWI >< AspI          > < MseI
    ATGTTGCAAG TGCTGCTATG ACCATGTGAT TTCAACATCA CACAAATTAG TGTTGTCTGT TAATCCCTAT
    16250      16260      16270      16280      16290      16300      16310

                >< ScrFI
                >< MvaI

```

FIGURE 13.37

```

>< EcoRII
>< Ecl136I
>< DsaV
>< BstOI
>< BstNI
>< BsiLI
>< BsaJI
>< ApyI
>< MaeIII >< MaeIII
>< MaeI
>< AluI
GTTTGCAATG CCCAGGTTG TGATGTCAC TATGTGACAC AACTGTATCT AGGAGGTATG AGCTATTATT
16320 16330 16340 16350 16360 16370 16380

>< MaeIII >< MnlI
GCAAGTCACA TAAGCCTCCC ATTAGTTTTC CATTATGTGC TAATGGTCAG GTTTTGGTT TATACAAAAA
16390 16400 16410 16420 16430 16440 16450

>< NspI >< NspI
>< NspHI >< Tth111I >< NspHI
>< NlaIII>< MaeIII>< MaeIII >< NlaIII
>< AflIII >< AspI >< AflIII
CACATGTGTA GGCAGTGACA ATGTCAC TTTCAATGCG ATAGCAACAT GTGATTGGAC TAATGCTGGC
16460 16470 16480 16490 16500 16510 16520

>< RsaI
>< P1eI
>< DdeI
>< Csp6I
>< BsmAI >< HinfI >< MnlI
>< Alw26I >< HindIII DdeI ><
>< AfaI >< AluI >< Fnu4HI >< BbvI
GATTACATAC TTGCCAACAC TTGTACTGAG AGACTCAAGC TTTTCGCAGC AGAAACGCTC AAAGCCACTG
16530 16540 16550 16560 16570 16580 16590

>< Thai
>< ScaI
>< RsaI >< RsaI
>< MvnI
>< Csp6I >< Csp6I
>< BstUI
>< Bsp50I
>< Tru9I
>< MseI >< NdeI >< AfaI >< AfaI
>< AluI >< AccII MnlI >
AGGAAACATT TAAGCTGTCA TATGGTATTG CCACTGTACG CGAAGTACTC TCTGACAGAG AATTGCATCT
16600 16610 16620 16630 16640 16650 16660

MaeIII ><
>< MaeIII
>< EcoO65I
>< Eco9II
>< BstPI
>< BstEII
>< BsrI
>< SfaNI >< RsaI
>< NlaIII >< MaeI
TTCATGGGAG GTTGGAAAAC CTAGACCACC ATTGAACAGA AACTATGTCT TTACTGGTTA CCGTGTAACT
16670 16680 16690 16700 16710 16720 16730

RsaI ><
>< MnlI
>< RsaI >< RsaI
>< Csp6I >< Csp6I >< SfaNI Csp6I ><
>< AfaI >< AfaI >< MaeIII >< HphI AfaI ><
AAAAATAGTA AAGTACAGAT TGGAGAGTAC ACCTTTGAAA AAGGTGACTA TGGTGATGCT GTTGTGTACA
16740 16750 16760 16770 16780 16790 16800

```

FIGURE 13.38


```

    >< RsaI
    >< Csp6I
    >< AfaI
GAGGTACTAC GACATACAAG TTGAATGTTG GTGATTACTT TGTGTTGACA TCTCACACTG TAATGCCACT
16810      16820      16830      16840      16850      16860      16870

    >< HphI
    >< HindII
    >< HincII
    DdeI ><
    BfrI ><

    >< VneI
    >< SnaI
    >< SduI
    >< NspII
    >< HgiAI
    >< SduI
    >< NspII
    >< HgiAI
    >< DraIII
    >< Bsp1286I
    >< BmyI
    >< BspWI >< DraIII
    >< RsaI
    >< Csp6I
    >< ApaLI >< RmaI
    >< Alw44I >< MaeI
    >< Alw21I
    >< BmyI
    >< BsrI
    >< AfaI
    DdeI >
TAGTGCACCT ACTCTAGTGC CACAAGAGCA CTATGTGAGA ATTACTGGCT TGTACCCAAC ACTCAACATC
16880      16890      16900      16910      16920      16930      16940

    StyI ><
    SniI >
    Sau96I >
    NspIV >
    EcoT14I ><
    Eco47I >
    Eco130I ><
    >< ScaI Cfr13I >
    BssT1I ><
    >< SphI >< RsaI Bsi2I >
    >< PaeI BsaJI ><
    >< NlaIII Bme18I >
    >< NspI>< Csp6I AvaII >
    >< NspHI>< AfaI AsuI >
TCAGATGAGT TTTCTAGCAA TGTTGCAAAT TATCAAAAGG TCGGCATGCA AAAGTACTCT AACTCCAAG
16950      16960      16970      16980      16990      17000      17010

    >< ScrFI
    >< RsaI
    >< MvaI
    >< EcoRII
    >< Ecl136I
    >< Csp6I
    >< BstOI
    >< BstNI
    >< XcmI >< BslI
    >< NspHII >< BsiYI
    >< BsiLI
    >< ApyI >< BsrI
    >< DsaV>< AfaI >< HinfI>< PleI
GACCACCTGG TACTGGTAAG AGTCATTTTG CCATCGGACT TGCTCTCTAT TACCCATCTG CTCGCATAGT
17020      17030      17040      17050      17060      17070      17080

    >< SfaNI
    >< SphI >< PvuII
    >< PaeI >< Psp5I
    >< NspI >< NspBII
    >< NspHI >< Fnu4HI
    >< Tru9I
    >< Bst1107I >< NlaIII>< BspWI >< SspI
    >< AccI >< NlaIII >< AluI >< BbvI >< MseI
GTATACGGCA TGCTCTCATG CAGCTGTTGA TGCCCTATGT GAAAAGGCAT TAAATATTT GCCCATAGAT
17090      17100      17110      17120      17130      17140      17150

```

FIGURE 13.39

```

> < ThaI
>< ThaI
> < MvnI
>< MvnI >< ThaI
> < HinPII
>< HinPII
>< HinPII >< MvnI
> < Hin6I
>< Hin6I
> < HhaI
>< HhaI >< HhaI
> < CfoI
>< CfoI >< CfoI
> < BstUI
>< BstUI >< BstUI
>< BssHII
>< BspMI
> < Bsp50I
>< Bsp50I>< Bsp50I
>< TfiI >< Hin6I> < AccII
>< HinfI >< AccII >< AccII
> < EcoRI
AAATGTAGTA GAATCATACC TGC GCGTGCG CGCGTAGAGT GTTTGTGATAA ATTCAAAGTG AATTCAACAC
17160 17170 17180 17190 17200 17210 17220

>< Zsp2I
>< Ppu10I
>< NsiI
>< Mph1103I
>< EcoT22I
>< BsqI > < AvaIII >< DrdI
TAGAACAGTA TGTTTCTGCG ACTGTAAATG CATTGCCAGA AACAAGTCT GACATTGTAG TCTTTGATGA
17230 17240 17250 17260 17270 17280 17290

>< RmaI
>< MaeI >< MaeII
AATCTCTATG GCTACTAATT ATGACTTGAG TGTTGTCAAT GCTAGACTTC GTGCAAAACA CTACGTCTAT
17300 17310 17320 17330 17340 17350 17360

>< Sau3AI
>< NdeII
>< MboI
>< DpnII
>< DpnI
>< BspAI
>< AlwI>< Bsp143I > < AclI >< RmaI
>< MaeI SspI ><
ATTGGCGATC CTGCTCAATT ACCAGCCCC CGCACATTGC TGACTAAAGG CACACTAGAA CCAGAATATT
17370 17380 17390 17400 17410 17420 17430

>< SniI
>< Sau96I
>< NspIV >< StyI
>< NspHII >< NspI
>< Eco47I >< NspHI
>< Cfr13I >< NlaIII
>< BsiZl >< EcoT14I
>< BsgI >< Eco130I
>< Bme18I >< BssT1I
>< AvalI >< BsaJI
>< AsuI> < AflIII
>< Tru9I
>< MseI
TTAATTCAGT GTGCAGACTT ATGAAAACAA TAGGTCCAGA CATGTTCCCTT GGAAGTTGTC GCCGTTGTCC
17440 17450 17460 17470 17480 17490 17500

```

FIGURE 13. 40

```

    >< HindII
    >< HincII
    >< AluI
TGCTGAAATT GTTGACACTG TGAGTGCTTT AGTTTATGAC AATAAGCTAA AAGCACACAA GGATAAGTCA
17510      17520      17530      17540      17550      17560      17570

>< AluI
GCTCAATGCT TCAAAATGTT CTACAAAGGT GTTATTACAC ATGATGTTTC ATCTGCAATC AACAGACCTC
17580      17590      17600      17610      17620      17630      17640

    >< MnlI
>< EcoNI
    >< BslI
    >< BsiYI
    >< HphI
    >< AluI
AAATAGGCGT TGTAAGAGAA TTTCTTACAC GCAATCCTGC TTGGAGAAAA GCTGTTTTTA TCTCACCTTA
17650      17660      17670      17680      17690      17700      17710

    >< SfcI
    >< DdeI
    >< TfiI
    >< AluI
    >< BfrI
    >< HinfI
TAATTCACAG AACGCTGTAG CTTCAAAAAT CTTAGGATTG CCTACGCAGA CTGTTGATTC ATCACAGGGT
17720      17730      17740      17750      17760      17770      17780

    >< Tth111I
    >< AspI
    >< HindII
    >< HincII
    >< AclI
TCTGAATATG ACTATGTCAT ATTCACACAA ACTACTGAAA CAGCACACTC TTGTAATGTC AACCGCTTCA
17790      17800      17810      17820      17830      17840      17850

    >< XhoII
    >< Sau3AI
    >< NdeII
    >< MflI
    >< MboI
    >< MamI
    >< DpnII
    >< DpnI
    >< BstYI
    >< BspAI
    >< Bsp143I
    >< BsiBI
    >< BsaBI
    >< BspWI
    >< BglII
ATGTGGCTAT CACAAGGGCA AAAATTGGCA TTTTGTGCAT AATGTCTGAT AGAGATCTTT ATGACAAACT
17860      17870      17880      17890      17900      17910      17920

    >< XbaI
    >< RmaI
    >< MaeI
    >< MaeII
    >< MaeIII
    >< BsrI
GCAATTTACA AGTCTAGAAA TACCACGTCG CAATGTGGCT ACATTACAAG CAGAAAATGT AACTGGACTT
17930      17940      17950      17960      17970      17980      17990

    >< Sau3AI
    >< NdeII
    >< MboII
    >< MboI
    >< FokI
    >< DpnII
    >< DpnI
    >< BspAI
    >< Bsp143I
    >< NlaIV
    >< Eco64I
    >< BscBI
    >< BanI
    >< MnlI
    >< Tru9I
    >< MseI
    >< SfcI
    >< BbsI
    >< BsrI
    >< Acc81I
    >< DdeI

```

FIGURE 13. 41

```

TTTAAGGACT GTAGTAAGAT CATTACTGGT CTTTCATCCTA CACAGGCACC TACACACCTC AGCGTTGATA
18000      18010      18020      18030      18040      18050      18060

      >> ScrFI
      >> MvaI
      >> EcoRII
      >> Eco57I
      >> Ecl136I
      >> DsaV
      >> BstOI
      >> BstNI
      >> HindII >> BsiLI
      >> HincII >> ApyI
      >> PleI
      >> NlaIII
      HinfI >>
      AccI >>
TAAAGTTCAA GACTGAAGGA TTATGTGTTG ACATACCAGG CATAACAAAG GACATGACCT ACCGTAGACT
18070      18080      18090      18100      18110      18120      18130

      >> MaeIII
      >> EcoO65I
      >> Eco91I
      >> BstXI
      >> BstPI
      >> BstEII
      >> HphI
      >> AccII
      >> ThaI
      >> MvnI
      >> BstUI
      >> Bsp50I
      >> AciI
CATCTCTATG ATGGGTTTCA AAATGAATTA CCAAGTCAAT GGTACCCTA ATATGTTTAT CACCCGCGAA
18140      18150      18160      18170      18180      18190      18200

      >> XmnI
      >> MboII
      >> MaeIII
      >> Asp700I
      >> AluI
      >> MaeII
      >> MnlI
      >> SfaNI
      >> RmaI
      >> NlaIII
      >> MaeI
GAAGCTATTC GTCACGTTTCG TGCCTGGATT GGCTTTGATG TAGAGGGCTG TCATGCAACT AGAGATGCTG
18210      18220      18230      18240      18250      18260      18270

      >> Tru9I
      >> MseI
      >> HpaI
      >> RsaI
      >> GsuI
      >> RmaI
      >> HindII
      >> RsaI
      >> Csp6I
      >> MnlI
      >> HincII
      >> Csp6I
      >> BpmI
      >> MaeI
      >> DdeI
      >> AluI
      >> BsrI
      >> AfaI
      >> AluI
      >> SfcI
      >> BfrI
      >> AfaI
TGGGTACTAA CCTACCTCTC CAGCTAGGAT TTTCTACAGG TGTTAACTTA GTAGCTGTAC CGACTGGTTA
18280      18290      18300      18310      18320      18330      18340

      >> ScrFI
      >> MvaI
      >> MnlI
      >> MaeIII
      >> EcoRII
      >> EcoO65I
      >> EcoNI
      >> Eco91I
      >> Ecl136I
      >> DsaV
      >> Tru9I
      >> DraIII
      >> BstPI
      >> BstOI
      >> BstNI
      >> PmeI
      >> BstEII
      >> BslI
      >> MseI
      >> BsiYI
      >> HphI
      >> BsiLI
      >> DraI
      >> ApyI
      >> BsrI
      >> HindII
      >> HphI
      >> Tru9I
      >> HincII
      >> EcoRI
      >> MseI

```

FIGURE 13.42

```

TGTGACACT GAAAATAACA CAGAATTCAC CAGAGTTAAT GCAAAACCTC CACCAGGTGA CCAGTTTAAA
18350      18360      18370      18380      18390      18400      18410

      >< ScrFI
      >< MvaI
      >< EcoRII
      >< Ecl136I
      >< DsaV
      >< BstOI
      >< BstNI
      >< BsiLI
      >< BsaJI
      >< NlaIII
      >< ApyI
      >< Tru9I>< Csp6I
      >< MseI >< AfaI
CATCTTATAC CACTCATGTA TAAAGGCTTG CCCTGGAATG TAGTGCGTAT TAAGATAGTA CAAATGCTCA
18420      18430      18440      18450      18460      18470      18480

      >< NlaIII
      >< HinPII
      >< Tth111I
      >< Hin6I
      >< HinfI
      >< HhaI
      >< AspI >< PleI >< CfoI >< AluI
GTGATACACT GAAAGGATTG TCAGACAGAG TCGTGTTCGT CCTTGGGCG CATGGCTTTG AGCTTACATC
18490      18500      18510      18520      18530      18540      18550

      >< SinI
      >< Sau96I
      >< NspIV
      >< NspHII
      >< Eco47I
      >< Cfr13I
      >< ScaI
      >< BsiZI
      >< RsaI
      >< Bml18I
      >< Csp6I
      >< AvaII
      >< MaeII
      >< AfaI
      >< AsuI
      >< AflIII
      >< MaeIII>< MaeII
AATGAAGTAC TTTGTCAAGA TTGGACCTGA AAGAACGTGT TGTCTGTGTG ACAAACGTGC AACTTGCTTT
18560      18570      18580      18590      18600      18610      18620

      >< TfiI
      >< Tth111I
      >< HinfI
      >< AspI
TCTACTTCAT CAGATACTTA TGCCTGCTGG AATCATTCTG TGGGTTTTGA CTATGTCTAT AACCCATTTA
18630      18640      18650      18660      18670      18680      18690

      >< ScrFI
      RsaI ><
      >< MvaI
      >< EcoRII
      Ecl136I ><
      >< DsaV
      Csp6I ><
      BstXI ><
      >< MaeIII
      >< Eco65I
      >< Eco91I
      >< BstPI
      >< Eco57I> < BstEII
      >< MaeIII >< NlaIII
      >< AfaI ><
TGATTGATGT TCAGCAGTGG GGCTTTACGG GTAACCTTCA GAGTAACCAT GACCAACATT GCCAGGTACA
18700      18710      18720      18730      18740      18750      18760

      >< SfaNI
      >< RmaI
      >< NspI
      >< NspHI

```

FIGURE 13.43

```

                >< NlaIII                >< RmaI
                >< MaeI                >< NlaIII                Tru9I ><
    >< NlaIII    >< BspWI                >< MaeI                >< NlaIII
    > < AflIII    >< BspHI                MseI ><
    TGGAAATGCA CATGTGGCTA GTTGTGATGC TATCATGACT AGATGTTTAG CAGTCCATGA GTGCTTTGTT
    18770      18780      18790      18800      18810      18820      18830

    >< ThaI
    >< MvnI
    >< HinPII
    >< Hin6I
    >< HhaI
    >< CfoI
    >< BstUI                >< EcoNI> < MnlI
    >< Bsp50I                >< BslI                >< Tru9I
    >< AccII                >< BsiYI                >< DdeI >< MseI
    AAGCGCGTTG ATTGGTCTGT TGAATACCTT ATTATAGGAG ATGAAGTTAG GGTAAATTCT GCTTGCAGAA
    18840      18850      18860      18870      18880      18890      18900

    >< RsaI
    >< Csp6I
    >< AfaI                >< NlaIII                >< BspWI                >< MboII                > < NlaIII
    AAGTACAACA CATGGTTGTG AAGCTGTCAT TGCTTGCTGA TAAGTTTCCA GTTCTTCATG ACATTGGAAA
    18910      18920      18930      18940      18950      18960      18970

    >< SauI
    >< MstII
    >< Eco8II
    >< DdeI
    >< CvnI                NlaIII ><
    >< Bsu36I                >< EspI
    >< Bse2II                >< Eco57I MaeIII ><
    >< AxyI                >< DdeI
    >< AocI                >< CelII
    >< AocI                >< AocI    >< MnlI    >< SfaNI    >< Bpu1102I
    TCCAAAGGCT ATCAAGTGTG TGCCTCAGGC TGAAGTAGAA TGGAAGTTCT ACGATGCTCA GCCATGTAGT
    18980      18990      19000      19010      19020      19030      19040

    >< MnlI                >< Ksp632I
    >< HindIII                >< EarI
    >< AluI                >< MboII                >< Eam1104I
    GACAAAGCTT ACAAATAGA GGAAGTCTTC TATCTTATG CTACACATCA CGATAAATTC ACTGATGGTG
    19050      19060      19070      19080      19090      19100      19110

    >< Sau3AI
    >< NdeII
    >< MboI
    >< MaeII> < MaeIII
    >< DpnII
    >< DpnI
    >< BspAI
    >< MaeIII    >< Bsp143I                >< MunI                HinfI >
    TTTGTTTGTT TTGGAATTGT AACGTTGATC GTTACCCAGC CAATGCAATT GTGTGTAGGT TTGACACAAG
    19120      19130      19140      19150      19160      19170      19180

    >< ScrFI
    >< MvaI
    >< EcoRII                Zsp2I ><
                                >< SphI
                                > < Ppu10I
                                >< PaeI
                                >< NspI
                                >< NspHI
                                >< NlaIII
                                Mph1103I ><

```

FIGURE 1344

```

                >< Ecl136I
            >< DsaV
                >< BstOI
                >< BstNI
                >< BsiLI
                >< ApyI
        >< PleI
AGTCTTGTC AACTTGAAC TACCAGGCTG TGATGGTGGT AGTTTGTATG TGAATAAGCA TGCATTCCAC
19190      19200      19210      19220      19230      19240      19250

                >< Tru9I
                > < MunI
                >< TthHB8I
                >< MseI
>< BcgI/a >< TaqI
                >< DraI
                >< BcgI
                >< AluI
ACTCCAGCTT TCGATAAAAG TGCATTACT AATTAAAGC AATTGCCTTT CTTTACTAT TCTGATAGTC
19260      19270      19280      19290      19300      19310      19320

                >< PleI
                >< NlaIII
                >< BsmAI
                >< HinfI >< Alw26I
                SfaNI ><
                >< MaeII
                BsaAI ><
                AflIII ><
CTTGTGAGTC TCATGGCAAA CAAGTAGTGT CGGATATTGA TTATGTTCCA CTCAAATCTG CTACGTGTAT
19330      19340      19350      19360      19370      19380      19390

                Zsp2I >
                >< ScaI
                Ppu10I ><
                >< RsaINsiI >
                Mph1103I >
                >< SfaNIEcoT22I >
                > < RsaI >< Csp6I
                >< Csp6I
                >< NlaIII> < AfaI >< AfaI
                >< AvaIII ><
TACACGATGC AATTAGGTG GTGCTGTTTG CAGACACCAT GCAAATGAGT ACCGACAGTA CTTGGATGCA
19400      19410      19420      19430      19440      19450      19460

                >< FokI
TATAATATGA TGATTCTGCG TGGATTAGC CTATGGATT ACAAACAATT TGATACTTAT AACCTGTGGA
19470      19480      19490      19500      19510      19520      19530

                >< ScrFI
                >< MvaI
                >< MaeIII
                >< EcoRII
                >< Ecl136I
                >< DsaV
                >< BstOI
                >< BstNI
                >< BsiLI
                >< ApyI
                >< Tru9I
                >< MseI
ATACATTAC CAGGTTACAG AGTTAGAAA ATGTGGCTTA TAATGTTGTT AATAAAGGAC ACTTTGATGG
19540      19550      19560      19570      19580      19590      19600

                >< SgrAI
                >< NaeI
                >< MspI
                >< HpaII
                >< HapII
                >< Cfr10I
                >< BspWI
                > < VspI
                > < Tru9I
                > < MseI
                > < AsnI
                > < AseI
ACACGCCGGC GAAGCACCTG TTTCCATCAT TAATAATGCT GTTTACACAA AGGTAGATGG TATTGATGTG
19610      19620      19630      19640      19650      19660      19670

```

FIGURE 13. 45

```

>< XhoII
>< Sau3AI
>< NdeII
>< MflI
>< MboI
>< DpnII
  >< DpnI
>< BstYI
>< BspAI
  >< Bsp143I
>< BglIII
GAGATCTTTG AAAATAAGAC AACACTTCCT GTTAATGTTG CATTGAGCT TTGGGCTAAG CGTAACATTA
19680      19690      19700      19710      19720      19730      19740

                                >< MaeIII
                                >< EspI
                                >< DdeI>Tru9I ><
                                >< CelIIMseI ><
                                >< Bpu1102I
                                >< AluI
                                >< Tru9I
                                >< MseI
                                >< EcoRV
                                >< Fnu4HI
                                >< Eco32I
                                >< BbvI
                                >< BsrI
                                >< MseI
                                >< Tru9I
AACCAGTGCC AGAGATTAAG ATACTCAATA ATTTGGGTGT TGATATCGCT GCTAATACTG TAATCTGGGA
19750      19760      19770      19780      19790      19800      19810

                                >< NspI
                                >< NspHI
                                >< NlaIII
                                >< BsgI
                                >< AflIII
CTACAAAAGA GAAGCCCCAG CACATGTATC TACAATAGGT GTCTGCACAA TGA CTGACAT TGCCAAGAAA
19820      19830      19840      19850      19860      19870      19880

                                >< DdeI>> MboII
                                >< AccI
CCTACTGAGA GTGCTTGTTC TTCCTTACT GTCTTGTTC ATGGTAGAGT GGAAGGACAG GTAGACCTTT
19890      19900      19910      19920      19930      19940      19950

                                SinI ><
                                Sau96I ><
                                NspIV ><
                                NspHII ><
                                NlaIV ><
                                Eco47I ><
                                Cfr13I ><
                                >< BslI
                                Bsi2I ><
                                >< BsiYI
                                BscBI ><
                                Bmel8I ><
                                AvaII ><
                                AsuI ><
                                >< Tru9I
                                >< MseI
TTAGAAACGC CCGTAATGGT GTTTTAATAA CAGAAGGTTT AGTCAAAGGT CTAACACCTT CAAAGGGACC
19960      19970      19980      19990      20000      20010      20020

                                >< VspI
                                >< Tru9I
                                >< PleI
                                >< MseI
                                >< RmaI
                                >< NheI
                                >< MaeI
                                >< AsnI
                                >< TfiI
                                >< HinfI>> AseI
                                >< HinfI
                                >< MseI
                                >< Tru9I ><
                                >< Tru9I
                                >< MseI ><
                                >< MseI
                                >< HgaI>> AluI
                                >< HinfI>> AseI
                                >< HinfI
                                >< MseI
AGCACAAGCT AGCGTCAATG GAGTCACATT AATTGGAGAA TCAGTAAAAA CACAGTTTAA CTACTTTAAG
20030      20040      20050      20060      20070      20080      20090

                                >< DdeI
                                >< MnlI
                                >< Tru9I ><
                                >< BsmAI
                                >< DdeI

```

FIGURE 1346


```

>< AccI                               >< Alw26I >< BfrIMseI ><
AAAGTAGACG GCATTATTCA ACAGTTGCCT GAAACCTACT TTACTCAGAG CAGAGACTTA GAGGATTTTA
20100      20110      20120      20130      20140      20150      20160

                                >< TthHB8I
                                >< TaqI
                                    >< SstI
                                    >< SduI
                                    >< SacI
                                > < PaeR7I
                                > < NspIII
                                    >< NspII
                                    >< HgiAI
                                > < Eco88I
                                > < XhoI>< Eco24I
                                    >< Ecl136II
                                >< XcmI
                                >< Sau3AI
                                >< NdeII
                                >< MboI
                                >< DpnII
                                >< DpnI
                                >< BspAI
                                >< Bsp143I
AGCCCAGATC ACAAATGGAA ACTGACTTTC TCGAGCTCGC TATGGATGAA TTCATACAGC GATATAAGCT
20170      20180      20190      20200      20210      20220      20230

                                >< TthHB8I
                                >< TaqI
                                >< SfuI
                                >< NspV
                                >< LspI
                                >< Csp45I
                                >< BstBI
                                >< Bsp119I
                                >< BsiCI
                                >< Bpu14I
                                >< AsuII >< BcgI
                                >< MboII
                                >< BbsI Tru9I ><
                                >< NlaIII >< AciIMseI ><
CGAGGGCTAT GCCTTCGAAC ACATCGTTTA TGGAGATTTC AGTCATGGAC AACTTGGCGG TCTTCATTTA
20240      20250      20260      20270      20280      20290      20300

                                >< HphI
                                >< HinPII
                                >< Hin6I
                                >< EspI > < HhaI >< TfiI
                                >< DdeI >< HaeII
                                >< CelII >< Eco47III >< Tru9I
                                >< Bpu1102I >< CfoI >< HinfI >< MseI
                                >< BfrI >< Bsp143II >< MnlI
ATGATAGGCT TAGCCAAGCG CTCACAAGAT TCACCACTTA AATTAGAGGA TTTTATCCCT ATGGACAGCA
20310      20320      20330      20340      20350      20360      20370

                                >< MstI
                                >< HinPII
                                >< Hin6I
                                >< HhaI
                                >< FspI
                                >< FdiII
                                >< CfoI
                                >< SfaNI >< AviII
                                >< Sau3AI ><
                                >< NdeII ><
                                >< MboI ><
                                >< DpnII ><
                                >< DpnI ><
                                >< BspAI ><
                                >< Bsp143I ><
CAGTGAAAAA TTACTTCATA ACAGATGCGC AAACAGGTTC ATCAAAATGT GTGTGTCTG TGATTGATCT
20380      20390      20400      20410      20420      20430      20440

                                >< TthHB8I

```

FIGURE 13. 47

```

>< Tth111I
>< TaqI
>< AspI >< MaeIII MaeIII ><
TTTACTTGAT GACTTTGTCG AGATAATAAA GTCACAAGAT TTGTCAGTGA TTCAAAAAGT GGTC AAGGTT
20450 20460 20470 20480 20490 20500 20510

>< NspI
>< NspHI
>< NlaIII
>< FokI
>< MunI >< NlaIII >< AflIII
ACAATTGACT ATGCTGAAAT TTCATTATG CTTTGGTGTA AGGATGGACA TGTTGAAACC TTCTACCCAA
20520 20530 20540 20550 20560 20570 20580

>< SfaNI
>< ScrFI
>< MvaI
>< EcoRII
>< Ecl136I
>< DsaV
>< BstOI >< SfaNI
>< BstNI >< RsaI BspWI ><
>< BsiLI >< Csp6I BsmI >
>< BspWI >< ApyI >< AfaI BscCI ><
AACTACAAGC AAGTCAAGCG TGGCAACCAG GTGTTGCGAT GCCTAAGTTC TACAAGATGC AAAGAATGCT
20590 20600 20610 20620 20630 20640 20650

>< Eco57I >< MaeIII >< HphI
TCTTGAAAAG TGTGACCTTC AGAATTATGG TGAAATGCT GTTATACCAA AAGGAATAAT GATGAATGTC
20660 20670 20680 20690 20700 20710 20720

>< RsaI
>< Csp6I
>< Bst1107I >< Tru9I >< AluI
>< AccI >< MseI >< AfaINlaIII ><
GCAAAGTATA CTCAACTGTG TCAATACTTA AATACACTTA CTTTAGCTGT ACCCTACAAC ATGAGAGTTA
20730 20740 20750 20760 20770 20780 20790

>< ScrFI
>< RsaI
>< MvaI
>< EcoRII >< NspBII
>< Ecl136I >< SduI
>< Csp6I >< NspII
>< BstOI >< PvuII >< HgiAI
>< BstNI >< DdeI
>< BsiLI >< Psp5I >< Bsp1286I
>< ApyI >< AluI >< BmyI
>< DsaV >< AfaI >< Alw21I
TTCACTTTGG TGCTGGCTCT GATAAAGGAG TTGCACCAGG TACAGCTGTG CTCAGACAAT GGTGCGCAAC
20800 20810 20820 20830 20840 20850 20860

>< XhoII
>< Tru9I
>< Sau3AI
>< NdeII
>< TthHB8I >< MseI
>< MflI
>< MboI
>< MamI
>< DpnII
>< TfiI >< DpnI

```

FIGURE 13. 48

```

>< BstYI >< TfiI
>< BspAI >< HinfI
>< HinfI>< Bsp143I >< Esp3I >< Tru9I
>< BsiBI >< Tth111I >< BsmBI >< MseI
>< BsaBI >< BsmAI >< BsmAI
>< BsrI >< TaqI >< BglII >< AspI >< Alw26I >< HgaI> < Alw26I
TGGCACACTA CTTGTTCGATT CAGATCTTAA TGACTTCGTC TCCGACGCAG ATTCTACTTT AATTGGAGAC
20870 20880 20890 20900 20910 20920 20930

>< StyI
>< SinI
>< Sau96I
>< SinI
>< Sau96I
>< PssI
>< Psp5II
>< PpuMI
>< NspIV
>< NspHII
>< NlaIV
>< EcoO109I
>< Eco47I
>< DraII
>< Cfr13I
>< Bsi2I
>< BscBI
>< RsaI
>< Csp6I
>< AfaI
TGTGCAACAG TACATACGGC TAATAAATGG GACCTTATTA TTAGCGATAT GTATGACCCT AGGACCAAAC
20940 20950 20960 20970 20980 20990 21000

>< NspI
>< NspHI
>< NlaIII >< PleI
>< MaeIII >< HinfI
ATGTGACAAA AGAGAATGAC TCTAAAGAAG GGTTTTTTCAC TTATCTGTGT GGATTTATAA AGCAAAAAC
21010 21020 21030 21040 21050 21060 21070

>< ScrFI
>< MvaI
>< EcoRII
>< Ecl136I
>< DsaV
>< BstOI
>< BstNI
>< BsiLI
>< BsaJI
>< BsaJI >< SfcI >< BsmI >< BsmI >< BsmI
>< ApyI >< AluI >< BscCI >< BscCIHindIII ><>< AluI
AGCCCTGGGT GGTTCATAG CTGTAAAGAT AACAGAGCAT TCTTGGAATG CTGACCTTTA CAAGCTTATG
21080 21090 21100 21110 21120 21130 21140

>< Zsp2I
>< Ppu10I
>< Pali
>< HaeIII
>< BsuRI
>< BshI
>< Pali
>< HaeIII
>< BsuRI
>< BshI
>< NsiI
>< Mph1103I
>< EcoT22I
>< AvaIII >< SfaNIBcgI/a ><
GGCCATTTCT CATGGTGGAC AGCTTTTGT ACAAATGTAA ATGCATCATC ATCGGAAGCA TTTTAAATTG
21150 21160 21170 21180 21190 21200 21210

```

FIGURE 13.49

```

                                >< Zsp2I
                                >< SphI
                                >< Ppu10I
                                >< PaeI
                                >< NspI
                                >< NspHI
                                >< NsiI
                                >< NlaIII
                                > < NlaIII
                                >< Mph1103I
                                >< EcoT22I
                                > < AvaIII >< MnlI
GGGCTAACTA TCTTGGCAAG CCGAAGGAAC AAATTGATGG CTATACCATG CATGCTAACT ACATTTTCTG
 21220      21230      21240      21250      21260      21270      21280

                                Tru9I ><
                                >< Tru9I
                                >< GsuI
                                >< BsrI
                                >< BpmI
                                >< BbsI
                                >< NlaIII >< MnlI
GAGGAACACA AATCCTATCC AGTTGTCTTC CTATTCACCTC TTTGACATGA GCAAATTTCC TCTTAAATTA
 21290      21300      21310      21320      21330      21340      21350

                                >< Tru9I
                                >< MseI
                                >< Esp4I> < TfiI
                                >< BsmAI
                                >< Alw26I
                                >< AflIII> < HinfI
                                >< MboII >< EarI
AGAGGAACTG CTGTAATGTC TCTTAAGGAG AATCAAATCA ATGATATGAT TTATTCTCTT CTGGAAAAAG
 21360      21370      21380      21390      21400      21410      21420

                                >< Tru9I
                                >< MseI
                                >< HindII
                                >< HincII
                                >< HpaI AflIII >
GTAGGCTTAT CATTAGAGAA AACAACAGAG TTGTGGTTTC AAGTGATATT CTTGTTAACA ACTAAACGAA
 21430      21440      21450      21460      21470      21480      21490

                                >< VneI
                                >< SnaI
                                >< SduI
                                >< NspII
                                >< HpaII
                                >< HgiAI
                                >< HapII
                                >< Cfr10I
                                >< Bsp1286I
                                >< MspI>< BmyI
                                >< ApaLI
                                >< Alw44I
                                >< AgeI >< Alw21I
                                >< SpeI
                                >< RmaI
                                >< MaeI >< MaeIII >< AgeI >< Alw21I
CATGTTTATT TTCTTATTAT TTCTTACTCT CACTAGTGGT AGTGACCTTG ACCGGTGCAC CACTTTTGAT
 21500      21510      21520      21530      21540      21550      21560

                                > < AluI
                                >< MnlI
GATGTTCAAG CTCCTAATTA CACTCAACAT ACTTCATCTA TGAGGGGGGT TTACTATCCT GATGAAATTT
 21570      21580      21590      21600      21610      21620      21630

                                >< Sau3AI

```

FIGURE 13. 50

```

>< NdeII
>< MboI
>< DpnII
  >< DpnI          >< Tru9I
>< BspAI          >< MseI > < MboII
  >< Bsp143I       >< DdeI          >< MaeIII
TTAGATCAGA CACTCTTTAT TTAAGTCAGG ATTTATTTCT TCCATTTTAT TCTAATGTGA CAGGGTTTCA
21640      21650      21660      21670      21680      21690      21700

  >< VspI
  >< Tru9I
  >< MseI
  >< AsnI          >< Tru9I          >< FokI
  >< AseI >< MaeII >< MseI >< BbvI    > < Fnu4HI
TACTATTAAT CATACGTTTG GCAACCTGT CATACCTTTT AAGGATGGTA TTTATTTTGC TGCCACAGAG
21710      21720      21730      21740      21750      21760      21770

          >< BslI
          >< DsaI>< BsiYI          >< NlaIII
          >< BsaJI          > < MaeIII
AAATCAAATG TTGTCCGTGG TTGGGTTTTT GGTCTACCA TGAACAACAA GTCACAGTGG GTGATTATTA
21780      21790      21800      21810      21820      21830      21840

          >< NspI
>< Tru9I          >< NspHI
>< MseI          >< NlaIII
>< HphI          >< MaeIII          >< MaeIII
TTAACAATTC TACTAATGTT GTTATACGAG CATGTAACCT TGAATTGTGT GACAACCCCT TCTTTGCTGT
21850      21860      21870      21880      21890      21900      21910

  >< StyI          >< Zsp2I
  >< NlaIII          >< Tru9I
>< NcoI >< RsaI    >< Ppu10I TthHB8I ><
>< EcoT14I          >< NsiI          >< TaqI
>< Eco130I          >< MseI          SfaNI ><
>< DsaI>< Csp6I    >< Mph1103I RsaI ><
>< BssT1I          >< TthHB8I >< EcoT22I Csp6I ><
>< BsaJI>< AfaI    >< TaqI >< AfaIII AfaI ><
TTCTAAACCC ATGGGTACAC AGACACATAC TATGATATTC GATAATGCAT TTAATTGCAC TTTTCGAGTAC
21920      21930      21940      21950      21960      21970      21980

          >< Tru9I
          >< MseI
          >< DraI
ATATCTGATG CCTTTTCGCT TGATGTTTCA GAAAAGTCAG GTAATTTTAA ACACTTACGA GAGTTTGTGT
21990      22000      22010      22020      22030      22040      22050

          >< Sau3AI
          >< NdeII
          >< MboI
          >< DpnII
>< Tru9I          >< DpnI
>< MseI          >< BspAI
  >< DraI          >< SfcI Bsp143I ><
TTAAAAATAA AGATGGGTTT CTCTATGTTT ATAAGGGCTA TCAACCTATA GATGTAGTTC GTGATCTACC
22060      22070      22080      22090      22100      22110      22120

          >< Tru9I
          > < Tru9I
          >< MseI
          >< MseI          >< MnlI
TTCTGGTTTT AACACTTTGA AACCTATTTT TAAGTTGCCT CTTGGTATTA ACATTACAAA TTTTAGAGCC
22130      22140      22150      22160      22170      22180      22190

```

FIGURE 13.51

```

> < SduI>< SfcI
>< PvuII
>< Psp5I
> < NspII
>< NspBII
> < MaeII > < Fnu4HI
> < Bsp1286I >< PstI Tru9I >
> < BmyI>< Fnu4HI MseI >
>< HphI >< BbvI >< AluI >< BbvI
ATTCTTACAG CCTTTTCACC TGCTCAAGAC ATTTGGGGCA CGTCAGCTGC AGCCTATTTT GTTGGCTATT
22200 22210 22220 22230 22240 22250 22260

>< SfaNI
>< RsaI
> < Csp6I
>< DraI >< AfaI >< AlwNI
TAAAGCCAAC TACATTTATG CTCAAGTATG ATGAAAATGG TACAATCACA GATGCTGTTG ATTGTTCTCA
22270 22280 22290 22300 22310 22320 22330

> < Tru9I
> < MseI
>< AluI
AAATCCACTT GCTGAACTCA AATGCTCTGT TAAGAGCTTT GAGATTGACA AAGGAATTTA CCAGACCTCT
22340 22350 22360 22370 22380 22390 22400

>< SauI
>< MstII
>< Eco81I
>< DdeI
>< CvnI
>< Bsu36I
>< Bse21I
>< AxyI >< TfiI
>< MnlI >< AocI >< MnlI >< HinfI >< SspI >< MnlI
AATTTCAGGG TTGTTCCCTC AGGAGATGTT GTGAGATTCC CTAATATTAC AAACCTTGTT CCTTTTGGAG
22410 22420 22430 22440 22450 22460 22470

>< Zsp2I
>< Ppu10I
>< NsiI
> < NlaIII
>< Mph1103I
>< EcoT22I
>< Tru9I
>< MseI >< AvaIII
AGGTTTTTAA TGCTACTAAA TTCCCTTCTG TCTATGCATG GGAGAGAAAA AAAATTTCTA ATTGTGTTGC
22480 22490 22500 22510 22520 22530 22540

>< SduI
>< NspII
>< HgiAI
>< Bsp1286I
>< BmyI >< Tru9I
>< Alw21I >< MseI DdeI ><
TGATTACTCT GTGCTCTACA ACTCAACATT TTTTCAACC TTAAAGTGCT ATGGCGTTTC TGCCACTAAG
22550 22560 22570 22580 22590 22600 22610

>< Sau3AI
>< NdeII
>< MboI
>< DpnII
>< DpnI

```

FIGURE 13.52

```

    >< BspAI
    >< BspI43I
    TTGAATGATC TTTGCTTCTC CAATGTCTAT GCAGATTCTT TTGTAGTCAA GGGAGATGAT GTAAGACAAA
    22620      22630      22640      22650      22660      22670      22680

    >< ScrFI
    >< MvaI
    >< HinPII
    >< Hin6I
    >< HhaI
    >< HaeII
    >< EcoRII
    >< Ecl136I
    >< DsaV
    >< CfoI
    >< BstOI
    >< BstNI
    >< BspI43II
    >< BsiLI
    >< ApyI
    >< BsrI
    TAGCGCCAGG ACAAACCTGGT GTTATTGCTG ATTATAATTA TAAATTGCCA GATGATTTCa TGGGTTGTGT
    22690      22700      22710      22720      22730      22740      22750

    >< SfaNI
    >< RmaI
    >< MaeI
    CCTTGCTTGG AATACTAGGA ACATTGATGC TACTTCAACT GGTAATTATA ATTATAAATA TAGGTATCTT
    22760      22770      22780      22790      22800      22810      22820

    >< Sau96I
    >< Pali
    >< NspIV
    >< HindIII
    >< HaeIII
    >< EcoO109I
    >< DraII
    >< DdeI
    >< Cfr13I
    >< BsuRI
    >< BsiZI
    >< BshI
    >< BfrI >< PssI
    >< NlaIII >< AsuI>< BsmAI
    >< AluI >< Alw26I
    AGACATGGCA AGCTTAGGCC CTTTGAGAGA GACATATCTA ATGTGCCTTT CTCCCCTGAT GGCAAACCTT
    22830      22840      22850      22860      22870      22880      22890

    >< Tru9I
    >< Pali
    >< MscI
    >< HaeIII
    >< EaeI>< MseI
    >< Tru9I >< BsuRI
    >< MseI >< BshI
    >< BspMI >< BalI
    GCACCCACCC TGCTCTTAAT TGTTATTGGC CATTAAATGA TTATGGTTTT TACACCACTA CTGGCATTGG
    22900      22910      22920      22930      22940      22950      22960

    Sau96I ><
    >< PalINspIV ><
    >< MspI NspHII ><
    >< HaeIII

```

FIGURE 13.53

FIGURE 13. 54


```

                >< BspWI                >< Hin6I
                    >< BspAI                > < HhaI                PleI ><
>< SfcI                >< Bsp143I                >< AluI> < CfoI                >< BsrI
CTACAGCAAT TCATGCAGAT CAACTCACAC CAGCTTGGCG CATATATTCT ACTGGAAACA ATGTATTCCA
23320      23330      23340      23350      23360      23370      23380

                >< TthHB8I
                >< TaqI
                >< SalI
                >< RtrI
                >< NspI
                >< EspI >< NspHI
                >< DdeI >< NlaIII
                >< CelII >< HindII
                >< Bpu1102I>< HincII
>< HinfI                >< AluI                >< AccI
GACTCAAGCA GGCTGTCTTA TAGGAGCTGA GCATGTCGAC ACTTCTTATG AGTGCGACAT TCCTATTGGA
23390      23400      23410      23420      23430      23440      23450

                > < SnaBI
                >< ScaI
                >< RsaI
                >< RmaI
                >< MaeII >< MaeI
                > < Eco105I
                >< Csp6I
                > < BsaAI
                >< AfaI
>< AluI                >< MaeI
GCTGGCATTT GTGCTAGTTA CCATACAGTT TCTTTATTAC GTAGTACTAG CCAAAAATCT ATTGTGGCTT
23460      23470      23480      23490      23500      23510      23520

                >< MniI
ATACTATGTC TTTAGGTGCT GATAGTTCAA TTGCTTACTC TAATAACACC ATTGCTATAC CTACTAACTT
23530      23540      23550      23560      23570      23580      23590

                RsaI ><
                >< MnlI
                Csp6I ><
                AfaI ><
>< SfcI
TTCAATTAGC ATTACTACAG AAGTAATGCC TGTTCCTATG GCTAAACCT CCGTAGATTG TAATATGTAC
23600      23610      23620      23630      23640      23650      23660

                > < TfiI
                > < HinfI
                >< AciI                > < AluI
ATCTGCGGAG ATTCTACTGA ATGTGCTAAT TTGCTTCTCC AATATGGTAG CTTTGCACA CAACTAAATC
23670      23680      23690      23700      23710      23720      23730

>< VneI
>< SduI
>< NspII
>< HgiAI
>< SnaI>< DdeI                >< Sau3AI                >< PmlI
>< Bsp1286I                >< NdeII                >< PmaCI
>< BmyI                >< MboI                >< MaeII
>< BbvI                >< DpnI                >< Eco72I
>< ApaI                >< Bsp143I                >< BsaAI
>< Alw44I                >< DpnII >< AlwI
>< Alw21I                >< Fnu4HI                >< BspAI                >< AflIII
GTGCACTCTC AGGTATTGCT GCTGAACAGG ATCGCAACAC ACGTGAAGTG TTCGCTCAAG TCAAACAAAT
23740      23750      23760      23770      23780      23790      23800

```

FIGURE 13.55

```

>< RsaI
>< Csp6I
>< AfaI
GTACAAAACC CCAACTTTGA AATATTTTGG TGGTTTAAAT TTTTCACAAA TATTACCTGA CCCTCTAAAG
23810 23820 23830 23840 23850 23860 23870

>< MnlI
>< MnlI
>< DdeI >< MnlI
>< Tru9I >< SfaNI >< HphI NlaIII ><
>< MseI >< MaeIII BspHI ><
CCAACTAAGA GGTCTTTTAT TGAGGACTTG CTCITTAATA AGGTGACACT CGCTGATGCT GGCTTCATGA
23880 23890 23900 23910 23920 23930 23940

>< XhoII
>< Sau3AI
>< StyI >< RmaI
>< RmaI >< NdeII
>< MaeI >< MflI
>< EcoT14I >< MboI >< MstI
>< Eco130I >< MaeI >< HinPII
>< BssTII >< VspI >< DpnII >< Hin6I
>< BsmI >< HphI> < DpnI >< HhaI
>< BscCI >< Tru9I >< BstYI >< FspI
>< BsaJI >< MseI >< BspAI >< FdiII
>< BlnI >< AsnI >< Bsp143I >< CfoI
>< AvrII >< AseI >< BglII >< AviII
AGCAATATGG CGAATGCCTA GGTGATATTA ATGCTAGAGA TCTCATTTGT GCGCAGAAGT TCAATGGACT
23950 23960 23970 23980 23990 24000 24010

>< RmaIRsaI ><
>< MnlI >< Fnu4HI >< Fnu4HI Csp6I ><
>< BspWI >< BbvI >< BbvI >< BspWI >< MaeIAfaI ><
TACAGTGTG CCACCTCTGC TCACTGATGA TATGATTGCT GCCTACACTG CTGCTCTAGT TAGTGGTACT
24020 24030 24040 24050 24060 24070 24080

>< MboII
>< HinPII
>< Hin6I
>< HhaI
>< HaeII
>< Fnu4HI >< Ksp632I
>< CfoI >< EarI
>< FokI >< BspWI >< Eam1104I
>< BbvI >< Bsp143II
GCCACTGCTG GATGGACATT TGGTGCTGGC GCTGCTCTTC AAATACCTTT TGCTATGCAA ATGGCATATA
24090 24100 24110 24120 24130 24140 24150

>< MaeIII
>< Tru9I ><
>< MseI ><
GGTTCAATGG CATTGGAGTT ACCCAAAATG TTCTCTATGA GAACCAAAAA CAAATCGCCA ACCAATTTAA
24160 24170 24180 24190 24200 24210 24220

>< MaeII ><
>< TfiI >< Fnu4HI
>< HinfI >< BbvI >< AluI
CAAGGCGATT AGTCAAATTC AAGAATCACT TACAACAACA TCAACTGCAT TGGGCAAGCT GCAAGACGTT
24230 24240 24250 24260 24270 24280 24290

>< Tru9I
>< MseI
>< HpaI
>< HindII >< BsmI >< Tru9I >< DdeI
>< HincII>< BscCI >< MseI >< Tru9I >< BfrI
>< MseI >< AluI

```

FIGURE 13. 56

```

GTTAACCAGA ATGCTCAAGC ATTAAACACA CTTGTTAAAC AACTTAGCTC TAATTTGGT GCAATTTCAA
24300      24310      24320      24330      24340      24350      24360

      >< Thal
      >< SpoI
      >< NruI
      >< MvnI
      >< BstUI      >< TthHB8I
      >< Bsp68I      >< TaqI      >< RsaI
      >< EcoRV      >< Bsp50I      >< MnlI      >< Csp6I      >< Tru9I
      >< Eco32I      >< AccII      >< MnlI      >< AciI      >< AfaI      >< MseI
GTGTGCTAAA TGATATCCTT TCGCGACTTG ATAAAGTCGA GGCGGAGGTA CAAATTGACA GGTTAATTAC
24370      24380      24390      24400      24410      24420      24430

      >< MaeIII      >< BbvI      >< Fnu4HI      BbvI      ><
AGGCAGACTT CAAAGCCTTC AACCTATGT AACACAACAA CTAATCAGGG CTGCTGAAAT CAGGGCTTCT
24440      24450      24460      24470      24480      24490      24500

      >< Fnu4HI
      >< BspWI      >< DdeI      >< HindII
      >< BspWI      >< DdeI      >< HincII
GCTAATCTTG CTGCTACTAA AATGTCTGAG TGTGTTCTTG GACAATCAAA AAGAGTTGAC TTTGTGGAA
24510      24520      24530      24540      24550      24560      24570

      > < NspI
      > < NspHI
      > < NlaIII
      >< MaeIII
      >< NlaIII
      >< MboII
      >< FokI
      >< Fnu4HI      >< BbsI      BsaAI      ><
      >< AciI      >< BbvI      >< AflIII
AGGGCTACCA CCTTATGTCC TTCCACAAAG CAGCCCCGCA TGGTGTGTC TTCCTACATG TCACGTATGT
24580      24590      24600      24610      24620      24630      24640

      >< ScrFI
      >< MvaI
      >< EcoRII
      >< Ecl136I
      >< BstOI
      >< BstNI      >< HinPII
      >< MnlI      >< BslI      >< Hin6I
      >< DsaV      >< BsiYI      >< HhaI
      >< BsiLI      >< HaeII
      >< BsaJI      >< HphI      >< CfoI      >< NlaIII
      >< ApyI      >< Bsp143II      >< BspHI      EcoNI      ><
GCCATCCCAG GAGAGGAACT TCACCACAGC GCCAGCAATT TGTCATGAAG GCAAAGCATA CTCCCTCGT
24650      24660      24670      24680      24690      24700      24710

      >< MnlI
      >< BslI      >< Tru9I
      >< BsiYI      >< MseI      >< MnlI
GAAGGTGTTT TTGTGTTTAA TGGCACTTCT TGGTTTATTA CACAGAGGAA CTTCTTTTCT CCACAAATAA
24720      24730      24740      24750      24760      24770      24780

      >< DdeI      >< Tru9I
      >< BsmAI      >< SfaNI
      >< SfcI      >< Alw26I      >< MseI      >< AlwI      ><
TTACTACAGA CAATACATTT GTCTCAGGAA ATTGTGATGT CGTTATTGGC ATCATTAAACA ACACAGTTTA
24790      24800      24810      24820      24830      24840      24850

      >< Sau3AI
      >< NdeII

```

FIGURE 13.57

```

>< MboI          >< PfuI          > < ScaI
>< DpnII         >< MnlI          > < Ksp632I      > < RsaI
>< DpnI          >< DdeI   >< HinfI          >< MboII
>< BspAI         >< BspWI          > < Eam1104I      >< Csp6I
>< Bsp143I       >< AluI           > < EarI   > < AluI   > < AfaI   > < HphI
TGATCCTCTG CAACCTGAGC TTGACTCATT CAAAGAAGAG CTGGACAAGT ACTTCAAAAA TCATACATCA
24860      24870      24880      24890      24900      24910      24920

>< Sau3AI
>< NdeII
>< MboI
>< MmI
>< DpnII
>< DpnI
>< BspAI
>< Bsp143I
>< BsiBI          >< Tru9I          >< HindII
>< BsaBI          >< MseI           >< HincII      AciI ><
CCAGATGTTG ATCTTGGCGA CATTTTCAGGC ATTAACGCTT CTGTCGTCAA CATTCAAAAA GAAATTGACC
24930      24940      24950      24960      24970      24980      24990

>< Tru9I
> < TfiI
>< MnlI          >< SmaI
>< EcoNI         >< MseI
>< BslI          > < HinfI
>< MnlI>< BsiYI   >< DraI
GCCTCAATGA GGTGCTTAAA AATTTAAATG AATCACTCAT TGACCTTCAA GAATTGGGAA AATATGAGCA
25000      25010      25020      25030      25040      25050      25060

>< StyI
>< PstI
>< HaeIII
>< EcoT14I
>< Eco130I
>< BsuRI
>< BssTII
>< Tru9I>< BshI          NlaIII ><
>< MseI >< BsaJI          MaeIII ><
ATATATTAAA TGGCCTTGGT ATGTTTGGCT CGGCTTCATT GCTGGACTAA TTGCCATCGT CATGGTTACA
25070      25080      25090      25100      25110      25120      25130

> < SphI
> < PaeI
>< SpeI          > < NspI
> < RmaI          > < NspHI
>< NlaIII        > < NlaIII
> < MaeI          >< MnlI>< BbvI Fnu4HI ><
ATCTTGCTTT GTTGCATGAC TAGTTGTTGC AGTTGCCTCA AGGGTGCATG CTCTTGTTGGT TCTTGCTGCA
25140      25150      25160      25170      25180      25190      25200

>< FokI
>< DdeI
>< MnlI >< PfuI>< HinfI >< BsrI
AGTTTGATGA GGATGACTCT GAGCCAGTTC TCAAGGGTGT CAAATTACAT TACACATAAA CGAACTTATG
25210      25220      25230      25240      25250      25260      25270

>< Sau3AI
>< NdeII
>< MboI
>< DpnII
> < DpnI

```

FIGURE 13.58

```

                >< BspAI
                > < Bsp143I
                >< BsgI                >< AlwI                >< BsrI                BspWI >
GATTTGTTTA TGAGATTTT TACTCTTGA TCAATTACTG CACAGCCAGT AAAAATTGAC AATGCTTCTC
25280      25290      25300      25310      25320      25330      25340

                >< ScaI
                >< RsaI
                >< Csp6I                >< SfcI
                >< AfaI                >< NlaIII                >< AciI                >< MnlI                FokI >
CTGCAAGTAC TGTTCATGCT ACAGCAACGA TACCGCTACA AGCCTCACTC CCTTTCGGAT GGCTTGTTAT
25350      25360      25370      25380      25390      25400      25410

                > < HinPII
                > < Hin6I
                >< HhaI                >< HaeII                >< HinPII                RmaI ><
                >< Eco47III                >< Hin6I                NheI ><
                >< CfoI                >< HhaI                MaeI ><
                >< BspWI                >< Bsp143II                >< CfoI                Fnu4HI ><
                >< CfoI                AluI ><
TGCGCTTGCA TTTCTTGCTG TTTTTCAGAG CGCTACCAAA ATAATTGCGC TCAATAAAAG ATGGCAGCTA
25420      25430      25440      25450      25460      25470      25480

                >< EcoNI
                >< BslI
                >< BsiYI                >< MaeIII
                >< BbvI                >< BsrI                >< BbvI                > < Fnu4HI                BbvI ><
GCCCTTTATA AGGGCTTCCA GTTCATTTGC AATTTACTGC TGCTATTTGT TACCATCTAT TCACATCTTT
25490      25500      25510      25520      25530      25540      25550

                >< SfcI                >< HinPII
                >< PstI                >< Hin6I                >< RsaI                Zsp2I ><
                > < Fnu4HI                >< HhaI                >< Csp6I                Ppu10I ><
                >< BspMI                >< MnlI                >< CfoI                NsiI ><
                >< MnlI                >< AfaI                >< MnlI                Mph1103I ><
                >< CfoI                >< AfaI                >< MnlI                EcoT22I ><
                >< BspMI                >< MnlI                >< CfoI                AvaIII ><
TGCTTGTCGC TGCAGGTATG GAGGCGCAAT TTTTGTACCT CTATGCCTTG ATATATTTTC TACAATGCAT
25560      25570      25580      25590      25600      25610      25620

                >< SfaNI
                >< NspI
                >< NspHI
                >< NlaIII                >< SfaNI
CAACGCATGT AGAATTATTA TGAGATGTTG GCTTTGTTGG AAGTGCAAAT CCAAGAACCC ATTACTTTAT
25630      25640      25650      25660      25670      25680      25690

                >< Bst1107I
                >< AccI                >< MaeIII ><
GATGCCAACT ACTTTGTTTG CTGGCACACA CATAACTATG ACTACTGTAT ACCATATAAC AGTGTACAG
25700      25710      25720      25730      25740      25750      25760

                >< MboII
                >< BstXI ><
                >< MunI >< MaeIII >< MaeIII                >< Eco57I                >< BbsI MnlI >
ATACAATTGT CGTTACTGAA GGTGACGGCA TTTCAACACC AAAACTCAAA GAAGACTACC AAATTGGTGG
25770      25780      25790      25800      25810      25820      25830

                >< RsaI
                > < NlaIII
                >< HphI
                >< Tru9I >< Tth111I>< Csp6I
                >< DdeI                >< DdeI                >< MseI>< AspI                >< AfaI

```

FIGURE 13.59

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TTATTCTGAG GATAGGCACT CAGGTGTAA AGACTATGTC GTTGATCATG GCTATTTTAC CGAAGTTTAC
25840      25850      25860      25870      25880      25890      25900

      > < HinfI>< P1eI      >< BsrI      Tru9I ><
      >< AluI >< AccI      >< SfcI >< AlwNI      MseI ><
TACCAGCTTG AGTCTACACA AATTACTACA GACACTGGTA TTGAAAATGC TACATTCTTC ATCTTTAACA
25910      25920      25930      25940      25950      25960      25970

      > < TthHB8I
      >< Tru9I      > < TaqI      >< Ksp632I
      >< MseI      > < MboII      >< EarI BspWI ><
>< AluI      >< Eco57I      >< Eam1104I AlwI ><
AGCTTGTTAA AGACCCACCG AATGTGCAAA TACACACAAT CGACGGCTCT TCAGGAGTTG CTAATCCAGC
25980      25990      26000      26010      26020      26030      26040

      >< XhoII
      >< Sau3AI
      >< NlaIV
      >< NdeII
      >< MflI
      >< MboI
      >< DpnII
      >< DpnI
      >< BstYI
      >< BstI
      >< BspAI
      >< Bsp143I
      >< BscBI      >< RmaI      RsaI ><
      >< BamHI >< AlwI      >< MaeI      Csp6I ><
AATGGATCCA ATTTATGATG AGCCGACGAC GACTACTAGC GTGCCTTTGT AAGCACAAGA AAGTGAGTAC
26050      26060      26070      26080      26090      26100      26110

      > < Tru9I
      >< RsaI
      > < MseI
      >< MboII
      >< MaeII      >< RsaI
      >< Csp6I      >< Csp6I      >< Tru9I >< Csp6I
      > < AfaI      >< AfaI      >< MseI >< AfaI
GAACTTATGT ACTCATTCGT TTCGGAAGAA ACAGGTACGT TAATAGTTAA TAGCGTACTT CTTTTTCTTG
26120      26130      26140      26150      26160      26170      26180

      >< TthHB8I
      >< TaqI
      >< RmaI      >< HinfI      > < RsaI
      > < MaeIII      >< HinfI      Fnu4HI ><
      >< MaeI >< RmaI      >< HhaI      >< Csp6I
      >< FokI >< MaeI      >< CfoI >< BbvI > < AfaI
CTTTCGTGGT ATTCTTGCTA GTCACACTAG CCATCCTTAC TGCGCTTCGA TTGTGTGCGT ACTGCTGCAA
26190      26200      26210      26220      26230      26240      26250

      >< Tru9I
      >< MseI      >< ThaI
      >< SspI >< MaeII      >< MvnI
      >< HpaI      >< MseI
      >< HindII      >< BstUI      Ksp632I >
      >< KincII      >< MaeII >< Bsp50I >< MboII EarI >
      >> AccI >> AccII      >> Eam1104I >
TATTGTTAAC GTGAGTTTAG TAAACCAAC GGTTTACGTC TACTCGCGTG TTAAAAATCT GAACTCTTCT
26260      26270      26280      26290      26300      26310      26320

```

FIGURE 13.60

```

>< Sau3AI
>< NdeII
>< MboI
>< DpnII
>< MboII>< DpnI
>< XmnI >< BspAI> < Eco57I
>< Asp700I>< Bsp143I
GAAGGAGTTC CTGATCTTCT GGTCTAAACG AACTAATAT TATTATTATT CTGTTTGGA CTTTAACATT
26330      26340      26350      26360      26370      26380      26390

>< ScrFI
>< MvaI
>< EcoRII
>< Ecl136I
>< DsaV NlaIV ><
>< RsaI
>< MnlI
>< Tru9I
>< BstNI RmaI ><
>< Csp6I
>< MseI
>< BsiLI MaeI ><
> < NlaIII >< AfaI > < AluI >< ApyIBscBI ><
GCTTATCATG GCAGACAACG GTACTATTAC CGTTGAGGAG CTTAAACAAC TCCTGGAACA ATGGAACCTA
26400      26410      26420      26430      26440      26450      26460

>< ScrFI
>< RmaI
>< MvaI
>< MaeI
>< EcoRII
>< Ecl136I
>< DsaV
>< BstOI
>< BstNI
>< BsiLI
>< ApyI >< MaeIII
GTAATAGGTT TCCTATTCCT AGCCTGGATT ATGTTACTAC AATTTGCCTA TTCTAATCGG AACAGGTTTT
26470      26480      26490      26500      26510      26520      26530

>< Pali
>< MscI
>< MnlI >< MaeIII
>< HaeIII
>< EaeI
>< BsuRI
>< BsrI
>< RsaI
>< Csp6I >< HindIII
>< AfaI >< AluI
>< BspWI
>< BshI
>< Bali
>< BbvI Fnu4HI ><
TGACATAAT AAAGCTTGTT TTCCTCTGGC TCTTGCGGCC AGTAACACTT GCTTGTTTTG TGCTTGCTGC
26540      26550      26560      26570      26580      26590      26600

>< VspI
>< Tru9I
>< MseI
>< SfcI >< AsnI >< HphI
>< AccI >< AseI>< MaeIII>< AciI
>< BsrI
TGTCTACAGA ATTAATTGGG TGACTGGCGG GATTGCGATT GCAATGGCTT GTATTGTAGG CTTGATGTGG
26610      26620      26630      26640      26650      26660      26670

>< EspI
>< Eco57I
>< DdeI
>< CelII
>< Bpu1102I
>< RsaI
>< Csp6I

```

FIGURE 13.61

FIGURE 13.62

FIGURE 13.62


```

CCACGCCGGT AGCAACGACA ATATTGCTTT GCTAGTACAG TAAGTGACAA CAGATGTTTC ATCTTGTTGA
 27030      27040      27050      27060      27070      27080      27090

>< ScrFI
>< MvaI
  >< MaeIII
>< EcoRII
  >< Ecl136I
>< DsaV
  >< BstOI
  >< BstNI
  >< BsiLI
  >< ApyI
                                >< MnlI
                                HinfI ><
CTTCCAGGTT ACAATAGCAG AGATATTGAT TATCATTATG AGGACTTTCA GGATTGCTAT TTGGAATCTT
 27100      27110      27120      27130      27140      27150      27160

>< MaeII          >< BsmAI          >< Tru9I          > < MnlI
>< Alw26I          >< MseI          >< DdeI          >< MboII
GACGTTATAA TAAGTTCAAT AGTGAGACAA TTATTTAAGC CTCTAACTAA GAAGAATTAT TCGGAGTTAG
 27170      27180      27190      27200      27210      27220      27230

                                >< Ksp632I
                                >< EarI
                                >< MboII
                                >< NlaIII Eam1104I ><
ATGATGAAGA ACCTATGGAG TTAGATTATC CATAAACGA ACATGAAAAT TATTCTCTTC CTGACATTGA
 27240      27250      27260      27270      27280      27290      27300

                                > < RsaI >< RsaI
                                >< Csp6I >< Csp6I
                                > < AluI          >< MnlI          > < AfaI >< AfaI
TTGTATTTAC ATCTTGCGAG CTATATCACT ATCAGGAGTG TGTTAGAGGT ACGACTGTAC TACTAAAAGA
 27310      27320      27330      27340      27350      27360      27370

                                >< MnlI          >< HphI          >< HphI          >< MnlI
ACCTTGCCCA TCAGGAACAT ACGAGGGCAA TTCACCATT T CAGCCTCTTG CTGACAATAA ATTTGCACTA
 27380      27390      27400      27410      27420      27430      27440

                                Sau3AI >
                                > < PvuII
                                > < Psp5I
                                > < NspBII
                                >< TthHB8I          NdeII >
                                >< TaqI          MboI >
                                >< RsaI          >< Fnu4HI
                                >< Csp6I          DpnII >
                                >< BbvI          BspAI >
                                >< AfaI          > < AluI
ACTTGCACTA GCACACACTT TGCTTTTGCT TGTGCTGACG GTACTCGACA TACCTATCAG CTGCGTGCAA
 27450      27460      27470      27480      27490      27500      27510

                                >< SstI
                                >< SduI
                                >< SacI
                                >< NspII
                                >< HgiAI
                                >< Eco24I
                                > < Ecl136II
                                >< BspWI
                                >< Bsp1286I
                                >< BmyI
                                >< BanII
                                >< Alw21I

>< HphI
>< DpnI
                                >< MnlI

```

FIGURE 13. 63

```

>< Bsp143I          >< MnlI          > < AluI      BbvI ><
GATCAGTTTC ACCAAACTT TTCATCAGAC AAGAGGAGGT TCAACAAGAG CTCTACTCGC CACTTTTTCT
27520      27530      27540      27550      27560      27570      27580

SstI ><
SduI ><
SacI ><
NspII ><
HgiAI ><
Eco24I ><
Ecl136II ><
Bsp1286I ><
BmyI ><
BanII ><
Alw21I ><
AluI ><

>< RmaI >< Tru9I
>< MaeI >< MseI          >< Tru9I
>< Fnu4HI >< HphI          >< MseI
CATTGTTGCT GCTCTAGTAT TTTTAATACT TTGCTTCACC ATTAAGAGAA AGACAGAATG AATGAGCTCA
27590      27600      27610      27620      27630      27640      27650

>< Tru9I          >< Tru9I
>< MseI          >< MseI
CTTTAATTGA CTTCTATTG TGCTTTTGTAG CCTTCTGCT ATTCCTTGTT TTAATAATGC TTATTATATT
27660      27670      27680      27690      27700      27710      27720

>< XhoII
>< XbaI
> < ScrFI
>< Sau3AI
>< RmaI
>< NdeII
> < MvaI
>< MflI
>< MboI
>< EcoRII>< MaeI
> < Ecl136I
>< DpnII
>< DpnI
>< BstYI
> < BstOI
> < BstNI
>< TthHB8I >< BspAI          > < RsaI
>< DsaV>< Bsp143I          >< MboII
> < BsiLI          >< Csp6I
>< TaqI > < ApyI > < AlwI > < AfaI          >< NlaIII
TTGGTTTTC CCGAAATCC AGGATCTAGA AGAACCTGT ACCAAAGTCT AAACGAACAT GAAACTTCTC
27730      27740      27750      27760      27770      27780      27790

>< HinP1I
>< Hin6I
>< HhaI
>< RsaI >< HaeII
>< SfcI          >< Eco47III
>< Csp6I>< CfoI SfaNI ><
>< AfaI >< Bsp143II
ATTGTTTTGA CTTGTATTTT TCTATGCAGT TGCATATGCA CTGTAGTACA GCGCTGTGCA TCTAATAAAC
27800      27810      27820      27830      27840      27850      27860

>< XhoII
>< Sau3AI
>< NdeII
> < MnlI
>< MflI

```

FIGURE 13.64

```

    >< MboI
    >< DpnII
        >< DpnI        >< RsaI
    >< BstYI    >< MboII
    >< NlaIII>< BspAI    >< Csp6I >< RmaI
    >< AlwI >< Bsp143I    >< AfaI >< MaeI
CTCATGTGCT TGAAGATCCT TGTAAGGTAC AACACTAGGG GTAATACTTA TAGCACTGCT TGGCTTTGTG
    27870      27880      27890      27900      27910      27920      27930

    >< SduI
    >< RmaI
    >< NspII
    >< MaeI
    >< HgiAI
    >< Bsp1286I
    >< BmyI
    >< Alw21I
    >< NspI
    >< NspHI
    >< NlaIII    >< MaeIII
CTCTAGGAAA GGTTTTACCT TTTCATAGAT GGCACACTAT GGTTCAAACA TGCACACCTA ATGTTACTAT
    27940      27950      27960      27970      27980      27990      28000

    > < XhoII
    > < Sau3AI > < Van91I
        >< PvuII
        >< Psp5I
    > < NdeII > < PflMI
    > < MflI>< NspBII
    > < DpnII    >< HinPII
        >< Bsp143I    >< Hin6I
    > < BstYI > < BslI >< HhaI >< RmaI
    > < BspAI > < BsiYI>< CfoI >< MaeI
    > < MboI>< AluI>< BspWI >< BspWI
    >< AlwI >< DpnI > < AccB7I    >< AluI
CAACTGTCAA GATCCAGCTG GTGGTGCGCT TATAGCTAGG TGGTGGTACC TTCATGAAGG TCACCAAAC
    28010      28020      28030      28040      28050      28060      28070

    >< SinI
    >< Sau96I
    >< NspIV
    NspHII ><
    NlaIV ><
        >< Eco47I
        >< Cfr13I
        >< BsiZI
    BscBI ><
        >< Bmel8I
        >< AvaII
        >< AsuI
    >< Fnu4HI    >< RsaI
    >< Esp3I    >< Csp6I    >< Tru9I
    >< BsmAI    >< BsmBI    >< MseI
    >< Alw26I    >< AfaI    >< DraI
    >< MseI
GCTGCATTTA GAGACGTACT TGTTGTTTAA AATAAACGAA CAAATTAAAA TGTCTGATAA TGGACCCCAA
    28080      28090      28100      28110      28120      28130      28140

    >< SinI
    >< Sau96I
    >< NspIV
    >< NspHII
    >< NlaIV
    >< Eco47I
    >< Cfr13I
    >< BsiZI
    >< SduI
    >< NspII
    >< Bsp1286I
    >< BmyI
    >< MaeII    >< AciI
    >< SinI
    >< Sau96I
    >< NspIV
    >< NspHII
    >< NlaIV
    >< Eco47I
    >< Cfr13I
    >< BsiZI
    >< BscBI
    >< Bmel8I
    >< AvaII    >< TfiI
    >< AsuI    >< HinfI
    >< MnlI

```

FIGURE 13.65

```

TCAAACCAAC GTAGTGCCCC CCGCATTACA TTTGGTGGAC CCACAGATTC AACTGACAAT AACCAGAATG
28150      28160      28170      28180      28190      28200      28210

      >< HinPII >< StyI
      >< HaeII
      > < Pali >< Hin6I >< EcoT14I
      > < HaeIII >< HhaI>< Eco130I
      >< BspWI >< BssTII
      > < BsuRI >< Bsp143II
      >< HgaI> < BshI >< CfoI>< BsaJI >< HgaI
GAGGACGCAA TGGGGCAAGG CCAAAACAGC GCCGACCCCA AGGTTTACCC AATAATACTG CGTCTTGGTT
28220      28230      28240      28250      28260      28270      28280

      >< TthHB8I
      > < ScrFI
      >< Pali
      >< PaeR7I
      >< NspIII
      > < MvaI
      >< HaeIII
      >< EcoRII
      >< Eco88I
      >< XhoI > < Ecl136I
      >< DsaV
      >< BsuRI
      >< SlaI > < BstOI
      >< MnlI>< TaqI> < BstNI
      >< CcrI > < BsiLI
      >< HinfI >< BshI
      >< TfiI>< BcoI>< BsaJI
      >< MnlI >< DdeI >< Aval > < ApyI
      >< AluI >< DdeI > < NlaIII >< BfrI >< Ama87I >< MnlI
CACAGCTCTC ACTCAGCATG GCAAGGAGGA ACTTAGATTC CCTCGAGGCC AGGGCGTTCC AATCAACACC
28290      28300      28310      28320      28330      28340      28350

      >< SinI
      >< Sau96I
      >< NspIV
      >< NspHII
      >< Eco47I
      >< Cfr13I
      >< Bsi2I
      >< Bme18I >< Ksp632I
      >< AvaII >< Eam1104I
      >< AsuI >< EarI > < AluI>< MboII >< MaeIII
AATAGTGGTC CAGATGACCA AATTGGCTAC TACCGAAGAG CTACCGGACG AGTTCGTGGT GGTGACGGCA
28360      28370      28380      28390      28400      28410      28420

      >< SstI
      >< SduI
      >< SacI
      >< NspII
      >< HgiAI
      >< EspI
      >< Eco24I
      >< Ecl136II >< StyI >< Sau96I
      >< DdeI >< RmaI >< NspIV
      >< CelII >< MaeI >< HaeIII
      >< Bsp1286I >< EcoT14I >< Cfr13I
      >< Bpu1102I >< Eco130I >< BsuRI
      >< BmyI >< BssTII > < BsrI
      >< BanII >< RsaI >< BsaJI >< Bsi2I

```

FIGURE 13.66

FIGURE 13.67

```

> < BshI > < BbvI >> MnlI >> BspWI >> SfaNI AfaI ><
AACAAACAAGG CCAAAGTGTG ACTAAGAAAT CTGCTGCTGA GGCATCTAAA AAGCCTCGCC AAAAACGTAC
28850 28860 28870 28880 28890 28900 28910

>> Tth111I
>> SniI
>> Sau96I
>> NspIV
>> NspHII
> < MaeII
>> Eco47I
>> Cfr13I
>> BsmBI
>> Bsi2I >> StyI
>> MaeIII >> Bme18I >> EcoT14I
>> MaeII >> Esp3I >> AvalI >> Eco130I
>> Csp6I >> BsmAI >> AsuI >> BssT1I
>> AfaI >> Alw26I> < AspI >> BsaJI
TGCCACAAAA CAGTACAACG TCACTCAAGC ATTTGGGAGA CGTGGTCCAG AACAAACCCA AGGAAATTTC
28920 28930 28940 28950 28960 28970 28980

>> SniI
>> Sau96I
>> NspIV
>> NspHII
>> NlaIV
>> Eco47I
>> Cfr13I
>> Bsi2I
>> BscBI
>> Bme18I
>> AvalI
>> AsuI
>> Pali
>> HaeIII
>> GdiII
>> Fnu4HI
>> EaeI
>> BsuRI
>> BshI
>> AciI
>> BspWI >
>> BspWI
GGGGACCAAG ACCTAATCAG ACAAGGAACT GATTACAAAC ATTGGCCGCA AATTGCACAA TTGCTCCAA
28990 29000 29010 29020 29030 29040 29050

>> BsmI
>> BscCI >> MnlI >> MaeIII >> MaeIII >> NlaIII
GTGCCTCTGC ATTCTTTGGA ATGTCACGCA TTGGCATGGA AGTCACACCT TCGGGAACAT GGCTGACTTA
29060 29070 29080 29090 29100 29110 29120

>> XhoII
>> Sau3AI
>> NdeII
>> MflI
>> MboI
>> FokI
>> Tru9I
>> NlaIV
>> NlaIII
>> MseI
>> BscBI >> BstXI>> AlwI> < Bsp143I >> AspI BspWI ><
TCATGGAGCC ATTAATTGG ATGACAAAGA TCCACAATTC AAAGACAACG TCATACTGCT GAACAAGCAC
29130 29140 29150 29160 29170 29180 29190

>> HgaI
ATTGACGCAT ACAAACATT CCCACCAACA GAGCCTAAAA AGGACAAAAA GAAAAAGACT GATGAAGCTC
29200 29210 29220 29230 29240 29250 29260

EspI ><
DdeI ><
CeiII ><
Bpu1102I ><
AluI ><

```

FIGURE 13.68

```

                >< P1eI
    >< Fnu4HI          >< MboII
    >< BspWI           >< MboII >< Ksp632I >< GsuI
    >< BsmAI           >< MaeIII >< EarI>< Fnu4HI
    >< Alw26I          >< HinfI >< Eam1104I>< BpmI
    >< AciI            >< Fnu4HI >< BbvI          >< AciI >< NlaIII
AGCCTTTGCC GCAGAGACAA AAGAAGCAGC CCACTGTGAC TCTTCTTCCT GCGGCTGACA TGGATGATTT
29270      29280      29290      29300      29310      29320      29330

                >< NlaIII >< HinfI          NlaIII ><
    >< FokI            >< AluI >< TfiI>< DdeI          >< BspHI
CTCCAGACAA CTTCAAAT CCATGAGTGG AGCTTCTGCT GATTCAACTC AGGCATAAAC ACTCATGATG
29340      29350      29360      29370      29380      29390      29400

                >< MaeII          >< AccI
ACCACACAAG GCAGATGGGC TATGTAAACG TTTTCGCAAT TCCGTTTACG ATACATAGTC TACTCTTGTC
29410      29420      29430      29440      29450      29460      29470

                >< Tru9I
                >< Tru9I
                >< MseI
    >< XmnI            >< MseI
    >< EcoRI>< MaeIII >< HpaI
    >< Asp700I >< BsgI >< HindII          Tru9I ><
    CAGAATGAAT TCTCGTAACT AAACAGCACA AGTAGGTTTA GTTAACTTTA ATCTCACATA GCAATCTTTA
29480      29490      29500      29510      29520      29530      29540

                XorII >
                TthHB8I >
                TaqI >
                Sau3AI ><
                RsaI ><
    >< ThaIPvuI >
                NdeII ><
                >< MnlI
    >< MvnIMcrI >
                MboI ><
                DpnII ><
                DpnI ><
                Csp6I ><
                >< BstUI
                >< HaeIII BspCI >
                BspAI ><
    >< TthHB8I >< Bsp50I
                >< Pali Bsp143I ><
                >< BsuRI BsiEI >
                >< BshIAfaI ><
    >< MnlI
    >< MaeIII          >< TaqI >< AciI
    ATCAATGTGT AACATTAGGG AGGACTTGAA AGAGCCACCA CATTTCATC GAGGCCACGC GGAGTACGAT
29550      29560      29570      29580      29590      29600      29610

                >< SduI
                >< NspII
                >< MboII >< VspI
    >< RsaI            >< RmaI >< Fnu4HI >< Eco24I >< Tru9I
    >< Csp6I            >< MaeI >< EarI >< Bsp1286I >< MseI
    >< AfaI            >< BbvI >< AluI>< Eam1104I >< BmyI >< AsnI
                >< BanII >< AseI

```

FIGURE 13.69

```
CGAGGGTACA GTGAATAATG CTAGGGAGAG CTGCCTATAT GGAAGAGCCC TAATGTGTAA AATTAATTTT
29620      29630      29640      29650      29660      29670      29680

                >< Tru9I    >< DdeI
                >< MseI    >< BfrI
                >< NlaIII   > < AluI
AGTAGTGCTA TCCCCATGTG ATTTTAATAG CTTCTTAGGA GAATGACAAA AAAAAAAAAA AAAAAA
29690      29700      29710      29720      29730      29740
```

FIGURE 13. 70

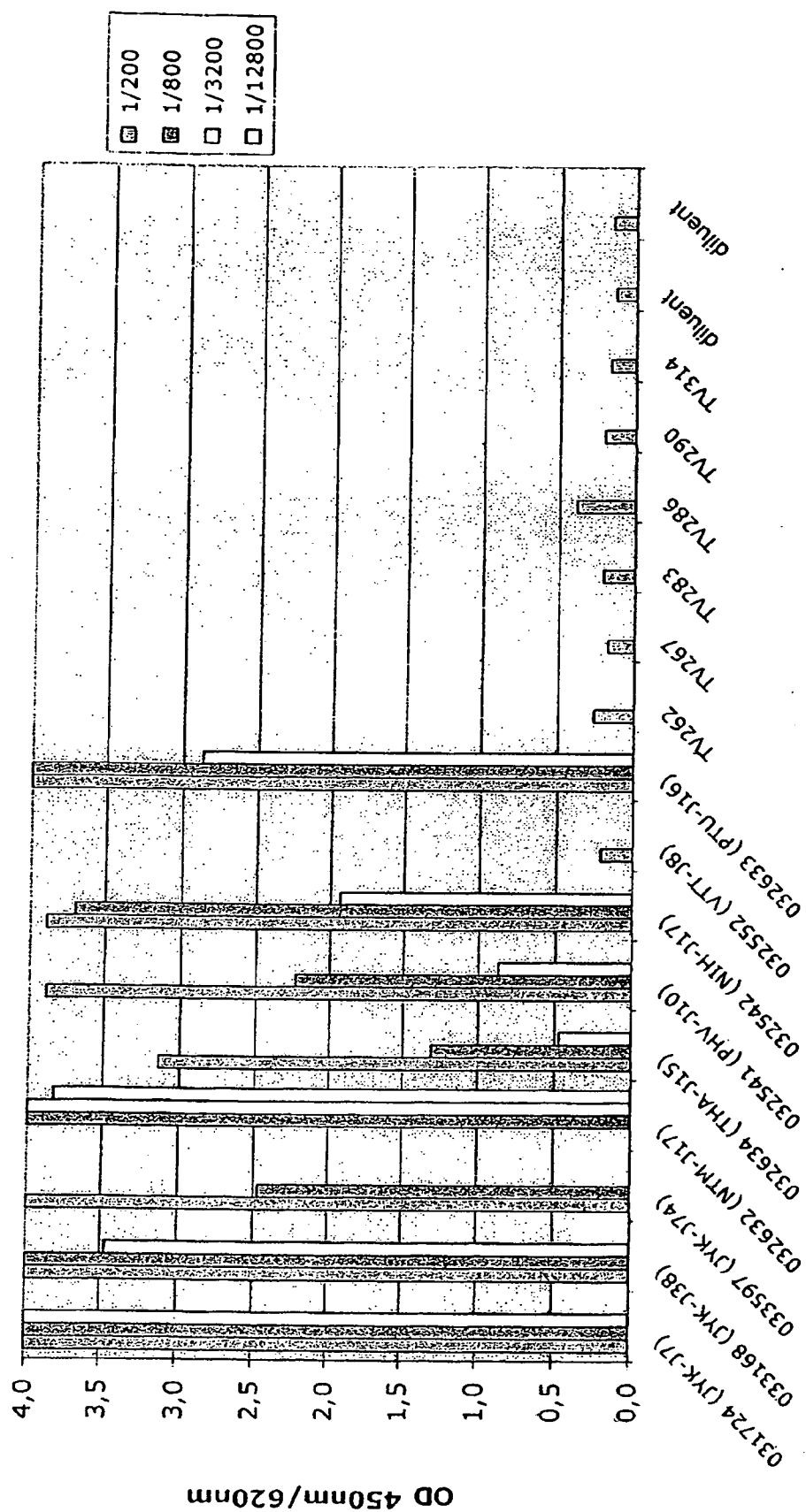


FIGURE 14

SRAS serology: Indirect N Technique (Second set)

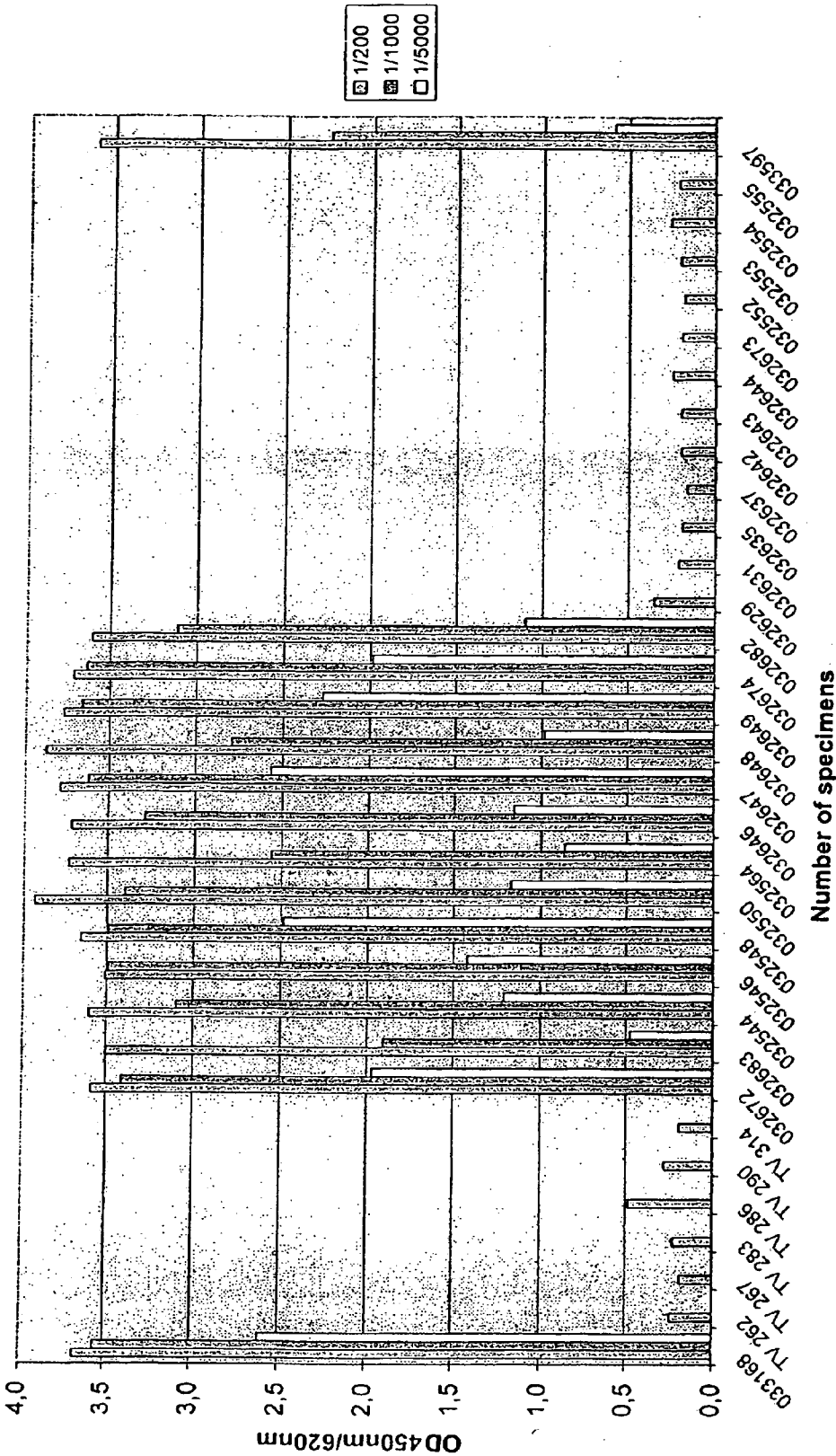
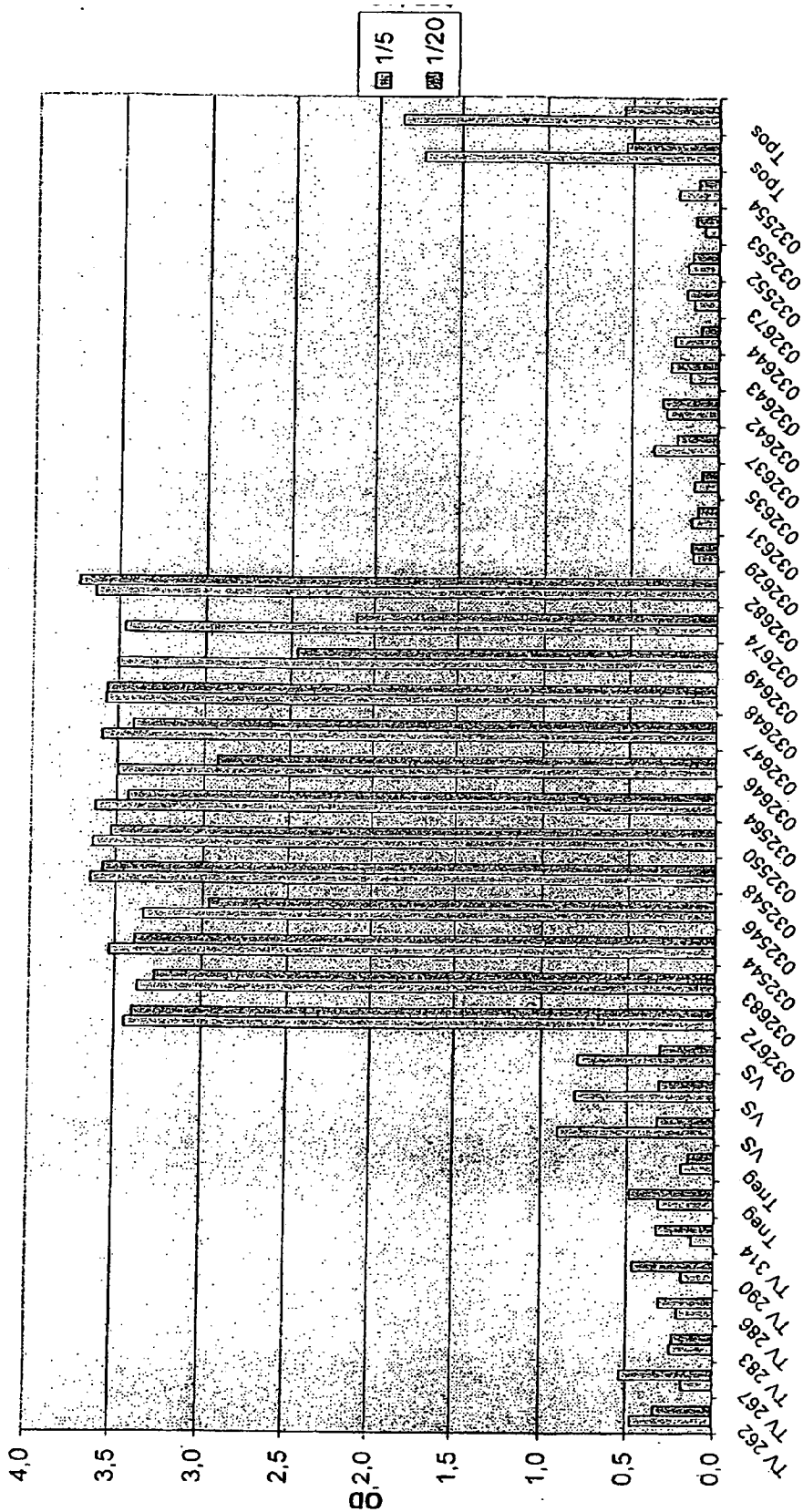


FIGURE 15

SRAS serology: Double Epitope Technique (Second set)



Number of specimens

FIGURE 17

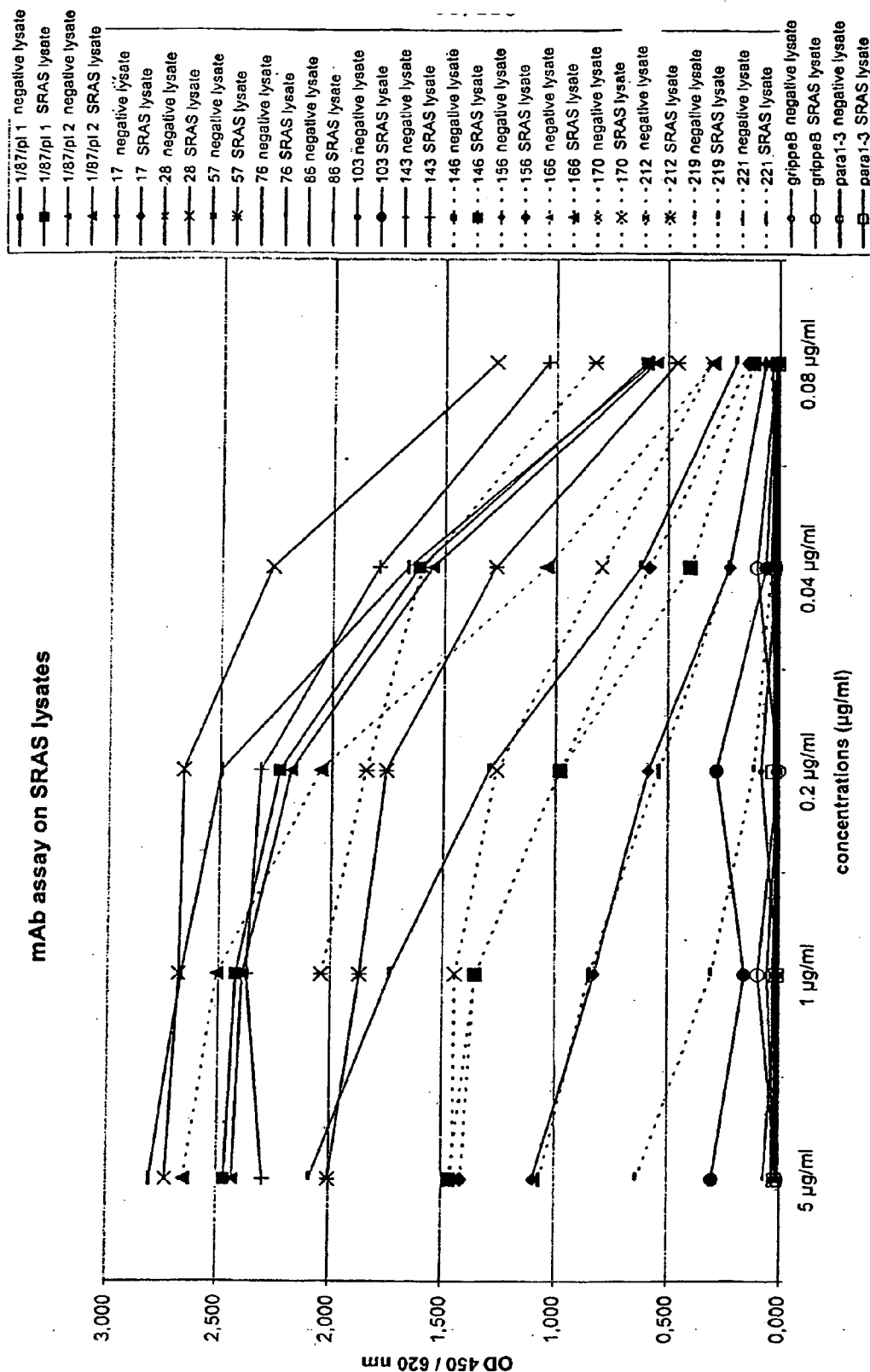


FIGURE 18

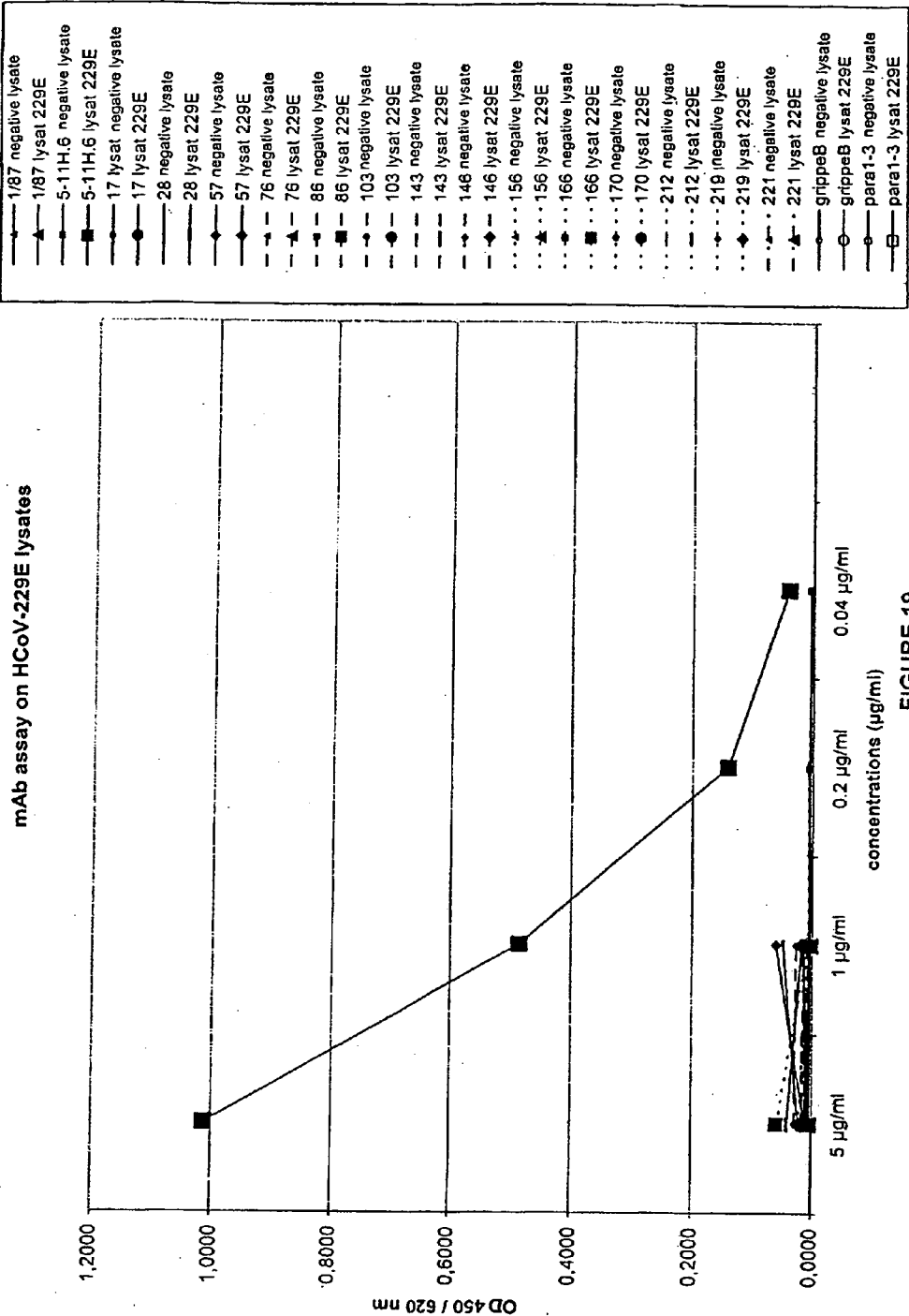


FIGURE 19

#para1-3

#grippeB

#221

#219

#212

#170

#166

#156

#146

#143

#103

#86

#76

#57

#28

#17

1/87

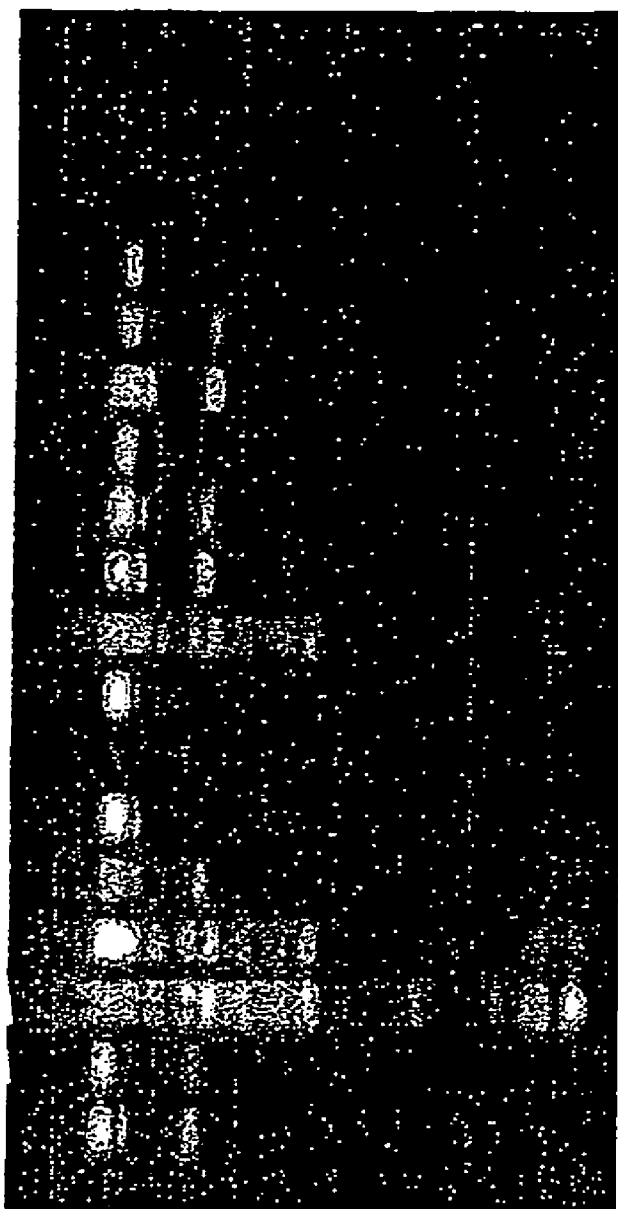


FIGURE 20

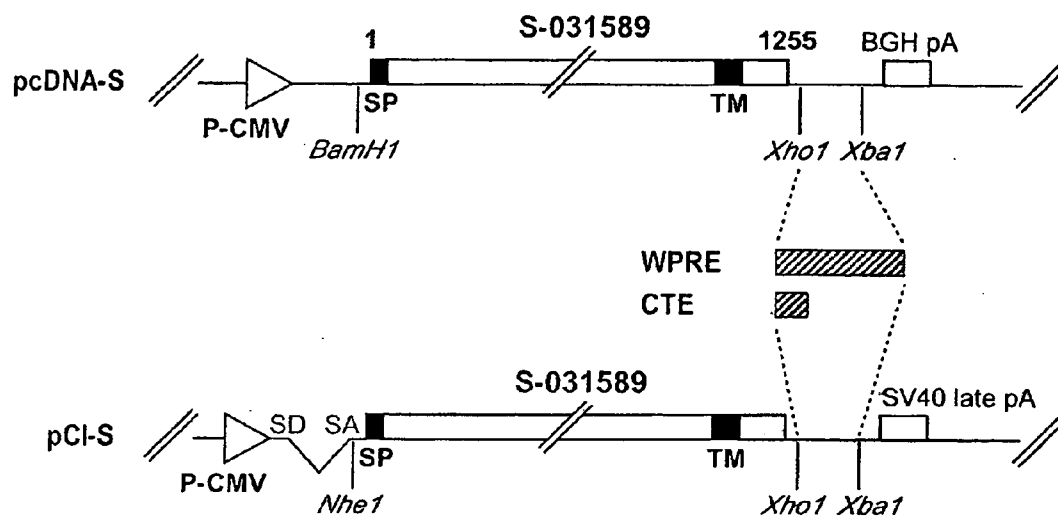


FIGURE 21

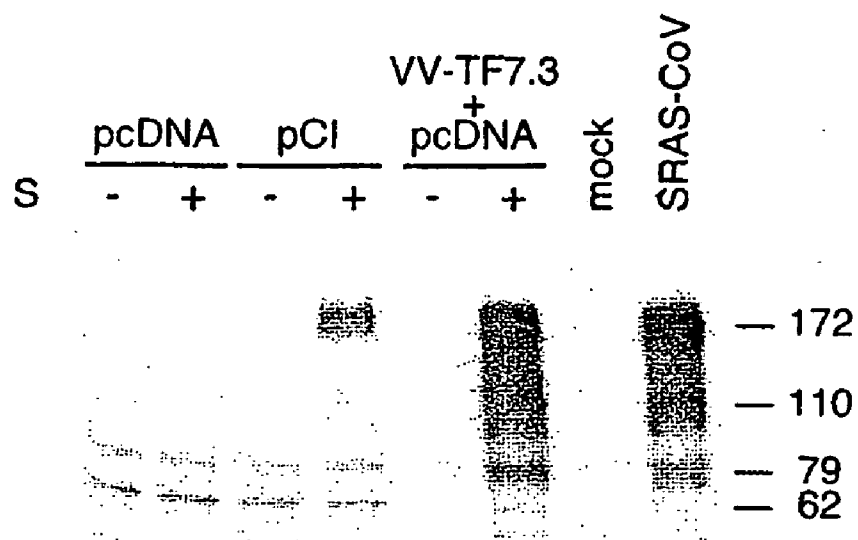
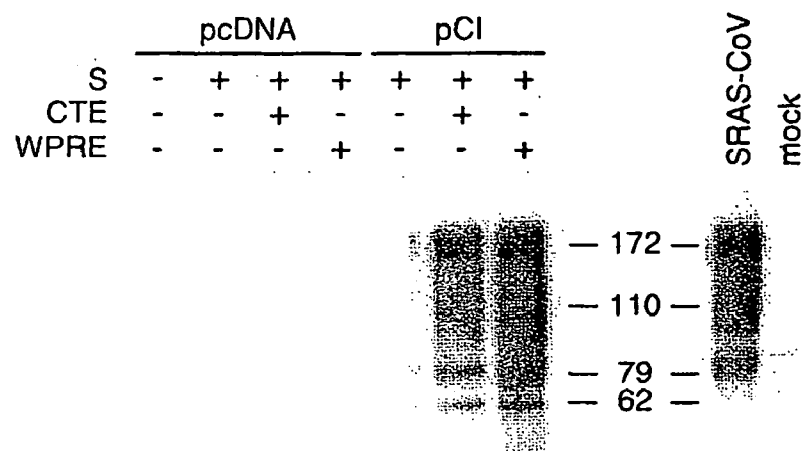


FIGURE 22

A.



B.

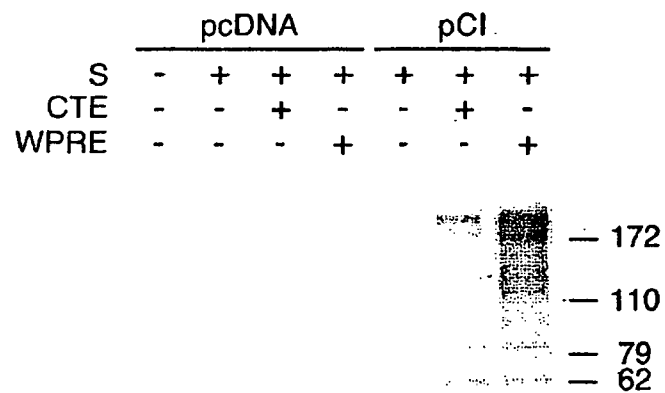


FIGURE 23

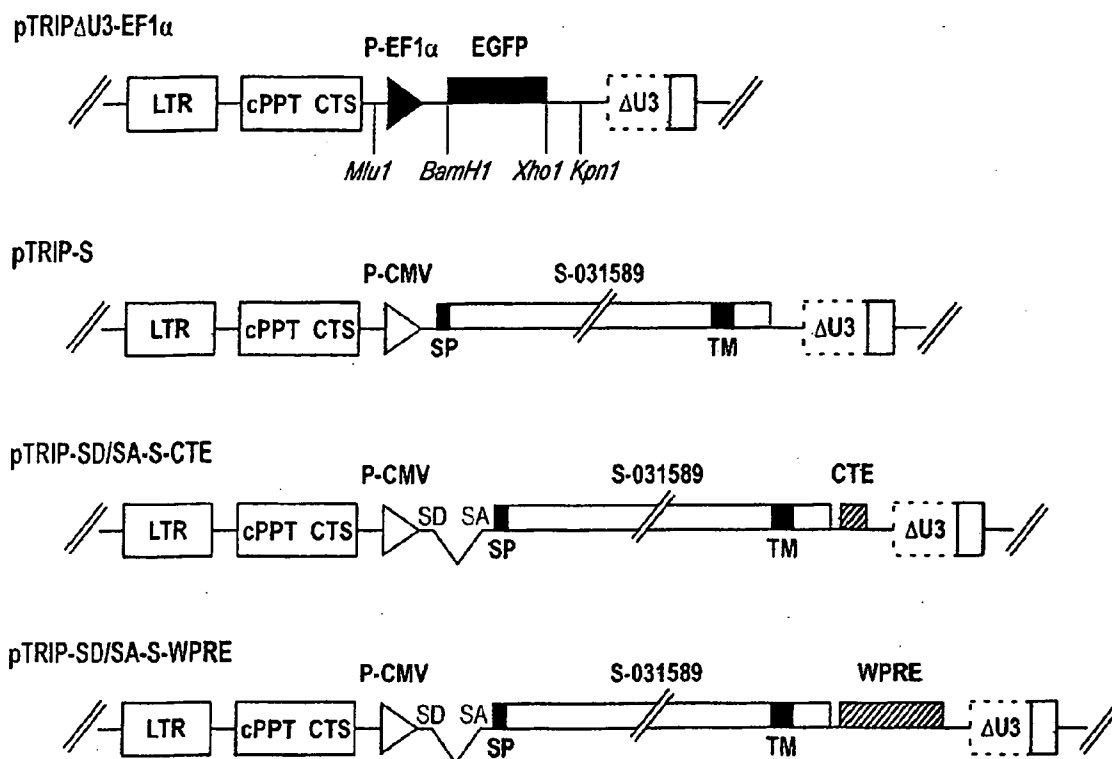


FIGURE 24

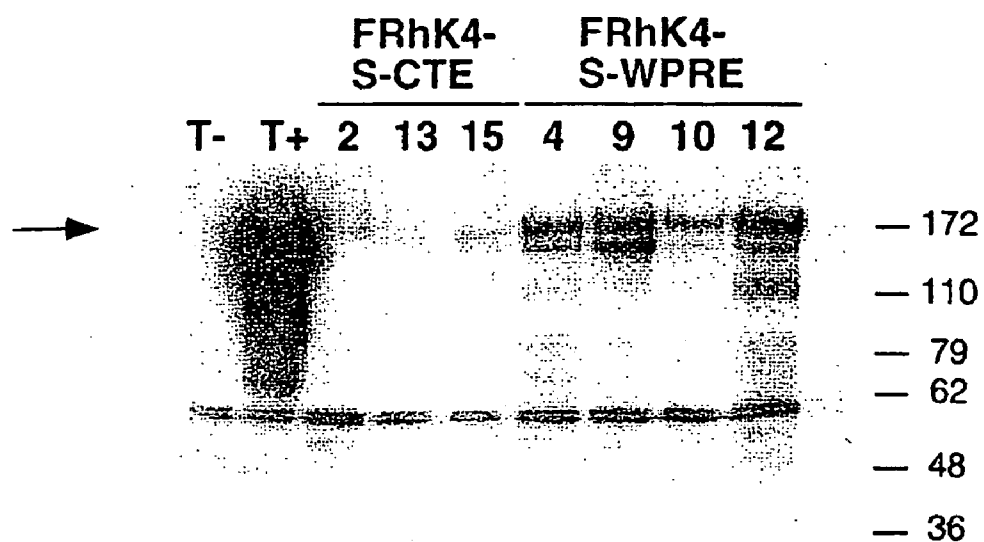


FIGURE 25

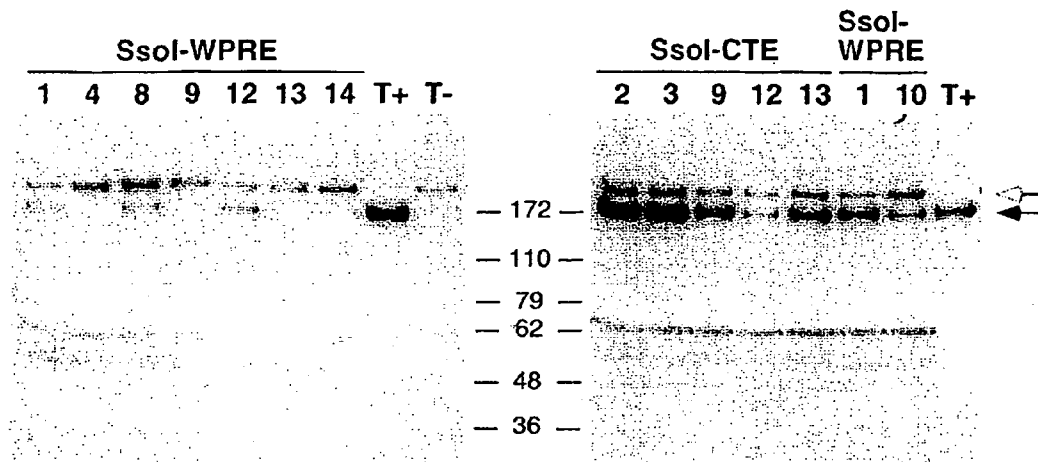
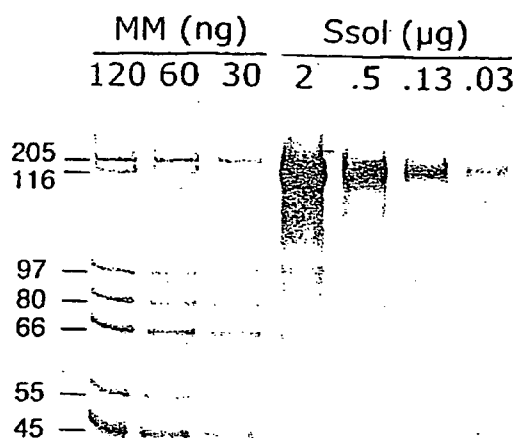
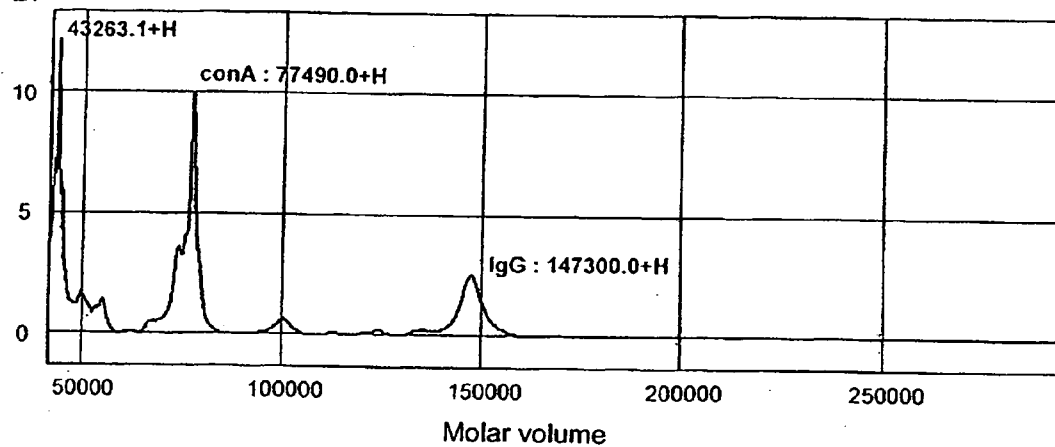


FIGURE 26

A.



B.



C.

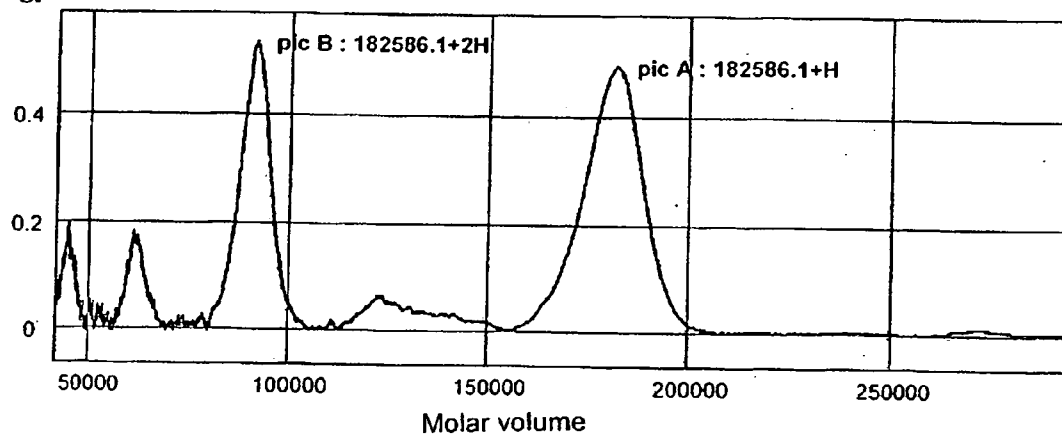


FIGURE 27 A-C

D.

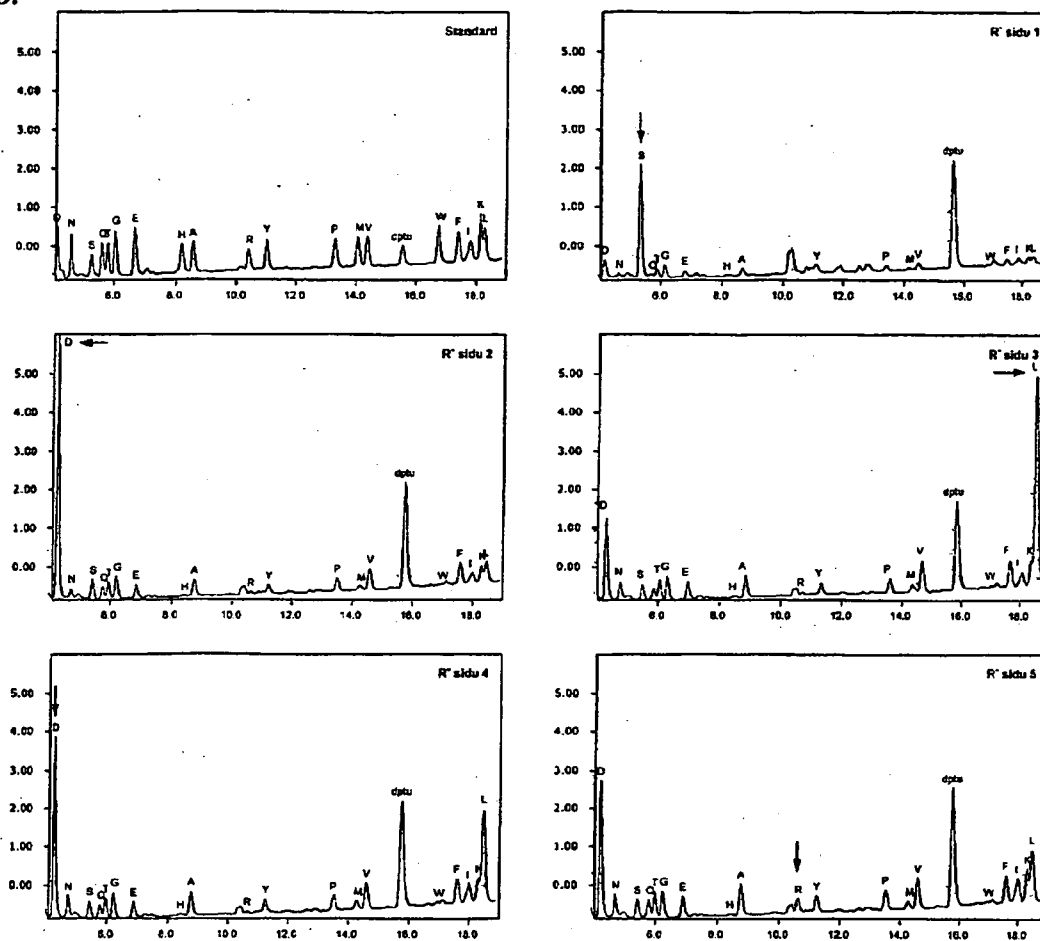
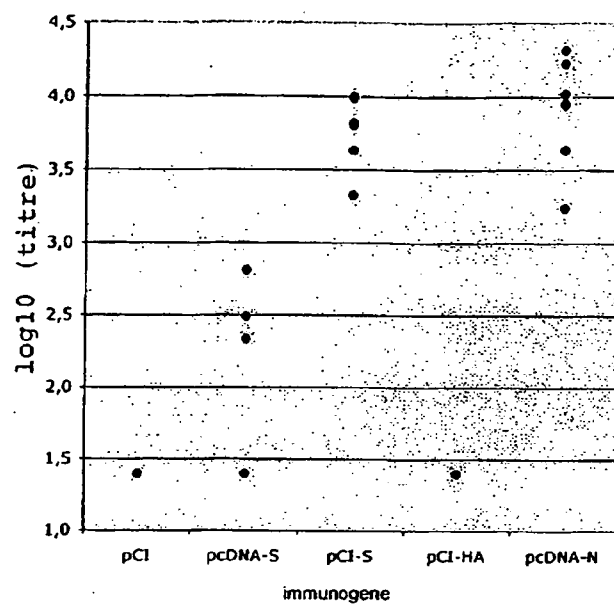


FIGURE 27 D

A.



B.

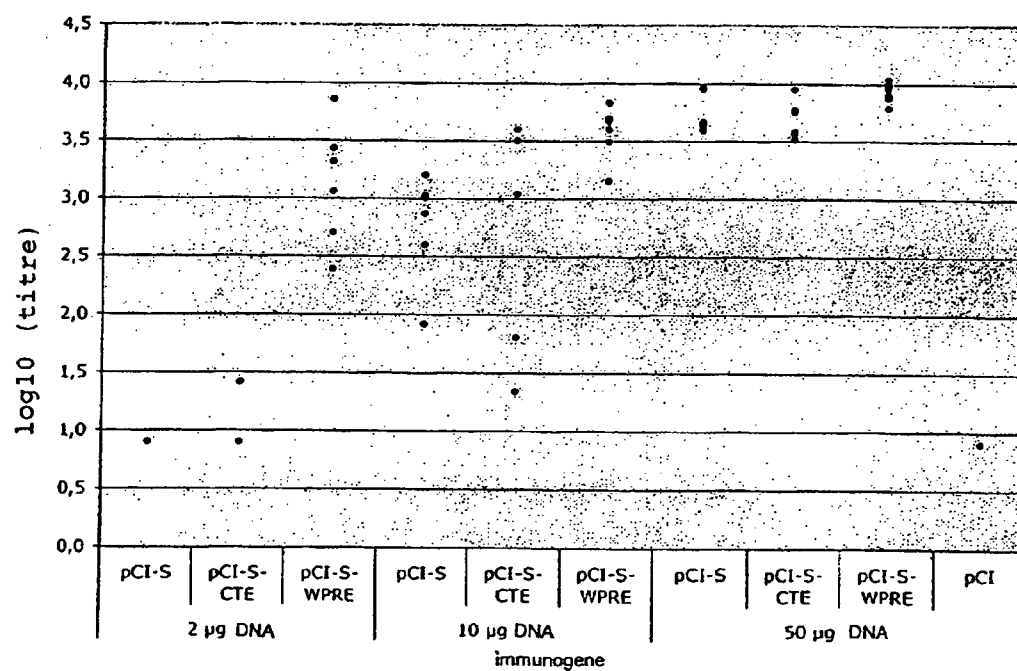


FIGURE 28

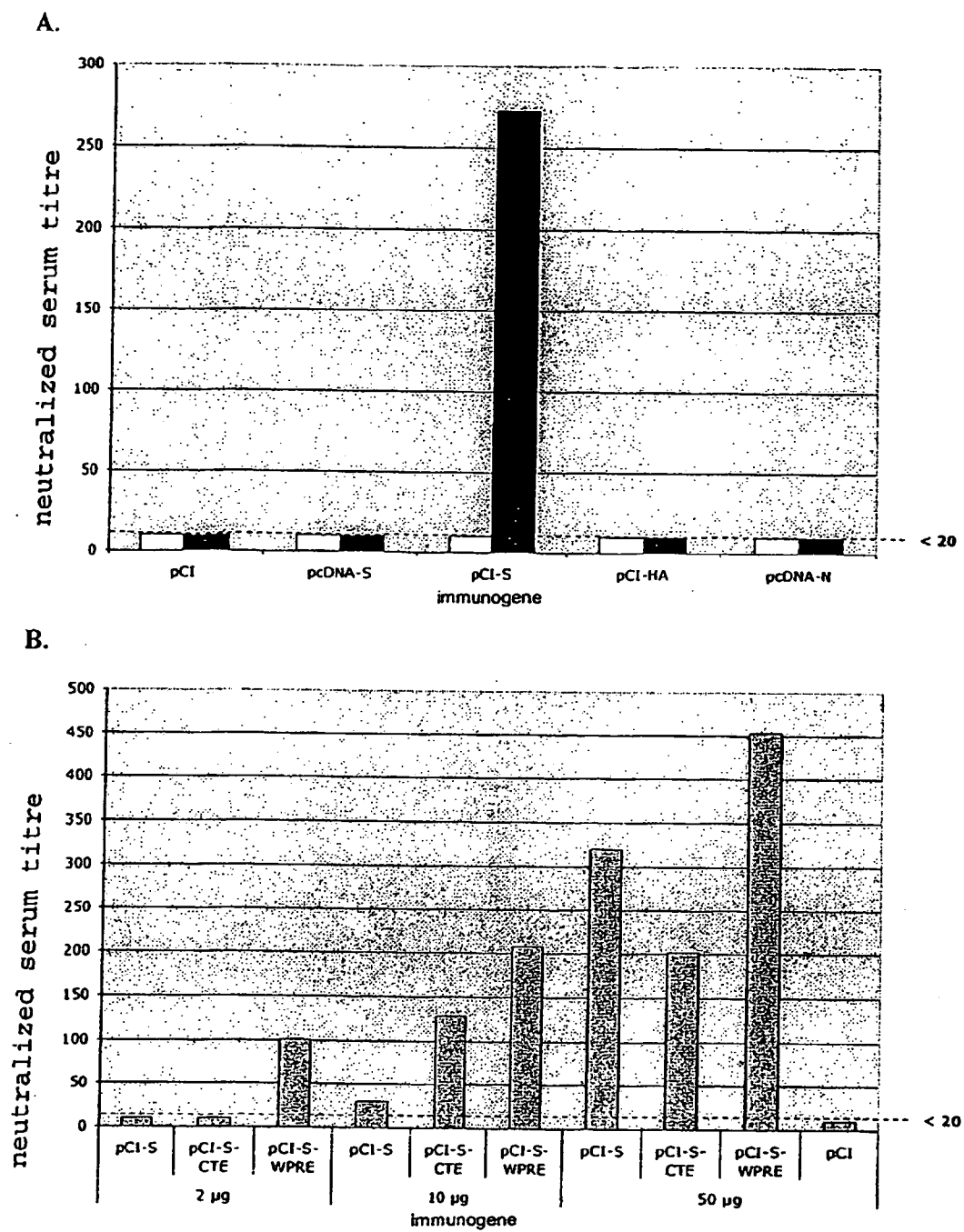


FIGURE 29

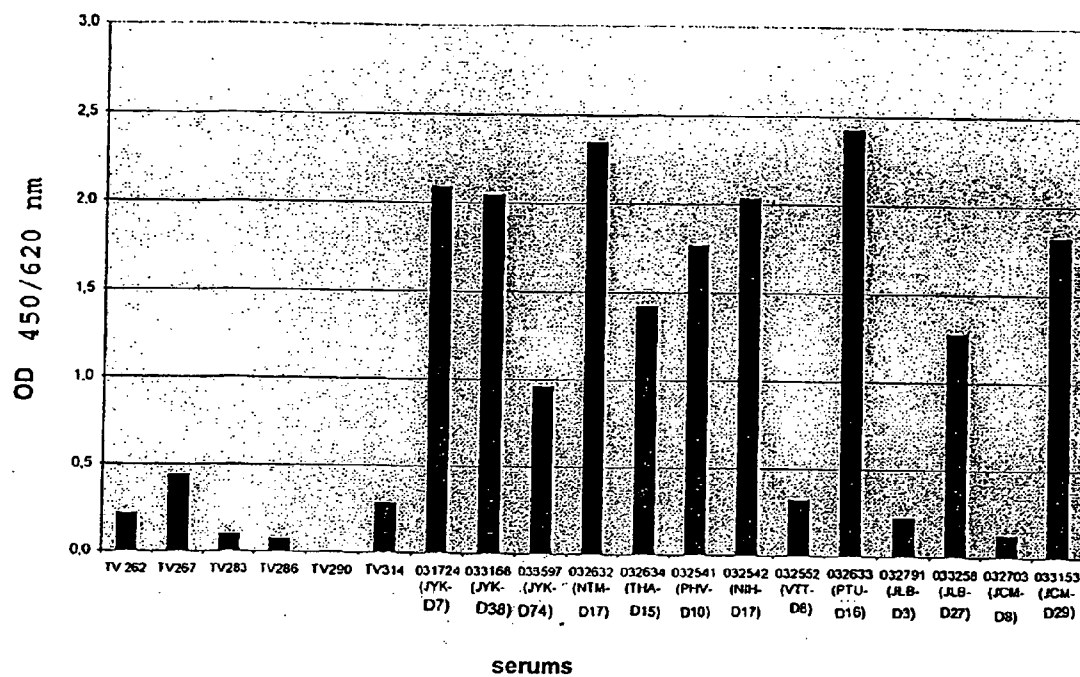


FIGURE 30

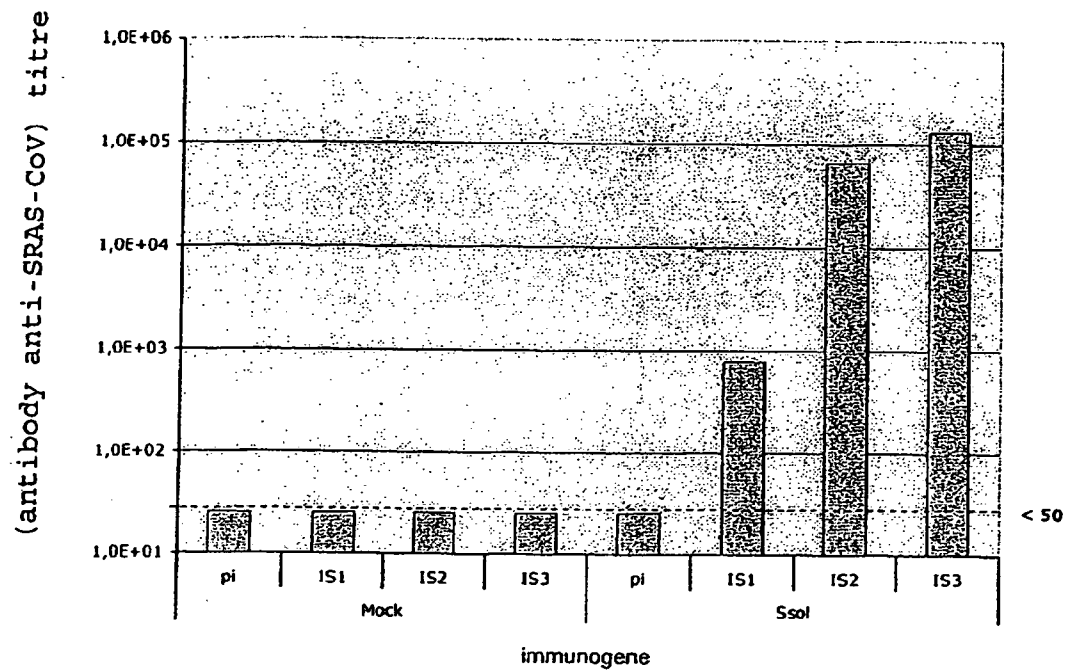


FIGURE 31

[illegible]

FIGURE 32.1

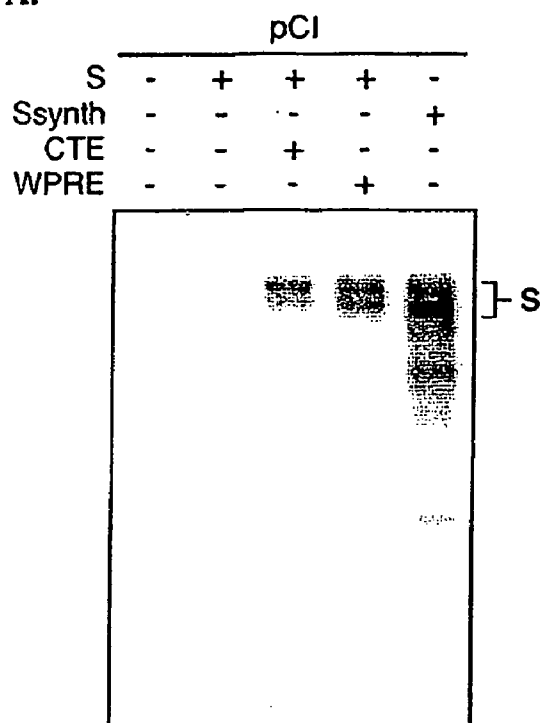
[illegible]

FIGURE 32.2

I-3059	2697	GGTGCTGGCGCTGCTCTTCAAATACCTTTTGCTATGCAAATGGCATATAGGTTCAATGGC
S-040530	2620	"A"C"A"C"C"G"G"C"C"C"C"C"C"C"C"C"C"C"C"C"C"
I-3059	2757	ATTGGAGTTACCCAAAATGTTCTCTATGAGAACCACAAAACAAATCGCCAAACCAATTAAAC
S-040530	2680	"C"C"C"G"G"G"C"G"G"C"G"G"G"G"G"G"G"G"G"G"G"G"G"G"
I-3059	2817	AAGGCGATTAGTCAAATTCAGAATCACTTACACCAACATCAACTGCATTGGGCAAGCTG
S-040530	2740	"C"C"C"C"G"C"G"GAGC"G"C"C"C"CAGC"C"CC"CCCCCCCC
I-3059	2877	CAAGACGTTGTTAACCAGAATGCTCAAGCATTAACACACTTGTTAAACAACTTAGCTCT
S-040530	2800	"G"G"G"G"G"G"CCCCC"C"C"G"CC"G"CCCC"C"G"G"G"G"G"AGC
I-3059	2937	AATTTTGGTGCAATTTCAAGTGTGCTAAATGATATCCTTTGCGGACTTGATAAAGTCGAG
S-040530	2860	"C"C"C"C"C"CAGCTC"CCCCG"C"C"CCCCGAGCA"G"C"C"CCCCG"
I-3059	2997	GCGGAGGTACAAATTGACAGGCTAATTACAGGCAGACTTCAAAGCCTTCAAACCTATGTA
S-040530	2920	"C"A"G"G"C"C"C"G"C"C"AC"C"G"GTG"G"G"CCCCC"
I-3059	3057	ACACAACAACTAATCAGGGCTGCTGAAATCAGGGCTTCTGCTAATCTTGCTGCTACTAAA
S-040530	2980	"C"G"G"G"AAAA"C"C"G"C"CCCCAGC"CCCCG"C"C"C"C"
I-3059	3117	ATGCTGAGTGTGTTCTTGACAATCAAAAAGAGTTGACTTTTGTGGAAAGGGCTACCAC
S-040530	3040	"AGC"CCCC"G"G"C"GAGC"GGGG"CCCCC"CCCCCCCCCT"
I-3059	3177	CTTATGTCCTTCCACAAGCAGCCCGCATGGTGTGCTTCTACATGTCACGTATGTG
S-040530	3100	"G"AG"CCCCC"C"G"C"CCCCC"C"C"G"G"GGGGG"C"G"C"CCCC"
I-3059	3237	CCATCCAGGAGAGGAACCTCACCACAGCGCCAGCAATTTGTCATGAAGGCAAAGCATAC
S-040530	3160	"TAG"CCCCC"CCCCCCCCCCCCC"CCCCC"CCCCC"GG"GG"CCCC"
I-3059	3297	TTCCCTCGTGAAGGTGTTTGTGTTAATGGCACTTCTTGGTTTATTACACAGAGGAAC
S-040530	3220	"CCCCG"G"C"G"C"CCCCCCCCC"CCCCCAGC"CCCCC"CCCCC"
I-3059	3357	TTCTTTCTCCACAAATAATTACTACAGACAATACATTTGTCTCAGGAAATTGTGATGTC
S-040530	3280	"CCCCAGC"C"G"C"C"C"CCCCCCCCC"CCCC"GG"C"C"CCCCCCCCG"
I-3059	3417	GTTATTGGCATCATTAACAACACAGTTTATGATCCTCTGCAACCTGAGCTTGACTCATTC
S-040530	3340	"G"C"CCCCC"C"TTTT"CCCCG"C"C"CCCCG"C"GGGG"AGC"
I-3059	3477	AAAGAAGAGCTGGACAAGTACTTCAAAAATCATACATCACCAGATGTTGATCTTGCCGAC
S-040530	3400	"G"G"AAAAA"AAAA"GG"C"C"C"C"C"C"C"G"C"G"TTTT
I-3059	3537	ATTTCAAGGCATTAACGCTTCTGTCGTCAACATTCAAAAAGAAATTGACCGCCTCAATGAG
S-040530	3460	"CAGC"CCCCC"CCCCG"G"GGGG"GGGG"GA"AA"GC"CA
I-3059	3597	GTCGCTAAAAATTTAAATGAATCACTCATTGACCTTCAAGAATTGGGAAATATGAGCAA
S-040530	3520	"G"C"G"CC"G"C"GAGC"G"C"GGGG"GGGG"CCCCG"C"GGGG
I-3059	3657	TATATTAAATGGCCTTGGTATGTTTGGCTCGGCTTCAATGCTGGACTAATTGCCATCGTC
S-040530	3580	"C"C"G"CCCCC"CCCCG"GGGG"CCCCC"CCCCG"C"GGGGG"
I-3059	3717	ATGGTTACAATCTTGCTTTGTTGCATGACTAGTTGTTGCAGTTGCCTCAAGGGTGCATGC
S-040530	3640	"GG"C"C"GG"C"CCCCCCCCC"CCCC"TT"CCCCG"AA"CC"CC"
I-3059	3777	TCTTGTGGTTCTTGCTGCAAGTTTGATGAGGATGACTCTGAGCCAGTTCTCAAGGGTGTG
S-040530	3700	AGC"CCCCAGC"CCCCCCCCC"CCCC"AGC"CCCCG"G"GGGG"CCG
I-3059	3837	AAATTACATTACACATAACGAACCTTATGGATTTGTTTATGAGATTTTTTACTCTTGAT
S-040530	3760	"GC"G"C"CCCC"GT" "CGA"
I-3059	3897	CAATTACTGCACAGCCAGTAAAAATTGACAAATGCTTCTCCTGCAAGT
S-040530		

FIGURE 32.3

A.



B.

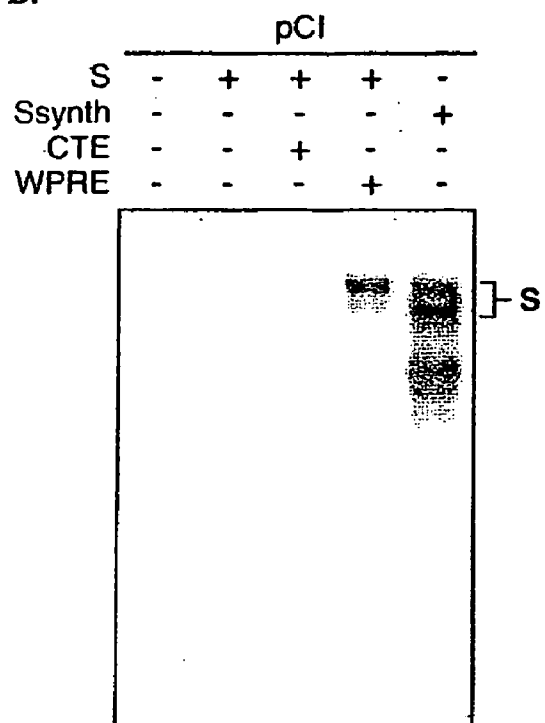
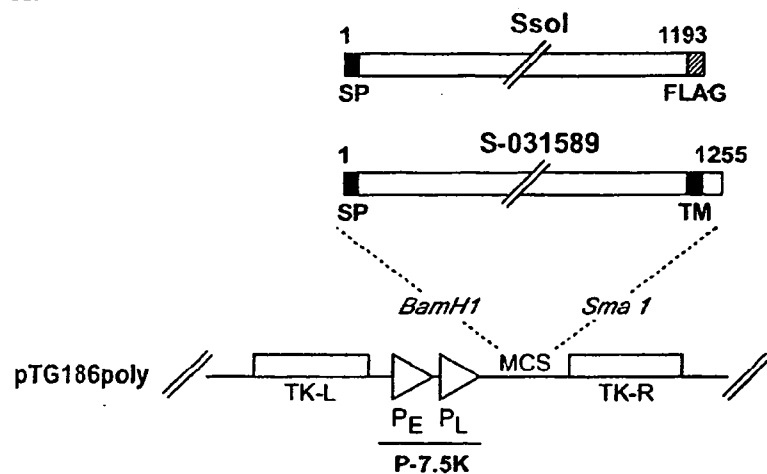
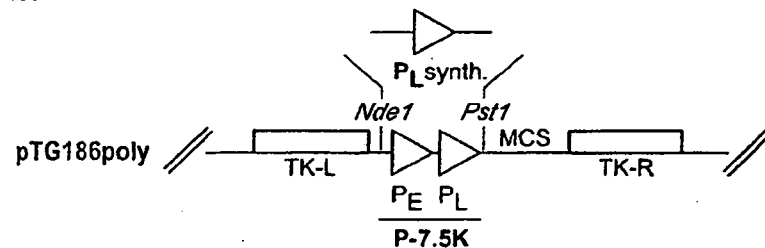


FIGURE 33

A.



B.



C.

CATATG AGC [T]₂₀GGCATATAAATA GACTC GGCGCGCC AT CTGCAG
NdeI promoteur 480 *AscI* *PstI*

FIGURE 34 A-C

D.

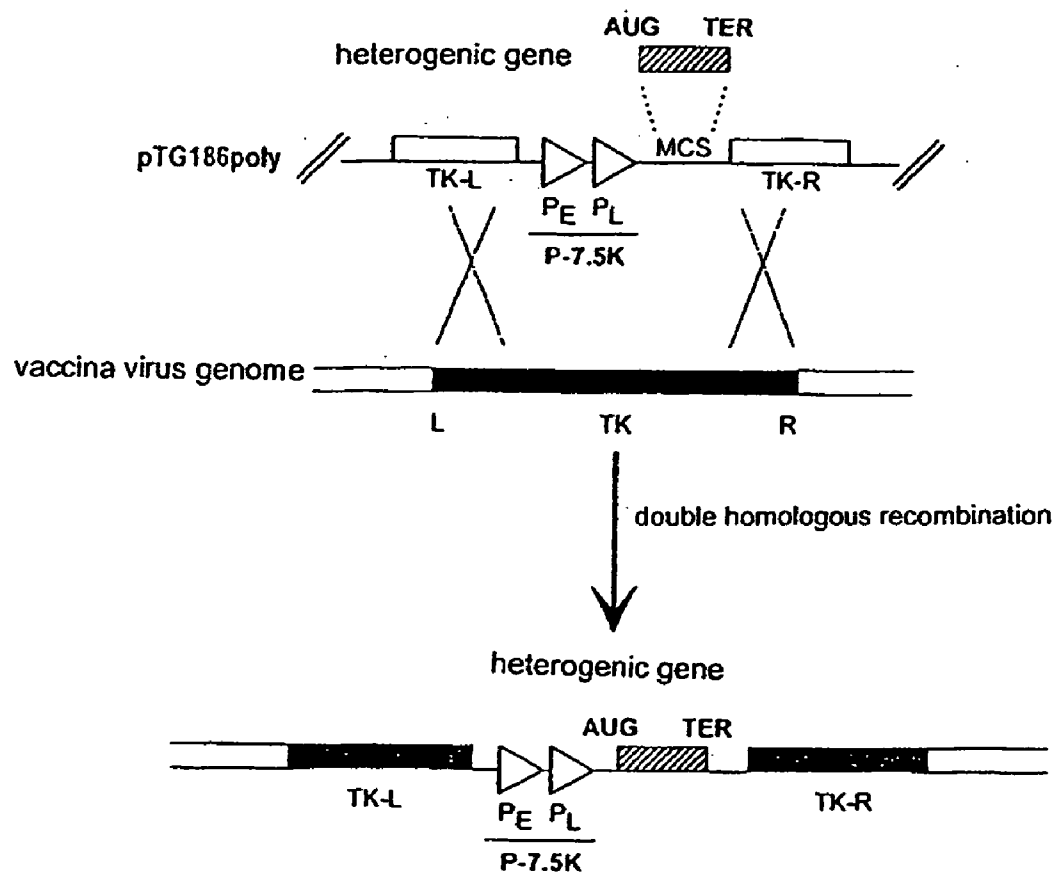


FIGURE 34 D

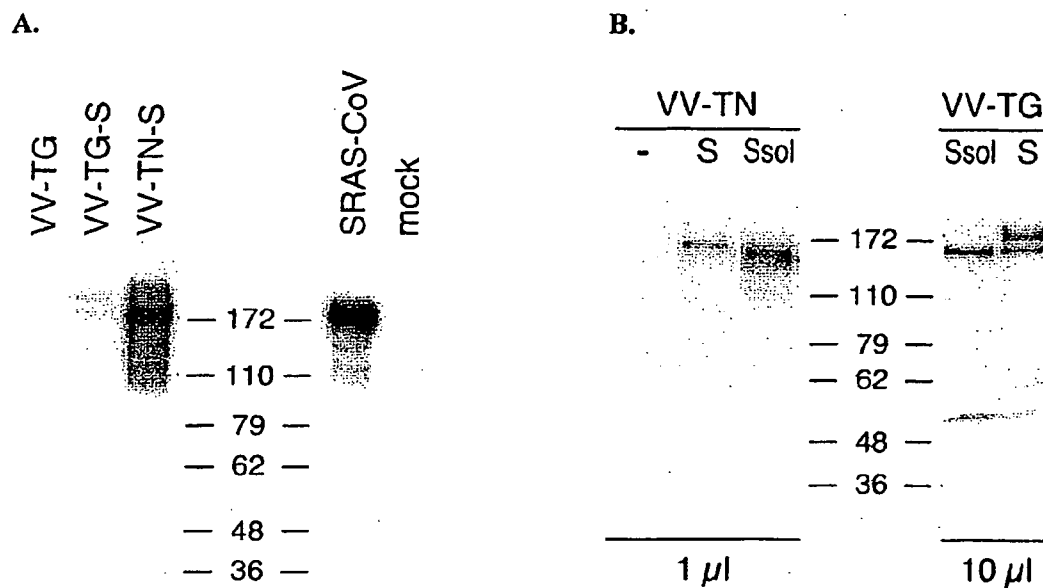
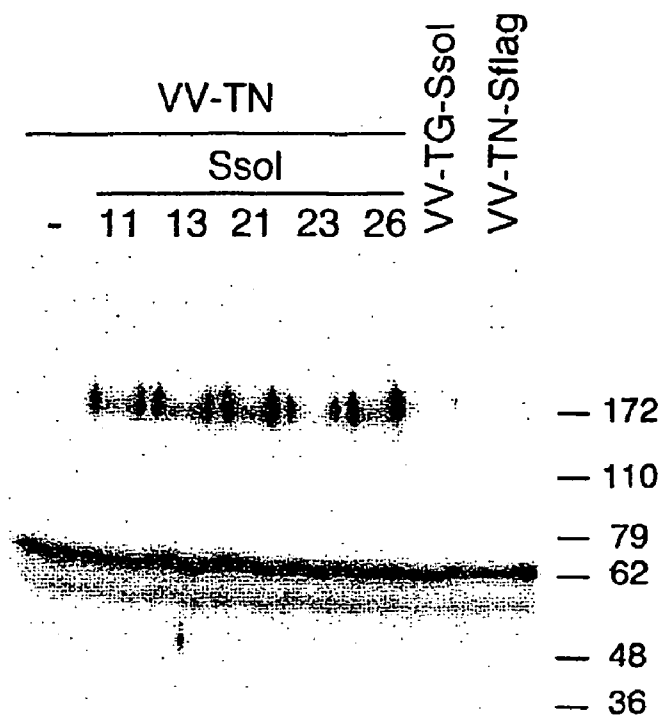


FIGURE 35

A.



B.

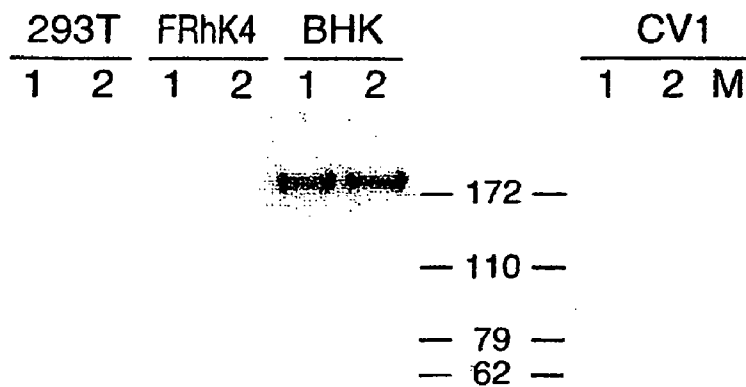


FIGURE 36



FIGURE 37

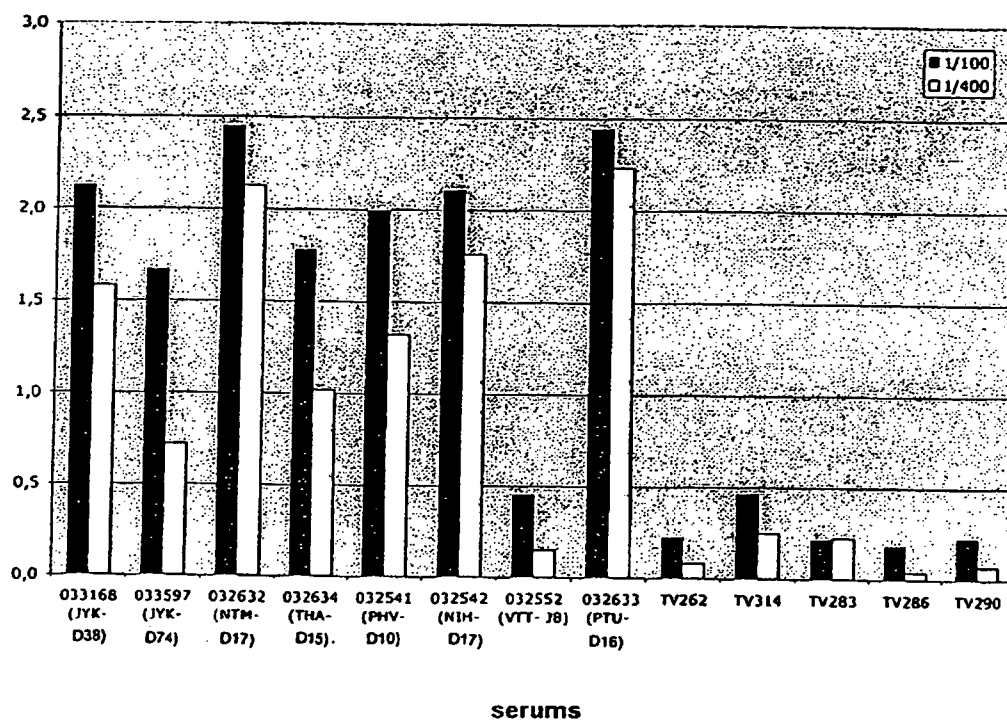
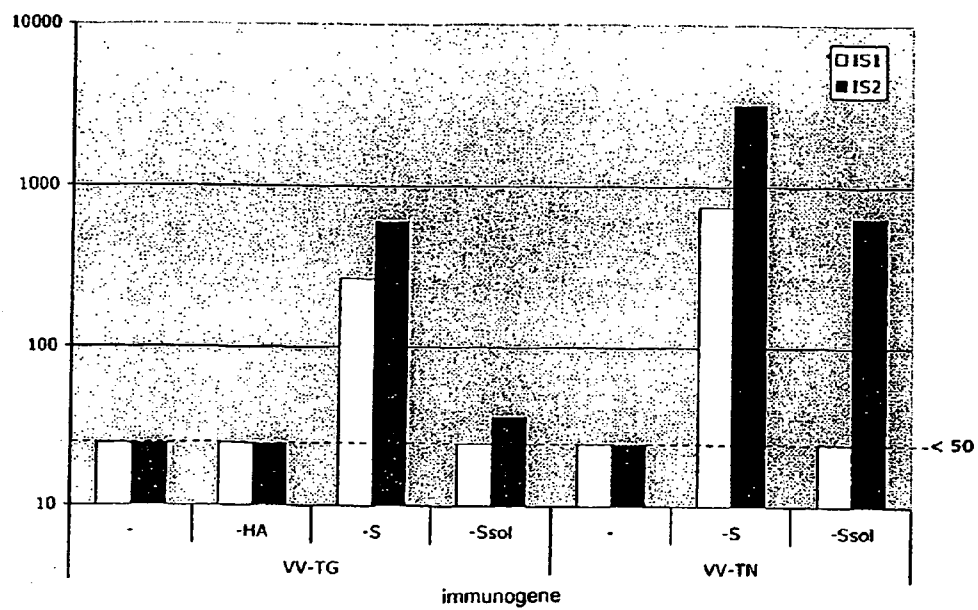


FIGURE 38

A.



B.

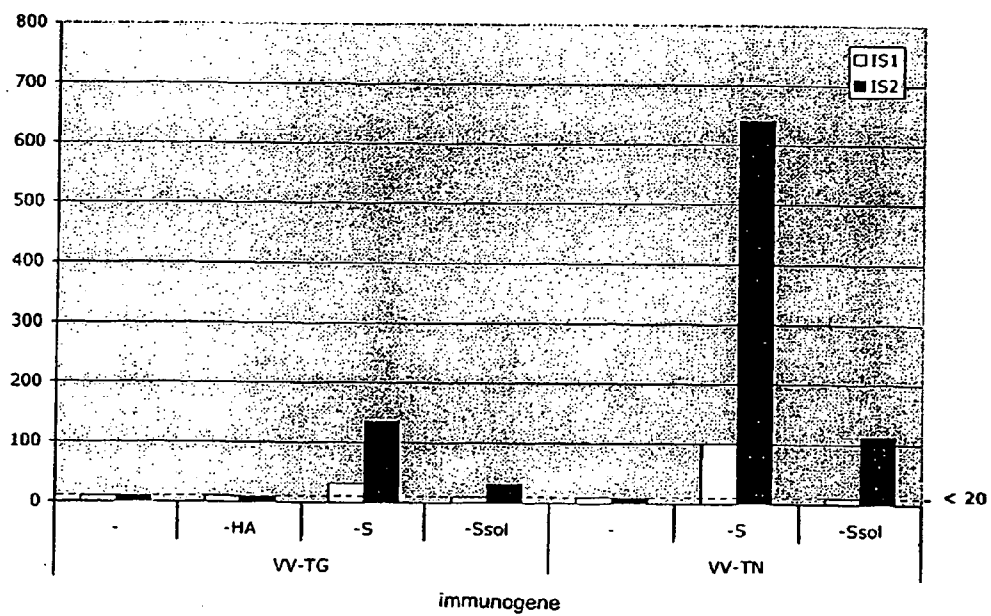


FIGURE 39

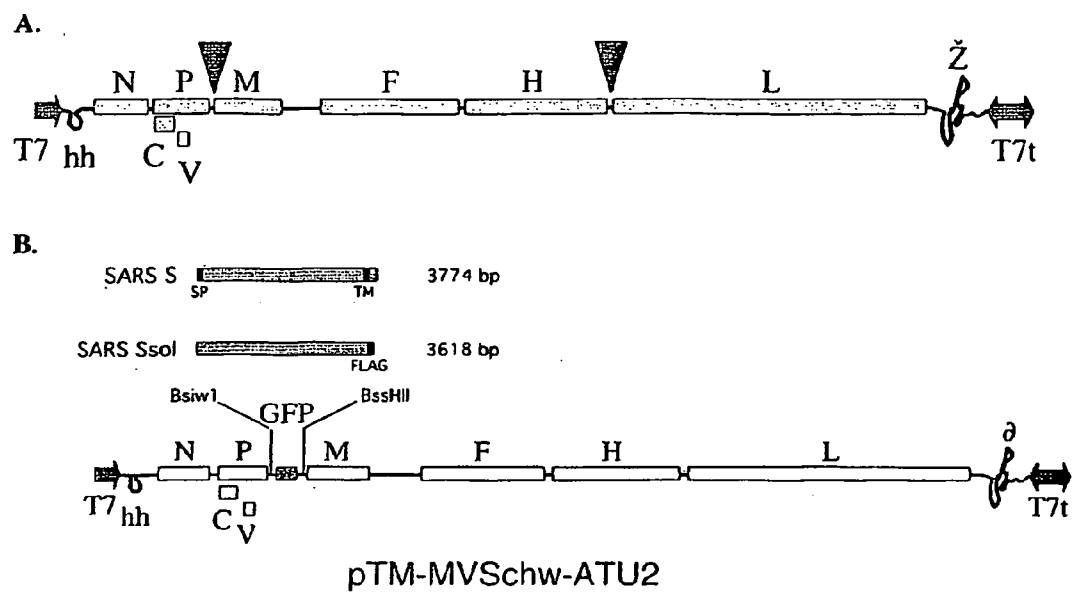


FIGURE 40

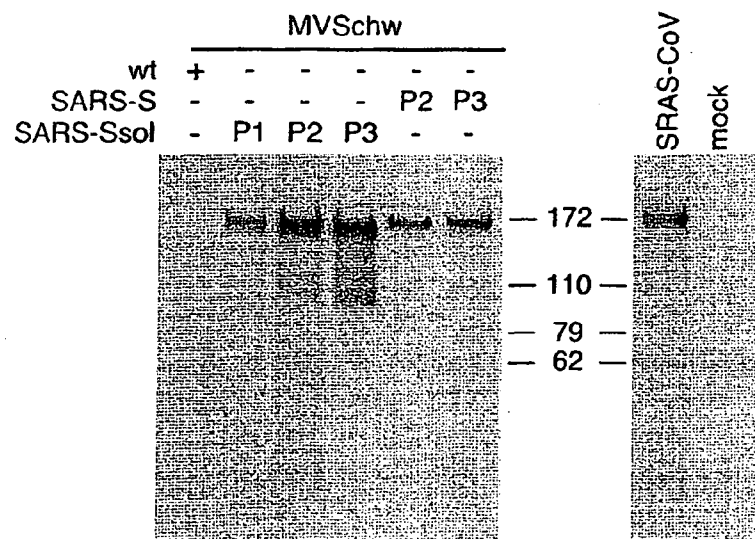


FIGURE 41

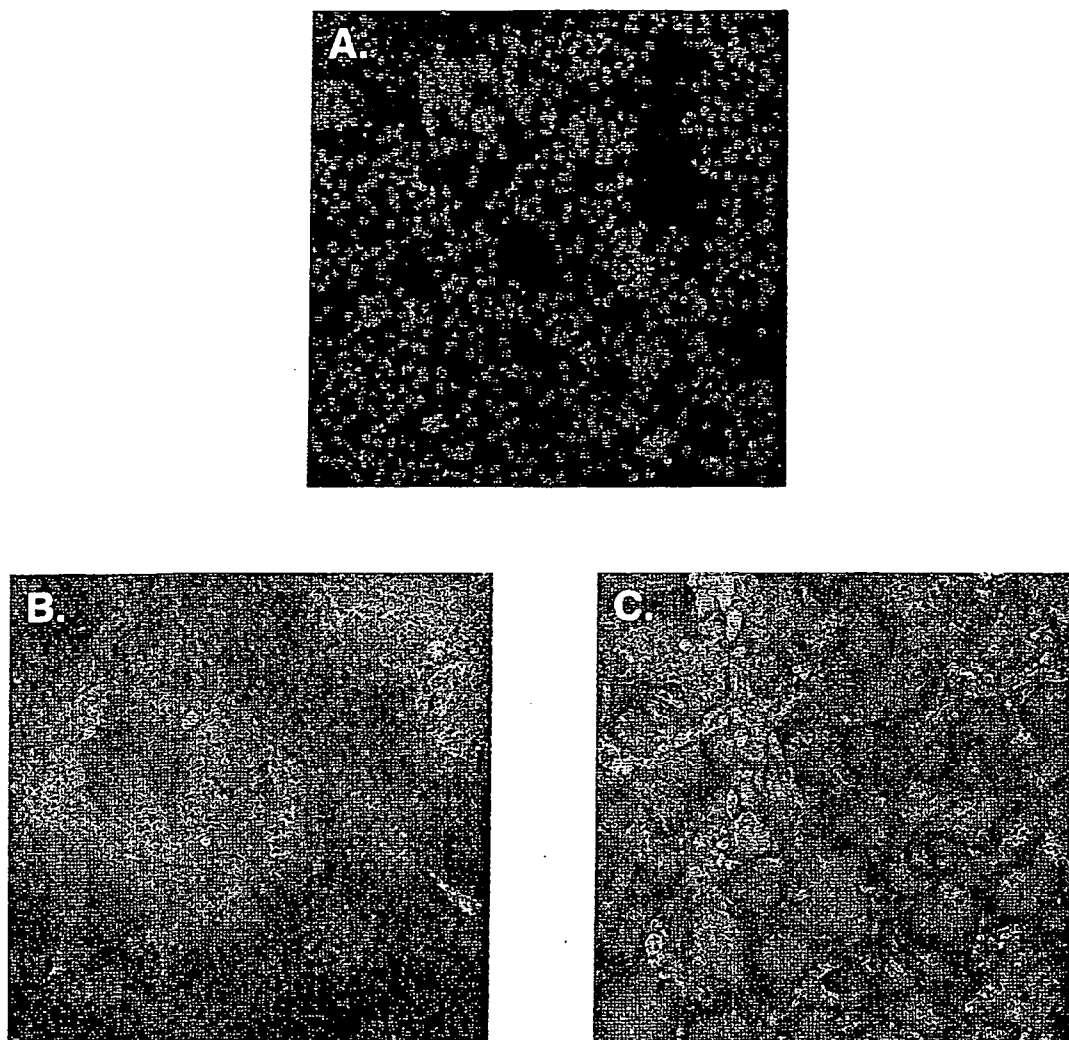


FIGURE 42

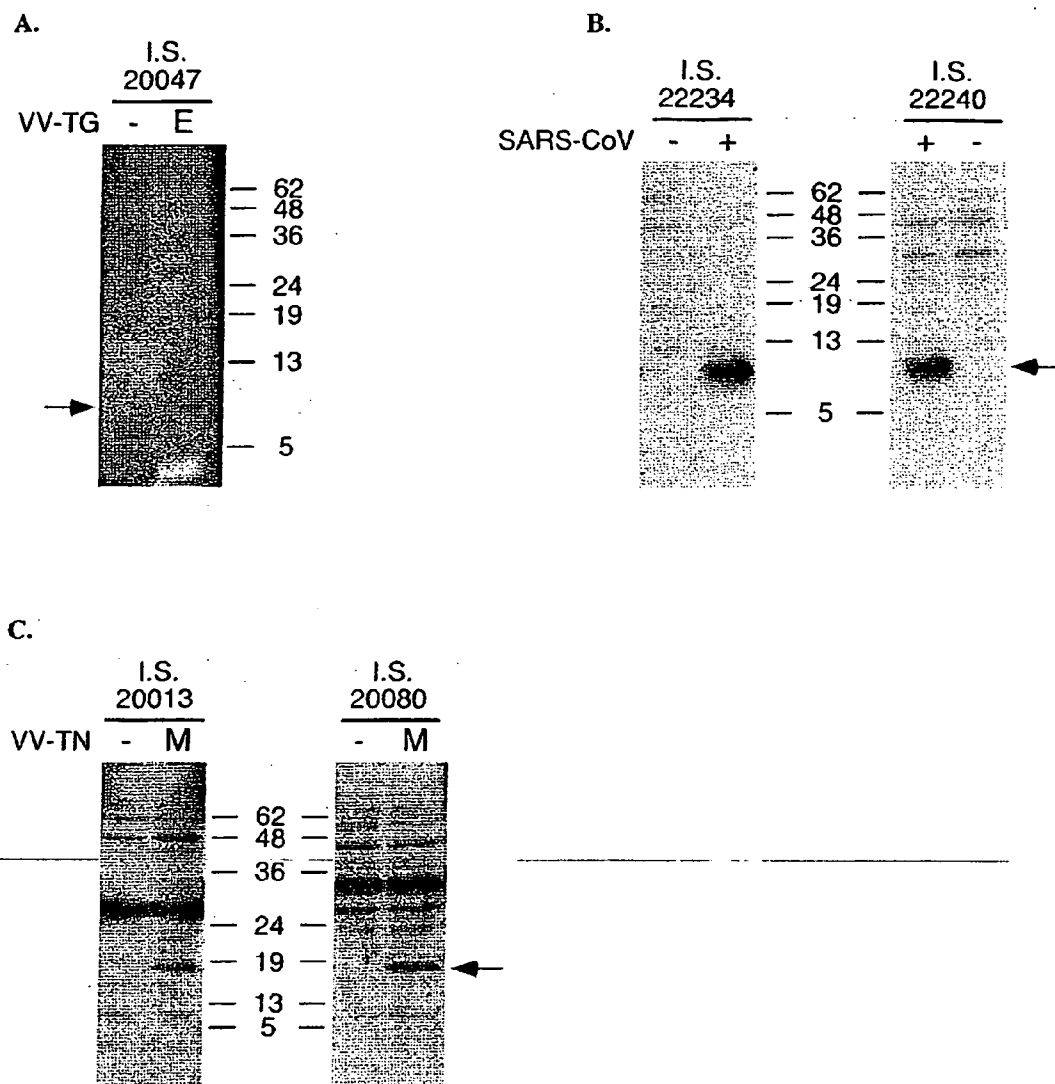


FIGURE 43

NOVEL STRAIN OF SARS-ASSOCIATED CORONAVIRUS AND APPLICATIONS THEREOF

[0001] The present invention relates to a novel strain of severe acute respiratory syndrome (SARS)-associated coronavirus derived from a sample recorded under No. 031589 and collected in Hanoi (Vietnam), to nucleic acid molecules derived from its genome, to the proteins and peptides encoded by said nucleic acid molecules and to their applications, in particular as diagnostic reagents and/or as vaccine.

[0002] Coronavirus is a virus containing single-stranded RNA, of positive polarity, of approximately 30 kilobases which replicates in the cytoplasm of the host cells; the 5' end of the genome has a capped structure and the 3' end contains a polyA tail. This virus is enveloped and comprises, at its surface, peplomer structures called spicules.

[0003] The genome comprises the following open reading frames or ORFs, from its 5' end to its 3' end: ORF1a and ORF1b corresponding to the proteins of the transcription-replication complex, and ORF-S, ORF-E, ORF-M and ORF-N corresponding to the structural proteins S, E, M and N. It also comprises ORFs corresponding to proteins of unknown function encoded by: the region situated between ORF-S and ORF-E and overlapping the latter, the region situated between ORF-M and ORF-N, and the region included in ORF-N.

[0004] The S protein is a membrane glycoprotein (200-220 kDa) which exists in the form of spicules or spikes emerging from the surface of the viral envelope. It is responsible for the attachment of the virus to the receptors of the host cell and for inducing the fusion of the viral envelope with the cell membrane.

[0005] The small envelope protein (E), also called sM (small membrane), which is a nonglycosylated transmembrane protein of about 10 kDa, is the protein present in the smallest quantity in the virion. It plays a powerful role in the coronavirus budding process which occurs at the level of the intermediate compartment in the endoplasmic reticulum and the Golgi apparatus.

[0006] The M protein or matrix protein (25-30 kDa) is a more abundant membrane glycoprotein which is integrated into the viral particle by an M/E interaction, whereas the incorporation of S into the particles is directed by an S/M interaction. It appears to be important for the viral maturation of coronaviruses and for the determination of the site where the viral particles are assembled.

[0007] The N protein or nucleocapsid protein (45-50 kDa) which is the most conserved among the coronavirus structural proteins is necessary for encapsidating the genomic RNA and then for directing its incorporation into the virion. This protein is probably also involved in the replication of the RNA.

[0008] When the host cell is infected, the reading frame (ORF) situated in 5' of the viral genome is translated into a polypeptide which is cleaved by the viral proteases and then releases several nonstructural proteins such as the RNA-dependent RNA polymerase (Rep) and the ATPase helicase (Hel). These two proteins are involved in the replication of the viral genome and in the generation of transcripts which are used in the synthesis of the viral proteins. The mechanisms

by which these subgenomic mRNAs are produced are not completely understood; however, recent facts indicate that the sequences for regulation of transcription at the 5' end of each gene represent signals which regulate the discontinuous transcription of the subgenomic mRNAs.

[0009] The proteins of the viral membrane (S, E and M proteins) are inserted into the intermediate compartment, whereas the replicated RNA (+ strand) is assembled with the N (nucleocapsid) protein. This protein-RNA complex then combines with the M protein contained in the membranes of the endoplasmic reticulum and the viral particles form when the nucleocapsid complex buds into the endoplasmic reticulum. The virus then migrates across the Golgi complex and eventually leaves the cell, for example by exocytosis. The site of attachment of the virus to the host cell is at the level of the S protein.

[0010] Coronaviruses are responsible for 15 to 30% of colds in humans and for respiratory and digestive infections in animals, especially cats (FIPV: Feline infectious peritonitis virus), poultry (IBV: Avian infectious bronchitis virus), mice (MHV: Mouse hepatitis virus), pigs (TGEV: Transmissible gastroenteritis virus, PEDV: Porcine Epidemic diarrhoea virus, PRCoV: Porcine Respiratory Coronavirus, HEV: Hemagglutinating encephalomyelitis Virus) and bovines (BCoV: Bovine coronavirus).

[0011] In general, each coronavirus affects only one species; in immunocompetent individuals, the infection induces optionally neutralizing antibodies and cell immunity, capable of destroying the infected cells.

[0012] An epidemic of atypical pneumonia, called severe acute respiratory syndrome (SARS) has spread in various countries (Vietnam, Hong Kong, Singapore, Thailand and Canada) during the first quarter of 2003, from an initial focus which appeared in China in the last quarter of 2002. The severity of this disease is such that its mortality rate is about 3 to 6%. The determination of the causative agent of this disease is underway by numerous laboratories worldwide.

[0013] In March 2003, a new coronavirus (SARS-CoV or SARS virus) was isolated, in association with cases of severe acute respiratory syndrome (T. G. KSIAZEK et al., *The New England Journal of Medicine*, 2003, 348, 1319-1330; C. DROSTEN et al., *The New England Journal of Medicine*, 2003, 348, 1967-1976; Peiris et al., *Lancet*, 2003, 361, 1319).

[0014] Genomic sequences of this new coronavirus have thus been obtained, in particular those of the Urbani isolate (Genbank accession No. AY274119.3 and A. MARRA et al., *Science*, May 1, 2003, 300, 1399-1404) and the Toronto isolate (Tor2, Genbank accession No. AY278741 and A. ROTA et al., *Science*, 2003, 300, 1394-1399).

[0015] The organization of the genome is comparable with that of other known coronaviruses, thus making it possible to confirm that SARS-CoV belongs to the Coronaviridae family; open reading frames ORF1a and 1b and open reading frames corresponding to the S, E, M and N proteins, and to proteins encoded by: the region situated between ORF-S and ORF-E (ORF3), the region situated between ORF-S and ORF-E and overlapping ORF-E (ORF4), the region situated between ORF-M and ORF-N (ORF7 to

ORF11) and the region corresponding to ORF-N (ORF13 and ORF14), have in particular been identified.

[0016] Seven differences have been identified between the sequences of the Tor2 and Urbani isolates; 3 correspond to silent mutations (c/t at position 16622 and a/g at position 19064 of ORF1b, t/c at position 24872 of ORF-S) and 4 modify the amino acid sequence of respectively: the proteins encoded by ORF1a (c/t at position 7919 corresponding to the A/V mutation), the S protein (g/t at position 23220 corresponding to the A/S mutation), the protein encoded by ORF3 (a/g at position 25298 corresponding to the R/G mutation) and the M protein (t/c at position 26857 corresponding to the S/P mutation).

[0017] In addition, phylogenetic analysis shows that SARS-CoV is distant from other coronaviruses and that it did not appear by mutation of human respiratory coronaviruses nor by recombination between known coronaviruses (for a review, see Holmes, J. C. I., 2003, 111, 1605-1609).

[0018] The determination and the taking into account of new variants are important for the development of reagents for the detection and diagnosis of SARS which are sufficiently sensitive and specific, and immunogenic compositions capable of protecting populations against epidemics of SARS.

[0019] The inventors have now identified another strain of SARS-associated coronavirus which is distinguishable from the Tor2 and Urbani isolates.

[0020] The subject of the present invention is therefore an isolated or purified strain of severe acute respiratory syndrome-associated human coronavirus, characterized in that its genome has, in the form of complementary DNA, a serine codon at position 23220-23222 of the gene for the S protein or a glycine codon at position 25298-25300 of the gene for ORF3, and an alanine codon at position 7918-7920 of ORF1a or a serine codon at position 26857-26859 of the gene for the M protein, said positions being indicated in terms of reference to the Genbank sequence AY274119.3.

[0021] According to an advantageous embodiment of said strain, the DNA equivalent of its genome has a sequence corresponding to the sequence SEQ ID No: 1; this coronavirus strain is derived from the sample collected from the bronchoalveolar washings from a patient suffering from SARS, recorded under the No. 031589 and collected at the Hanoi (Vietnam) French hospital.

[0022] In accordance with the invention, said sequence SEQ ID No: 1 is that of the deoxyribonucleic acid corresponding to the ribonucleic acid molecule of the genome of the isolated coronavirus strain as defined above.

[0023] The sequence SEQ ID No: 1 is distinguishable from the Genbank sequence AY274119.3 (Tor2 isolate) in that it possesses the following mutations:

[0024] g/t at position 23220; the alanine codon (gct) at position 577 of the amino acid sequence of the Tor2 S protein is replaced by a serine codon (tct),

[0025] a/g at position 25298; the arginine codon (aga) at position 11 of the amino acid sequence of the protein encoded by the Tor2 ORF3 is replaced by a glycine codon (gga).

[0026] In addition, the sequence SEQ ID No: 1 is distinguishable from the Genbank sequence AY278741 (Urbani isolate) in that it possesses the following mutations:

[0027] t/c at position 7919; the valine codon (ggt) in position 2552 of the amino acid sequence of the protein encoded by ORF1a is replaced by an alanine codon (gct),

[0028] t/c at position 16622: this mutation does not modify the amino acid sequence of the proteins encoded by ORF1b (silent mutation),

[0029] g/a at position 19064: this mutation does not modify the amino acid sequence of the proteins encoded by ORF1b (silent mutation),

[0030] c/t at position 24872: this mutation does not modify the amino acid sequence of the S protein, and

[0031] c/t at position 26857: the proline codon (ccc) at position 154 of the amino acid sequence of the M protein is replaced by a serine codon (tcc).

[0032] Unless otherwise stated, the positions of the nucleotide and peptide sequences are indicated with reference to the Genbank sequence AY274119.3.

[0033] The subject of the present invention is also an isolated or purified polynucleotide, characterized in that its sequence is that of the genome of the isolated coronavirus strain as defined above.

[0034] According to an advantageous embodiment of said polynucleotide, it has the sequence SEQ ID No: 1.

[0035] The subject of the present invention is also an isolated or purified polynucleotide, characterized in that its sequence hybridizes under high stringency conditions with the sequence of the polynucleotide as defined above.

[0036] The terms "isolated or purified" mean modified "by the hand of humans" from the natural state; in other words if an object exists in nature, it is said to be isolated or purified if it is modified or extracted from its natural environment or both. For example, a polynucleotide or a protein/peptide naturally present in a living organism is neither isolated nor purified; on the other hand, the same polynucleotide or protein/peptide separated from coexisting molecules in its natural environment, obtained by cloning, amplification and/or chemical synthesis is isolated for the purposes of the present invention. Furthermore, a polynucleotide or a protein/peptide which is introduced into an organism by transformation, genetic manipulation or by any other method, is "isolated" even if it is present in said organism. The term purified as used in the present invention means that the proteins/peptides according to the invention are essentially free of association with the other proteins or polypeptides, as is for example the product purified from the culture of recombinant host cells or the product purified from a nonrecombinant source.

[0037] For the purposes of the present invention, high stringency hybridization conditions are understood to mean temperature and ionic strength conditions chosen such that they make it possible to maintain the specific and selective hybridization between complementary polynucleotides.

[0038] By way of illustration, high stringency conditions for the purposes of defining the above polynucleotides are

advantageously the following: the DNA-DNA or DNA-RNA hybridization is performed in two steps: (1) prehybridization at 42° C. for 3 hours in phosphate buffer (20 mM, pH 7.5) containing 5×SSC (1×SSC corresponds to a 0.15 M NaCl+0.015 M sodium citrate solution), 50% formamide, 7% sodium dodecyl sulfate (SDS), 10× Denhardt's, 5% dextran sulfate and 1% salmon sperm DNA; (2) hybridization for 20 hours at 42° C. followed by 2 washings of 20 minutes at 20° C. in 2×SSC+2% SDS, 1 washing of 20 minutes at 20° C. in 0.1×SSC+0.1% SDS. The final washing is performed in 0.1×SSC+0.1% SDS for 30 minutes at 60° C.

[0039] The subject of the present invention is also a representative fragment of the polynucleotide as defined above, characterized in that it is capable of being obtained either by the use of restriction enzymes whose recognition and cleavage sites are present in said polynucleotide as defined above, or by amplification with the aid of oligonucleotide primers specific for said polynucleotide as defined above, or by transcription in vitro, or by chemical synthesis.

[0040] According to an advantageous embodiment of said fragment, it is selected from the group consisting of: the cDNA corresponding to at least one open reading frame (ORF) chosen from: ORF1a, ORF1b, ORF-S, ORF-E, ORF-M, ORF-N, ORF3, ORF4, ORF7 to ORF11, ORF13 and ORF14 and the cDNA corresponding to the noncoding 5' or 3' ends of said polynucleotide.

[0041] According to an advantageous feature of this embodiment, said fragment has a sequence selected from the group consisting of:

[0042] the sequences SEQ ID NO: 2 and 4 representing the cDNA corresponding to the ORF-S which encodes the S protein,

[0043] the sequences SEQ ID NO: 13 and 15 representing the cDNA corresponding to the ORF-E which encodes the E protein,

[0044] the sequences SEQ ID NO: 16 and 18 representing the cDNA corresponding to the ORF-M which encodes the M protein,

[0045] the sequences SEQ ID NO: 36 and 38 representing the cDNA corresponding to the ORF-N which encodes the N protein,

[0046] the sequences representing the cDNA corresponding respectively: to ORF1a and ORF1b (SEQ ID NO: 31), to ORF3 and ORF4 (SEQ ID NO: 7, 8), to ORF7 to 11 (SEQ ID NO: 19, 20) to ORF13 (SEQ ID NO: 32) and to ORF14 (SEQ ID NO: 34), and

[0047] the sequences representing the cDNAs corresponding respectively to the noncoding 5' (SEQ ID NO: 39 and 72) and 3' (SEQ ID NO: 40, 73) ends of said polynucleotide.

[0048] The subject of the present invention is also a cDNA fragment encoding the S protein, as defined above, characterized in that it has a sequence selected from the group consisting of the sequences SEQ ID NO: 5 and 6 (Sa and Sb fragments).

[0049] The subject of the present invention is also a cDNA fragment corresponding to ORF1a and ORF1b as defined

above, characterized in that it has a sequence selected from the group consisting of the sequences SEQ ID NO: 41 to 54 (L0 to L12 fragments).

[0050] The subject of the present invention is also a polynucleotide fragment as defined above, characterized in that it has at least 15 consecutive bases or base pairs of the sequence of the genome of said strain including at least one of those situated in position 7979, 16622, 19064, 23220, 24872, 25298 and 26857. Preferably this is a fragment of 20 to 2500 bases or base pairs, preferably from 20 to 400.

[0051] According to an advantageous embodiment of said fragment, it includes at least one pair of bases or base pairs corresponding to the following positions: 7919 and 23220, 7919 and 25298, 16622 and 23220, 19064 and 23220, 16622 and 25298, 19064 and 25298, 23220 and 24872, 23220 and 26857, 24872 and 25298, 25298 and 26857.

[0052] The subject of the present invention is also primers of at least 18 bases capable of amplifying a fragment of the genome of a SARS-associated coronavirus or of the DNA equivalent thereof.

[0053] According to an embodiment of said primers, they are selected from the group consisting of:

[0054] the pair of primers No. 1 corresponding respectively to positions 28507 to 28522 (sense primer, SEQ ID NO: 60) and 28774 to 28759 (antisense primer, SEQ ID NO: 61) of the sequence of the polynucleotide as defined above,

[0055] the pair of primers No. 2 corresponding respectively to positions 28375 to 28390 (sense primer, SEQ ID NO: 62) and 28702 to 28687 (antisense primer, SEQ ID NO: 63) of the sequence of the polynucleotide as defined above, and

[0056] the pair of primers consisting of the primers SEQ ID Nos: 55 and 56.

[0057] The subject of the present invention is also a probe capable of detecting the presence of the genome of a SARS-associated coronavirus or of a fragment thereof, characterized in that it is selected from the group consisting of: the fragments as defined above and the fragments corresponding to the following positions of the polynucleotide sequence as defined above: 28561 to 28586, 28588 to 28608, 28541 to 28563 and 28565 to 28589 (SEQ ID NO: 64 to 67).

[0058] The probes and primers according to the invention may be labeled directly or indirectly with a radioactive or nonradioactive compound by methods well known to persons skilled in the art so as to obtain a detectable and/or quantifiable signal. Among the radioactive isotopes used, there may be mentioned ³²P, ³³P, ³⁵S, ³H or ¹²⁵I. The nonradioactive entities are selected from ligands such as biotin, avidin, streptavidin, digoxigenin, haptens, dyes, luminescent agents such as radioluminescent, chemoluminescent, bioluminescent, fluorescent and phosphorescent agents.

[0059] The invention encompasses the labeled probes and primers derived from the preceding sequences.

[0060] Such probes and primers are useful for the diagnosis of infection by a SARS-associated coronavirus.

[0061] The subject of the present invention is also a method for the detection of a SARS-associated coronavirus, from a biological sample, which method is characterized in that it comprises at least:

[0062] (a) the extraction of nucleic acids present in said biological sample,

[0063] (b) the amplification of a fragment of ORF-N by RT-PCR with the aid of a pair of primers as defined above, and

[0064] (c) the detection, by any appropriate means, of the amplification products obtained in (b).

[0065] The amplification products (amplicons) in (b) are 268 bp for the pair of primers No. 1 and 328 bp for the pair of primers No. 2.

[0066] According to an advantageous embodiment of said method, the step (b) of detection is carried out with the aid of at least one probe corresponding to positions 28561 to 28586, 28588 to 28608, 28541 to 28563 and 28565 to 28589 of the sequence of the polynucleotide as defined above.

[0067] Preferably, the SARS-associated coronavirus genome is detected and optionally quantified by PCR in real time with the aid of the pair of primers No. 2 and probes corresponding to positions 28541 to 28563 and 28565 to 28589 labeled with different compounds, in particular different fluorescent agents.

[0068] The real time RT-PCR which uses this pair of primers and this probe is very sensitive since it makes it possible to detect 10^2 copies of RNA and up to 10 copies of RNA; it is in addition reliable and reproducible.

[0069] The invention encompasses the single-stranded, double-stranded and triple-stranded polydeoxyribonucleotides and polyribonucleotides corresponding to the sequence of the genome of the isolated strain of coronavirus and its fragments as defined above, and to their sense or antisense complementary sequences, in particular the RNAs and cDNAs corresponding to the sequence of the genome and of its fragments as defined above.

[0070] The present invention also encompasses the amplification fragments obtained with the aid of primers specific for the genome of the purified or isolated strain as defined above, in particular with the aid of primers or pairs of primers as defined above, the restriction fragments formed by or comprising the sequence of fragments as defined above, the fragments obtained by transcription in vitro from a vector containing the sequence SEQ ID NO: 1 or a fragment as defined above, and fragments obtained by chemical synthesis. Examples of restriction fragments are deduced from the restriction map of the sequence SEQ ID NO: 1 illustrated by FIG. 13. In accordance with the invention, said fragments are either in the form of isolated fragments, or in the form of mixtures of fragments. The invention also encompasses fragments modified, in relation to the preceding ones, by removal or addition of nucleotides in a proportion of about 15%, relative to the length of the above fragments and/or modified in terms of the nature of the nucleotides, as long as the modified nucleotide fragments retain a capacity for hybridization with the genomic or antigenomic RNA sequences of the isolate as defined above.

[0071] The nucleic acid molecules according to the invention are obtained by conventional methods, known per se, following standard protocols such as those described in *Current Protocols in Molecular Biology* (Frederick M. AUSUBEL, 2000, Wiley and son Inc., Library of Congress, USA). For example, they may be obtained by amplification of a nucleic sequence by PCR or RT-PCR or alternatively by total or partial chemical synthesis.

[0072] The subject of the present invention is also a DNA or RNA chip or filter, characterized in that it comprises at least one polynucleotide or one of its fragments as defined above.

[0073] The DNA or RNA chips or filters according to the invention are prepared by conventional methods, known per se, such as for example chemical or electrochemical grafting of oligonucleotides on a glass or nylon support.

[0074] The subject of the present invention is also a recombinant cloning and/or expression vector, in particular a plasmid, a virus, a viral vector or a phage comprising a nucleic acid fragment as defined above. Preferably, said recombinant vector is an expression vector in which said nucleic acid fragment is placed under the control of appropriate elements for regulating transcription and translation. In addition, said vector may comprise sequences (tags) fused in phase with the 5' and/or 3' end of said insert, which are useful for the immobilization and/or detection and/or purification of the protein expressed from said vector.

[0075] These vectors are constructed and introduced into host cells by conventional recombinant DNA and genetic engineering methods which are known per se. Numerous vectors into which a nucleic acid molecule of interest may be inserted in order to introduce it and to maintain it in a host cell are known per se; the choice of an appropriate vector depends on the use envisaged for this vector (for example replication of the sequence of interest, expression of this sequence, maintenance of the sequence in extrachromosomal form or alternatively integration into the chromosomal material of the host), and on the nature of the host cell.

[0076] In accordance with the invention, said plasmid is selected in particular from the following plasmids:

[0077] the plasmid, called SARS-S, contained in the bacterial strain deposited under the No. I-3659, on Jun. 20, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA sequence encoding the S protein of the SARS-CoV strain derived from the sample recorded under the No. 031589, said sequence corresponding to the nucleotides at positions 21406 to 25348 (SEQ ID NO: 4), with reference to the Genbank sequence AY274119.3,

[0078] the plasmid, called SARS-S1, contained in the bacterial strain deposited under the No. I-3020, on May 12, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains a 5' fragment of the cDNA sequence encoding the S protein of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above, said fragment corresponding to the nucleotides at positions 21406 to 23454 (SEQ ID NO: 5), with reference to the Genbank sequence AY274119.3 Tor2,

- [0079] the plasmid, called SARS-S2, contained in the bacterial strain deposited under the No. I-3019, on May 12, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains a 3' fragment of the cDNA sequence encoding the S protein of the SARS-CoV strain derived from the sample recorded under the number No. 031589, as defined above, said fragment corresponding to the nucleotides at positions 23322 to 25348 (SEQ ID NO: 6), with reference to the Genbank sequence accession No. AY274119.3,
- [0080] the plasmid, called SARS-SE, contained in the bacterial strain deposited under the No. I-3126, on Nov. 13, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA corresponding to the region situated between ORF-S and ORF-E and overlapping ORF-E of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above, said region corresponding to the nucleotides at positions 25110 to 26244 (SEQ ID NO: 8), with reference to the Genbank sequence accession No. AY274119.3,
- [0081] the plasmid, called SARS-E, contained in the bacterial strain deposited under the No. I-3046, on May 28, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA sequence encoding the E protein of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above, said sequence corresponding to the nucleotides at positions 26082 to 26413 (SEQ ID NO: 15), with reference to the Genbank sequence accession No. AY274119.3,
- [0082] the plasmid, called SARS-M, contained in the bacterial strain deposited under the No. I-3047, on May 28, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA sequence encoding the M protein of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above; said sequence corresponding to the nucleotides at positions 26330 to 27098 (SEQ ID NO: 18), with reference to the Genbank sequence accession No. AY274119.3,
- [0083] the plasmid, called SARS-MN, contained in the bacterial sequence deposited under the No. I-3125, on Nov. 13, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA sequence corresponding to the region situated between ORF-M and ORF-N of the SARS-CoV strain derived from the sample recorded under the No. 031589 and collected in Hanoi, as defined above, said sequence corresponding to the nucleotides at positions 26977 to 28218 (SEQ ID NO: 20), with reference to the Genbank accession No. AY274119.3,
- [0084] the plasmid, called SARS-N, contained in the bacterial strain deposited under the No. I-3048, on Jun. 5, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA encoding the N protein of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above, said sequence corresponding to the nucleotides at positions 28054 to 29430 (SEQ ID NO: 38), with reference to the Genbank sequence accession No. AY274119.3; thus, this plasmid comprises an insert of sequence SEQ ID NO: 38 and is contained in a bacterial strain which was deposited under the No. I-3048, on Jun. 5, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15,
- [0085] the plasmid, called SARS-5'NC, contained in the bacterial strain deposited under the No. I-3124, on Nov. 7, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA corresponding to the noncoding 5' end of the genome of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above, said sequence corresponding to the nucleotides at positions 1 to 204 (SEQ ID NO: 39), with reference to the Genbank sequence accession No. AY274119.3,
- [0086] the plasmid called SARS-3'NC, contained in the bacterial strain deposited under the No. I-3123 on Nov. 7, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA sequence corresponding to the noncoding 3' end of the genome of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above, said sequence corresponding to that situated between the nucleotide and position 28933 to 29727 (SEQ ID NO: 40), with reference to the Genbank sequence accession No. AY274119.3, ends with a series of nucleotides a.,
- [0087] the expression plasmid, called pIV2.3N, containing a cDNA fragment encoding a C-terminal fusion of the N protein (SEQ ID NO: 37) with a polyhistidine tag,
- [0088] the expression plasmid, called pIV2.3S_C, containing a cDNA fragment encoding a C-terminal fusion of the fragment corresponding to positions 475 to 1193 of the amino acid sequence of the S protein (SEQ ID NO: 3) with a polyhistidine tag,
- [0089] the expression plasmid, pIV2.3S_L, containing a cDNA fragment encoding a C-terminal fusion of the fragment corresponding to positions 14 to 1193 of the amino acid sequence of the S protein (SEQ ID NO: 3) with a polyhistidine tag,
- [0090] the expression plasmid, called pIV2.4N, containing a cDNA fragment encoding a N-terminal fusion of the N protein (SEQ ID NO: 3) with a polyhistidine tag,
- [0091] the expression plasmid, called pIV2.4S_C or pIV2.4S_L, containing an insert encoding a N-terminal fusion of the fragment corresponding to positions 475 to 1193 of the amino acid sequence of the S protein (SEQ ID NO: 3) with a polyhistidine tag, and
- [0092] the expression plasmid, called pIV2.4S_L, containing a cDNA fragment encoding an N-terminal fusion of the fragment corresponding to positions 14 to

1193 of the amino acid sequence of the S protein (SEQ ID NO: 3) with a polyhistidine tag.

[0093] According to an advantageous feature of the expression plasmid as defined above, it is contained in a bacterial strain which was deposited under the No. I-3117, on Oct. 23, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15.

[0094] According to another advantageous feature of the expression plasmid as defined above, it is contained in a bacterial strain which was deposited under the No. I-3118, on Oct. 23, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15.

[0095] According to another feature of the expression plasmid as defined above, it is contained in a bacterial strain which was deposited at the CNCM, 25 rue du Docteur Roux, 75724 Paris Cedex 15 under the following numbers:

- [0096] a) strain No. I-3118, deposited on Oct. 23, 2003,
- [0097] b) strain No. I-3019, deposited on May 12, 2003,
- [0098] c) strain No. I-3020, deposited on May 12, 2003,
- [0099] d) strain No. I-3059, deposited on Jun. 20, 2003,
- [0100] e) strain No. I-3323, deposited on Nov. 22, 2004,
- [0101] f) strain No. I-3324, deposited on Nov. 22, 2004,
- [0102] g) strain No. I-3326, deposited on Dec. 1, 2004,
- [0103] h) strain No. I-3327, deposited on Dec. 1, 2004,
- [0104] i) strain No. I-3332, deposited on Dec. 1, 2004,
- [0105] j) strain No. I-3333, deposited on Dec. 1, 2004,
- [0106] k) strain No. I-3334, deposited on Dec. 1, 2004,
- [0107] l) strain No. I-3335, deposited on Dec. 1, 2004,
- [0108] m) strain No. I-3336, deposited on Dec. 1, 2004,
- [0109] n) strain No. I-3337, deposited on Dec. 1, 2004,
- [0110] o) strain No. I-3338, deposited on Dec. 2, 2004,
- [0111] p) strain No. I-3339, deposited on Dec. 2, 2004,
- [0112] q) strain No. I-3340, deposited on Dec. 2, 2004,
- [0113] r) strain No. I-3341, deposited on Dec. 2, 2004.

[0114] The subject of the present invention is also a nucleic acid insert of viral origin, characterized in that it is contained in any of the strains as defined above in a)-r).

[0115] The subject of the present invention is also a nucleic acid containing a synthetic gene allowing optimized expression of the S protein in eukaryotic cells, characterized in that it possesses the sequence SEQ ID NO: 140.

[0116] The subject of the present invention is also an expression vector containing a nucleic acid containing a synthetic gene allowing optimized expression of the S protein, which vector is contained in the bacterial strain deposited at the CNCM, on Dec. 1, 2004, under the No. I-3333.

[0117] According to one embodiment of said expression vector, it is a viral vector, in the form of a viral particle or in the form of a recombinant genome.

[0118] According to an advantageous feature of this embodiment, this is a recombinant viral particle or a recombinant viral genome capable of being obtained by transfection of a plasmid according to paragraphs g), h) and k) to r) as defined above, in an appropriate cellular system, that is to say, for example, cells transfected with one or more other plasmids intended to transcomplement certain functions of the virus that are deleted in the vector and that are necessary for the formation of the viral particles.

[0119] The expression "S protein family" is understood here to mean the complete S protein, its ectodomain and fragments of this ectodomain which are preferably produced in a eukaryotic system.

[0120] The subject of the present invention is also a lentiviral vector encoding a polypeptide of the S protein family, as defined above.

[0121] The subject of the present invention is also a recombinant measles virus encoding a polypeptide of the S protein family, as defined above.

[0122] The subject of the present invention is also a recombinant vaccinia virus encoding a polypeptide of the S protein family, as defined above.

[0123] The subject of the present invention is also the use of a vector according to paragraphs e) to r) as defined above, or of a vector containing a synthetic gene for the S protein, as defined above, for the production, in a eukaryotic system, of the SARS-associated coronavirus S protein or of a fragment of this protein.

[0124] The subject of the present invention is also a method for producing the S protein in a eukaryotic system, comprising a step of transfecting eukaryotic cells in culture with a vector chosen from the vectors contained in the bacterial strains mentioned in paragraphs e) to r) above or a vector containing a synthetic gene allowing optimized expression of the S protein.

[0125] The subject of the present invention is also a cDNA library characterized in that it comprises fragments as defined above, in particular amplification fragments or restriction fragments, cloned into a recombinant vector, in particular an expression vector (expression library).

[0126] The subject of the present invention is also cells, in particular prokaryotic cells, modified by a recombinant vector as defined above.

[0127] The subject of the present invention is also a genetically modified eukaryotic cell expressing a protein or a polypeptide as defined above. Quite obviously, the terms "genetically modified eukaryotic cell" do not denote a cell modified with a wild-type virus.

[0128] According to an advantageous embodiment of said cell, it is capable of being obtained by transfection with any of the vectors mentioned in paragraphs i) to l) above.

[0129] According to an advantageous feature of this embodiment, this is the cell FRhK4-Sso1-30, deposited at the CNCM on Nov. 22, 2004, under the No. I-3325.

[0130] The recombinant vectors as defined above and the cells transformed with said expression vectors are advantageously used for the production of the corresponding proteins and peptides. The expression libraries derived from

said vectors, and the cells transformed with said expression libraries are advantageously used to identify the immunogenic epitopes (B and T epitopes) of the SARS-associated coronavirus proteins.

[0131] The subject of the present invention is also the purified or isolated proteins and peptides, characterized in that they are encoded by the polynucleotide or one of its fragments as defined above.

[0132] According to an advantageous embodiment of the invention, said protein is selected from the group consisting of:

[0133] the S protein having the sequence SEQ ID NO: 3 or its ectodomaine

[0134] the E protein having the sequence SEQ ID NO: 14

[0135] the M protein having the sequence SEQ ID NO: 17

[0136] the N protein having the sequence SEQ ID NO: 37

[0137] the proteins encoded by the ORFs: ORF1a, ORF1b, ORF3, ORF4 and ORF7 to ORF11, ORF13 and ORF14 and having the respective sequence, SEQ ID NO: 74, 75, 10, 12, 22, 24, 26, 28, 30, 33 and 35.

[0138] The terms "ectodomaine of the S protein" and "soluble form of the S protein" will be used interchangeably below.

[0139] According to an advantageous embodiment of the invention, said polypeptide consists of the amino acids corresponding to positions 1 to 1193 of the amino acid sequence of the S protein.

[0140] According to another advantageous embodiment of the invention, said peptide is selected from the group consisting of:

[0141] a) the peptides corresponding to positions 14 to 1193 and 475 to 1193 of the amino acid sequence of the S protein,

[0142] b) the peptides corresponding to positions 2 to 14 (SEQ ID NO: 69) and 100 to 221 of the amino acid sequence of the M protein; these peptides correspond respectively to the ectodomaine and to the endodomaine of the M protein, and

[0143] c) the peptides corresponding to positions 1 to 12 (SEQ ID NO: 70) and 53 to 76 (SEQ ID NO: 71) of the amino acid sequence of the E protein; these peptides correspond respectively to the ectodomaine and to the C-terminal end of the E protein, and

[0144] d) the peptides of 5 to 50 consecutive amino acids, preferably of 10 to 30 amino acids, inclusive or partially or completely overlapping the sequence of the peptides as defined in a), b) or c).

[0145] The subject of the present invention is also a peptide, characterized in that it has a sequence of 7 to 50 amino acids including an amino acid residue selected from the group consisting of:

[0146] the alanine situated at position 2552 of the amino acid sequence of the protein encoded by ORF1a,

[0147] the serine situated at position 577 of the amino acid sequence of the S protein of the SARS-CoV strain as defined above,

[0148] the glycine at position 11 of the amino acid sequence of the protein encoded by ORF3 of the SARS-CoV strain as defined above,

[0149] the serine at position 154 of the amino acid sequence of the M protein of the SARS-CoV strain as defined above.

[0150] The subject of the present invention is also an antibody or a polyclonal or monoclonal antibody fragment which can be obtained by immunization of an animal with a recombinant vector as defined above, a cDNA library as defined above or alternatively a protein or a peptide as defined above, characterized in that it binds to at least one of the proteins encoded by SARS-CoV as defined above.

[0151] The invention encompasses the polyclonal antibodies, the monoclonal antibodies, the chimeric antibodies such as the humanized antibodies, and fragments thereof (Fab, Fv, scFv).

[0152] A subject of the present invention is also a hybridoma producing a monoclonal antibody against the N protein, characterized in that it is chosen from the following hybridomas:

[0153] the hybridoma producing the monoclonal antibody 87, deposited at the CNCM on Dec. 1, 2004 under the number I-3328,

[0154] the hybridoma producing the monoclonal antibody 86, deposited at the CNCM on Dec. 1, 2004 under the number I-3329,

[0155] the hybridoma producing the monoclonal antibody 57, deposited at the CNCM on Dec. 1, 2004 under the number I-3330, and

[0156] the hybridoma producing the monoclonal antibody 156, deposited at the CNCM on Dec. 1, 2004 under the number I-3331.

[0157] The subject of the present invention is also a polyclonal or monoclonal antibody or antibody fragment directed against the N protein, characterized in that it is produced by a hybridoma as defined above.

[0158] For the purposes of the present invention, the expression chimeric antibody is understood to mean, in relation to an antibody of a particular animal species or of a particular class of antibody, an antibody comprising all or part of a heavy chain and/or of a light chain of an antibody of another animal species or of another class of antibody.

[0159] For the purposes of the present invention, the expression humanized antibody is understood to mean a human immunoglobulin in which the residues of the CDRs (Complementary Determining Regions) which form the antigen-binding site are replaced by those of a nonhuman monoclonal antibody possessing the desired specificity, affinity or activity. Compared with the nonhuman antibodies, the humanized antibodies are less immunogenic and possess a prolonged half-life in humans because they possess only a small proportion of nonhuman sequences given that practically all the residues of the FR (Framework) regions and of

the constant (Fc) region of these antibodies are those of a consensus sequence of human immunoglobulins.

[0160] A subject of the present invention is also a protein chip or filter, characterized in that it comprises a protein, a peptide or alternatively an antibody as defined above.

[0161] The protein chips according to the invention are prepared by conventional methods known per se. Among the appropriate supports on which proteins may be immobilized, there may be mentioned those made of plastic or glass, in particular in the form of microplates.

[0162] The subject of the present invention is also reagents derived from the isolated strain of SARS-associated coronavirus, derived from the sample recorded under the No. 031589, which are useful for the study and diagnosis of the infection caused by a SARS-associated coronavirus, said reagents are selected from the group consisting of:

[0163] (a) a pair of primers, a probe or a DNA chip as defined above,

[0164] (b) a recombinant vector or a modified cell as defined above,

[0165] (c) an isolated coronavirus strain or a polynucleotide as defined above,

[0166] (d) a protein or a peptide as defined above,

[0167] (e) an antibody or an antibody fragment as defined above, and

[0168] (f) a protein chip as defined above.

[0169] These various reagents are prepared and used according to conventional molecular biology and immunology techniques following standard protocols such as those described in *Current Protocols in Molecular Biology* (Frederick M. AUSUBEL, 2000, Wiley and Son Inc., Library of Congress, USA), in *Current Protocols in Immunology* (John E. Cologan, 2000, Wiley and Son Inc., Library of Congress, USA) and in *Antibodies: A Laboratory Manual* (E. Howell and D. Lane, Cold Spring Harbor Laboratory, 1988).

[0170] The nucleic acid fragments according to the invention are prepared and used according to conventional techniques as defined above. The peptides and proteins according to the invention are prepared by recombinant DNA techniques, known to persons skilled in the art, in particular with the aid of the recombinant vectors as defined above. Alternatively, the peptides according to the invention may be prepared by conventional techniques of solid or liquid phase synthesis, known to persons skilled in the art.

[0171] The polyclonal antibodies are prepared by immunizing an appropriate animal with a protein or a peptide as defined above, optionally coupled to KLH or to albumin and/or combined with an appropriate adjuvant such as (complete or incomplete) Freund's adjuvant or aluminum hydroxide; after obtaining a satisfactory antibody titer, the antibodies are harvested by collecting serum from the immunized animals and enriched with IgG by precipitation, according to conventional techniques, and then the IgGs specific for the SARS-CoV proteins are optionally purified by affinity chromatography on an appropriate column to which said peptide or said protein is attached, as defined above, so as to obtain a monospecific IgG preparation.

[0172] The monoclonal antibodies are produced from hybridomas obtained by fusion of B lymphocytes from an animal immunized with a protein or a peptide as defined above with myelomas, according to the Köhler and Milstein technique (Nature, 1975, 256, 495-497); the hybridomas are cultured in vitro, in particular in fermenters or produced in vivo, in the form of ascites; alternatively, said monoclonal antibodies are produced by genetic engineering as described in American U.S. Pat. No. 4,816,567.

[0173] The humanized antibodies are produced by general methods such as those described in International application WO 98/45332.

[0174] The antibody fragments are produced from the cloned V_H and V_L regions, from the mRNAs of hybridomas or splenic lymphocytes of an immunized mouse; for example, the Fv, scFv or Fab fragments are expressed at the surface of filamentous phages according to the Winter and Milstein technique (Nature, 1991, 349, 293-299); after several selection steps, the antibody fragments specific for the antigen are isolated and expressed in an appropriate expression system, by conventional techniques for cloning and expression of recombinant DNA.

[0175] The antibodies or fragments thereof as defined above are purified by conventional techniques known to persons skilled in the art, such as affinity chromatography.

[0176] The subject of the present invention is additionally the use of a product selected from the group consisting of: a pair of primers, a probe, a DNA chip, a recombinant vector, a modified cell, an isolated coronavirus strain, a polynucleotide, a protein or a peptide, an antibody or an antibody fragment and a protein chip as defined above, for the preparation of a reagent for the detection and optionally genotyping/serotyping of a SARS-associated coronavirus.

[0177] The proteins and peptides according to the invention, which are capable of being recognized and/or of inducing the production of antibodies specific for the SARS-associated coronavirus, are useful for the diagnosis of infection with such a coronavirus; the infection is detected, by an appropriate technique—in particular EIA, ELISA, RIA, immunofluorescence—in a biological sample collected from an individual capable of being infected.

[0178] According to an advantageous feature of said use, said proteins are selected from the group consisting of the S, E, M and/or N proteins and the peptides as defined above.

[0179] The S, E, M and/or N proteins and the peptides derived from these proteins as defined above, for example the N protein, are used for the indirect diagnosis of a SARS-associated coronavirus infection (serological diagnosis; detection of an antibody specific for SARS-CoV), in particular by an immunoenzymatic method (ELISA).

[0180] The antibodies and antibody fragments according to the invention, in particular those directed against the S, E, M and/or N proteins and the derived peptides as defined above, are useful for the direct diagnosis of a SARS-associated coronavirus infection; the detection of the protein(s) of SARS-CoV is carried out by an appropriate technique, in particular EIA, ELISA, RIA, immunofluorescence, in a biological sample collected from an individual capable of being infected.

[0181] The subject of the present invention is also a method for the detection of a SARS-associated coronavirus, from a biological sample, which method is characterized in that it comprises at least:

[0182] (a) bringing said biological sample into contact with at least one antibody or one antibody fragment, one protein, one peptide or alternatively one protein or peptide chip or filter as defined above, and

[0183] (b) visualizing by any appropriate means antigen-antibody complexes formed in (a), for example by EIA, ELISA, RIA, or by immunofluorescence.

[0184] According to one advantageous embodiment of said process, step (a) comprises:

[0185] (a₁) bringing said biological sample into contact with at least a first antibody or an antibody fragment which is attached to an appropriate support, in particular a microplate,

[0186] (a₂) washing the solid phase, and

[0187] (a₃) adding at least a second antibody or an antibody fragment, different from the first, said antibody or antibody fragment being optionally appropriately labeled.

[0188] This method, which makes it possible to capture the viral particles present in the biological sample, is also called immunocapture method.

[0189] For example:

[0190] step (a₁) is carried out with at least a first monoclonal or polyclonal antibody or a fragment thereof, directed against the S, M and/or E protein, and/or a peptide corresponding to the ectodomain of one of these proteins (M2-14 or E1-12 peptides)

[0191] step (a₃) is carried out with at least one antibody or an antibody fragment directed against another epitope of the same protein or preferably against another protein, preferably against an inner protein such as the N nucleoprotein or the endodomain of the E or M protein, more preferably still these are antibodies or antibody fragments directed against the N protein which is very abundant in the viral particle; when an antibody or an antibody fragment directed against an inner protein (N) or against the endodomain of the E or M proteins is used, said antibody is incubated in the presence of detergent, such as Tween 20 for example, at concentrations of the order of 0.1%.

[0192] step (b) for visualizing the antigen-antibody complexes formed is carried out, either directly with the aid of a second antibody labeled for example with biotin or an appropriate enzyme such as peroxidase or alkaline phosphatase, or indirectly with the aid of an anti-immunoglobulin serum labeled as above. The complexes thus formed are visualized with the aid of an appropriate substrate.

[0193] According to a preferred embodiment of this aspect of the invention, the biological sample is mixed with the visualizing monoclonal antibody prior to its being brought into contact with the capture monoclonal antibodies. Where appropriate, the serum-visualizing antibody mixture is incubated for at least 10 minutes at room temperature before being applied to the plate.

[0194] The subject of the present invention is also an immunocapture test intended to detect an infection by the SARS-associated coronavirus by detecting the native nucleoprotein (N protein), in particular characterized in that the antibody used for the capture of the native viral nucleoprotein is a monoclonal antibody specific for the central region and/or for a conformational epitope.

[0195] According to one embodiment of said test, the antibody used for the capture of the N protein is the monoclonal antibody mAb87, produced by the hybridoma deposited at the CNCM on Dec. 1, 2004 under the number I-3328.

[0196] According to another embodiment of said immunocapture test, the antibody used for the capture of the N protein is the monoclonal antibody mAb86, produced by the hybridoma deposited at the CNCM on Dec. 1, 2004 under the number I-3329.

[0197] According to another embodiment of said immunocapture test, the monoclonal antibodies mAb86 and mAb87 are used for the capture of the N protein.

[0198] In the immunocapture tests according to the invention, it is possible to use, for visualizing the N protein, the monoclonal antibody mAb57, produced by the hybridoma deposited at the CNCM on Dec. 1, 2004 under the number I-3330, said antibody being conjugated with a visualizing molecule or particle.

[0199] In accordance with said immunocapture test, a combination of the antibodies mAb57 and mAb87, conjugated with a visualizing molecule or particle, is used for the visualization of the N protein.

[0200] A visualizing molecule may be a radioactive atom, a dye, a fluorescent molecule, a fluorophore, an enzyme; a visualizing particle may be for example: colloidal gold, a magnetic particle or a latex bead.

[0201] The subject of the present invention is also a reagent for detecting a SARS-associated coronavirus, characterized in that it is selected from the group consisting of:

[0202] (a) a pair of primers or a probe as defined above,

[0203] (b) a recombinant vector as defined above or a modified cell as defined above,

[0204] (c) an isolated coronavirus strain as defined above or a polynucleotide as defined above,

[0205] (d) an antibody or an antibody fragment as defined above,

[0206] (e) a combination of antibodies comprising the monoclonal antibodies mAb86 and/or mAb87, and the monoclonal antibody mAb57, as defined above,

[0207] (f) a chip or a filter as defined above.

[0208] The subject of the present invention is also a method for the detection of a SARS-associated coronavirus infection, from a biological sample, by indirect IgG ELISA using the N protein, which method is characterized in that the plates are sensitized with an N protein solution at a concentration of between 0.5 and 4 µg/ml, preferably to 2 µg/ml, in a 10 mM PBS buffer pH 7.2, phenol red at 0.25 ml/l.

[0209] The subject of the present invention is additionally a method for the detection of a SARS-associated coronavirus infection, from a biological sample, by double epitope ELSA, characterized in that the serum to be tested is mixed with the visualizing antigen, said mixture then being brought into contact with the antigen attached to a solid support.

[0210] According to one variant of the tests for detecting SARS-associated coronaviruses, these tests combine an ELSA using the N protein, and another ELSA using the S protein, as described below.

[0211] The subject of the present invention is also an immune complex formed of a polyclonal or monoclonal antibody or antibody fragment as defined above, and of a SARS-associated coronavirus protein or peptide.

[0212] The subject of the present invention is additionally a SARS-associated coronavirus detection kit, characterized in that it comprises at least one reagent selected from the group consisting of: a pair of primers, a probe, a DNA or RNA chip, a recombinant vector, a modified cell, an isolated coronavirus strain, a polynucleotide, a protein or a peptide, an antibody, and a protein chip as defined above.

[0213] The subject of the present invention is additionally an immunogenic composition, characterized in that it comprises at least one product selected from the group consisting of:

- [0214] a) a protein or a peptide as defined above,
- [0215] b) a polynucleotide of the DNA or RNA type or one of its representative fragments as defined above, having a sequence chosen from:
 - [0216] (i) the sequence SEQ ID NO: 1 or its RNA equivalent
 - [0217] (ii) the sequence hybridizing under high stringency conditions with the sequence SEQ ID NO: 1,
 - [0218] (iii) the sequence complementary to the sequence SEQ ID NO: 1 or to the sequence hybridizing under high stringency conditions with the sequence SEQ ID NO: 1,
 - [0219] (iv) the nucleotide sequence of a representative fragment of the polynucleotide as defined in (i), (ii) or (iii),
 - [0220] (v) the sequence as defined in (i), (ii), (iii) or (iv), modified, and
- [0221] c) a recombinant expression vector comprising a polynucleotide as defined in b), and
- [0222] d) a cDNA library as defined above,

said immunogenic composition being capable of inducing protective humoral or cellular immunity specific for the SARS-associated coronavirus, in particular the production of an antibody directed against a specific epitope of the SARS-associated coronavirus.

[0223] The proteins and peptides as defined above, in particular the S, M, E and/or N proteins and the derived peptides, and the nucleic acid (DNA or RNA) molecules encoding said proteins or said peptides are good candidate vaccines and may be used in immunogenic compositions for the production of a vaccine against the SARS-associated coronavirus.

[0224] According to an advantageous embodiment of the compositions according to the invention, they additionally contain at least one pharmaceutically acceptable vehicle and optionally carrier substances and/or adjuvants.

[0225] The pharmaceutically acceptable vehicles, the carrier substances and the adjuvants are those conventionally used.

[0226] The adjuvants are advantageously chosen from the group consisting of oily emulsions, saponin, mineral substances, bacterial extracts, aluminum hydroxide and squalene.

[0227] The carrier substances are advantageously selected from the group consisting of unilamellar liposomes, multilamellar liposomes, micelles of saponin or solid microspheres of a saccharide or auriferous nature.

[0228] The compositions according to the invention are administered by the general route, in particular by the intramuscular or subcutaneous route or alternatively by the local, in particular nasal (aerosol) route.

[0229] The subject of the present invention is also the use of an isolated or purified protein or peptide having a sequence selected from the group consisting of the sequences SEQ ID NO: 3, 10, 12, 14, 17, 22, 24, 26, 28, 30, 33, 35, 37, 69, 70, 71, 74 and 75 to form an immune complex with an antibody specifically directed against an epitope of the SARS-associated coronavirus.

[0230] The subject of the present invention is also an immune complex consisting of an isolated or purified protein or peptide having a sequence selected from the group consisting of the sequences SEQ ID NO: 3, 10, 12, 14, 17, 22, 24, 26, 28, 30, 33, 35, 37, 69, 70, 71, 74 and 75, and of an antibody specifically directed against an epitope of the SARS-associated coronavirus.

[0231] The subject of the present invention is also the use of an isolated or purified protein or peptide having a sequence selected from the group consisting of the sequences SEQ ID NO: 3, 10, 12, 14, 17, 22, 24, 26, 28, 30, 33, 35, 37, 69, 70, 71, 74 and 75 to induce the production of an antibody capable of specifically recognizing an epitope of the SARS-associated coronavirus.

[0232] The subject of the present invention is also the use of an isolated or purified polynucleotide having a sequence selected from the group consisting of the sequences SEQ ID NO: 1, 2, 4, 7, 8, 13, 15, 16, 18, 19, 20, 31, 36 and 38 to induce the production of an antibody directed against the protein encoded by said polynucleotide and capable of specifically recognizing an epitope of the SARS-associated coronavirus.

[0233] The subject of the present invention is also monoclonal antibodies recognizing the native S protein of a SARS-associated coronavirus.

[0234] The subject of the present invention is also the use of a protein or a polypeptide of the S protein family, as defined above, or of an antibody recognizing the native S protein, as defined above, to detect an infection by a SARS-associated coronavirus, in a biological sample.

[0235] The subject of the present invention is also a method for detecting an infection by a SARS-associated coronavirus, in a biological sample, characterized in that the

detection is carried out by ELISA using the recombinant S protein, expressed in a eukaryotic system.

[0236] According to an advantageous embodiment of said method, it is a double epitope ELISA method, and the serum to be tested is mixed with the visualizing antigen, said mixture then being brought into contact with the antigen attached to a solid support.

[0237] The subject of the present invention is also an immune complex consisting of a monoclonal antibody or antibody fragment recognizing the native S protein, and of a protein or a peptide of the SARS-associated coronavirus.

[0238] The subject of the present invention is also an immune complex consisting of a protein or a polypeptide of the S protein family, as defined above, and of an antibody specifically directed against an epitope of the SARS-associated coronavirus.

[0239] The subject of the present invention is additionally a SARS-associated coronavirus detection kit or box, characterized in that it comprises at least one reagent selected from the group consisting of: a protein or polypeptide of the S protein family, as defined above, a nucleic acid encoding a protein or peptide of the S protein family, as defined above,

a cell expressing a protein or polypeptide of the S protein family, as defined above, or an antibody recognizing the native S protein of a SARS-associated coronavirus.

[0240] The subject of the present invention is an immunogenic and/or vaccine composition, characterized in that it comprises a polypeptide or a recombinant protein of the S protein family, as defined above, obtained in a eukaryotic expression system.

[0241] The subject of the present invention is also an immunogenic and/or vaccine composition, characterized in that it comprises a vector or recombinant virus, expressing a protein or a polypeptide of the S protein family, as defined above.

[0242] In addition to the preceding features, the invention further comprises other features, which will emerge from the description which follows, which refers to examples of use of the polynucleotide representing the genome of the SARS-CoV strain derived from the sample recorded under the number 031589, and derived cDNA fragments which are the subject of the present invention, and to Table I presenting the sequence listing:

TABLE I

Identification number	Sequence	Sequence listing	
		Position of the cDNA with reference to Genbank AY274119.3	Deposit number at the of the CNM corresponding plasmid
SEQ ID NO: 1	genome of the strain derived from the sample 031589	—	—
SEQ ID NO: 2	ORF-S*	21406–25348	—
SEQ ID NO: 3	S protein	—	—
SEQ ID NO: 4	ORF-S**	21406–25348	I-3059
SEQ ID NO: 5	Sa fragment	21406–23454	I-3020
SEQ ID NO: 6	Sb fragment	23322–25348	I-3019
SEQ ID NO: 7	ORF-3 + ORF-4*	25110–26244	—
SEQ ID NO: 8	ORF-3 + ORF-4**	25110–26244	I-3126
SEQ ID NO: 9	ORF3	—	—
SEQ ID NO: 10	ORF-3 protein	—	—
SEQ ID NO: 11	ORF4	—	—
SEQ ID NO: 12	ORF-4 protein	—	—
SEQ ID NO: 13	ORF-E*	26082–26413	—
SEQ ID NO: 14	E protein	—	—
SEQ ID NO: 15	ORF-E**	26082–26413	I-3046
SEQ ID NO: 16	ORF-M*	26330–27098	—
SEQ ID NO: 17	M protein	—	—
SEQ ID NO: 18	ORF-M**	26330–27098	I-3047
SEQ ID NO: 19	ORF7 to 11*	26977–28218	—
SEQ ID NO: 20	ORF7 to 11**	26977–28218	I-3125
SEQ ID NO: 21	ORF7	—	—
SEQ ID NO: 22	ORF7 protein	—	—
SEQ ID NO: 23	ORF8	—	—
SEQ ID NO: 24	ORF8 protein	—	—
SEQ ID NO: 25	ORF9	—	—
SEQ ID NO: 26	ORF9 protein	—	—
SEQ ID NO: 27	ORF10	—	—
SEQ ID NO: 28	ORF10 protein	—	—
SEQ ID NO: 29	ORF11	—	—
SEQ ID NO: 30	ORF11 protein	—	—
SEQ ID NO: 31	OrFlab	265–21485	—
SEQ ID NO: 32	ORF13	28130–28426	—
SEQ ID NO: 33	ORF13 protein	—	—
SEQ ID NO: 34	ORF14	—	—
SEQ ID NO: 35	ORF14 protein	28583–28795	—

TABLE I-continued

Sequence listing			
Identification number	Sequence	Position of the cDNA with reference to Genbank AY274119.3	Deposit number at the of the CNCM corresponding plasmid
SEQ ID NO: 36	ORF-N*	28054-29430	—
SEQ ID NO: 37	N protein	—	—
SEQ ID NO: 38	ORF-N**	28054-29430	I-3048
SEQ ID NO: 39	noncoding 5'**	1-204	I-3124
SEQ ID NO: 40	noncoding 3'**	28933-29727	I-3123
SEQ ID NO: 41	ORF1ab	30-500	—
	Fragment L0		
SEQ ID NO: 42	Fragment L1	211-2260	—
SEQ ID NO: 43	Fragment L2	2136-4187	—
SEQ ID NO: 44	Fragment L3	3892-5344	—
SEQ ID NO: 45	Fragment L4b	4932-6043	—
SEQ ID NO: 46	Fragment L4	5305-7318	—
SEQ ID NO: 47	Fragment L5	7275-9176	—
SEQ ID NO: 48	Fragment L6	9032-11086	—
SEQ ID NO: 49	Fragment L7	10298-12982	—
SEQ ID NO: 50	Fragment L8	12815-14854	—
SEQ ID NO: 51	Fragment L9	14745-16646	—
SEQ ID NO: 52	Fragment L10	16514-18590	—
SEQ ID NO: 53	Fragment L11	18500-20602	—
SEQ ID NO: 54	Fragment L12	20319-22224	—
SEQ ID NO: 55	Sense N primer	—	—
SEQ ID NO: 56	Antisense N primer	—	—
SEQ ID NO: 57	Sense S _C primer	—	—
SEQ ID NO: 58	Sense S _L primer	—	—
SEQ ID NO: 59	Antisense S _C and S _L primer	—	—
SEQ ID NO: 60	Sense primer series 1	28507-28522	—
SEQ ID NO: 61	Antisense primer series 1	28774-28759	—
SEQ ID NO: 62	Sense primer series 2	28375-28390	—
SEQ ID NO: 63	Antisense primer series 2	28702-28687	—
SEQ ID NO: 64	Probe 1/series 1	28561-28586	—
SEQ ID NO: 65	Probe 2/series 1	28588-28608	—
SEQ ID NO: 66	Probe 1/series 2	28541-28563	—
SEQ ID NO: 67	Probe 2/series 2	28565-28589	—
SEQ ID NO: 68	Anchor primer 14T	—	—
SEQ ID NO: 69	Peptide M2-14	—	—
SEQ ID NO: 70	Peptide E1-12	—	—
SEQ ID NO: 71	Peptide E53-76	—	—
SEQ ID NO: 72	Noncoding 5'*	1-204	—
SEQ ID NO: 73	Noncoding 3'*	28933-29727	—
SEQ ID NO: 74	ORF1a protein	—	—
SEQ ID NO: 75	ORF1b protein	—	—
SEQ ID NO: 76-139	Primers	—	—
SEQ ID NO: 140	Pseudogene of S	—	—
SEQ ID NO: 141-148	Primers	—	—
SEQ ID NO: 149	Aa1-13 of S	—	—
SEQ ID NO: 150	Polypeptide	—	—
SEQ ID NO: 151-158	Primers	—	—

* PCR amplification product (amplicon)

** Insert cloned into the plasmid deposited at the CNCM and to the appended drawings in which:

[0243] FIG. 1 illustrates Western-blot analysis of the expression in vitro of the recombinant proteins N, S_C and S_L from the expression vectors pIVEX. Lane 1: pIV2.3N. Lane 2: pIV2.3S_C. Lane 3: pIV2.3S_L. Lane 4: pIV2.4N. Lane 5: pIV2.4S₁ or pIV2.4S_C. Lane 6: pIV2.4S_L. The expression of the GFP protein expressed from the same vector is used as a control.

[0244] FIG. 2 illustrates the analysis, by polyacrylamide gel electrophoresis under denaturing conditions (SDS-PAGE) and staining with Coomassie blue, of the expression in vivo of the N protein from the expression vectors pIVEX. The *E. coli* BL21(DE3)pDIA17 strain transformed with the recombinant vectors pIVEX is cultured at 30° C. in LB medium, in the presence or in the absence of inducer (IPTG 1 mM). Lane 1: pIV2.3N. Lane 2: pIV2.4N.

[0245] FIG. 3 illustrates the analysis, by polyacrylamide gel electrophoresis under denaturing conditions (SDS-

PAGE) and staining with Coomassie blue, of the expression in vivo of the S_L and S_C polypeptides from the expression vectors pIVEX. The *E. coli* BL21(DE3)pDIA17 strain transformed with the recombinant vectors pIVEX is cultured at 30° C. in LB medium, in the presence or in the absence of inducer (IPTG 1 mM). Lane 1: pIV2.3 S_C . Lane 2: pIV2.3 S_L . Lane 3: pIV2.4 S_L . Lane 4: pIV2.4 S_L .

[0246] FIG. 4 illustrates the antigenic activity of the recombinant N, S_L and S_C proteins produced in the *E. coli* BL21(DE3)pDIA17 strain transformed with the recombinant vectors pIVEX. A: electrophoresis (SDS-PAGE) of the bacterial lysates. B and C: Western-blot with the sera, obtained from the same patient infected with SARS-CoV, collected 6 days (B: serum M12) and 29 days (C: serum M13) respectively after the onset of the SARS symptoms. Lane 1: pIV2.3N. Lane 2: pIV2.4N. Lane 3: pIV2.3 S_C . Lane 4: pIV2.4 S_L . Lane 5: pIV2.3 S_L . Lane 6: pIV2.4 S_L .

[0247] FIG. 5 illustrates the purification on an Ni-NTA agarose column of the recombinant N protein produced in the *E. coli* BL21(DE3)pDIA17 strain from the vector pIV2.3N. Lane 1: total bacterial extract. Lane 2: soluble extract. Lane 3: insoluble extract. Lane 4: extract deposited on the Ni-NTA column. Lane 5: unbound proteins. Lane 6: fractions of peak 1. Lane 7: fractions of peak 2.

[0248] FIG. 6 illustrates the purification of the recombinant S_C protein from the inclusion bodies produced in the *E. coli* BL21(DE3)pDIA17 strain transformed with pIV2.4 S_L . A: Treatment with Triton X-100 (2%): Lane 1: total bacterial extract. Lane 2: soluble extract. Lane 3: insoluble extract. Lane 4: supernatant after treatment with Triton X-100 (2%). Lanes 5 and 6: pellet after treatment with Triton X-100 (2%). B: Treatment with 4 M, 5 M, 6 M and 7 M urea of the soluble and insoluble extracts.

[0249] FIG. 7 represents the immunoblot produced with the aid of a lysate of cells infected with SARS-CoV and a serum from a patient suffering from atypical pneumopathy.

[0250] FIG. 8 represents immunoblots produced with the aid of a lysate of cells infected with SARS-CoV and rabbit immunosera specific for the nucleoprotein N (A) and for the spicule protein S (B). I.S.: immune serum. p.i.: preimmune serum. The anti-N immune serum was used at 1/50 000 and the anti-S immune serum at 1/10 000.

[0251] FIG. 9 illustrates the ELISA reactivity of the rabbit monospecific polyclonal sera directed against the N protein or the short fragment of the S protein (S_C), toward the corresponding recombinant proteins used for immunization. A: rabbits P13097, P13081 and P13031 immunized with the purified recombinant N protein. B: rabbits P11135, P13042 and P14001 immunized with a preparation of inclusion bodies corresponding to the short fragment of the S protein (S_C). I.S.: immune serum. p.i.: preimmune serum.

[0252] FIG. 10 illustrates the ELISA reactivity of the purified recombinant N protein, toward sera from patients suffering from atypical pneumonia caused by SARS-CoV. FIG. 10A: ELISA plates prepared with the N protein at the concentration of 4 µg/ml and 2 µg/ml. FIG. 10B: ELISA plate prepared with the N protein at the concentration of 1 µg/ml. The sera designated A, B, D, E, F, G, H correspond to those of Table IV.

[0253] FIG. 11 illustrates the amplification by RT-PCR of decreasing quantities of synthetic RNA of the SARS-CoV N

gene (10^7 to 1 copy), with the aid of pairs of primers No. 1 (N/+28507, N/-28774) (A) and No. 2 (N/+28375, N/-28702) (B). T: amplification performed in the absence of RNA. MW: DNA marker.

[0254] FIG. 12 illustrates the amplification by RT-PCR in real time of synthetic RNA for the SARS-CoV N gene: decreasing quantities of synthetic RNA as replica (repli.; lanes 16 to 29) and of viral RNA diluted $1/20 \times 10^4$ (lane 32) were amplified by RT-PCR in real time with the aid of the kit "Light Cycler RNA Amplification Kit Hybridization Probes" and pairs of primers and probes of the No. 2 series, under the conditions described in Example 8.

[0255] FIG. 13 (FIGS. 13.1 to 13.7) represents the restriction map of the sequence SEQ ID NO: 1 corresponding to the DNA equivalent of the genome of the SARS-CoV strain derived from the sample recorded under the number 031589.

[0256] FIG. 14 shows the result of the SARS serology test by indirect N ELISA (1st series of sera tested).

[0257] FIG. 15 shows the result of the SARS serology test by indirect N ELISA (2nd series of sera-tested).

[0258] FIG. 16 presents the result of the SARS serology test by double epitope N ELISA (1st series of sera tested).

[0259] FIG. 17 shows the result of the SARS serology test by double epitope N ELISA (2nd series of sera tested).

[0260] FIG. 18 illustrates the test of reactivity of the anti-N monoclonal antibodies by ELISA on the native nucleoprotein N of SARS-CoV. The antibodies were tested in the form of hybridoma culture supernatants by indirect ELISA using an irradiated lysate of VeroE6 cells infected with SARS-CoV as antigen (SARS lysate curves). A negative control for reactivity is performed for each antibody on a lysate of uninfected VeroE6 cells (negative lysate curves). Several monoclonal antibodies of known specificity were used as negative control antibodies: para1-3 directed against the antigens of the parainfluenza viruses type 1-3 (Bio-Rad) and influenza B directed against the antigens of the influenza virus type B (Bio-Rad).

[0261] FIG. 19 illustrates the test of reactivity of the anti-N of SARS-CoV monoclonal antibodies by ELISA on the native antigens of the human coronavirus 229E (HCoV-229E). The antibodies were tested in the form of hybridoma culture supernatants by an indirect ELISA test using a lysate of MRC-5 cells infected with the human coronavirus 229E as antigen (229E lysate curves). A negative control for immunoreactivity was performed for each antibody on a lysate of noninfected MRC-5 cells (negative lysate curves). The monoclonal antibody 5-11H.6 directed against the S protein of the human coronavirus 229E (Sizun et al. 1998, J. Virol. Met. 72: 145-152) is used as positive control antibody. The antibodies para1-3 directed against the antigens of the parainfluenza virus type 1-3 (Bio-Rad) and influenza B directed against the antigens of the influenza virus type B (Bio-Rad) were added to the panel of monoclonal antibodies tested.

[0262] FIG. 20 shows a test of reactivity of the anti-N of SARS-CoV monoclonal antibodies by Western blotting on the denatured native nucleoprotein N of SARS-CoV. A lysate of VeroE6 cells infected with SARS-CoV was prepared in the loading buffer according to Laemmli and caused to migrate in a 12% SDS polyacrylamide gel and then the

proteins were transferred onto PVDF membrane. The anti-N monoclonal antibodies tested were used for the immunoassay at the concentration of 0.05 µg/ml. The visualization is carried out with anti-mouse IgG(H+L) antibodies coupled to peroxidase (NA93IV, Amersham) and the ECL+ system. Two monoclonal antibodies were used as negative controls for reactivity: influenza B directed against the antigens of the influenza virus type B (Bio-Rad) and para1-3 directed against the antigens of the parainfluenza virus type 1-3 (Bio-Rad).

[0263] FIG. 21 presents the plasmids for expression in mammalian cells of the SARS-CoV S protein. The cDNA for the SARS-CoV S was inserted between the BamH1 and Xho1 sites of the expression plasmid pcDNA3.1(+) (Clontech) in order to obtain the plasmid pcDNA-S and between the Nhe1 and Xho1 sites of the expression plasmid pCI (Promega) in order to obtain the plasmid pCI-S. The WPRE and CTE sequences were inserted between each of the two plasmids pcDNA-S and pCI-S between the Xho1 and Xba1 sites in order to obtain the plasmids pcDNA-S-CTE, pcDNA-S-WPRE, pCI-S-CTE and pCI-S-WPRE, respectively.

[0264] SP: signal peptide predicted (aa 1-13) with the software signalP v2.0 (Nielsen et al., 1997, Protein Engineering, 10: 1-6)

[0265] TM: transmembrane region predicted (aa 1196-1218) with the software TMHMM v2.0 (Sonnhammer et al., 1998, Proc. of Sixth Int. Conf. on Intelligent Systems for Molecular Biology, pp. 175-182, AAAI Press). It should be noted that the amino acids W-1194 and P1195 are possibly part of the transmembrane region with the respective probabilities of 0.13 and 0.42

[0266] P-CMV: cytomegalovirus immediate/early promoter. BGH pA: polyadenylation signal of the bovine growth hormone gene

[0267] SV40 late pA: SV40 virus late polyadenylation signal

[0268] SD/SA: splice donor and acceptor sites

[0269] WPRE: sequences of the "Woodchuck Hepatitis Virus posttranscriptional regulatory element" of the woodchuck hepatitis virus

[0270] CTE: sequences of the "constitutive transport element" of the Mason-Pfizer simian retrovirus

[0271] FIG. 22 illustrates the expression of the S protein after transfection of VeroE6 cells. Cellular extracts were prepared 48 hours after transfection of VeroE6 cells with the plasmids pcDNA, pcDNA-S, pCI and pCI-S. Cellular extracts were also prepared 18 hours after infection with the recombinant vaccinia virus VV-TF7.3 and transfection with the plasmids pcDNA or pcDNA-S. As a control, extracts of VeroE6 cells were prepared 8 hours after infection with SARS-CoV at a multiplicity of infection of 3. They were separated on an 8% SDS acrylamide gel and analyzed by Western blotting with the aid of an anti-S rabbit polyclonal antibody and an anti-rabbit IgG(H+L) polyclonal antibody coupled to peroxidase (NA934V, Amersham). A molecular mass ladder (kDa) is presented in the figure.

[0272] SARS-CoV: extract of VeroE6 cells infected with SARS-CoV

[0273] Mock: control extract of noninfected cells

[0274] FIG. 23 illustrates the effect of the CTE and WPRE sequences on the expression of the S protein after transfection of VeroE6 and 293T cells. Cellular extracts were prepared 48 hours after transfection of VeroE6 cells (A) or 293T cells (B) with the plasmids pcDNA, pcDNA-S, pcDNA-S-CTE, pcDNA-S-WPRE, pCI-S, pCI-S-CTE and pCI-S-WPRE separated on 8% SDS polyacrylamide gel and analyzed by Western blotting with the aid of an anti-S rabbit polyclonal antibody and an anti-rabbit IgG(H+L) polyclonal antibody coupled to peroxidase (NA934V, Amersham). A molecular mass ladder (kDa) is presented in the figure.

[0275] SARS-CoV: extract of VeroE6 cells prepared 8 hours after infection with SARS-CoV at a multiplicity of infection of 3.

[0276] Mock: control extract of noninfected VeroE6 cells

[0277] FIG. 24 presents defective lentiviral vectors with central DNA flap for the expression of SARS-CoV S. The cDNA for the SARS-CoV S protein was cloned in the form of a BamH1-Xho1 fragment into the plasmid pTRIPΔU3-CMV containing a defective lentiviral vector TRIP with central DNA flap (Sirven et al., 2001, Mol. Ther., 3: 438-448) in order to obtain the plasmid pTRIP-S. The optimum expression cassettes consisting of the CMV virus immediate/early promoter, a splice signal, cDNA for S and either of the posttranscriptional signals CTE or WPRE were substituted for the cassette EF1α-EGFP of the defective lentiviral expression vector with central DNA flap TRIPΔU3-EF1α (Sirven et al., 2001, Mol. Ther., 3: 438-448) in order to obtain the plasmids pTRIP-SD/SA-S-CTE and pTRIP-SD/SA-S-WPRE.

[0278] SP: signal peptide

[0279] TM: transmembrane region

[0280] P-CMV: cytomegalovirus immediate/early promoter

[0281] P-EF1α: EF1α gene promoter

[0282] SD/SA: splice donor and acceptor sites

[0283] WPRE: sequences of the "Woodchuck Hepatitis Virus posttranscriptional regulatory element" of the woodchuck hepatitis virus

[0284] CTE: sequences of the "constitutive transport element" of the Mason-Pfizer simian retrovirus

[0285] LTR: long terminal repeat

[0286] ΔU3: LTR deleted for the "promoter/enhancer" sequences

[0287] cPPT: "polypurine tract cis-active sequence"

[0288] CTS: "central termination sequence"

[0289] FIG. 25 shows the Western-blot analysis of the expression of the SARS-CoV S by cell lines transduced with the lentiviral vectors TRIP-SD/SA-S-WPRE and TRIP-SD/SA-S-CTE. Cellular extracts were prepared from established lines FrhK4-S-CTE and FrhK4-S-WPRE after transduction with the lentiviral vectors TRIP-SD/SA-S-CTE and TRIP-SD/SA-S-WPRE respectively. They were separated on an 8% SDS acrylamide gel and analyzed by Western blotting

with the aid of an anti-S rabbit polyclonal antibody and an anti-rabbit IgG(H+L) conjugate coupled to peroxidase. A molecular mass ladder (kDa) is presented in the figure.

[0290] T-: control extract of FrhK-4 cells

[0291] T+: extract of FrhK-4 cells prepared 24 hours after infection with SARS-CoV at a multiplicity of infection of 3.

[0292] FIG. 26 relates to the analysis of the expression of Ssol polypeptide by cell lines transduced with the lentiviral vectors TRIP-SD/SA-Ssol-WPRE and TRIP-SD/SA-Ssol-CTE. The secretion of the Ssol polypeptide was determined in the supernatant of a series of cell clones isolated after transduction of FrhK-4 cells with the lentiviral vectors TRIP-SD/SA-Ssol-WPRE and TRIP-SD/SA-Ssol-CTE. 5 μ l of supernatant, diluted 1/2 in loading buffer according to Laemmli, were analyzed by Western blotting, visualized with an anti-FLAG monoclonal antibody (M2, Sigma) and an anti-mouse IgG(H+L) conjugate coupled to peroxidase. T-: supernatant of the parental FRhK-4 line. T+: supernatant of BHK cells infected with a recombinant vaccinia virus expressing the Ssol polypeptide. The solid arrow indicates the Ssol polypeptide, while the empty arrow indicates a cross reaction with a protein of cellular origin.

[0293] FIG. 27 shows the results relating to the analysis of the purified Ssol polypeptide

[0294] A. 8, 2, 0.5 and 0.125 μ g of recombinant Ssol polypeptide purified by anti-FLAG affinity chromatography and gel filtration (G75) were separated on 8% SDS polyacrylamide gel. The Ssol polypeptide and variable quantities of molecular mass markers (MM) were visualized by staining with silver nitrate (Gelcode SilverSNAP stain kit II, Pierce).

[0295] B. Standard markers for analysis by SELDI-TOF mass spectrometry

[0296] IgG: bovine IgG of MM 147300

[0297] ConA: conalbumin of MM 77490

[0298] HRP: horseradish peroxidase analyzed as a control and of MM 43240

[0299] C. Analysis by mass spectrometry (SELDI-TOF) of the recombinant Ssol polypeptide.

[0300] The peaks A and B correspond to the single and double charged Ssol polypeptide.

[0301] D. Sequencing of the N-terminal end of the recombinant Ssol polypeptide. 5 Edman degradation cycles in liquid phase were carried out on an ABI494 sequencer (Applied Biosystems).

[0302] FIG. 28 illustrates the influence of a splicing signal and of the CTE and WPRE sequences on the efficacy of the gene immunization with the aid of plasmid DNA encoding the SARS-CoV S

[0303] A. Groups of 7 BALB/c mice were immunized twice at 4 weeks' interval with the aid of 50 μ g of plasmid DNA of pCI, pcDNA-S, pCI-S, pcDNA-N and pCI-HA.

[0304] B. Groups of 6 BALB/c mice were immunized twice at 4 weeks' interval with the aid of 2 μ g, 10 μ g or 50 μ g of plasmid DNA of pCI, pCI-S, pCI-S-CTE and pCI-S-WPRE.

[0305] The immune sera collected 3 weeks after the second immunization were analyzed by indirect ELISA using a lysate of VeroE6 cells infected with SARS-CoV as antigen. The anti-SARS-CoV antibody titers are calculated as the reciprocal of the dilution producing a specific OD of 0.5 after visualization with an anti-mouse IgG polyclonal antibody coupled to peroxidase (NA931V, Amersham) and TMB (KPL).

[0306] FIG. 29 shows the seroneutralization of the infectivity of SARS-CoV with the antibodies induced in mice after gene immunization with the aid of plasmid DNA encoding SARS-CoV S. Pools of immune sera collected 3 weeks after the second immunization were prepared for each of the groups of experiments described in FIG. 28 and evaluated for their capacity to seroneutralize the infectivity of 100 TCID₅₀ of SARS-CoV on FRhK-4 cells. 4 points are produced for each of the 2-fold dilutions tested from 1/20. The seroneutralizing titer is calculated according to the Reed and Munch method as the reciprocal of the dilution neutralizing the infectivity of 2 wells out of 4.

[0307] A. Groups by BALB/c mice immunized twice at 4 weeks' interval with the aid of 50 μ g of plasmid DNA of pCI, pcDNA-S, pCI-S, pcDNA-N and pCI-HA. □: pre-immune serum. ■: immune serum.

[0308] B. Groups of BALB/c mice immunized twice at 4 weeks' interval with the aid of 2 μ g, 10 μ g or 50 μ g of plasmid DNA of pCI, pCI-S, pCI-S-CTE and pCI-S-WPRE.

[0309] FIG. 30 illustrates the immunoreactivity of the recombinant Ssol polypeptide toward sera from patients suffering from SARS. The reactivity of sera from patients was analyzed by indirect ELISA test against solid phases prepared with the aid of the purified recombinant Ssol polypeptide. The antibodies from patients reacting with the solid phase at a dilution of 1/400 are visualized with a human anti-IgG(H+L) polyclonal antibody coupled to peroxidase (Amersham NA933V) and TMB plus, H202 (KPL). The sera of probable SARS cases are identified by a National Reference Center for Influenza Viruses serial number and by the initials of the patient and the number of days elapsed since the onset of symptoms, where appropriate. The TV sera are control sera from subjects which were collected in France before the SARS epidemic which occurred in 2003.

[0310] FIG. 31 shows the induction of antibodies directed against SARS-CoV after immunization with the recombinant Ssol polypeptide. Two groups of 6 mice were immunized at 3 weeks' interval with 10 μ g of recombinant Ssol polypeptide (Ssol group) adjuvanted with aluminum hydroxide or, as a control, of adjuvant alone (mock group). Three successive immunizations were performed and the immune sera were collected 3 weeks after each of the three immunizations (IS1, IS2, IS3). The immune sera were analyzed per pool for each of the 2 groups by indirect ELISA using a lysate of VeroE6 cells infected with SARS CoV as antigen. The anti-SARS-CoV antibody titers are calculated as the reciprocal of the dilution producing a specific OD of 0.5 after visualization with an anti-mouse IgG polyclonal antibody coupled to peroxidase (Amersham) and TMB (KPL).

[0311] FIG. 32 presents the nucleotide alignment of the sequences of the synthetic gene 040530 with the sequence of the wild-type gene of the SARS-CoV isolate 031589. I-3059

corresponds to nucleotides 21406-25348 of the SARS-CoV isolate 031589 deposited at the C.N.C.M. under the number I-3059 (SEQ ID NO: 4, plasmid pSARS-S) S-040530 is the sequence of the synthetic gene 040530.

[0312] FIG. 33 illustrates the use of a synthetic gene for the expression of the SARS-CoV S. Cellular extracts prepared 48 hours after transfection of VeroE6 cells (A) or 293T cells (B) with the plasmids pCI, pCI-S, pCI-S-CTE, pCI-S-WPRE and pCI-Ssynth were separated on 8% SDS acrylamide gel and analyzed by Western blotting with the aid of an anti-S rabbit polyclonal antibody and an anti-rabbit IgG(H+L) polyclonal antibody coupled to peroxidase (NA934V, Amersham). The Western blot is visualized by luminescence (ECL+, Amersham) and acquisition on a digital imaging device (FluorS, BioRad). The levels of expression of the S protein were measured by quantifying the 2 predominant bands identified on the image.

[0313] FIG. 34 presents a diagram for the construction of recombinant vaccinia viruses VV-TG-S, VV-TG-Ssol, VV-TN-S and VV-TN-Ssol

[0314] A. The cDNAs for the S protein and the Ssol polypeptide of SARS-CoV were inserted between the BamHI and SmaI sites of the transfer plasmid pTG186 in order to obtain the plasmids pTG-S and pTG-Ssol.

[0315] B. The sequences of the synthetic promoter 480 were then substituted for those of the 7.5 promoter by exchange of the NdeI-PstI fragments of the plasmids pTG186poly, pTG-S and pTG-Ssol in order to obtain the transfer plasmids pTN480, pTN-S and pTN-Ssol.

[0316] C. Sequence of the synthetic promoter 480 as contained between the NdeI and PstI sites of the transfer plasmids of the pTN series. An AscI site was inserted in order to facilitate subsequent handling. The restriction sites and the promoter sequence are underlined.

[0317] D. The recombinant vaccinia viruses are obtained by double homologous recombination in vivo between the TK cassette of the transfer plasmids of the pTG and pTN series and the TK gene of the Copenhagen strain of the vaccinia virus.

[0318] SP: signal peptide predicted (aa 1-13) with the software signalP v2.0 (Nielsen et al., 1997, Protein Engineering, 10: 1-6)

[0319] TM: transmembrane region predicted (aa 1196-1218) with the software TMHMM v2.0 (Sonnhammer et al., 1998, Proc. of Sixth Int. Conf. on Intelligent Systems for Molecular Biology, pp. 175-182, AAAI Press). It should be noted that the amino acids W1194 and P1195 possibly form part of the transmembrane region with respective probabilities of 0.13 and 0.42.

[0320] TK-L, TK-R: left- and right-hand parts of the vaccinia virus thymidine kinase gene

[0321] MCS: multiple cloning site

[0322] PE: early promoter

[0323] PL: late promoter

[0324] PL synth: synthetic late promoter 480

[0325] FIG. 35 illustrates the expression of the S protein by recombinant vaccinia viruses, analyzed by Western blot-

ting. Cellular extracts were prepared 18 hours after infection of CV1 cells with the recombinant vaccinia viruses VV-TG, VV-TG-S and VV-TN-S at an M.O.I. of 2 (A). As a control, extracts of VeroE6 cells were prepared 8 hours after infection with SARS-CoV at a multiplicity of infection of 2. Cellular extracts were also prepared 18 hours after infection of CV1 cells with the recombinant vaccinia viruses VV-TG-S, VV-TG-Ssol, VV-TN, VV-TN-S and VV-TN-Ssol (B). They were separated on 8% SDS acrylamide gels and analyzed by Western blotting with the aid of an anti-S rabbit polyclonal antibody and an anti-rabbit IgG(H+L) polyclonal antibody coupled to peroxidase (NA934V, Amersham). "1 μ l" and "10 μ l" indicates the quantities of cellular extracts deposited on the gel. A molecular mass ladder (kDa) is presented in the figure.

[0326] SARS-CoV: extract of VeroE6 cells infected with SARS-CoV

[0327] Mock: control extract of noninfected cells

[0328] FIG. 36 shows the result of a Western-blot analysis of the secretion of the Ssol polypeptide by the recombinant vaccinia viruses.

[0329] A. Supernatants of CV1 cells infected with the recombinant vaccinia virus VV-TN, various clones of the VV-TN-Ssol virus and with the viruses VV-TG-Ssol or VV-TN-Sflag were harvested 18 hours after infection of CV1 cells at an M.O.I. of 2.

[0330] B. Supernatants of 293T, FRhK-4, BHK-21 and CV1 cells infected in duplicate (1.2) with the recombinant vaccinia virus VV-TN-Ssol at an M.O.I. of 2 were harvested 18 hours after infection. The supernatant of CV1 cells infected with the virus VV-TN was also harvested as a control (M).

[0331] All the supernatants were separated on 8% SDS acrylamide gel according to Laemmli and analyzed by Western blotting with the aid of an anti-FLAG mouse monoclonal antibody and an anti-mouse IgG(H+L) polyclonal antibody coupled to peroxidase (NA931V, Amersham) (A) or with the aid of an anti-S rabbit polyclonal antibody and an anti-rabbit IgG(H+L) polyclonal antibody coupled to peroxidase (NA934V, Amersham) (B).

[0332] A molecular mass ladder (kDa) is presented in the figure.

[0333] FIG. 37 shows the analysis of the Ssol polypeptide, purified on SDS polyacrylamide gel

[0334] 10, 5 and 2 μ l of recombinant Ssol polypeptide purified by anti-FLAG affinity chromatography were separated on 4 to 15% gradient SDS polyacrylamide gel. The Ssol polypeptide and variable quantities of molecular mass markers (MM) were visualized by staining with silver nitrate (Gelcode SilverSNAP stain kit II, Pierce).

[0335] FIG. 38 illustrates the immunoreactivity of the recombinant Ssol polypeptide produced by the recombinant vaccinia virus VV-TN-Ssol toward sera of patients suffering from SARS. The reactivity of sera from patients was analyzed by indirect ELISA test against solid phases prepared with the aid of the purified recombinant Ssol polypeptide. The antibodies from patients reacting with the solid phase at a dilution of 1/100 and 1/400 are visualized with a human anti-IgG(H+L) polyclonal antibody coupled to peroxidase

(Amersham NA933V) and TMB plus H202 (KPL). The sera of probable SARS cases are identified by a National Reference Center for Influenza Virus serial number and by the initials of the patient and the number of days elapsed since the onset of symptoms, where appropriate. The TV sera are control sera from subjects which were collected in France before the SARS epidemic which occurred in 2003.

[0336] FIG. 39 shows the anti-SARS-CoV antibody response in mice after immunization with the recombinant vaccinia viruses. Groups of 7 BALB/c mice were immunized by the i.v. route twice at 4 weeks' interval with 106 pfu of recombinant vaccinia viruses VV-TG, VV-TG-HA, VV-TG-S, W-TG-Ssol, VV-TN, VV-TN-S, VV-TN-Ssol.

[0337] A. Pools of immune sera collected 3 weeks after each of the two immunizations were prepared for each of the groups and were analyzed by indirect ELISA using a lysate of VeroE6 cells infected with SARS-CoV as antigen. The anti-SARS-CoV antibody titers are calculated as the reciprocal of the dilution producing a specific OD of 0.5 after visualization with an anti-mouse IgG polyclonal antibody coupled to peroxidase (NA931V, Amersham) and TMB (KPL).

[0338] B. The pools of immune sera were evaluated for their capacity to seroneutralize the infectivity of 100 TCID₅₀ of SARS-CoV on FRhK-4 cells. 4 points are produced for each of the 2-fold dilutions tested from 1/20. The seroneutralizing titer is calculated according to the Reed and Munsch method as the reciprocal of the dilution neutralizing the infectivity of 2 wells out of 4.

[0339] FIG. 40 describes the construction of the recombinant viruses MVSchw2-SARS-S and MVSchw2-SARS-Ssol.

[0340] A. The measles vector is a complete genome of the Schwarz vaccine strain of the measles virus (MV) into which an additional transcription unit has been introduced (Combretet, 2003, Journal of Virology, 77: 11546-11554). The expression of the additional open reading frames (ORF) is controlled by cis-acting elements necessary for the transcription, for the formation of the cap and for the polyadenylation of the transgene which were copied from the elements present at the N/P junction. 2 different vectors allow the insertion between the P (phosphoprotein) and M (matrix) genes on the one hand and the H (hemagglutinin) and L (polymerase) genes on the other hand.

[0341] B. The recombinant genomes MVSchw2-SARS-S and MVSchw2-SARS-Ssol of the measles virus were constructed by inserting the ORFs of the S protein and of the Ssol polypeptide into an additional transcription unit located between the P and M genes of the vector.

[0342] The various genes of the measles virus (MV) are indicated: N (nucleoprotein), PVC (V/C phosphoprotein and protein), M (matrix), F (fusion), H (hemagglutinin), L (polymerase). T7=T7 RNA polymerase promoter, hh=hammerhead ribozyme, T7t=T7 phage RNA polymerase terminator sequence, δ=ribozyme of the hepatitis δ virus, (2), (3)=additional transcription units (ATU).

[0343] Size of the MV genome: 15 894 nt.

[0344] SP: signal peptide

[0345] TM: transmembrane region

[0346] FLAG: FLAG tag

[0347] FIG. 41 illustrates the expression of the S protein by the recombinant measles viruses, analyzed by Western blotting.

[0348] Cytoplasmic extracts were prepared after infection of Vero cells by different passages of the viruses MVSchw2-SARS-S and MVSchw2-SARS-Ssol and the wild-type virus MWSchw as control. Cellular extracts in loading buffer according to Laemmli were also prepared 8 hours after infection of VeroE6 cells with SARS-CoV at a multiplicity of infection of 3. They were separated on 8% SDS acrylamide gel and analyzed by Western blotting with the aid of an anti-S rabbit polyclonal antibody and an anti-rabbit IgG(H+L) polyclonal antibody coupled to peroxidase (NA934V, Amersham).

[0349] A molecular mass ladder (kDa) is presented in the figure.

[0350] Pn: nth passage of the virus after coculture of 293-3-46 and Vero cells

[0351] SARS-CoV: extract of VeroE6 cells infected with SARS-CoV

[0352] Mock: control extract of noninfected VeroE6 cells

[0353] FIG. 42 shows the expression of the S protein by the recombinant measles viruses, analyzed by immunofluorescence

[0354] Vero cells in monolayers on glass slides were infected with the wild-type virus MWSchw (A) or the viruses MVSchw2-SARS-S (B) and MVSchw2-SARS-Ssol (C). When the syncytia have reached 30 to 40% confluence (A., B.) or 90-100% (C), the cells were fixed, permeabilized and labeled with anti-SARS-CoV rabbit polyclonal antibodies and an anti-rabbit IgG(H+L) conjugate coupled to FITC (Jackson).

[0355] FIG. 43 illustrates the Western-blot analysis of the immunoreactivity of rabbit sera directed against the peptides E1-12, E53-76 and M2-14. The rabbit 20047 was immunized with the peptide E1-12 coupled to KLH. The rabbits 22234 and 22240 were immunized with the peptide E53-76 coupled to KLH. The rabbits 20013 and 20080 were immunized with the peptide M2-14 coupled to KLH. The immune sera were analyzed by Western blotting with the aid of extracts of cells infected with SARS-CoV (B) or with the aid of extracts of cells infected with a recombinant vaccinia virus expressing the protein E (A) or M (C) of the SARS-CoV 031589 isolate. The immunoblots were visualized with the aid of an anti-rabbit IgG(H+L) conjugate coupled to peroxidase (NA934V, Amersham).

[0356] The position of the E and M proteins is indicated by an arrow.

[0357] A molecular mass ladder (kDa) is presented in the figure.

[0358] It should be understood, however, that these examples are given solely by way of illustration of the subject of the invention, and do not constitute in any manner a limitation thereto.

EXAMPLE 1

Cloning and Sequencing of the Genome of the
SARS-CoV Strain Derived from the Sample
Recorded Under the Number 031589

[0359] The RNA of the SARS-CoV strain was extracted from the sample of bronchoalveolar washing recorded under the number 031589, performed on a patient at the Hanoi (Vietnam) French hospital suffering from SARS.

[0360] The isolated RNA was used as template to amplify the cDNAs corresponding to the various open reading frames of the genome (ORF1a, ORF1b, ORF-S, ORF-E, ORF-M, ORF-N (including ORF-13 and ORF-14), ORF3, ORF4, ORF7 to ORF11), and at the noncoding 5' and 3' ends. The sequences of the primers and of the probes used for the amplification/detection were defined based on the available SARS-CoV nucleotide sequence.

[0361] In the text which follows, the primers and the probes are identified by: the letter S, followed by a letter which indicates the corresponding region of the genome (L for the 5' end including ORF1a and ORF1b; S, M and N for ORF-S, ORF-M, ORF-N, SE and MN for the corresponding intergene regions), and then optionally by Fn, Rn, with n between 1 and 6 corresponding to the primers used for the nested PCR (F1+R1 pair for the first amplification, F2+R2 pair for the second amplification, and the like), and then by +/- or -/- corresponding to a sense or antisense primer and finally by the positions of the primers with reference to the Genbank sequence AY27411.3; for the sense and antisense S and N primers and the other sense primers only, when a single position is indicated, it corresponds to that of the 5' end of a probe or of a primer of about 20 bases; for the antisense primers other than the S and N primers, when a single position is indicated, it corresponds to that of the 3' end of a probe or of a primer of about 20 bases.

[0362] The amplification products thus generated were sequenced with the aid of specific primers in order to determine the complete sequence of the genome of the SARS-CoV strain derived from the sample recorded under the number 031589. These amplification products, with the exception of those corresponding to ORF1a and ORF1b, were then cloned into expression vectors in order to produce the corresponding viral proteins and the antibodies directed against these proteins, in particular by DNA-based immunization.

1. Extraction of the RNAs

[0363] The RNAs were extracted with the aid of the QIamp viral RNA extraction mini kit (QIAGEN) according to the manufacturer's recommendations. More specifically: 140 μ l of the sample and 560 μ l of AVL buffer were vigorously mixed for 15 seconds, incubated for 10 minutes at room temperature and then briefly centrifuged at maximum speed. 560 μ l of 100% ethanol were added to the supernatant and the mixture thus obtained was very vigorously stirred for 15 sec. 630 μ l of the mixture were then deposited on the column.

[0364] The column was placed on a 2 ml tube, centrifuged for 1 min at 8000 rpm, and then the remainder of the preceding mixture was deposited on the same column, centrifuged again, for 1 min at 8000 rpm, and the column was transferred over a clean 2 ml tube. Next, 500 μ l of AW1

buffer were added to the column, and then the column was centrifuged for 1 min at 8000 rpm and the eluate was discarded. 500 μ l of AW2 buffer were added to the column which was then centrifuged for 3 min at 14 000 rpm and transferred onto a 1.5 ml tube. Finally, 60 μ l of AVE buffer were added to the column which was incubated for 1 to 2 min at room temperature and then centrifuged for 1 min at 8000 rpm. The eluate corresponding to the purified RNA was recovered and frozen at -20° C.

2. Amplification, Sequencing and Cloning of the cDNAs

2.1) cDNA Encoding the S Protein

[0365] The RNAs extracted from the sample were subjected to reverse transcription with the aid of random sequence hexameric oligonucleotides (pdN6), so as to produce cDNA fragments.

[0366] The sequence encoding the SARS-CoV S glycoprotein was amplified in the form of two overlapping DNA fragments: 5' fragment (SARS-Sa, SEQ ID NO: 5) and 3' fragment (SARS-Sb, SEQ ID NO: 6), by carrying out two successive amplifications with the aid of nested primers. The amplicons thus obtained were sequenced, cloned into the PCR plasmid vector 2.1-TOPO™ (INVITROGEN), and then the sequence of the cloned cDNAs was determined.

a) Cloning and Sequencing of the Sa and Sb Fragments

a.1) Synthesis of the cDNA

[0367] The reaction mixture containing: RNA (5 μ l), H₂O for injection (3.5 μ l), 5 \times reverse transcriptase buffer (4 μ l), 5 mM dNTP (2 μ l), pdN6 100 μ g/ml (4 μ l), RNasin 40 IU/ μ l (0.5 μ l) and reverse transcriptase AMV-RT, 10 IU/ μ l, PROMEGA (1 μ l) was incubated in a thermocycler under the following conditions: 45 min at 42° C., 15 min at 55° C., 5 min at 95° C., and then the cDNA obtained was kept at $+4^{\circ}$ C.

a.2) First PCR Amplification

[0368] The 5' and 3' ends of the S gene were respectively amplified with the pairs of primers S/F1+/21350-21372 and S/R1-/23518-23498, S/F3+/23258-23277 and S/R3-/25382-25363. The 50 μ l reaction mixture containing: cDNA (2 μ l), 50 μ M primers (0.5 μ l), 10 \times buffer (5 μ l), 5 mM dNTP (2 μ l), Taq Expand High Fidelity, Roche (0.75 μ l) and H₂O (39, 75 μ l) was amplified in a thermocycler, under the following conditions: an initial step of denaturation at 94° C. for 2 min was followed by 40 cycles comprising: a step of denaturation at 94° C. for 30 sec, a step of annealing at 55° C. for 30 sec and then a step of extension at 72° C. for 2 min 30 sec, with 10 sec of additional extension at each cycle, and then a final step of extension at 72° C. for 5 min.

a.3) Second PCR Amplification

[0369] The products of the first PCR amplification (5' and 3' amplicons) were subjected to a second PCR amplification step (nested PCR) under conditions identical to those of the first amplification, with the pairs of primers S/F2+/21406-21426 and S/R2-/23454-23435 and S/F4+/23322-23341 and S/R4-/25348-25329, respectively for the 5' amplicon and the 3' amplicon.

a.4) Cloning and Sequencing of the Sa and Sb Fragments

[0370] The Sa (5' end) and Sb (3' end) amplicons thus obtained were purified with the aid of the QIAquick PCR

purification kit (QIAGEN), following the manufacturer's instructions, and then they were cloned into the vector PCR2.1-TOPO (Invitrogen kit), to give the plasmids called SARS-S1 and SARS-S2.

[0371] The DNA of the Sa and Sb clones was isolated and then the corresponding insert was sequenced with the aid of the Big Dye kit, Applied Biosystem® and universal primers M13 forward and M13 reverse, and primers: S/S/+21867, S/S/+22353, S/S/+22811, S/S/+23754, S/S/+24207, S/S/+24699, S/S/+24348, S/S/-24209, S/S/-23630, S/S/-23038, S/S/-22454, S/S/-21815, S/S/-24784, S/S/+21556, S/S/+23130 and S/S/+24465 following the manufacturer's instructions; the sequences of the Sa and Sb fragments thus obtained correspond to the sequences SEQ ID NO: 5 and SEQ ID NO: 6 in the sequence listing appended as an annex.

[0372] The plasmid, called SARS-S1, was deposited under the No. I-3020, on May 12, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains a 5' fragment of the sequence of the S gene of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above, said fragment called Sa corresponding to the nucleotides at positions 21406 to 23454 (SEQ ID NO: 5), with reference to the Genbank sequence AY274119.3 Tor2.

[0373] The plasmid, called TOP10F'-SARS-S2, was deposited under the No. I-3019, on May 12, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains a 3' fragment of the sequence of the S gene of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above, said fragment called Sb corresponding to the nucleotides at positions 23322 to 25348 (SEQ ID NO: 6), with reference to the Genbank sequence accession No. AY274119.3.

b) Cloning and Sequencing of the Complete cDNA (SARS-S Clone of 4 kb)

[0374] The complete S cDNA was obtained from the abovementioned clones SARS-S1 and SARS-S2, in the following manner:

[0375] 1) A PCR amplification reaction was carried out on a SARS-S2 clone in the presence of the abovementioned primer S/R4/-25348-25329 and of the primer S/S/+24696-24715: an amplicon of 633 bp was obtained,

[0376] 2) Another PCR amplification reaction was carried out on another SARS-S2 clone, in the presence of the primers S/F4/+23322-23341 mentioned above and S/S/-24803-24784: an amplicon of 1481 bp was obtained.

[0377] The amplification reaction was carried out under the conditions as defined above for the amplification of the Sa and Sb fragments, with the exception that 30 amplification cycles comprising a step of denaturation at 94° C. for 20 sec and a step of extension at 72° C. for 2 min 30 sec were carried out.

[0378] 3) The 2 amplicons (633 bp and 1481 bp) were purified under the conditions as defined above for the Sa and Sb fragments.

[0379] 4) Another PCR amplification reaction with the aid of the abovementioned primers S/F4/+23322-23341 and

S/R4/-25348-25329 was carried out on the purified amplicons obtained in 3). The amplification reaction was carried out under the conditions as defined above for the amplification of the Sa and Sb fragments, except that 30 amplification cycles were performed.

[0380] The 2026 bp amplicon thus obtained was purified, cloned into the vector PCR2.1-TOPO and then sequenced as above, with the aid of the primers as-defined above for the Sa and Sb fragments. The clone thus obtained was called clone 3'.

[0381] 5) The clone SARS-S1 obtained above and the clone 3"were digested with EcoR I, the bands of about 2 kb thus obtained were gel purified and then amplified by PCR with the abovementioned primers S/F2/+21406-21426 and S/R4/-25348-25329. The amplification reaction was carried out under the conditions as defined above for the amplification of the Sa and Sb fragments, except that 30 amplification cycles were performed. The amplicon of about 4 kb was purified and sequenced. It was then cloned into the vector PCR2.1-TOPO in order to give the plasmid, called SARS-S, and the insert obtained in this plasmid was sequenced as above, with the aid of the primers as defined above for the Sa and Sb fragments. The cDNA sequences of the insert and of the amplicon encoding the S protein correspond respectively to the sequences SEQ ID NO: 4 and SEQ ID NO: 2 in the sequence listing appended as an annex, they encode the S protein (SEQ ID NO: 3).

[0382] The sequence of the amplicon corresponding to the cDNA encoding the S protein of the SARS-CoV strain derived from the sample No. 031589 has the following two mutations compared with the corresponding sequences of respectively the Tor2 and Urbani isolates, the positions of the mutations being indicated with reference to the complete sequence of the genome of the Tor2 isolate (Genbank AY274119.3):

[0383] g/t in position 23220; the alanine codon (gct) in position 577 of the amino acid sequence of the S protein of Tor2 is replaced with a serine codon (tct),

[0384] c/t in position 24872: this mutation does not modify the amino acid sequence of the S protein, and

the plasmid, called SARS-S, was deposited under the No. I-3059, on Jun. 20, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA sequence encoding the S protein of the SARS-CoV strain derived from the sample recorded under the No. 031589, said sequence corresponding to the nucleotides at positions 21406 to 25348 (SEQ ID NO: 4), with reference to the Genbank sequence AY274119.3.

2.2) cDNA Encoding the M and E Proteins

[0385] The RNAs derived from the sample 031589, extracted as above, were subjected to a reverse transcription, combined, during the same step (Titan One Step RT-PCR® kit, Roche), with a PCR amplification reaction, with the aid of the pairs of primers:

[0386] S/E/F1/+26051-26070 and S/E/R1/-26455-26436 in order to amplify ORF-E, and

[0387] S/M/F1/+26225-26244 and S/M/R1/-27148-27129 in order to amplify ORF-M.

[0388] A first reaction mixture containing: 8.6 µl of H₂O for injection, 1 µl of dNTP (5 mM), 0.2 µl of each of the primers (50 µM), 1.25 µl of DTT (100 mM) and 0.25 µl of RNAsin (40 IU/µl) was combined with a second reaction mixture containing: 1 µl of RNA, 7 µl of H₂O for injection, 5 µl of 5×RT-PCR buffer and 0.5 µl of enzyme mixture and the combined mixtures were incubated in a thermocycler under the following conditions: 30 min at 42° C., 10 min at 55° C., 2 min at 94° C. followed by 40 cycles comprising a step of denaturation at 94° C. for 10 sec, a step of annealing at 55° C. for 30 sec and a step of extension at 68° C. for 45 sec, with 3 sec increment per cycle and finally a step of terminal extension at 68° C. for 7 min.

[0389] The amplification products thus obtained (M and E amplicons) were subjected to a second PCR amplification (nested PCR) using the Expand High-Fi® kit, Roche), with the aid of the pairs of primers:

[0390] S/E/F2/+26082-26101 and S/E/R2/-26413-26394 for the amplicon E, and

[0391] S/M/F2/+26330-26350 and S/M/R2/-27098-27078 for the amplicon M.

[0392] The reaction mixture containing: 2 µl of the product of the first PCR., 39.25 µl of H₂O for injection, 5 µl of 10× buffer containing MgCl₂, 2 µl of dNTP (5 mM), 0.5 µl of each of the primers (50 µM) and 0.75 µl of enzyme mixture was incubated in a thermocycler under the following conditions: a step of denaturation at 94° C. for 2 min was followed by 30 cycles comprising a step of denaturation at 94° C. for 15 sec, a step of annealing at 60° C. for 30 sec and a step of extension at 72° C. for 45 sec, with 3 sec increment per cycle, and finally a step of terminal extension at 72° C. for 7 min. The amplification products obtained corresponding to the cDNAs encoding the E and M proteins were sequenced as above, with the aid of the primers: S/E/F2/+26082 and S/E/R2/-26394, S/M/F2/+26330, S/M/R2/-27078 cited above and the primers S/M/+26636-26655 and S/M/-26567-26548. They were then cloned, as above, in order to give the plasmids called SARS-E and SARS-M. The DNA of these clones was then isolated and sequenced with the aid of the universal primers M13 forward and M13 reverse and the primers S/M/+26636 and S/M/-26548 mentioned above.

[0393] The sequence of the amplicon representing the cDNA encoding the E protein (SEQ ID NO: 13) of the SARS-CoV strain derived from the sample No. 031589 does not contain differences in relation to the corresponding sequences of the isolates AY274119.3-Tor2 and AY278741-Urbani. The sequence of the E protein of the SARS-CoV 031589 strain corresponds to the sequence SEQ ID NO: 14 in the sequence listing appended as an annex.

[0394] The plasmid, called SARS-E, was deposited under the No. I-3046, on May 28, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA sequence encoding the E protein of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above, said sequence corresponding to the nucleotides at positions 26082 to 26413 (SEQ ID NO: 15), with reference to the Genbank sequence accession No. AY274119.3.

[0395] The sequence of the amplicon representing the cDNA encoding M (SEQ ID NO: 16) from the SARS-CoV strain derived from the sample No. 031589 does not contain differences in relation to the corresponding sequence of the isolate AY274119.3-Tor2. By contrast, at position 26857, the

isolate AY278741-Urbani contains a c and the sequence of the SARS-CoV strain derived from the sample recorded under the No. 031589 contains a t. This mutation results in a modification of the amino acid sequence of the corresponding protein: at position 154, a proline (AY278741-Urbani) is changed to serine in the SARS-CoV strain derived from the sample recorded under the No. 031589. The sequence of the M protein of the SARS-CoV strain derived from the sample recorded under the No. 031589 corresponds to the sequence SEQ ID NO: 17 in the sequence listing appended as an annex.

[0396] The plasmid, called SARS-M, was deposited under the No. I-3047, on May 28, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA sequence encoding the M protein of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above; said sequence corresponding to the nucleotides at positions 26330 to 27098 (SEQ ID NO: 18), with reference to the Genbank sequence accession No. AY274119.3.

2.3) cDNA Corresponding to ORF3, ORF4, ORF7 to ORF11

[0397] The same amplification, cloning and sequencing strategy was used to obtain the cDNA fragments corresponding respectively to the following ORFs: ORF3, ORF4, ORF7, ORF8, ORF9, ORF10 and ORF11. The pairs of primers used for the first amplification are:

[0398] ORF3 and ORF4: S/SE/F1/+25069-25088 and S/SE/R1/-26300-26281

[0399] ORF7 to ORF11: S/MN/F1/+26898-26917 and S/MN/R1/-28287-28266

[0400] The pairs of primers used for the second amplification are:

[0401] ORF3 and ORF4: S/SE/F2/+25110-25129 and S/SE/R2/-26244-26225

[0402] ORF7 to ORF11: S/MN/F2/+26977-26996 and S/MN/R2/-28218-28199

[0403] The conditions for the first amplification (RT-PCR) are the following: 45 min at 42° C., 10 min at 55° C., 2 min at 94° C. followed by 40 cycles comprising a step of denaturation at 94° C. for 15 sec, a step of annealing at 58° C. for 30 sec and a step of extension at 68° C. for 1 min, with 5 sec increment per cycle and finally a step of terminal extension at 68° C. for 7 min. The conditions for the nested PCR are the following: a step of denaturation at 94° C. for 2 min was followed by 40 cycles comprising a step of denaturation at 94° C. for 20 sec, a step of annealing at 58° C. for 30 sec and a step of extension at 72° C. for 50 sec, with 4 sec increment per cycle and finally a step of terminal extension at 72° C. for 7 min.

[0404] The amplification products obtained corresponding to the cDNAs containing respectively ORF3 and 4 and ORF7 to 11 were sequenced with the aid of the primers: S/SE/+25363, S/SE/+25835, S/SE/-25494, S/SE/-25875, S/MN/+27839, S/MN/+27409, S/MN/-27836, S/MN/-27799 and cloned as above for the other ORFs, to give the plasmids called SARS-SE and SARS-MN. The DNA of these clones was isolated and sequenced with the aid of these same primers and of the universal primers M13 sense and M13 antisense.

[0405] The sequence of the amplicon representing the cDNA of the region containing ORF3 and ORF4 (SEQ ID NO: 7) of the SARS-CoV strain derived from the sample No.

031589 contains a nucleotide difference in relation to the corresponding sequence of the isolate AY274119-Tor2. This mutation at position 25298 results in a modification of the amino acid sequence of the corresponding protein (ORF3): at position 11, an arginine (AY274119-Tor2) is changed to glycine in the SARS-CoV strain derived from the sample No. 031589. By contrast, no mutation was identified in relation to the corresponding sequence of the isolate AY278741-Urbani. The sequences of ORF3 and 4 of the SARS-CoV strain derived from the sample No. 031589 correspond respectively to the sequences SEQ ID NO: 10 and 12 in the sequence listing appended as an annex.

[0406] The plasmid, called SARS-SE, was deposited under the No. I-3126, on Nov. 13, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA corresponding to the region situated between ORF-S and ORF-E and overlapping ORF-E of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above, said region corresponding to the nucleotides at positions 25110 to 26244 (SEQ ID NO: 8), with reference to the Genbank sequence accession No. AY274119.3.

[0407] The sequence of the amplicon representing the cDNA corresponding to the region containing ORF7 to ORF11 (SEQ ID NO: 19) of the SARS-CoV strain derived from the sample No. 031589 does not contain differences in relation to the corresponding sequences of the isolates AY274119-Tor2 and AY278741-Urbani. The sequences of ORF7 to 11 of the SARS-CoV strain derived from the sample No. 031589 correspond respectively to the sequences SEQ ID NO: 22, 24, 26, 28 and 30 in the sequence listing appended as an annex.

[0408] The plasmid, called SARS-MN, was deposited under the No. I-3125, on Nov. 13, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA sequence corresponding to the region situated between ORF-M and ORF-N of the SARS-CoV strain derived from the sample recorded under the No. 031589 and collected in Hanoi, as defined above, said sequence corresponding to the nucleotides at positions 26977 to 28218 (SEQ ID NO: 20), with reference to the Genbank sequence accession No. AY274119.3.

[0409] The sequence of the amplicon representing the cDNA corresponding to the region containing ORF7 to ORF11 (SEQ ID NO: 19) of the SARS-CoV strain derived from the sample No. 031589 does not contain differences in relation to the corresponding sequences of the isolates AY274119-Tor2 and AY278741-Urbani. The sequences of ORF7 to 11 of the SARS-CoV strain derived from the sample No. 031589 correspond respectively to the sequences SEQ ID NO: 22, 24, 26, 28 and 30 in the sequence listing appended as an annex.

2.4) cDNA Encoding the N Protein and Including ORF13 and ORF14

[0410] The cDNA was synthesized and amplified as described above for the fragments Sa and Sb. More specifically, the reaction mixture containing: 5 µl of RNA, 5 µl of H₂O for injection, 4 µl of 5× reverse transcriptase buffer, 2 µl of dNTP (5 mM), 2 µl of oligo 20T (5 µM), 0.5 µl of RNasin (40 IU/µl) and 1.5 µl of AMV-RT (10 IU/µl Promega) was incubated in a thermocycler under the fol-

lowing conditions: 45 min at 42° C., 15 min at 55° C., 5 min at 95° C., and it was then kept at +4° C.

[0411] A first PCR amplification was performed with the pair of primers S/N/F3/+28023 and S/N/R3/-29480.

[0412] The reaction mixture as above for the amplification of the S1 and S2 fragments was incubated in a thermocycler, under the following conditions: an initial step of denaturation at 94° C. for 2 min was followed by 40 cycles comprising a step of denaturation at 94° C. for 20 sec, a step of annealing at 55° C. for 30 sec and then a step of extension at 72° C. for 1 min 30 sec with 10 sec of additional extension at each cycle, and then a final step of extension at 72° C. for 5 min.

[0413] The amplicon obtained at the first PCR amplification was subjected to a second PCR amplification step (nested PCR) with the pairs of primer S/N/F4/+28054 and S/N/R4/-29430 under conditions identical to those of the first amplification.

[0414] The amplification product obtained, corresponding to the cDNA encoding the N protein of the SARS-CoV strain derived from the sample No. 031589, was sequenced with the aid of the primers: S/N/F4/+28054, S/N/R4/-29430, S/N/+28468, S/N/+28918 and S/N/-28607 and cloned as above for the other ORFs, to give the plasmid called SARS-N. The DNA of these clones was isolated and sequenced with the aid of the universal primers M13 sense and M13 antisense, and the primers S/N/+28468, S/N/+28918 and S/N/-28607.

[0415] The sequence of the amplicon representing the cDNA corresponding to ORF-N and including ORF13 and ORF14 (SEQ ID NO: 36) of the SARS-CoV strain derived from the sample No. 031589 does not contain differences in relation to the corresponding sequences of the isolates AY274119.3-Tor2 and AY278741-Urbani. The sequence of the N protein of the SARS-CoV strain derived from the sample No. 031589 corresponds to the sequence SEQ ID NO: 37 in the sequence listing appended as an annex.

[0416] The sequences of ORF13 and 14 of the SARS-CoV strain derived from the sample No. 031589 correspond respectively to the sequences SEQ ID NO: 32 and 34 in the sequence listing appended as an annex.

[0417] The plasmid, called SARS-N, was deposited under the No. I-3048, on Jun. 5, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA encoding the N protein of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above, said sequence corresponding to the nucleotides at positions 28054 to 29430 (SEQ ID NO: 38), with reference to the Genbank sequence accession No. AY274119.3.

2.5) Noncoding 5' and 3' Ends

a) Noncoding 5' end (5'NC)

a₁) Synthesis of the cDNA

[0418] The RNAs derived from the sample 031589, extracted as above, were subjected to reverse transcription under the following conditions:

[0419] The RNA (15 µl) and the primer S/L/-443 (3 µl at the concentration of 5 µM) were incubated for 10 min at 75° C.

[0420] Next, the 5× reverse transcriptase buffer (6 µl, INVITROGEN), 10 mM dNTP (1 µl), 0.1 M DTT (3 µl) were added and the mixture was incubated at 50° C. for 3 min.

[0421] Finally, the reverse transcriptase (3 µl of Superscript®, INVITROGEN) was added to the preceding mixture which was incubated at 50° C. for 1 h 30 min and then at 90° C. for 2 min.

[0422] The cDNA thus obtained was purified with the aid of the QIAquick PCR purification kit (QIAGEN) according to the manufacturer's recommendations.

b₁) Terminal Transferase Reaction (TdT)

[0423] The cDNA (10 µl) is incubated for 2 min at 100° C., stored in ice, and the following are then added: H₂O (2.5 µl), 5× TdT buffer (4 µl, AMERSHAM), 5 mM dATP (2 µl) and TdT (1.5 µl, AMERSHAM). The mixture thus obtained is incubated for 45 min at 37° C. and then for 2 min at 65° C.

[0424] The product obtained is amplified by a first PCR reaction with the aid of the primers: S/L/-225-206 and anchor 14T: 5'-AGATGAATTCGGTAC-CTTTTTTTTTTTTTTT-3' (SEQ ID NO: 68). The amplification conditions are the following: an initial step of denaturation at 94° C. for 2 min is followed by 10 cycles comprising a step of denaturation at 94° C. for 10 sec, a step of annealing at 45° C. for 30 sec and then a step of extension at 72° C. for 30 sec and then by 30 cycles comprising a step of denaturation at 94° C. for 10 sec, a step of annealing at 50° C. for 30 sec and then a step of extension at 72° C. for 30 sec, and then a final step of extension at 72° C. for 5 min.

[0425] The product of the first PCR amplification was subjected to a second amplification step with the aid of the primers: S/L/-204-185 and anchor 14T mentioned above under conditions identical to those of the first amplification. The amplicon thus obtained was purified, sequenced with the aid of the primer S/L/-182-163 and it was then cloned as above for the different ORFs, to give the plasmid called SARS-5'NC. The DNA of this clone was isolated and sequenced with the aid of the universal primers M13 sense and M13 antisense and the primer S/L/-182-163 mentioned above.

[0426] The amplicon representing the cDNA corresponding to the 5'NC end of the SARS-CoV strain derived from the sample recorded under the No. 031589 corresponds to the sequence SEQ ID NO: 72 in the sequence listing appended as an annex; this sequence does not contain differences in relation to the corresponding sequences of the isolates AY274119.3-Tor2 and AY278741-Urbani.

[0427] The plasmid, called SARS-5'NC, was deposited under the No. I-3124, on Nov. 7, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA corresponding to the noncoding 5' end of the genome of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above, said sequence corresponding to the nucleotides at positions 1 to 204 (SEQ ID NO: 39), with reference to the Genbank sequence accession No. AY274119.3.

b) Noncoding 3' end (3'NC)

a₁) Synthesis of the cDNA

[0428] The RNAs derived from the sample 031589, extracted as above, were subjected to reverse transcription, according to the following protocol: the reaction mixture containing: RNA (5 µl), H₂O (5 µl), 5× reverse transcriptase buffer (4 µl), 5 mM dNTP (2 µl), 5 µM Oligo 20T (2 µl), 40 U/µl RNasin (0.5 µl) and 10 IU/µl RT-AMV (1.5 µl, PROMEGA) was incubated in a thermocycler, under the following conditions: 45 min at 42° C., 15 min at 55° C., 5 min at 95° C., and it was then kept at +4° C.

[0429] The cDNA obtained was amplified by a first PCR reaction with the aid of the primers S/N/+28468-28487 and anchor 14T mentioned above. The amplification conditions are the following: an initial step of denaturation at 94° C. for 2 min is followed by 10 cycles comprising a step of denaturation at 94° C. for 20 sec, a step of annealing at 45° C. for 30 sec and then a step of extension at 72° C. for 50 sec and then 30 cycles comprising a step of denaturation at 94° C. for 20 sec, a step of annealing at 50° C. for 30 sec and then a step of extension at 72° C. for 50 sec, and then a final step of extension at 72° C. for 5 min.

[0430] The product of the first PCR amplification was subjected to a second amplification step with the aid of the primers S/N/+28933-28952 and anchor 14T mentioned above, under conditions identical to those of the first amplification. The amplicon thus obtained was purified, sequenced with the aid of the primer S/N/+29257-29278 and cloned as above for the different ORFs, to give the plasmid called SARS-3'NC. The DNA of this clone was isolated and sequenced with the aid of the universal primers M13 sense and M13 antisense and the primer S/N/+29257-29278 mentioned above.

[0431] The amplicon representing the cDNA corresponding to the 3'NC end of the SARS-CoV strain derived from the sample recorded under the No. 031589 corresponds to the sequence SEQ ID NO: 73 in the sequence listing appended as an annex; this sequence does not contain differences in relation to the corresponding sequences of the isolates AY274119.3-Tor2 and AY278741-Urbani.

[0432] The plasmid called SARS-3'NC was deposited under the No. I-3123 on Nov. 7, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA sequence corresponding to the noncoding 3' end of the genome of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above, said sequence corresponding to that situated between the nucleotide at positions 28933 to 29277 (SEQ ID NO: 40), with reference to the Genbank sequence accession No. AY274119.3, ends with a series of nucleotides a.

2.6) ORF1a and ORF1b

[0433] The amplification of the 5' region containing ORF1a and ORF1b of the SARS-CoV genome derived from the sample 031589 was performed by carrying out RT-PCR reactions followed by nested PCRs according to the same principles as those described above for the other ORFs. The amplified fragments overlap over several tenths of bases, thus allowing computer reconstruction of the complete sequence of this part of the genome. On average, the amplified fragments are of two kilobases.

[0434] 14 overlapping fragments, called L0 to L12, were thus amplified with the aid of the following primers:

TABLE II

Primers used for the amplification of the 5' region (ORF1a and ORF1b)				
REGION AMPLIFIED AND SEQUENCED (does not include the primers)	RT-PCR sense primer	RT-PCR antisense primer	Nested PCR sense primer	Nested PCR antisense primer
L0 50-480	S/L0/F1/+30	S/L0/R1/-481		
L1 231-2240	S/L1/F1/+147	S/L1/R1/-2336	S/L1/F2/+211	S/L1/R2/-2241
L2 2156-4167	S/L2/F1/+2033	S/L2/R1/-4192	S/L2/F2/+2136	S/L2/R2/-4168
L3 3913-5324	S/L3bis/F1/+3850	S/L3bis/R1/-5365	S/L3bis/F2/+3892	S/L3bis/R2/-5325
L4b 4952-6023	S/L4b/F1/+4878	S/L4b/R1/-6061	S/L4b/F2/+4932	S/L4b/R2/-6024
L4 5325-7318	S/L4/F1/+5272	S/L4/R1/-7392	S/L4/F2/+5305	S/L4/R2/-7323
L5 7296-9156	S/L5/F1/+7111	S/L5/R1/-9253	S/L5/F2/+7275	S/L5/R2/-9157
L6 9053-11066	S/L6/F1/+8975	S/L6/R1/-11151	S/L6/F2/+9032	S/L6/R2/-11067
L7 10928-12962	S/L7/F1/+10883	S/L7/R1/-13050	S/L7/F2/+10928	S/L7/R2/-12963
L8 12835-14834	S/L8/F1/+12690	S/L8/R1/-14857	S/L8/F2/+12815	S/L8/R2/-14835
L9 14765-16624	S/L9/F1/+14688	S/L9/R1/-16678	S/L9/F2/+14745	S/L9/R2/-16625
L10 16534-18570	S/L10/F1/+16451	S/L10/R1/-18594	S/L10/F2/+16514	S/L10/R2/-18571
L11 18521-20582	S/L11/F1/+18441	S/L11/R1/-20612	S/L11/F2/+18500	S/L11/R2/-20583
L12 20338-22205	S/L12/F1/+20279	S/L12/R1/-22229	S/L12/F2/+20319	S/L12/R2/-22206

All the fragments were amplified under the following conditions, except fragment L0 which was amplified as described above for ORF-M:

RT-PCR: 30 min at 42° C., 15 min at 55° C., 2 min at 94° C., and then the cDNA obtained is amplified under the following conditions: 40 cycles comprising: a step of denaturation at 94° C. for 15 sec, a step of annealing at 58° C. for 30 sec and then a step of extension at 68° C. for 1 min 30 sec, with 5 sec additional extension at each cycle, and then a final step of extension at 68° C. for 7 min.

Nested PCR: An initial step of denaturation at 94° C. for 2 min is followed by 35 cycles comprising: a step of denaturation at 94° C. for 15 sec, a step of annealing at 60° C. for 30 sec and then a step of extension at 72° C. for 1 min 30 sec, with 5 sec of additional extension at each cycle, and then a final step of extension at 72° C. for 7 min.

[0435] The amplification products were sequenced with the aid of the primers defined in table III below:

TABLE III

Primers used for the sequencing of the 5' region (ORF1a and ORF1b)	
Names	Sequences (SEQ ID NO: 76 to 139)
S/L3/+4932	5'-CCACACACAGCTTGTGGATA-3'
S/L4/+6401	5'-CCGAAGTTGTAGGCAATGTC-3'
S/L4/+6964	5'-TTGGTGTCTCTTCTTATTG-3'
S/L4/-6817	5'-CCGGCATCCAAACATAATTT-3'
S/L5/-7633	5'-TGGTCAGTAGGGTTGATTGG-3'
S/L5/-8127	5'-CATCCTTTGTGTCAACATCG-3'
S/L5/-8633	5'-GTCACGAGTGACACCAATCCT-3'
S/L5/+7839	5'-ATGCGACGAGTCTGCTTCTA-3'
S/L5/+8785	5'-TTCATAGTGCCTGGCTTACC-3'
S/L5/+8255	5'-ATCTTGGCGCATGTAITGAC-3'
S/L6/-9422	5'-TGCATTAGCAGCAACAACAT-3'

TABLE III-continued

Primers used for the sequencing of the 5' region (ORF1a and ORF1b)	
Names	Sequences (SEQ ID NO: 76 to 139)
S/L6/-9966	5'-TCTGCAGAACAGCAGAAAGTG-3'
S/L6/-10542	5'-CCTGTGTCAGTTTGTCTGTCA-3'
S/L6/+10677	5'-CCTGTGTCAGTGAAGTACA-3'
S/L6/+10106	5'-ATGTCATTGTCACAGCAGAA-3'
S/L6/+9571	5'-CTTCAATGGTTTGCCATGTT-3'
S/L7/-11271	5'-TGCGAGCTGTCATGAGAATA-3'
S/L7/-11801	5'-AACCAGAGCAGTACCACAG-3'
S/L7/-12383	5'-TTTGGCTGCTGTAGTCAATG-3'
S/L7/+12640	5'-CTACGACAGATGCTCTGTGC-3'
S/L7/+12088	5'-GAGCAGGCTGTAGCTAATGG-3'
S/L7/+11551	5'-TTAGGCTATTGTTGCTGCTG-3'
S/L8/-13160	5'-CAGACAACATGAAGCACCAC-3'
S/L8/-13704	5'-CGCTGACGTGATATATGTGG-3'

TABLE III-continued

Primers used for the sequencing of the 5' region (ORF1a and ORF1b)	
Names	Sequences (SEQ ID NO: 76 to 139)
S/L8/-14284	5'-TGCACAATGAAGGATACACC-3'
S/L8/+14453	5'-ACATAGCTCGCGTCTCAGTT-3'
S/L8/+13968	5'-GGCATTGTAGGCGTACTGAC-3'
S/L8/+13401	5'-GTTTGCGGGTGAAGTGCAG-3'
S/L9/-15098	5'-TAGTGCGCGCTATTGACTTC-3'
S/L9/-15677	5'-CTAAACCTTGAGCCGCATAG-3'
S/L9/-16247	5'-CATGGTCATAGCAGCACTTG-3'
S/L9/+16323	5'-CCAGGTTGTGATGCTACTGAT-3'
S/L9/+15858	5'-CCTTACCCAGATCCATCAAG-3'
S/L9/+15288	5'-CGCAAACATAACACTTGCTG-3'
S/L10/-16914	5'-AGTGTTGGGTACAAGCCAGT-3'
S/L10/-17466	5'-GTTCCAAGGAACATGTCTGG-3'
S/L10/-18022	5'-AGGTGCCTGTGTAGGATGAA-3'
S/L10/+18245	5'-GGGCTGTCTCATGCAACTAGAG-3'
S/L10/+17663	5'-TCTTACACGCAATCTGCTT-3'
S/L10/+17061	5'-TACCAATCTGCTCGCATAGT-3'
S/L11/-18877	5'-GCAAGCAGAAATTAACCTCA-3'
S/L11/-19396	5'-AGCACCACCTAAATTGCATC-3'
S/L11/-20002	5'-TGGTCCCTTTGAAGGTGTTA-3'
S/L11/+20245	5'-TCGAACACATCGTTTATGGA-3'
S/L11/+19611	5'-GAAGCACCTGTTTCCATCAT-3'
S/L11/+19021	5'-ACGATGCTCAGCCATGTAGT-3'
SARS/L1/F3/+800	5'-GAGGTGCAGTCACTCGCTAT-3'
SARS/L1/F4/+1391	5'-CAGAGATTGGACCTGAGCAT-3'
SARS/L1/F5/+1925	5'-CAGCAAACCACTCAATTCCT-3'
SARS/L1/R3/-1674	5'-AAATGATGGCAACCTCTTCA-3'
SARS/L1/R4/-1107	5'-CACGTGGTTGAATGACTTTG-3'
SARS/L1/R5/-520	5'-ATTCTGCAACCAAGTCAAC-3'
SARS/L2/F3/+2664	5'-CGCATTGTCTCCTGGTTTAC-3'
SARS/L2/F4/+3232	5'-GAGATTGAGCCAGAACCAGA-3'
SARS/L2/F5/+3746	5'-ATGAGCAGGTTGTCTATGGAT-3'
SARS/L2/R3/-3579	5'-CTGCCCTTAAAGAAGCTGGATG-3'
SARS/L2/R4/-2991	5'-TTTCTTACCAGCATCATCA-3'
SARS/L2/R5/-2529	5'-CACCGTTCTTGAGAAACAACC-3'
SARS/L3/F3/+4708	5'-TCTTTGGCTGGCTCTTACAG-3'
SARS/L3/F4/+5305	5'-GCTGGTGATGTGCTAACTT-3'
SARS/L3/F5/+5822	5'-CCATCAAGCCTGTGTGCTAT-3'
SARS/L3/R3/-5610	5'-CAGGTGGTGCAGACATCATA-3'
SARS/L3/R4/-4988	5'-AACATCAGCACCATCCAAGT-3'
SARS/L3/R5/-4437	5'-ATCGGACACCATAGTCAACG-3'

[0436] The sequences of the fragments L0 to L12 of the SARS-CoV strain derived from the sample recorded under the No. 031589 correspond respectively to the sequences SEQ ID NO: 41 to SEQ ID NO: 54 in the sequence listing appended as an annex. Among these sequences, only that corresponding to the fragments L5 contains a nucleotide difference in relation to the corresponding sequence of the isolate AY278741-Urbani. This t/c mutation at position 7919 results in a modification of the amino acid sequence of the corresponding protein, encoded by ORF1a: at position 2552, a valine (gtt codon; AY278741) is changed to alanine (gct codon) in the SARS-CoV strain 031589. By contrast, no mutation was identified in relation to the corresponding sequence of the isolate AY274119.3-Urbani. The other fragments do not exhibit differences in relation to the corresponding sequences of the isolates Tor2 and Urbani.

EXAMPLE 2

Production and Purification of the Recombinant N and S Proteins of the SARS-CoV Strain Derived from the Sample Recorded Under the Number 031589

[0437] The entire N protein and two polypeptide fragments of the S protein of the SARS-CoV strain derived from

the sample recorded under the number 031589 were produced in *E. coli*, in the form of fusion proteins comprising an N- or C-terminal polyhistidine tag. In the two S polypeptides, the N- and C-terminal hydrophobic sequences of the S protein (signal peptide: positions 1 to 13 and transmembrane helix: positions 1196 to 1218) were deleted whereas the β helix (positions 565 to 687) and the two motifs of the coiled-coil type (positions 895 to 980 and 1155 to 1186) of the S protein were preserved. These two polypeptides consist of: a long fragment (S_L) corresponding to positions 14 to 1193 of the amino acid sequence of the S protein and a short fragment (S_C) corresponding to positions 475 to 1193 of the amino acid sequence of the S protein.

1) Cloning of the cDNAs N, S_L and S_C into the Expression Vectors pIVEX2.3 and pIVEX2.4

[0438] The cDNAs corresponding to the N protein and to the S_L and S_C fragments were amplified by PCR under standard conditions, with the aid of the DNA polymerase Platinum Pfx® (INVITROGEN). The plasmids SRAS-N and SRAS-S were used as template and the following oligonucleotides as primers:

5'- <u>CCCATATG</u> TCTGATAATGGACCCCAATCAAA C-3'	(N sense, SEQ ID NO: 55)
5'- <u>CCCCCGGGTGCCTGAGTTGAATCAGCAGAAG</u> C-3'	(N antisense, SEQ ID NO: 56)
5'- <u>CCCATATGAGTGACCTTGACCGGTGCACCA</u> C-3'	(S_C sense, SEQ ID NO: 57)
5'- <u>CCCATATGAAACCTTGACCCACCTGCT</u> C-3'	(S_L sense, SEQ ID NO: 58)
5'- <u>CCCCCGGGTTTAATATATTGCTCATATTTTCC</u> C-3'.	(S_C and S_L antisense, SEQ ID NO: 29)

[0439] The sense primers introduce an NdeI site (underlined) while the antisense primers introduce an XmaI site (underlined). The 3 amplification products were column purified (QIAquick PCR Purification kit, QIAGEN) and cloned into an appropriate vector. The plasmid DNA purified from the 3 constructs (QIAfilter Midi Plasmid kit, QIAGEN) was verified by sequencing and digested with the enzymes NdeI and XmaI. The 3 fragments corresponding to the cDNAs N, S_L and S_C were purified on agarose gel and then inserted into the plasmids pIVEX2.3MCS (C-terminal polyhistidine tag) and pIVEX2.4d (N-terminal polyhistidine tag) digested beforehand with the same enzymes. After verification of the constructs, the 6 expression vectors thus obtained (pIV2.3N, pIV2.3 S_C , pIV2.3 S_L , pIV2.4N, pIV2.4 S_C also called pIV2.4 S_1 , pIV2.4 S_L) were then used, on the one hand to test the expression of the proteins in vitro, and on the other hand to transform the bacterial strain BL21(DE3)pDIA17 (NOVAGEN). These constructs encode proteins whose expected molecular mass is the following: pIV2.3N (47174 Da), pIV2.3 S_C (82897 Da), pIV2.3 S_L (132056 Da), pIV2.4N (48996 Da), pIV2.4 S_1 (81076 Da) and pIV2.4 S_L (133877 Da). Bacteria transformed with pIV2.3N were deposited at the CNCM on Oct. 23, 2003, under the number I-3117, and bacteria transformed with pIV2.4 S_1 were deposited at the CNCM on Oct. 23, 2003, under the number I-3118.

2) Analysis of the Expression of the Recombinant Proteins In Vitro and In Vivo

[0440] The expression of recombinant proteins from the 6 recombinant vectors was tested, in a first instance, in a

system in vitro (RTS100, Roche). The proteins produced in vitro, after incubation of the recombinant vectors pIVEX for 4 h at 30° C., in the RTS100 system, were analyzed by Western blotting with the aid of an anti-(his)₆ antibody coupled to peroxidase. The result of expression in vitro (FIG. 1) shows that only the N protein is expressed in large quantities, regardless of the position, N- or C-terminal, of the polyhistidine tag. In a second step, the expression of the N and S proteins was tested in vivo at 30° C. in LB medium in the presence or in the absence of inducer (1 mM IPTG). The N protein is very well produced in this bacterial system (FIG. 2) and is found mainly in a soluble fraction after lysis of the bacteria. By contrast, the long version of S (S_L) is very weakly produced and is completely insoluble (FIG. 3). The short version (S_C) also exhibits a very weak solubility, but an expression level that is much higher than that of the long version. Moreover, the construct S_C fused with a polyhistidine tag at the C-terminal position has a smaller size than that expected. An immunodetection experiment with an anti-polyhistidine antibody has shown that this construct was incomplete. In conclusion, the two constructs, pIV2.3N and pIV2.4S₁, which express respectively the entire N protein fused with the C-terminal polyhistidine tag and the short S protein fused with the N-terminal polyhistidine tag, were selected in order to produce the two proteins in a large quantity so as to purify them. The plasmids pIV2.3N and pIV2.4S₁ were deposited respectively under the No. I-3117 and I-3118 at the CNCM, 25 rue du Docteur Roux, 75724 PARIS 15, on Oct. 23, 2003.

3) Analysis of the Antigenic Activity of the Recombinant Proteins

[0441] The antigenic activity of the N, S_L and S_C proteins was tested by Western blotting with the aid of two serum samples, obtained from the same patient infected with SARS-CoV, collected 8 days (M12) and 29 days (M13) after the onset of the SARS symptoms. The experimental protocol is as described in example 3. The results illustrated by FIG. 4 show (i) the seroconversion of the patient, and (ii) that the N protein possesses a higher antigenic reactivity than the short S protein.

4) Purification of the N Protein from pIV2.3N

[0442] Several experiments for purifying the N protein, produced from the vector pIV2.3N, were carried out according to the following protocol. The bacteria BL21(DE3)pDIA17, transformed with the expression vector pIV2.3N, were cultured at 30° C. in 1 liter of culture medium containing 0.1 mg/ml of ampicillin, and induced with 1 mM IPTG when the cell density equivalent to A₆₀₀=0.8 is reached (about 3 hours). After 2 hours of culture in the presence of inducer, the cells were recovered by centrifugation (10 min at 5000 rpm), resuspended in the lysis buffer (50 mM NaH₂PO₄, 0.3 M NaCl, 20 mM imidazole, pH 8, containing the mixture of protease inhibitors Complete®, Roche), and lysed with the French press (12 000 psi). After centrifugation of the bacterial lysate (15 min at 12 000 rpm), the supernatant (50 ml) was deposited at a flow rate of 1 ml/min on a metal chelation column (15 ml) (Ni-NTA superflow, Qiagen), equilibrated with the lysis buffer. After washing the column with 200 ml of lysis buffer, the N protein was eluted with an imidazole gradient (20→250 mM) in 10 column volumes. The fractions containing the N protein were assembled and analyzed by polyacrylamide gel electrophoresis under denaturing conditions followed by staining with Coomassie blue. The results illus-

trated by FIG. 5 show that the protocol used makes it possible to purify the N protein with a very satisfactory homogeneity (95%) and a mean yield of 15 mg of protein per liter of culture.

5) Purification of the S_C Protein from pIV2.4S_C (pIV2.4S₁)

[0443] The protocol followed for purifying the short S protein is very different from that described above because the protein is highly aggregated in the bacterial system (inclusion bodies). The bacteria BL21(DE3)pDIA17, transformed with the expression vector pIV2.4S₁, were cultured at 30° C. in 1 liter of culture medium containing 0.1 mg/ml of ampicillin, and induced with 1 mM IPTG when the cell density equivalent to A₆₀₀=0.8 is reached (about 3 hours). After 2 hours of culture in the presence of inducer, the cells were recovered by centrifugation (10 min at 5000 rpm), resuspended in the lysis buffer (0.1 M Tris-HCl, 1 mM EDTA, pH 7.5), and lysed with the French press (1200 psi). After centrifugation of the bacterial lysate (15 min at 12 000 rpm), the pellet was resuspended in 25 ml of lysis buffer containing 2% Triton X100 and 10 mM β-mercaptoethanol, and then centrifuged for 20 min at 12 000 rpm. The pellet was resuspended in 10 mM Tris-HCl buffer containing 7 M urea, and gently stirred for 30 min at room temperature. This final washing of the inclusion bodies with 7 M urea is necessary in order to remove most of the *E. coli* membrane proteins which co-sediment with the aggregated S_C protein. After a final centrifugation for 20 min at 12 000 rpm, the final pellet is resuspended in the 10 mM Tris-HCl buffer. The electrophoretic analysis of this preparation (FIG. 6) shows that the short S protein may be purified with a satisfactory homogeneity (about 90%) from the inclusion bodies (insoluble extract).

EXAMPLE 3

Immunodominance of the N Protein

[0444] The reactivity of the antibodies present in the serum of patients suffering from atypical pneumopathy caused by the SARS-associated coronavirus (SARS-CoV), toward the various proteins of this virus, was analyzed by Western blotting under the conditions described below.

1) Materials

a) Lysate of Cells Infected with SARS-CoV

[0445] Vero E6 cells (2×10⁶) were infected with SARS-CoV (isolate recorded under the number FFM/MA104) at a multiplicity of infection (M.O.I.) of 10⁻¹ or 10⁻² and then incubated in DMEM medium containing 2% FCS, at 35° C. in an atmosphere containing 5% CO₂. 48 hours later, the cellular lawn was washed with PBS and then lysed with 500 μl of loading buffer prepared according to Laemmli and containing β-mercaptoethanol. The samples were then boiled for 10 minutes and then sonicated for 3 times 20 seconds.

b) Antibodies

b₁) Serum from a Patient Suffering from Atypical Pneumopathy

[0446] The serum designated by a reference at the National Reference Center for Influenza Viruses (Northern region) under the No. 20033168 is that from a French patient suffering from atypical pneumopathy caused by SARS-CoV

collected on day 38 after the onset of the symptoms; the diagnosis of SARS-CoV infection was performed by nested RT-PCR and quantitative PCR.

b₂) Monospecific Rabbit Polyclonal Sera Directed Against the N Protein or the S Protein

[0447] The sera are those produced from the recombinant N and S_C proteins (example 2), according to the immunization protocol described in example 4; they are the rabbit P13097 serum (anti-N serum) and the rabbit P11135 serum (anti-S serum).

2) Method

[0448] 20 µl of lysate of cells infected with SARS-CoV at M.O.I. values of 10⁻¹ and 10⁻² and, as a control, 20 µl of a lysate of noninfected cells (mock) were separated on 10% SDS polyacrylamide gel and then transferred onto a nitrocellulose membrane. After blocking in a solution of PBS/5% milk/0.1% Tween and washing in PBS/0.1% Tween, this membrane was hybridized overnight at 4° C. with: (i) the immune serum No. 20033168 diluted 1/300, 1/1000 and 1/3000 in the buffer PBS/1% BSA/0.1% Tween, (ii) the rabbit P13097 serum (anti-N serum) diluted 1/50 000 in the same buffer and (iii) the rabbit P11135 serum (anti-S serum) diluted 1/10 000 in the same buffer. After washing in PBS/Tween, a secondary hybridization was performed with the aid of either sheep polyclonal antibodies directed against the heavy and light chains of human G immunoglobulins and coupled with peroxidase (NA933V, Amersham), or of donkey polyclonal antibodies directed against the heavy and light chains of the rabbit G immunoglobulins and coupled with peroxidase (NA934V, Amersham). The bound antibodies were visualized with the aid of the ECL+ kit (Amersham) and of Hyperfilm MP autoradiography films (Amersham). A molecular mass ladder (kDa) is presented in the figure.

3) Results

[0449] FIG. 7 shows that three polypeptides of apparent molecular mass 35, 55 and 200 kDa are specifically detected in the extracts of cells infected with SARS-CoV.

[0450] In order to identify these polypeptides, two other immunoblots (FIG. 8) were prepared on the same samples and under the same conditions with rabbit polyclonal antibodies specific for the nucleoprotein N (rabbit P13097, FIG. 8A) and for the spike protein S (rabbit P11135, FIG. 8B). This experiment shows that the 200 kDa polypeptide corresponds to the SARS-CoV spike glycoprotein S, that the 55 kDa polypeptide corresponds to the nucleoprotein N while the 35 kDa polypeptide probably represents a truncated or degraded form of N.

[0451] The data presented in FIG. 7 therefore show that the serum 20033168 strongly reacts with N and a lot more weakly with the SARS-CoV S since the 35 and 55 kDa polypeptides are visualized in the form of intense bands for 1/300, 1/1000 and 1/3000 dilutions of the immunosera whereas the 200 kDa polypeptide is only weakly visualized for a dilution of 1/300. It is also possible to note that no other SARS-CoV polypeptide is detected for dilutions greater than 1/300 of the serum 20033168.

[0452] This experiment indicates that the antibody response specific for the SARS-CoV N dominates the antibody responses specific for the other SARS-CoV polypeptides and in particular the antibody response directed against

the S glycoprotein. It indicates an immuno-dominance of the nucleoprotein N during human infections with SARS-CoV.

EXAMPLE 4

Preparation of Monospecific Polyclonal Antibodies Directed Against the SRAS-associated Coronavirus (SARS-CoV) N and S Proteins

1) Materials and Method

[0453] Three rabbits (P13097, P13081, P13031) were immunized with the purified recombinant polypeptide corresponding to the entire nucleoprotein (N), prepared according to the protocol described in example 2. After a first injection of 0.35 mg per rabbit of protein emulsified in complete Freund's adjuvant (intradermal route), the animals received 3 booster injections at 3 and then 4 weeks' interval, of 0.35 mg of recombinant protein emulsified in incomplete Freund's adjuvant.

[0454] Three rabbits (P11135, P13042, P14001) were immunized with the recombinant polypeptide corresponding to the short fragment of the S protein (S_C) produced as described in example 2. As this polypeptide is found mainly in the form of inclusion bodies in the bacterial cytoplasm, the animals received 4 intradermal injections at 3-4 weeks' interval of a preparation of inclusion bodies corresponding to 0.5 mg of recombinant protein emulsified in incomplete Freund's adjuvant. The first 3 injections were made with a preparation of inclusion bodies prepared according to the protocol described in example 2, while the fourth injection was made with a preparation of inclusion bodies which were prepared according to the protocol described in example 2 and then purified on sucrose gradient and washed in 2% Triton X100.

[0455] For each rabbit, a preimmune (p.i.) serum was prepared before the first immunization and an immune serum (I.S.) 5 weeks after the fourth immunization.

[0456] In a first instance, the reactivity of the sera was analyzed by ELISA test on preparations of recombinant proteins similar to those used for the immunizations; the ELISA tests were carried out according to the protocol and with the reagents as described in example 6.

[0457] In a second instance, the reactivity of the sera was analyzed by preparing an immunoblot (Western blot) of a lysate of cells infected with SARS-CoV, according to the protocol as described in example 3.

2) Results

[0458] The ELISA tests (FIG. 9) demonstrate that the preparations of recombinant N protein and of inclusion bodies of the short fragment of the S protein (S_C) are immunogenic in animals and that the titer of the immune sera is high (more than 1/25 000).

[0459] The immunoblot (FIG. 8) shows that the rabbit P13097 immune serum recognizes two polypeptides present in the lysates of cells infected with SARS-CoV: a polypeptide whose apparent molecular mass (50-55 kDa based on experiments) is compatible with that of the nucleoprotein N (422 residues, predicted molecular mass of 46 kDa) and a polypeptide of 35 kDa, which probably represents a truncated or degraded form of N.

[0460] This experiment also shows that the rabbit P11135 serum mainly recognizes a polypeptide whose apparent molecular mass (180-220 kDa based on experiments) is compatible with a glycosylated form of S (1255 residues, nonglycosylated polypeptide chain of 139 kDa), as well as lighter polypeptides, which probably represent truncated and/or nonglycosylated forms of S.

[0461] In conclusion, all these experiments demonstrate that the recombinant polypeptides expressed in *E. coli* and corresponding to the SARS-CoV N and S proteins make it possible to induce, in animals, polyclonal antibodies capable of recognizing the native forms of these proteins.

EXAMPLE 5

Preparation of Monospecific Polyclonal Antibodies Directed Against the SARS-associated Coronavirus (SARS-CoV) M and E Proteins

1) Analysis of the Structure of the M and E Proteins

a) E Protein

[0462] The structure of the SARS-CoV E protein (76 amino acids) was analyzed in silico, with the aid of various software packages such as signalP v1.1, NetNGlyc 1.0, THMM 1.0 and 2.0 (Krogh et al., 2001, J. Mol. Biol., 305(3):567-580) or alternatively TOPPED (von Heijne, 1992, J. Mol. Biol. 225, 487-494). The analysis shows that this nonglycosylated polypeptide is a type 1 membrane protein, containing a single transmembrane helix (aa 12-34 according to THMM), and in which the majority of the hydrophilic domain (42 residues) is located at the C-terminal end and probably inside the viral particle (endodomain). It is possible to note an inversion in the topology predicted by versions 1.0 (N-ter is external) and 2.0 (N-ter is internal) of the THMM software, but that other algorithms, in particular TOPPED and THUMBUP (Zhou et Zhou, 2003, Protein Science 12:1547-1555) confirm an external location of the N-terminal end of E.

b) M Protein

[0463] A similar analysis carried out on the SARS-CoV M protein (221 amino acids) shows that this polypeptide does not possess a signal peptide (according to the software signalP v1.1) but three transmembrane domains (residues 15-37, 50-72, 77-99 according to THMM2.0) and a large hydrophilic domain (aa 100-221) located inside the viral particle (endodomain). It is probably glycosylated on the asparagine at position 4 (according to NetNGlyc 1.0).

[0464] Thus, in agreement with the experimental data known for the other coronaviruses, it is remarkable that the two M and E proteins exhibit endodomains corresponding to the majority of the polypeptides and of the ectodomains that are very small in size.

[0465] The ectodomain of E probably corresponds to residues 1 to 11 or 1 to 12 of the protein: MYSFVSEETGT(L), SEQ ID NO: 70. Indeed, the probability associated with the transmembrane location of residue 12 is intermediate (0.56 according to THMM 2.0).

[0466] The ectodomain of M probably corresponds to residues 2 to 14 of the protein: ADNGTITVEELKQ, SEQ ID NO: 69. Indeed, the N-terminal methionine of M is very probably cleaved from the mature polypep-

tide because the residue at position 2 is an alanine (Varshavsky, 1996, 93:12142-12149).

[0467] Moreover, the analysis of the hydrophobicity (Kyte & Doolittle, Hopp & Woods) of the E protein demonstrates that the C-terminal end of the endodomain of E is hydrophilic and therefore probably exposed at the surface of this domain. Thus, a synthetic peptide corresponding to this end is a good immunogenic candidate for inducing, in animals, antibodies directed against the endodomain of E. Consequently, a peptide corresponding to 24 C-terminal residues of E was synthesized.

2) Preparation of Antibodies Directed Against the Ectodomain of the M and E Proteins and the Endodomain of the E Protein

[0468] The peptides M2-14 (ADNGTITVEELKQ, SEQ ID NO: 69), E1-12 (MYSFVSEETGT(L), SEQ ID NO: 70) and E53-76 (KPTVYVYSRV KNLNSSEGVP DLLV, SEQ ID NO: 71) were synthesized by Neosystem. They were coupled with KLH (Keyhole Limpet Hemocyanin) with the aid of MBS (m-maleimido-benzoyl-N-hydroxysuccinimide ester) via a cysteine added during the synthesis either at the N-terminus of the peptide (case for E53-76) or at the C-terminus (case of M2-14 and E1-12).

[0469] Two rabbits were immunized with each of the conjugates, according to the following immunization protocol: after a first injection of 0.5 mg of peptide coupled with KLH and emulsified in complete Freund's adjuvant (intradermal route), the animals receive 2 to 4 booster injections at 3 or 4 weeks' interval of 0.25 mg of peptide coupled to KLH and emulsified in incomplete Freund's adjuvant.

[0470] For each rabbit, a preimmune (p.i.) serum was prepared before the first immunization and an immune serum (I.S.) is prepared 3 to 5 weeks after the booster injections.

[0471] The reactivity of the sera was analyzed by Western blotting with the aid of extracts of cells infected with SARS-CoV (FIG. 43B) or with the aid of extracts of cells infected with a recombinant vaccinia virus expressing the protein E (VV-TG-E, FIG. 43A) or M (VV-TN-M, FIG. 43C) of the SARS-CoV 031589 isolate.

[0472] The immune sera of the rabbits 22234 and 22240, immunized with the conjugate KLH-E53-76, recognize a polypeptide of about 9 to 10 kD, which is present in the extracts of cells infected with SARS-CoV but absent from the extracts of noninfected cells (FIG. 43B). The apparent mass of this polypeptide is compatible with the predicted mass of the E protein, which is 8.4 kD. Similarly, the immune serum of the rabbit 20047, immunized with the conjugate KLH-E1-12, recognizes a polypeptide present in the extracts of cells infected with the VV-TG-E virus, whose apparent molar mass is compatible with that of the E protein (FIG. 43A).

[0473] The immune serum of the rabbits 20013 and 20080, immunized with the conjugate KLH-M2-14, recognizes a polypeptide present in the extracts of cells infected with the VV-TN-M virus (FIG. 43C), whose apparent molar mass (about 18 kD) is compatible with that of the glycoprotein M, which is 25.1 kD and has a high iso-electric point (9.1 for the naked polypeptide).

[0474] These results demonstrate that the peptides E1-12 and E53-76, on the one hand, and the peptide M2-14, on the other hand, make it possible to induce, in animals, polyclonal antibodies capable of recognizing the native forms of the SARS-CoV E and M proteins, respectively.

EXAMPLE 6

Analysis of the ELISA Reactivity of the Recombinant N Protein Toward Sera from Patients Suffering from SARS

1) Materials

[0475] The antigen used to prepare the solid phases is the purified recombinant nucleoprotein N prepared according to the protocol described in example 2.

[0476] The sera to be tested (table IV) were chosen on the basis of the results of analysis of their reactivity by immunofluorescence (IF-SARS titer), toward cells infected with SARS-CoV.

TABLE IV

Sera tested by ELISA				
Reference	Serum No.	Type of serum	Date of the serum***	IF-SARS titer
3050	A	Control	na*	nt**
3048	B	Control	na	nt
033168	D	Patient 1-SARS	Apr. 27, 2003 (D38)	320
033397	E	Patient-1 SARS	May 11, 2005 (D52)	320
032632	F	Patient-2 SARS	Mar. 21, 2003 (D17)	2500
032791	G	Patient-3 SARS	Apr. 04, 2003 (D3)	<40
033258	H	Patient-3 SARS	Apr. 28, 2003 (D27)	160

*na: not applicable.

**nt: not tested.

***the dates indicated correspond to the number of days after the onset of the SARS symptoms.

2) Method

[0477] The N protein (100 μ l) diluted at various concentrations in 0.1 M carbonate buffer, pH 9.6 (1, 2 or 4 μ g/ml) is distributed into the wells of ELISA plates, and then the plates are incubated overnight at laboratory temperature. The plates are washed with PBS-Tween buffer saturated with PBS-skimmed milk-sucrose (5%) buffer. The test sera (100 μ l), diluted beforehand (1/50, 1/100, 1/200, 1/400, 1/800, 1/1600 and 1/3200) are added and then the plates are incubated for 1 h at 37° C. After 3 washings, the peroxidase-labeled anti-human IgG conjugate (reference 209-035-098, JACKSON) diluted 1/18 000 is added and then the plates are incubated for 1 h at 37° C. After 4 washings, the chromogen (TMB) and the substrate (H₂O₂) are added and the plates are incubated for 30 min at room temperature, protected from light. The reaction is then stopped and then the absorbance at 450 nm is measured with the aid of an automated reader.

3) Results

[0478] The ELISA tests (FIG. 10) demonstrate that the recombinant N protein preparation is specifically recognized by the antibodies of sera from patients suffering from SARS collected in the late phase of the infection (≥ 17 days after the onset of the symptoms) whereas it is not significantly recognized by the antibodies of a patient's serum collected

in the early phase of the infection (3 days after the onset of the symptoms) or by control sera from subjects not suffering from SARS.

EXAMPLE 7

ELISA Tests Prepared for a Very Specific and Sensitive Detection of a SARS-associated Coronavirus Infection, from Sera of Patients

1) Indirect ELISA IgG Test

a) Reagents

Preparation of the Plates

[0479] The plates are sensitized with a solution of N protein at 2 μ g/ml in a 10 mM PBS buffer, pH 7.2, phenol red at 0.25 ml/l. 100 μ l of solution are deposited in the wells and left to incubate at room temperature overnight. Saturation is obtained by prewashing in 10 mM PBS/0.1% Tween buffer, followed by washing with a saturation solution PBS, 25% milk/sucrose.

Diluent Sera

[0480] Buffer 0.48 g/l TRIS, 10 mM PBS, 3.7 g/l EDTA, 15% v/v milk, pH 6.7

Diluent Conjugate

[0481] Citrate buffer (15 g/l), 0.5% Tween, 25% bovine serum, 12% NaCl, 6% v/v skimmed milk pH 6.5

Conjugate

[0482] 50 \times anti-human IgG conjugate, marketed by Bio-Rad: *Platelia H. pylori* kit ref 72778

Other Solutions:

[0483] Washing solution R2, solutions for visualizing with TMB R8 diluent, R9 chromogen, R10 stopping solution: reagents marketed by Bio-Rad (e.g.: *Platelia pylori* kit, ref 72778)

b) Procedure

[0484] Dilute the sera 1/200 in the sample diluent

[0485] Distribute 100 μ l/well

[0486] Incubation 1 h at 37° C.

[0487] 3 washings in 10 \times WASHING solution R2 diluted beforehand 10-fold in demineralized water (i.e., 1 \times washing solution)

[0488] Distribute 100 μ l of conjugate (50 \times conjugate to be diluted immediately before use in the diluent conjugate provided)

[0489] Incubation 1 h at 37° C.

[0490] 4 washings in 1 \times washing solution

[0491] Distribute 200 μ l/well of visualization solution (to be diluted immediately before use e.g.: 1 ml of R9 in 10 ml of R8)

[0492] Incubation for 30 min at room temperature in the dark

[0493] Stop the reaction with 100 μ l/well of R10

[0494] READING at 450/620 nm

[0495] The results can be interpreted by taking a THRESHOLD serum giving a response above which the sera tested would be considered as positive. This serum is chosen and diluted so as to give a significantly higher signal than the background noise.

2) Double Epitope ELISA Test

a) Reagents

Preparation of the Plates

[0496] The plates are sensitized with a solution of N protein at 1 µg/ml in a 10 mM PBS buffer, pH 7.2, phenol red at 0.25 ml/l. 100 µl of solution are deposited in the wells and left to incubate at room temperature overnight. Saturation is obtained by prewashing in 10 mM PBS/0.1% Tween buffer, followed by washing with a saturation solution 10 mM PBS, 25% (V/V) milk.

Diluent Sera and Conjugate

[0497] Buffer 50 mM TRIS saline, pH 8, 2% milk

Conjugate

[0498] This is the purified recombinant N protein coupled with peroxidase according to the Nakane protocol (Nakane P. K. and Kawaoi A.; (1974): *Peroxydase-labeled antibody, a new method of conjugation. The Journal of Histochemistry and Cytochemistry* Vol. 22, N) 23, pp. 1084-1091), in respective molar ratios 1/2. This ProtN POD conjugate is used at a concentration of 2 µg/ml in serum/conjugate diluent.

Other Solutions:

[0499] Washing solution R2, solutions for visualization with TMB R8, diluent, R9 chromogen, R10 stopping solution: reagents marketed by Bio-Rad (e.g. *Platelia pylori* kit, ref 72778).

b) Procedure

1st Step in "Predilution" Plate

[0500] Dilute each serum 1/5 in the predilution plate (48 µl of diluent+12 µl of serum).

[0501] After having diluted all the sera, distribute 60 µl of conjugate.

[0502] Where appropriate, the serum+conjugate mix is left to incubate.

2nd Step in "Reaction" Plate

[0503] Transfer 100 µl of mixture/well into the reaction plate

[0504] Incubation 1 h 37° C.

[0505] 5 washings in 10× WASHING solution R2 diluted 10-fold beforehand in demineralized water (→1× washing solution)

[0506] Distribute 200 µl/well of visualization solution (to be diluted immediately before use e.g.: 1 ml of R9 in 10 ml of R8)

[0507] Incubation 30 min at room temperature and protected from light

[0508] Stop the reaction with 100 µl/well of R10

[0509] READING at 450/620 nm

[0510] Likewise as for the indirect ELISA test, the results can be interpreted using a "threshold value" serum. Any serum having a response greater than the threshold value serum will be considered as positive.

2) Results

[0511] The sera of patients classified as probable cases of SARS from the French hospital of Hanoi, Vietnam or in relation with the French hospital of Hanoi (JYK) were analyzed using the indirect IgG-N test and the double epitope N test.

[0512] The results of the indirect IgG-N test (FIGS. 14 and 15) and double epitope N test (FIGS. 16 and 17) show an excellent correlation between them and with an indirect ELISA test comparing the reactivity of the sera toward a lysate of VeroE6 cells infected or not infected with SARS-CoV (ELISA-SARS-CoV lysate; see table V below). All the sera collected 12 days or more after the onset of the symptoms were found to be positive, including in patients for whom it had not been possible to document the SARS-CoV virus infection by analyzing respiratory samples by RT-PCR, probably because of a sample being collected too late during the infection (\geq D12). In the case of the patient TTH for whom a nasal sample collected on D7 was found to be negative by RT-PCR, the quality of the sample may be in question.

[0513] Some sera were found to be negative whereas the presence of SARS-CoV was detected by RT-PCR. They are in all cases early sera collected less than 10 days after the onset of the symptoms (e.g.: serum # 032637). In the case of a patient PTTH (serum # 032673), only a suspicion of SARS was raised at the time the samples were collected.

[0514] In conclusion, the indirect IgG-N and N-double epitope serological tests make it possible to document the SARS-CoV infection in all the patients for the sera collected 12 days or more after the infection.

TABLE V

Results of the ELISA tests						
Sample Num	Patient	Day	PCR-SARS (1)	ELISA SARS-CoV lysate (2)	IgG-N (2nd series)	2Xepitope (2nd series)
033168	JYK	38	POS	+++	>5000	NT
033597	JYK	74	POS	NT	~5000	NT
032552	VTT	8	NEG-D3&D8&D12	NEG	<200	<5
032544	CTP	16	NEG-D16&D20	++	>5000	>>20

TABLE V-continued

Results of the ELISA tests						
Sample Num	Patient	Day	PCR-SARS (1)	ELISA SARS-CoV lysate (2)	IgG-N (2nd series)	2Xepitope (2nd series)
032546	CJF	15	NEG	++	>5000	>>20
			D15&D19			
032548	PTL	17	NEG	++	>5000	>>20
			D17&D21			
032550	NTH	17	NEG-D17&D21	++	>5000	>>20
032553	VTT	8	NEG-	NEG	<200	<5
			D3&D8&D12			
032554	NTBV	4	POS	NEG	<200	<5
032555	NTBV	4	POS	NEG	<200	<5
032564	NTP	15	POS	++	>5000	>>20
032629	NVH	4	POS	NEG	<200	<5
032631	BTIX	9	POS	NEG	<200	<5
032635	NHH	4	POS	NEG	<200	<5
032637	NHB	10	POS	NEG	<200	<5
032642	BTIX	9	POS	NEG	<200	<5
032643	LTDH	1	POS	NEG	<200	<5
032644	NTBV	4	POS	NEG	<200	<5
032646	TTH	12	NEG	++	>5000	>>20
			D7&D12&D16			
032647	DTH	17	NEG	++	>5000	>>20
			D17&D21			
032648	NNT	15	NEG	++	>5000	>>20
			D15&D19			
032649	PTH	17	NEG	++	>5000	>>20
			D17&D21			
032672	LVV	16	NEG	+	>5000	>>20
			D16&D20			
032673	PTTH	NA	NEG	NEG	<200	<5
032674	PNB	17	NEG	++	>5000	>>20
			D17&D21			
032682	VTH	12	NEG	++	>5000	>>20
			D12&D16			
032683	DTV	17	NEG	+	>1000	>>20
			D17&D21			

Remarks:

(1): The RT-PCR analyses were carried out by nested RT-PCR BNI, LC Artus and LC-N on nasal or pharyngeal swabs; POS means that at least one sample was found to be positive in this patient.

(2): The reactivity of the sera in the ELISA test using a lysate of cells infected with SARS-CoV was classified as very highly reactive (+++), highly reactive (++), reactive (+) and negative according to the OD value obtained at the dilutions tested.

EXAMPLE 8

Detection of SARS-associated Coronavirus (SARS-CoV) by RT-PCR

1) Real Time Development of RT-PCR Conditions with the Aid of Primers Specific for the Gene for the Nucleocapsid Protein—"Light Cycler N" Test

a) Design of the Primers and Probes

[0515] The primers and probes were designed from the sequence of the genome of the SARS-CoV strain derived from the sample recorded under the number 031589, with the aid of the programme "Light Cycler Probe Design (Roche)". Thus, the following two series of primers and probes were selected:

series 1 (SEQ ID NO: 60, 61, 64, 65):

sense primer:

N/+28507:

5'-GGC ATC GTA TGG GTT G-3'

[28507-28522]

-continued

antisense primer:

N/-/28774:

5'-CAG TTT CAC CAC CTC C-3'

[28774-28759]

probe 1:

5'-GGC ACC CGC AAT CCT AAT AAC AAT

GC-fluorescein 3'

[28561-28586]

probe 2:

5' Red705-GCC ACC GTG CTA CAA CTT

CCT-phosphate

[28588-28608]

series 2 (SEQ ID NO: 62, 63, 66, 67)

sense primer:

N/+28375:

5'-GGC TAC TAC CGA AGA G-3'

[28375-28390]

antisense primer:

N/-/28702:

5'-AAT TAC CGC GAC TAC G-3'

[28702-28687]

-continued

probe 1:
SARS/N/FL:
5'-ATA CAC CCA AAG ACC ACA TTG GC- [28541-28563]
fluorescein 3'

probe 2:
SARS/N/LC705:
5' Red705-CCC GCA ATC CTA ATA ACA ATG [28565-28589]
CTG C-phosphate 3'

b) Analysis of the Efficacy of the Two Primer Pairs

[0516] In order to test the respective efficacy of the two pairs of primers, an RT-PCR amplification was carried out on a synthetic RNA corresponding to nucleotides 28054-29430 of the genome of the SARS-CoV strain derived from the sample recorded under the number 031589 and containing the sequence of the N gene.

[0517] More specifically:

[0518] This synthetic RNA was prepared by in vitro transcription with the aid of the T7 phage RNA polymerase, of a DNA template obtained by linearization of the plasmid SRAS-N with the enzyme Bam H1. After eliminating the DNA template by digestion with the aid of DNase 1, the synthetic RNAs are purified by a phenol-chloroform extraction, followed by two successive precipitations in ammonium acetate and isopropanol. They are then quantified by measuring the absorbance at 260 nm and their quality is checked by the ratio of the absorbances at 260 and 280 nm and by agarose gel electrophoresis. Thus, the concentration of the synthetic RNA preparation used for these studies is 1.6 mg/ml, which corresponds to 2.1×10^{15} copies/ml of RNA.

[0519] Decreasing quantities of synthetic RNA were amplified by RT-PCR with the aid of the "Superscript™ One-Step RT-PCR with Platinum® Taq" kit and the pairs of primers No. 1 (N/+28507, N/-28774) (FIG. 1A) and No. 2 (N/+28375, N/-28702) (FIG. 1B), according to the supplier's instructions. The amplification conditions used are the following: the cDNA was synthesized by incubation for 30 min at 45° C., 15 min at 55° C. and then 2 min at 94° C. and it was then amplified by 5 cycles comprising: a step of denaturation at 94° C. for 15 sec, a step of annealing at 45° C. for 30 sec and then a step of extension at 72° C. for 30 sec, followed by 35 cycles comprising: a step of denaturation at 94° C. for 15 sec, a step of annealing at 55° C. for 30 sec and then a step of extension at 72° C. for 30 sec, with 2 sec of additional extension at each cycle, and a final step of extension at 72° C. for 5 min. The amplification products obtained were then kept at 10° C.

[0520] The results presented in FIG. 11 show that the pair of primers No. 2 (N/+28375, N/-28702) makes it possible to detect up to 10 copies of RNA (band of weak intensity) or 10^2 copies (band of good intensity) against 10^4 copies for the pair of primers No. 1 (N/+28507, N/-28774). The amplicons are respectively 268 bp (pair 1) and 328 bp (pair 2).

c) Development of Real Time RT-PCR

[0521] A real time RT-PCR was developed with the aid of the pair of primers No. 2 and of the pair of probes consisting of SRAS/N/FL and SRAS/N/LC705 (FIG. 2).

[0522] The amplification was carried out on a LightCycler™ (Roche) with the aid of the "Light Cycler RNA Amplification Kit Hybridization Probes" kit (reference 2 015 145, Roche) under the following optimized conditions. A reaction mixture containing: H₂O (6.8 µl), 25 mM MgCl₂ (0.8 µl, 4 µM Mg²⁺ final), 5× reaction mixture (4 µl), 3 µM probe SRAS/N/FL (0.5 µl, 0.075 µM final), 3 µM probe SRAS/N/LC705 (0.5 µl, 0.075 µM final), 10 µM primer N/+28375 (1 µl, 0.5 µM final), 10 µM primer N/-28702 (1 µl, 0.5 µM final), enzyme mixture (0.4 µl) and sample (viral RNA, 5 µl) was amplified according to the following program:

<u>Reverse transcription:</u>			
50° C.	10:00 min	analysis mode: none	
<u>Denaturation:</u>			
95° C.	30 sec × 1	analysis mode: none	
<u>Amplification:</u>			
95° C.	2 sec	analysis mode: quantification* thermal ramp 2.0° C./sec	} ×45
50° C.	15 sec		
72° C.	13 sec		
<u>Annealing:</u>			
40° C.	30 sec × 1	analysis mode: none	

*The fluorescence is measured at the end of the annealing and at each cycle (in SINGLE mode).

[0523] The results presented in FIG. 12 show that this real time RT-PCR is very sensitive since it makes it possible to detect 10^2 copies of synthetic RNA in 100% of the 5 samples analyzed (29/29 samples in 8 experiments) and up to 10 copies of RNA in 100% of the 5 samples analyzed (40/45 samples in 8 experiments). It also shows that this RT-PCR makes it possible to detect the presence of the SARS-CoV genome in a sample and to quantify the number of genomes present. By way of example, the viral RNA of a SARS-CoV stock cultured on Vero E6 cells was extracted with the aid of the "QIamp viral RNA extraction" kit (Qiagen), diluted to 0.05×10^{-14} and analyzed by real time RT-PCR according to the protocol described above; the analysis presented in FIG. 12 shows that this virus stock contains 6.5×10^9 genome-equivalents/ml (geq/ml), which is entirely similar to the 1.0×10^{10} geq/ml value measured with the aid of the "RealArt™ HPA-Coronavirus LC RT PCR Reagents" kit marketed by Artus.

2) Development of Nested RT-PCR Conditions Targeting the Gene for RNA Polymerase—"CDC (Centers for Disease Control and Prevention)/IP Nested RT-PCR" Test

a) Extraction of the Viral RNA

[0524] Clinical sample: QIamp viral RNA Mini Kit (QIAGEN) according to the manufacturer's instructions, or an equivalent technique. The RNA is eluted in a volume of 60 µl.

b) "SNE/SAR" Nested RT-PCR

First Step: "SNE" Coupled RT-PCR

[0525] The Invitrogen "Superscript™ One-Step RT-PCR with Platinum® Taq" kit was used, but the "Titan" kit from Roche Boehringer can be used in its place with similar results.

[0526] Oligonucleotides:

SNE-S1
5' GGT TGG GAT TAT CCA AAA TGT GA 3'

SNE-AS1
5' GCA TCA TCA GAA AGA ATC ATC ATG 3'

→ Expected size: 440 bp

[0527] 1. Prepare a mix:

H2O	6.5 µl
Reaction mix 2X	12.5 µl
Oligo SNE-S1 50 µM	0.2 µl
Oligo SNE-AS1 50 µM	0.2 µl
RNAsin 40 U/µl	0.12 µl
RT/Platinum Taq mix	0.5 µl

[0528] 2. To 20 µl of the mix, add 5 µl of RNA and carry out the amplification on a thermocycler (ABI 9600 conditions):

2.1	45° C.	30 min.		
	55° C.	15 min.		
	94° C.	2 min.		
2.2.	94° C.	15 sec.	}	×5 cycles
	45° C.	30 sec.		
	72° C.	30 sec.		
2.3.	94° C.	15 sec.	}	×35 cycles
	55° C.	30 sec.		
	72° C.	30 sec. + 2 sec./cycle		
2.4.	72° C.	5 min.		
2.5	10° C.	∞		

Storage at +4° C.

[0529] The RNAsin (N2511/N2515) from Promega was used as RNase inhibitors.

[0530] Synthetic RNAs served as positive control. As the control, 10³, 10² and 10 copies of synthetic RNA_{SNE} were amplified in each experiment.

Second Step: "SAR" Nested PCR

[0531] Oligonucleotides:

SAR1-S
5' CCT CTC TTG TTC TTG CTC GCA 3'

SAR1-AS
5' TAT AGT GAG CCG CCA CAC ATG 3'

→ Expected size: 121 bp

[0532] 1. Prepare a mix:

H2O	35.8 µl
Taq buffer 10X	5 µl
MgCl ₂ 25 mM	4 µl
Mix dNTPs 5 mM	2 µl
Oligo SAR1-S 50 µM	0.5 µl
Oligo SAR1-AS 50 µM	0.5 µl
Taq DNA pol 5 U/µl	0.25 µl

[0533] AmpliTaq DNA Pol from Applied Biosystems was used (10× buffer without MgCl₂, ref 27216601).

[0534] 2. To 48 µl of the mix, add 2 µl of the product from the first PCR and carry out the amplification (ABI 9600 conditions):

2.1.	94° C.	2 min.		
2.2.	94° C.	30 sec.	}	×5 cycles
	45° C.	45 sec.		
	72° C.	30 sec.		
2.3.	94° C.	30 sec.	}	×35 cycles
	55° C.	30 sec.		
	72° C.	30 sec. + 1 sec./cycle		
2.4.	72° C.	5 min.		
2.5	10° C.	∞		

[0535] 3. Analyze 10 µl of the reaction product on "low-melting" gel (Seakem GTG type) containing 3% agarose.

[0536] The sensitivity of the nested test is routinely, under the conditions described, 10 copies of RNA.

[0537] 4. The fragments can then be purified on QIAquick PCR kit (QIAGEN) and sequenced with the oligos SAR1-S and SAR1-AS.

3) Detection of the SARS-CoV RNA by PCR from Respiratory Samples

a) First Comparative Study

[0538] A comparative study was carried out on a series of respiratory samples received by the National Reference Center for the Influenza Virus (Northern region) and likely to contain SARS-CoV. To do this, the RNA was extracted from the samples with the aid of the "Qiaamp viral RNA extraction" kit (Qiagen) and analyzed by real time RT-PCR, on the one hand with the aid of the pairs of primers and probes of the No. 2 series under the conditions described above on the one hand, and on the other hand with the aid of the kit "LightCycler SARS-CoV quantification kit" marketed by Roche (reference 03 604 438). The results are summarized in table VI below. They show that 18 of the 26 samples are negative and 5 of the 26 samples are positive for the two kits, while one sample is positive for the Roche kit alone and two for the "series 2" N reagents alone. Additionally, for 3 samples (20032701, 20032712, 20032714) the quantities of RNA detected are markedly higher with the reagents (probes and primers) of the No. 2 series. These results indicate that the "series 2" N primers and probes are more sensitive for the detection of the SARS-CoV genome in biological samples than those of the kit currently available.

TABLE VI

Real time RT-PCR analysis of the RNAs extracted from a series of samples from 5 patients with the aid of the pairs of primers and probes of the No. 2 series ("series 2" N) or of the kit "Lightcycler SARS-CoV quantification kit" (Roche). The type of sample is indicated as well as the number of copies of viral genome measured in each of the two tests. NEG: negative RT-PCR.				
Sample No.	Patient	Type of sample	ROCHE KIT	"Series 2" N
20033082	K	nasal	NEG	NEG
20033083	K	pharyngeal	NEG	NEG
20033086	K	nasal	NEG	NEG
20033087	K	pharyngeal	NEG	NEG
20032802	M	nasal	NEG	NEG
20032803	M	expectoration	NEG	NEG
20032806	M	nasal or pharyngeal	NEG	NEG
20031746ARN2	C	pharyngeal	NEG	NEG
20032711	C	nasal or pharyngeal	39	NEG
20032910	B	nasal	NEG	NEG
20032911	B	pharyngeal	NEG	NEG
20033356	V	expectoration	NEG	NEG
20033357	V	expectoration	NEG	NEG
20031725	K	endotracheal asp.	NEG	150
20032657	K	endotracheal asp.	NEG	NEG
20032698	K	endotracheal asp.	NEG	NEG
20032720	K	endotracheal asp.	3	5
20033074	K	stools	115	257
20032701	M	pharyngeal	443	1676
20032702	M	expectoration	NEG	249
20031747ARN2	C	pharyngeal	NEG	NEG
20032712	C	unknown	634	6914
20032714	C	pharyngeal	17	223
20032800	B	nasal	NEG	NEG
20033353	V	nasal	NEG	NEG
20033384	V	nasal	NEG	NEG

b) Second Comparative Study

[0539] The performance of various nested RT-PCR and real time RT-PCR methods were then compared for 121 respiratory samples from possible cases of SARS at the French hospital in Hanoi, Vietnam, taken between the 4th and the 17th day after the onset of the symptoms. Among these samples, 14 were found to be positive during a first test using the nested RT-PCR method targeting ORF1b (encoding replicase) as described initially by Bernhard Nocht Institute (BNI nested RT-PCR).

[0540] Information relating to this test is available on the internet, at the address <http://www15.bni-hamburg.de/bni2/neu2/getfile.acgi?area=engl&diagnostics&pid=4112>.

[0541] The various tests compared in this study are:

[0542] the quantitative RT-PCR method according to the invention, with the "series 2" N primers and probes described above (LightCycler N column),

[0543] the nested RT-PCR test targeting the RNA polymerase gene described above, developed by the CDC, BNI and Institut Pasteur <CDC/IP nested RT-PCR>),

[0544] the ARTUS kit with the reference "HPA Corona LC RT-PCR Kit # 5601-02", which is a real time RT-PCR test targeting the ORF1b gene,

[0545] the BNI nested RT-PCR test, also targeting the RNA polymerase gene mentioned above.

[0546] The inventors observed:

[0547] 1) an inter-test variability for the same technique, linked to the degradation of the RNA preparation during repeated thawing, in particular for the samples containing the lowest quantities of RNA,

[0548] 2) a reduced sensitivity of the CDC/IP nested RT-PCR compared with the BNI nested RT-PCR, and

[0549] 3) a comparable sensitivity of the quantitative RT-PCR test according to the invention (Lightcycler N) compared with the Artus LightCycler (LC) test.

[0550] These results, which are presented in table VII below, show that the quantitative RT-PCR test according to the invention constitutes an excellent addition—or an alternative—to the tests currently available. Indeed, the SARS-linked coronavirus is an emergent virus which is capable of changing rapidly. In particular, the gene for the RNA polymerase of the SARS-linked coronavirus, which is targeted in most of the tests currently available, can recombine with that of other coronaviruses not linked to SARS. The use of a test targeting this gene exclusively could then lead to the production of false-negatives.

[0551] The quantitative RT-PCR test according to the invention does not target the same genomic region as the ARTUS kit since it targets the gene encoding the N protein. By carrying out a diagnostic test targeting two different genes of the SARS-linked coronavirus, it can therefore be hoped to avoid false-negative type results which could be due to the genetic evolution of the virus.

[0552] Furthermore, it appears particularly advantageous to target the gene for the nucleocapsid protein because it is very stable because of the high selection pressure linked to the high structural constraints regarding this protein.

TABLE VII

Comparison of various methods of analysis by gene amplification, from 121 samples of probable cases of SARS at the French hospital in Hanoi, Vietnam (epidemic 2003)							
NRC No.	Sample type (1)	Sample collection day	Patient	CDC/IP nested RT-PCR	BNI nested RT-PCR	Artus LightCycler kit	LightCycler N (IP)
107 samples	N and P			Negative	Negative	Negative	Negative
032529	P	10	NHB	Negative	Positive	Negative	Negative
032530	N	10	NHB	Positive	Positive	3.10E+01	4.20E+01
032531	P	7	LP	Positive	Positive	7.70E+00	3.10E+00

TABLE VII-continued

Comparison of various methods of analysis by gene amplification, from 121 samples of probable cases of SARS at the French hospital in Hanoi, Vietnam (epidemic 2003)							
NRC No.	Sample type (1)	Sample collection day	Patient	CDC/IP nested RT-PCR	BNI nested RT-PCR	Artus Light Cycler kit	Light Cycler N (IP)
032534	N	15	BND	Positive	Positive	1.60E+00	Negative
032600	P	4	NHH	Negative	Positive	Negative	1.30E+02
032612	P	17	NTS	Negative	Positive	Negative	Negative
032688	P	9	BTX	Positive	Positive	Negative	Negative
032689	N	4	NVH	Positive	Positive	1.20E+01	2.30E+02
032690	P	4	NVH	Negative	Positive	1.60E+00	Negative
032727	P	8	NVH	Positive	Positive	2.30E+02	4.00E+02
032728	N	8	NVH	Positive	Positive	1.10E+03	1.60E+04
032729	P	14	NHB	Positive	Positive	5.90E+00	3.40E+01
032730	N	14	NHB	Positive	Positive	1.30E+02	4.80E+02
032741	P	8	NHH	Positive	Positive	2.10E+02	1.30E+02
positives				10	14	10	9
fraction detected from the 14 positives				71.4%	100.0%	71.4%	64.3%

(1) P = pharyngeal swab N = nasal swab

EXAMPLE 9

Production and Characterization of Monoclonal
Antibodies Directed Against the N Protein

[0553] Balb C mice were immunized with the purified recombinant N protein and their spleen cells fused with an appropriate murine myeloma according to the Köhler and Milstein techniques.

[0554] Nineteen anti-N antibody secreting hybridomas were preselected and their immunoreactivities determined. These antibodies do indeed recognize the recombinant N protein (in ELISA) with variable intensities, and the natural viral N protein in ELISA and/or in Western blotting. FIGS. 18 to 20 show the results of these tests for 15 of these 19 monoclonal antibodies.

[0555] The highly reactive clones 12, 17, 28, 57, 72, 76, 86, 87, 98, 103, 146, 156, 166, 170, 199, 212, 218, 219 and 222 were subcloned. Specificity studies were carried out with the appropriate tools in order to determine the epitopes recognized and verify the absence of reactivity toward other human coronaviruses and certain respiratory viruses.

[0556] Epitope mapping studies (performed on spot membrane with the aid of overlapping peptides of 15 aa) and additional studies performed on the natural N protein in Western blotting revealed the existence of 4 groups of monoclonal antibodies:

[0557] 1. Monoclonal antibodies specific for a major linear epitope at the N-ter position (75-81, sequence: INTNSVP).

[0558] The representative of this group is antibody 156. The hybridoma producing this antibody was deposited at the Collection Nationale de Cultures de Microorganismes (CNCM) of the Institut Pasteur (Paris, France) on Dec. 1, 2004, under the number I-3331. This same epitope is also recognized by a rabbit serum (anti-N polyclonal) obtained by conventional immunization with the aid of this same N protein.

[0559] 2. Monoclonal antibodies specific for a major linear epitope located in a central position (position 217-224, sequence: ETALALL); the representatives of this group are the monoclonal antibodies 87 and 166. The hybridoma producing antibody 87 was deposited at the CNCM on Dec. 1, 2004, under the number I-3328.

[0560] 3. Monoclonal antibodies specific for a major linear epitope located at the C-terminal position (position 403-408, sequence: DFFRQL), the representatives of this group are the antibodies 28, 57 and 143. The hybridoma producing antibody 57 was deposited at the CNCM on Dec. 1, 2004, under the number I-3330.

[0561] 4. Monoclonal antibodies specific for a discontinuous conformational epitope. This group of antibodies does not recognize any of the peptides spanning the sequence of the N protein, but react strongly on the non-denatured natural protein. The representative of this final group is the antibody 86. The hybridoma producing this antibody was deposited at the CNCM on Dec. 1, 2004, under the number I-3329.

[0562] Table VIII below summarizes the epitope mapping results obtained:

TABLE VIII

Epitope mapping of the monoclonal antibodies			
Antibody	Epitope	Position	Region
28	DFSRQL Q	403 . . . 408	C-Ter.
143	DFSRQL Q		
76	DFSRQL Q		
57	DFSRQL Q		
	FFGMS RI	315 . . . 319	
146	LPQRQ	383 . . . 387	
166	ETALALLL	217 . . . 224	central
87	ETALALL	217 . . . 224	
156	INTNSGP	75 . . . 81	N-Ter.
86	Conformational		
212	Conformational		
170	Conformational		

EXAMPLE 10

Combinations of the Monoclonal Antibodies for the Development of a Sensitive Immunocapture Test Specific for the Viral N Antigen in the Serum or Biological Fluids of Patients Infected with the SARS-CoV Virus

[0563] The antibodies listed below were selected because of their very specific properties for an additional capture and detection study of the viral N protein, in the serum of the subjects or patients.

[0564] These antibodies were produced in ascites on mice, purified by affinity chromatography and used alone or in combination, as capture antibodies and as signal antibodies.

[0565] List of the antibodies selected:

[0566] Ab anti-C-ter region (No. 28, 57, 143)

[0567] Ab anti-central region (No. 87, 166)

[0568] Ab anti-N-ter region (No. 15-6)

[0569] Ab anti-discontinuous conformational epitope 486)

1) Preparation of the Reagents:

a) Immunocapture ELISA plates

[0570] The plates are sensitized with the antibody solutions at 5 µg/ml in 0.1 M carbonate buffer, pH 9.6. The (monovalent or plurivalent) solutions are deposited in a volume of 100 µl in the wells and incubated overnight at room temperature. These plates are then washed with PBS buffer (10 mM pH 7.4 supplemented with 0.1% Tween 20) and then saturated with a PBS solution supplemented with 0.3% BSA and 5% sucrose). The plates are then dried and then packaged in a bag in the presence of a desiccant. They are ready to use.

b) Conjugates

[0571] The purified antibodies were coupled with peroxidase according to the Nakane protocol (Nakane et al.—1974, J. of Histo and cytochemistry, vol. 22, pp. 1084-1091) in a ratio of one molecule of IgG per 3 molecules of peroxidase. These conjugates were purified by exclusion chromatography and stored concentrated (concentration between 1 and 2 mg/ml) in the presence of 50% glycerol and at -20° C. They are diluted for their use in the assays at the final concentration of 1 or 2 µg/ml in PBS buffer (pH 7.4) supplemented with 1% BSA.

c) Other Reagents

[0572] Human sera negative for all the serum markers for the HIV, HBV, HCV and THLV viruses

[0573] Pool of negative human sera supplemented with 0.5% Triton X 100

[0574] Inactivated viral Ag: viral culture supernatant inactivated by irradiation and inactivation verified after placing in culture on sensitive cells—titer of the suspension before inactivation about 10⁷ infectious particles per ml or alternatively about 5×10⁹ physical viral particles per ml of antigen

[0575] The Ag samples diluted in negative human serum: these samples were prepared by diluting 1:100 and then by 5-fold serial dilution.

[0576] These noninfectious samples mimic human samples thought to contain low to very low concentrations of viral nucleoprotein N. Such samples are not available for routine work.

[0577] Washing solution R2, solution for visualization TMB R8, chromogen R9 and stop solution R10, are the generic reagents marketed by Bio-Rad in its ELISA kits (e.g.: *Platelia pylori* kit ref. 72778).

2) Procedure

[0578] The samples of human sera overloaded with inactivated viral Ag are distributed in an amount of 100 µl per well, directly in the ready-to-use sensitized plates, and then incubated for 1 hour at 37° C. (Bio-Rad IPS incubation).

[0579] The material not bound to the solid phase is removed by 3 washings (washing with dilute R2 solution, automatic LP 35 washer).

[0580] The appropriate conjugates, diluted to the final concentration of 1 or 2 µg/ml, are distributed in an amount of 100 µl per well and the plates are again incubated for one hour at 37° C. (IPS incubation).

[0581] The excess conjugate is removed by 4 successive washings (dilute R2 solution—LP 35 washer).

[0582] The presence of conjugate attached to the plates is visualized after adding 100 µl of visualization solution prepared before use (1 ml of R9 and 10 ml of R8) and after incubation for 30 minutes, at room temperature and protected from light.

[0583] The enzymatic reaction is finally blocked by adding 100 µl of R10 reagent (1 N H₂SO₄) to all the wells.

[0584] The reading is carried out with the aid of an appropriate microplate reader at double wavelength (450/620 nm).

[0585] The results can be interpreted by using, as provisional threshold value, the mean of at least two negative controls multiplied by a factor of 2 or alternatively the mean of 100 negative sera supplemented with an increment corresponding to 6 SD (standard deviation calculated on the 100 individual measurements).

3) Results

[0586] Various capture antibody and signal antibody combinations were tested based on the properties of the antibodies selected, and avoiding the combinations of antibodies specific for the same epitopes in solid phase and as conjugates.

[0587] The best results were obtained with the 4 combinations listed below. These results are reproduced in table IX below.

1. Combination F/28

[0588] Solid phase (Ab 166+87 central region): conjugate antibody 28 (C-ter)

2. Combination G/28

[0589] Solid phase (Ab 86—conformational epitope): conjugate antibody 28 (C-ter)

3. Combination H/28

[0590] Solid phase (Ab 86, 166 and 87 central region and conformational epitope): conjugate antibody 28 (C-ter)

4. Combination H/28+87

[0591] Solid phase (Ab 86, 166 and 87 central region and conformational epitope): mixed conjugate antibodies 28 (C-ter) and 87 (central)

5. Combination G/87

[0592] Solid phase (Ab 86—conformational epitope): conjugate antibody 87 (central region)

[0593] The first 4 combinations exhibit equivalent and reproduced performance levels, greater than the other combinations used (such as for example the combination G/87). Of course, in these combinations, a monoclonal antibody may be replaced with another antibody recognizing the same epitope. Thus, the following variants may be mentioned:

6. Variant of the combination F/28

[0594] Solid phase (Ab 87 only): conjugate antibody 57 (C-ter)

7. Variant of the combination G/28

[0595] Solid phase (Ab 86—conformational epitope): conjugate antibody 57 (C-ter)

8. Variant of the combination H/28

[0596] Solid phase (Ab 86 and 87 central region and conformational epitope): conjugate antibody 57 (C-ter)

9. Variant of the combination H/28+87

[0597] Solid phase (Ab 86 and 87 central region and conformational epitope): mixed conjugate antibodies 57 (C-ter) and 87 (central)

TABLE IX

Test of immunoreactivity of the anti-SARS-CoV nucleoprotein Abs: optical densities measured with each combination of antibodies according to the dilutions of the inactivated viral antigen.						
No.	Dilution	F/28	G/28	G/87	H/28	H/28 + 87
0	1/100	5	5	3.495	3.900	5
1	1/500	3.795	3.814	1.379	3.702	3.804
2	1/2 500	2.815	2.950	0.275	3.268	2.680
3	1/12 500	0.987	1.038	0.135	1.374	0.865
4	1/62 500	0.404	0.348	0.125	0.480	0.328
5	1/312 500	0.285	0.211	0.123	0.240	0.215
6	Control	0.210	0.200	0.098	0.186	0.156
7	Control	0.269	0.153	0.104	0.193	0.202

[0598] The detection limit for these 4 experimental trials corresponds to the antigen dilution in negative serum 1:62 500. A rapid extrapolation suggests the detection of less than 10³ infectious particles per ml of sera.

[0599] From this study, it is evident that the most appropriate antibodies for the capture of the native viral nucleoprotein are the antibodies specific for the central region and/or for a conformational epitope, both being antibodies also selected for their high affinity for the native antigen.

[0600] Having determined the best antibodies for the composition of the solid phase, the antibodies to be selected as a priority for the detection of the antigens attached to the solid phase are the complementary antibodies specific for a dominant epitope in the C-ter region. The use of any other complementary antibody specific for epitopes located in the N-ter region of the protein leads to average or poor results.

EXAMPLE 11

Eukaryotic Expression Systems for the SARS-associated Coronavirus (SARS-CoV) Spicule (S) Protein

1) Optimization of the Conditions for Expression of the SARS-CoV S in Mammalian Cells

[0601] The conditions for transient expression of the SARS-CoV spicule (S) protein were optimized in mammalian cells (293T, VeroE6).

[0602] For that, a DNA fragment containing the cDNA for SARS-CoV S was amplified by PCR with the aid of the oligonucleotides 5'-ATAGGATCCA CCAATGTTTAT TTTCTTATTA TTTCTTACTC TCACT-3' and 5'-ATACTC-GAGTT ATGTGTAATG TAATTTGACA CCCTTG-3' from the plasmid pSARS-S (C.N.C.M. No. I-3059) and then inserted between the BamH1 and Xho1 sites of the plasmid pTRIPAU3-CMV containing a lentiviral vector TRIP (Sirven, 2001, Mol. Ther., 3, 438-448) in order to obtain the plasmid pTRIP-S. The BamH1 and Xho1 fragment containing the cDNA for S was then subcloned between BamH1 and Xho1 of the eukaryotic expression plasmid pcDNA3.1(+) (Clontech) in order to obtain the plasmid pcDNA-S. The Nhe1 and Xho1 fragment containing the cDNA for S was then subcloned between the corresponding sites of the expression plasmid pCI (Promega) in order to obtain the plasmid pCI-S. The WPRE sequences of the woodchuck hepatitis virus ("Woodchuck Hepatitis Virus posttranscriptional regulatory element") and the CTE sequences ("constitutive transport element") of the simian retrovirus from Mason-Pfizer were inserted into each of the two plasmids pcDNA-S and pCI-S between the Xho1 and Xba1 sites in order to obtain respectively the plasmids pcDNA-S-CTE, pcDNA-S-WPRE, pCI-S-CTE and pCI-S-WPRE (FIG. 21). The plasmid pCI-S-WPRE was deposited at the CNCM, on Nov. 22, 2004, under the number I-3323. All the inserts were sequenced with the aid of a BigDye Terminator v1.1 kit (Applied Biosystems) and an automated sequencer ABI377.

[0603] The capacity of the plasmid constructs to direct the expression of SARS-CoV S in mammalian cells was assessed after transfection of VeroE6 cells (FIG. 22). In this experiment, monolayers of 5×10⁵ VeroE6 cells in 35 mm Petri dishes were transfected with 2 µg of plasmids pcDNA (as control), pcDNA-S, pCI and pCI-S and 6 µl of Fugene6 reagent according to the manufacturer's instructions (Roche). After 48 hours of incubation at 37° C. and under 5% CO₂, cellular extracts were prepared in loading buffer according to Laemmli, separated on 8% SDS polyacrylamide gel, and then transferred onto a PVDF membrane (BioRad). The detection of this immunoblot (Western blot) was carried out with the aid of an anti-S rabbit polyclonal serum (immune serum from the rabbit P11135: cf. example 4 above) and donkey polyclonal antibodies directed against rabbit IgGs and coupled with peroxidase (NA934V, Amersham). The bound antibodies were visualized by luminescence with the aid of the ECL+ kit (Amersham) and autoradiography films Hyperfilm MP (Amersham).

[0604] This experiment (FIG. 22) shows that the plasmid pcDNA-S does not make it possible to direct the expression of SARS-CoV S at detectable levels whereas the plasmid pCI-S allows a weak expression, close to the limit of detection, which may be detected when the film is overexposed. Similar results were obtained when the expression of S was sought by immunofluorescence (data not shown). This impossibility to detect effective expression of S cannot be

attributed to the detection techniques used since the S protein can be detected at the expected size (180 kDa) in an extract of cells infected with SARS-CoV or in an extract of VeroE6 cells infected with the recombinant vaccinia virus VV-TF7.3 and transfected with the plasmid pcDNA-S. In this latter experiment, the virus VV-TF7.3 expresses the RNA polymerase of the T7 phage and allows the cytoplasmic transcription of an uncapped RNA capable of being efficiently translated. This experiment suggests that the expression defects described above are due to an intrinsic inability of the cDNA for S to be efficiently expressed when the step for transcription to messenger RNA is carried out at the nuclear level.

[0605] In a second experiment, the effect of the CTE and WPRE signals on the expression of S was assessed after transfection of VeroE6 (FIG. 23A) and 293T (FIG. 23B) cells and according to a protocol similar to that described above. Whereas the expression of S cannot be detected after transfection of the plasmids pcDNA-S-CTE and pcDNA-S-WPRE derived from pcDNA-S, the insertion of the WPRE and CTE signals greatly improves the expression of S in the context of the expression plasmid pCI-S.

[0606] To specify this result, a second series of experiments were carried out where the immunoblot is quantitatively visualized by luminescence and acquisition on a digital imaging device (FluorS, BioRad). The analysis of the results obtained with the QuantityOne v4.2.3 software (BioRad) shows that the WPRE and CTE sequences increase respectively the expression of S by a factor of 20 to 42 and 10 to 26 in Vero E6 cells (table X). In 293T cells (table X), the effect of the CTE sequence is more moderate (4 to 5 times) whereas that of the WPRE sequence remains high (13 to 28 times).

TABLE X

Quantitative analysis of the effect of the CTE and WPRE signals on the expression of SARS-CoV S: Cellular extracts were prepared 48 hours after transfection of VeroE6 or 293T cells with the plasmid pCI-S, pCI-S-CTE and pCI-S-WPRE and analyzed by Western blotting as described in the legend to FIG. 22. The Western blot is visualized by luminescence (ECL+, Amersham) and acquisition on a digital imaging device (FluorS, BioRad). The expression levels are indicated according to an arbitrary scale where the value of 1 represents the level measured after transfection of the plasmid pCI-S. Two independent experiments were carried out for each of the two cell types. In experiment 1 on VeroE6 cells, the transfections were carried out in duplicate and the results are indicated in the form of the mean and standard deviation values for the expression levels measured.			
Plasmid	cell	exp. 1	exp. 2
PCI	VeroE6	0.0	0.0
pCI-S	VeroE6	1.0 ± 0.1	1.0
pCI-S-CTE	VeroE6	9.8 ± 0.9	26.4
pCI-S-WPRE	VeroE6	20.1 ± 2.0	42.3
PCI	293T	0.0	0.0
pCI-S	293T	1.0	1.0
pCI-S-CTE	293T	4.6	4.0
pCI-S-WPRE	293T	27.6	12.8

[0607] In summary, all these results show that the expression, in mammalian cells, of the cDNA for the SARS-CoV S under the control of the RNA polymerase II promoter

sequences requires, to be efficient, the expression of a splice signal and of either of the sequences WPRE and CTE.

2) Production of Stable Lines Allowing the Expression of SARS-CoV S

[0608] The cDNA for the SARS-CoV S protein was cloned in the form of a BamH1-Xho1 fragment into the plasmid pTRIPΔU3-CMV containing a defective lentiviral vector TRIP with central DNA flap (Sirven et al., 2001, Mol. Ther., 3: 438-448) in order to obtain the plasmid pTRIP-S (FIG. 24). Transient cotransfection according to Zennou et al. (2000, Cell, 101: 173-185) of this plasmid, of an encapsidation plasmid (p8.2) and of a plasmid for expression of the VSV envelope glycoprotein G (pHCMV-G) in 293T cells allowed the preparation of retroviral pseudoparticles containing the vector TRIP-S and pseudotyped with the envelope protein G. These pseudotyped TRIP-S vectors were used to translate 293T and FRhK-4 cells: no expression of the S protein could be detected by Western blotting and immunofluorescence in the transduced cells (data not presented).

[0609] The optimum expression cassettes consisting of the CMV virus immediate/early promoter, a splice signal, cDNA for S and either of the posttranscriptional signals WPRE or CTE described above were then substituted for the EF1α-EGFP cassette of the defective lentiviral expression vector with central DNA flap TRIPΔU3-EF1α (Sirven et al., 2001, Mol. Ther., 3: 438-448) (FIG. 25). These substitutions were carried out by a series of successive subclonings of the S expression cassettes which were excised from the plasmids PCT-S-CTE (BglII-ApaI) or respectively pCI-S-WPRE (BglII-SalI) and then inserted between the MluI and KpnI sites or respectively MluI or XhoI sites of the plasmid TRIPΔU3-EF1α in order to obtain the plasmids pTRIP-SD/SA-S-CTE and pTRIP-SD/SA-S-WPRE, deposited at the CNM, on Dec. 1, 2004, under the numbers I-3336 and I-3334, respectively. Pseudotyped vectors were produced according to Zennou et al. (2000, Cell, 101: 173-185) and used to transduce 293T cells (10 000 cells) and FRhK-4 cells (15 000 cells) according to a series of 5 successive transduction cycles with a quantity of vectors corresponding to 25 ng (TRIP-SD/SA-S-CTE) or 22 ng TRIP-SD/SA-S-WPRE) of p24 per cycle.

[0610] The transduced cells were cloned by limiting dilution and a series of clones were qualitatively analyzed for the expression of SARS-CoV S by immunofluorescence (data not shown), and then quantitatively by Western blotting (FIG. 25) with the aid of an anti-S rabbit polyclonal serum. The results presented in FIG. 25 show that clones 2 and 15 of FRhK4-s-CTE cells transduced with TRIP-SD/SA-S-CTE and clones 4, 9 and 12 of FRhK4-S-WPRE cells transduced with TRIP-SD/SA-S-WPRE allow the expression of the SARS-CoV S at respectively low or moderate levels if they are compared to those which can be observed during infection with SARS-CoV.

[0611] In summary, the vectors TRIP-SD/SA-S-CTE and TRIP-SD/SA-S-WPRE allow the production of stable clones of FRhK-4 cells and similarly 293T cells expressing SARS-CoV S, whereas the assays carried out with the "parent" vector TRIP-S remained unsuccessful, which demonstrates the need for a splice signal and for either of the sequences CTE and WPRE for the production of stable cell clones expressing the S protein.

[0612] In addition, these modifications of the vector TRIP (insertion of a splice signal and of a post-transcriptional signal like CTE and WPRE) could prove advantageous for improving the expression of other cDNAs than that for S.

3) Production of Stable Lines Allowing the Expression of a Soluble Form of SARS-CoV S. Purification of this Recombinant Antigen.

[0613] A cDNA encoding a soluble form of the S protein (Ssol) was obtained by fusing the sequences encoding the ectodomain of the protein (amino acids 1 to 1193) with those of a tag (FLAG: DYKDDDDK) via a BspE1 linker encoding the SG dipeptide. Practically, in order to obtain the plasmid pcDNA-Ssol, a DNA fragment encoding the ectodomain of SARS-CoV S was amplified by PCR with the aid of the oligonucleotides 5'-ATAGGATCCA CCATGTTTAT TTTCTTATTA TTTCTTACTC TCACT-3' and 5'-ACCTC-CGGAT TTAATATATT GCTCATATTT TCCCAA-3' from the plasmid pcDNA-S, and then inserted between the unique BamH1 and BspE1 sites of a modified eukaryotic expression plasmid pcDNA3.1(+) (Clontech) containing the tag sequence FLAG between its BamH1 and Xho1 sites:

```
// GGATCC . . . nnn . . . TCC GGA GAT TAT AAA GAT
   BamH1                S G D Y K D
```

```
GAC GAC GAT AAA TAA CTCGAG //
D D D K ter Xho1
```

[0614] The Nhe1-Xho1 and BamH1-Xho1 fragments, containing the cDNA for S, were then excised from the plasmid pcDNA-Ssol, and subcloned between the corresponding sites of the plasmid pTRIP-SD/SA-S-CTE and of the plasmid pTRIP-SD/SA-S-WPRE, respectively, in order to obtain the plasmids pTRIP-SD/SA-Ssol-CTE and pTRIP-SD/SA-Ssol-WPRE, deposited at the CNM, on Dec. 1, 2004, under the numbers I-3337 and I-3335, respectively.

[0615] Pseudotyped vectors were produced according to Zennou et al. (2000, Cell, 101:173-185) and used to transduce FRhK-4 cells (15 000 cells) according to a series of 5 successive transduction cycles (15 000 cells) with a quantity of vector corresponding to 24 ng (TRIP-SD/SA-Ssol-CTE) or 40 ng (TRIP-SD/SA-Ssol-WPRE) of p24 per cycle. The transduced cells were cloned by limiting dilution and a series of 16 clones transduced with TRIP-SD/SA-Ssol-CTE and of 15 clones with TRIP-SD/SA-Ssol-WPRE were analyzed for the expression of the Ssol polypeptide by Western blotting visualized with an anti-FLAG monoclonal antibody (FIG. 26 and data not presented), and by capture ELISA specific for the Ssol polypeptide which was developed for this purpose (table XI and data not presented). Part of the process for selecting the best secretory clones is shown in FIG. 26. Capture ELISA is based on the use of solid phases coated with polyclonal antibodies of rabbits immunized with purified and inactivated SARS-CoV. These solid phases allow the capture of the Ssol polypeptide secreted into the cellular supernatants, whose presence is then visualized with a series of steps successively involving the attachment of an anti-FLAG monoclonal antibody (M2, SIGMA), of anti-mouse IgG(H+L) biotinylated rabbit polyclonal antibodies (Jackson) and of a streptavidin-peroxidase conjugate (Amersham) and then the addition of chromogen and substrate (TMB+H₂O₂, KPL)

TABLE XI

Analysis of the expression of the Ssol polypeptide by cell lines transduced with the lentiviral vectors TRIP-SD/SA-Ssol-WPRE and TRIP-SD/SA-Ssol-CTE. The secretion of the Ssol polypeptide was assessed in the supernatant of a series of cell clones isolated after transduction of FRhK-4 cells with the lentiviral vectors TRIP-SD/SA-Ssol-WPRE and TRIP-SD/SA-Ssol-CTE. The supernatants diluted 1/50 were analyzed by a capture ELISA test specific for SARS-CoV S.		
Vector	Clone	OD (450 nm)
Control	—	0.031
TRIP-SD/SA-Ssol-CTE	CTE2	0.547
	CTE3	0.668
	CTE9	0.171
	CTE12	0.208
	CTE13	0.133
TRIP-SD/SA-Ssol-WPRE	WPRE1	0.061
	WPRE10	0.134

[0616] The cell line secreting the highest quantities of Ssol polypeptide in the culture supernatant is the FRhK4-Ssol-CTE3 line. It was subjected to a second series of 5 cycles of transduction with the vector TRIP-SD/SA-Ssol-CTE under conditions similar to those described above and then cloned. The subclone secreting the highest quantities of Ssol was selected by a combination of Western blot and capture ELISA analysis: it is the subclone FRhK4-Ssol-30, which was deposited at the CNM, on Nov. 22, 2004, under the name I-3325.

[0617] The FRhK4-Ssol-30 line allows the quantitative production and purification of the recombinant Ssol polypeptide. In a typical experiment where the experimental conditions for growth, production and purification are optimized, the cells of the FRhK4-Ssol-30 line are inoculated in standard culture medium (pyruvate-free DMEM containing 4.5 g/l of glucose and supplemented with 5% FCS, 100 U/ml of penicillin and 100 µg/ml of streptomycin) in the form of a subconfluent monolayer (1 million cells per each 100 cm² in 20 ml of medium). At confluence, the standard medium is replaced with the secretion medium where the quantity of FCS is reduced to 0.5% and the quantity of medium reduced to 16 ml per each 100 cm². The culture supernatant is removed after 4 to 5 days of incubation at 35° C. and under 5% CO₂. The recombinant polypeptide Ssol is purified from the supernatant by the succession of steps of filtration on 0.1 µm polyethersulfone (PES) membrane, concentration by ultrafiltration on a PES membrane with a 50 kD cut-off, affinity chromatography on anti-FLAG matrix with elution with a solution of FLAG peptide (DYKDDDDK) at 100 µg/ml in TBS (50 mM tris, pH 7.4, 150 mM NaCl) and then gel filtration chromatography in TBS on sephadex G-75 beads (Pharmacia). The concentration of the purified recombinant Ssol polypeptide was determined by micro-BCA test (Pierce) and then its biochemical characteristics analyzed.

[0618] Analysis by 8% SDS acrylamide gel stained with silver nitrate demonstrates a predominant polypeptide whose molecular mass is about 180 kD and whose degree of purity may be evaluated at 98% (FIG. 27A). Two main peaks are detected by SELDI-TOF mass spectrometry (Cypher-ger): they correspond to single and double charged forms of a predominant polypeptide whose molecular mass is thus determined at 182.6±3.7 kD (FIGS. 27B and C). After transfer onto Prosorb membrane and rinsing in 0.1% TFA,

the N-terminal end of the Ssol polypeptide was sequenced in liquid phase by Edman degradation on 5 residues (ABI494, Applied Biosystems) and determined as being SDLDR (FIG. 27D). This demonstrates that the signal peptide located at the N-terminal end of the SARS-CoV S protein, composed of aa 1 to 13 (MFIFLLFLTLTSG) according to an analysis carried out with the software signalP v2.0 (Nielsen et al., 1997, *Protein Engineering*, 10:1-6), is cleaved from the mature Ssol polypeptide. The recombinant Ssol polypeptide therefore consists of amino acids 14 to 1193 of the SARS-CoV S protein fused at the C-terminals with a sequence SGDYKDDDDK containing the sequence of the FLAG tag (underlined). The difference between the theoretical molar mass of the naked Ssol polypeptide (132.0 kD) and the real molar mass of the mature polypeptide (182.6 kD) suggests that the Ssol polypeptide is glycosylated.

[0619] A preparation of purified Ssol polypeptide, whose protein concentration was determined by micro-BCA test, makes it possible to prepare a calibration series in order to measure, with the aid of the capture ELISA test described above, the concentrations of Ssol present in the culture supernatants and to review the characteristics of the secretory lines. According to this test, the FRhK4-Ssol-CT3 line secretes 4 to 6 µg/ml of polypeptide Ssol while the FRhK4-Ssol-30 line secretes 9 to 13 µg/ml of Ssol after 4 to 5 days of culture at confluence. In addition, the purification scheme presented above makes it possible routinely to purify from 1 to 2 mg of Ssol polypeptide per liter of culture supernatant.

EXAMPLE 12

Gene Immunization Involving the SARS-associated Corona Virus (SARS-CoV) Spicule (S) Protein

[0620] The effect of a splice signal and of the posttranscriptional signals WPRE and CTE was analyzed after gene immunization of BALB/c mice (FIG. 28).

[0621] For that, BALB/c mice were immunized at intervals of 4 weeks by injecting into the tibialis anterior a saline solution of 50 µg of plasmid DNA of pcDNA-S and pCI-S and, as a control, 50 µg of plasmid DNA of pcDNA-N (directing the expression of SARS-CoV N) or of pCI-HA (directing the expression of the HA of the influenza virus A/PR/8/34) and the immune sera collected 3 weeks after the 2nd injection. The presence of antibodies directed against the SARS-CoV S was assessed by indirect ELISA using as antigen a lysate of VeroE6 cells infected with SARS-CoV and, as a control, a lysate of noninfected VeroE6 cells. The anti-SARS-CoV antibody titers (TI) are calculated as the reciprocal of the dilution producing a specific OD of 0.5 (difference between OD measured on a lysate of infected cells and OD measured on a lysate of noninfected cells) after visualization with an anti-mouse IgG polyclonal antibody coupled with peroxidase (NA931V, Amersham) and TMB supplemented with H₂O₂ (KPL) (FIG. 28A).

[0622] Under these conditions, the expression plasmid pcDNA-S only allows the induction of low antibody titers directed against SARS-CoV S in 3 mice out of 6 (LOG₁₀(TI)=1.9±0.6) whereas the plasmid pcDNA-N allows the induction of anti-N antibodies at high titers (LOG₁₀(TI)=3.9±0.3) in all the animals, and the control plasmids (pCI, pCI-HA) do not result in any detectable antibody (LOG₁₀(TI)<1.7). The plasmid pCI-S equipped with a splice signal allows the induction of antibodies at high titers (LOG₁₀(TI)=3.7±0.2), which are approximately 60 times higher than those observed after injection of the plasmid pcDNA-S (p<10⁻⁵).

[0623] The efficiency of the posttranscriptional signals was studied by carrying out a dose-response study of the anti-S antibody titers induced in the BALB/c mouse as a function of the quantity of plasmid DNA used as immunogen (2 µg, 10 µg and 50 µg). This study (FIG. 28B) demonstrates that the posttranscriptional signal WPRE greatly improves the efficiency of gene immunization when small doses of DNA are used (p<10⁻⁵ for a dose of 2 µg of DNA and p<10⁻² for a dose of 10 µg), whereas the effect of the CTE signal remains marginal (p=0.34 for a dose of 2 µg of DNA).

[0624] Finally, the antibodies induced in mice after gene immunization neutralize the infectivity of SARS-CoV in vitro (FIGS. 29A and 29B) at titers which are consistent with the titers measured by ELISA.

[0625] In summary, the use of a splice signal and of the posttranscriptional signal WPRE of the woodchuck hepatitis virus considerably improves the induction of neutralizing antibodies directed against SARS-CoV after gene immunization with the aid of plasmid DNA directing the expression of the cDNA for SARS-CoV S.

EXAMPLE 13

Diagnostic Applications of the S Protein

[0626] The ELISA reactivity of the recombinant Ssol polypeptide was analyzed with respect to sera from patients suffering from SARS.

[0627] The sera from probable cases of SARS tested were chosen on the basis of the results (positive or negative) of analysis of their specific reactivity toward the native antigens of SARS-CoV by immunofluorescence test on VeroE6 cells infected with SARS-CoV and/or by indirect ELISA test using as antigen a lysate of VeroE6 cells infected with SARS-CoV. The sera of these patients are identified by a serial number of the National Reference Center for Influenza Viruses and by the initials of the patient and the number of days elapsed since the onset of the symptoms. All the sera of probable cases (cf. Table XII) recognize the native antigens of SARS-CoV, with the exception of the serum 032552 of the patient VTT for whom infection with SARS-CoV could not be confirmed by RT-PCR performed on respiratory samples of days 3, 8 and 12. A panel of control sera was used as control (TV sera): they are sera collected in France before the SARS epidemic that occurred in 2003.

TABLE XII

Sera of probable cases of SARS		
Serum	Patient	Sample collection day
031724	JYK	7
033168	JYK	38
033597	JYK	74
032632	NTM	17
032634	THA	15
032541	PHV	10
032542	NIH	17
032552	VTT	8
032633	PTU	16
032791	JLB	3
033258	JLB	27
032703	JCM	8
033153	JCM	29

[0628] Solid phases sensitized with the recombinant Ssol polypeptide were prepared by adsorption of a solution of purified Ssol polypeptide at 2 µg/ml in PBS in the wells of an ELISA plate, and then the plates are incubated overnight at 4° C. and washed with PBS-Tween buffer (PBS, 0.1% Tween 20). After saturating the ELISA plates with a solution of PBS-10% skimmed milk (weight/volume) and washing in PBS-Tween, the sera to be tested (100 µl) are diluted 1/400 in PBS skimmed milk-Tween buffer (PBS, 3% skimmed milk, 0.1% Tween) and then added to the wells of the sensitized ELISA plate. The plates are incubated for 1 h at 37° C. After 3 washings with PBS-Tween buffer, the anti-human IgG conjugate labeled with peroxidase (ref. NA933V, Amersham) diluted 1/4000 in PBS-skimmed milk-Tween buffer is added, and then the plates are incubated for 1 hour at 37° C. After 6 washings with PBS-Tween buffer, the chromogen (TMB) and the substrate (H₂O₂) are added and the plates are incubated for 10 minutes protected from light. The reaction is stopped by adding a 1 N H₃PO₄ solution, and then the absorbance is measured at 450 nm with a reference at 620 nm.

[0629] The ELISA tests (FIG. 30) demonstrate that the recombinant Ssol polypeptide is specifically recognized by the serum antibodies of patients suffering from SARS collected at the medium or late phase of infection (≥10 days after the onset of the symptoms) whereas it is not significantly recognized by the serum antibodies of 2 patients (JLB and JCM) collected in the early phase of infection (3 to 8 days after the onset of the symptoms) or by control sera of subjects not suffering from SARS. The serum antibodies of patients JLB and JCM show a seroconversion between days 3 and 27 for the first and 8 and 29 for the second after the onset of the symptoms, which confirms the specificity of the reactivity of these sera toward the Ssol polypeptide.

[0630] In conclusion, these results demonstrate that the recombinant Ssol polypeptide may be used as an antigen for the development of an ELISA test for serological diagnosis of infection with SARS-CoV.

EXAMPLE 14

Vaccine Applications of the Recombinant Soluble S Protein

[0631] The immunogenicity of the recombinant Ssol polypeptide was studied in mice.

[0632] For that, a group of 6 mice was immunized at 3 weeks' interval with 10 µg of recombinant Ssol polypeptide adjuvanted with 1 mg of aluminum hydroxide (Alu-gel-S, Serva) diluted in PBS. Three successive immunizations were performed and the immune sera were collected 3 weeks after each of the immunizations (IS1, IS2, IS3). As a control, a group of mice (mock group) received aluminum hydroxide alone according to the same protocol.

[0633] The immune sera were analyzed per pool for each of the 2 groups by indirect ELISA using a lysate of VeroE6 cells infected with SARS-CoV as antigen and as a control a lysate of noninfected VeroE6 cells. The anti-SARS-CoV antibody titers are calculated as the reciprocal of the dilution producing a specific OD of 0.5 after visualization with an anti-mouse IgG (H+L) polyclonal antibody coupled with peroxidase (NA931V, Amersham) and TMB supplemented with H₂O₂ (KPL). This analysis (FIG. 31) shows that the

immunization with the Ssol polypeptide induces in mice, from the first immunization, antibodies directed against the native form of the SARS-CoV spicule protein present in the lysate of infected VeroE6 cells. After 2 then 3 immunizations, the anti-S antibody titers become very high.

[0634] The immune sera were analyzed per pool for each of the two groups for their capacity to seroneutralize the infectivity of SARS-CoV. 4 points of seroneutralization on FRhK-4 cells (100 TCID₅₀ of SARS-CoV) are produced for each of the 2-fold dilutions tested from 1/20. The seroneutralizing titer is calculated according to the Reed and Munsch method as the reciprocal of the dilution neutralizing the infectivity of 2 wells out of 4. This analysis shows that the antibodies induced in mice by the Ssol polypeptide are neutralizing: the titers observed are very high after 2 and then 3 immunizations (greater than 2560 and 5120 respectively, table XIII).

TABLE XIII

Induction of antibodies directed against SARS-CoV after immunization with the recombinant Ssol polypeptide. The immune sera were analyzed per pool for each of the two groups for their capacity to seroneutralize the infectivity of 100 TCID ₅₀ of SARS-CoV on FRhK-4 cells. 4 points are produced for each of the 2-fold dilutions tested from 1/20. The seroneutralizing titer is calculated according to the Reed and Munsch method as the reciprocal of the dilution neutralizing the infectivity of 2 wells out of 4.		
Group	Sera	Neutralizing Ab
Mock	pi	<20
	IS1	<20
	IS2	<20
	IS3	<20
Ssol	pi	<20
	IS1	57
	IS2	>2560
	IS3	>5120

[0635] The neutralizing titers observed in mice immunized with the Ssol polypeptide reach levels far greater than the titers observed by Yang et al. in mice (2004, Nature, 428:561-564) and those observed by Buchholz in the hamster (2004, PNAS 101:9804-9809) which protect respectively mice and hamsters from infection with SARS-CoV. It is therefore probable that the neutralizing antibodies induced in mice after immunization with the Ssol polypeptide protect these animals against infection with SARS-CoV.

EXAMPLE 15

Optimized Synthetic Gene for the Expression in Mammalian Cells of the SARS-associated Coronavirus (SARS-CoV) Spicule (S) Protein

1) Design of the Synthetic Gene

[0636] A synthetic gene encoding the SARS-CoV spicule protein was designed from the gene of the isolate 031589 (plasmid pSARS-S, C.N.C.M. No. I-3059) so as to allow high levels of expression in mammalian cells and in particular in cells of human origin.

[0637] For that:

[0638] the use of codons of the wild-type gene of the isolate 031589 was modified so as to become close to

the bias observed in humans and to improve the efficiency of translation of the corresponding mRNA

[0639] the overall GC content of the gene was increased so as to extend the half-life of the corresponding mRNA

[0640] the optionally cryptic motifs capable of interfering with an efficient expression of the gene were deleted (splice donor and acceptor sites, polyadenylation signals, sequences very rich (>80%) or very low (<30%) in GC, repeat sequences, sequences involved in the formation of secondary RNA structures, TATA boxes)

[0641] a second STOP codon was added to allow efficient termination of translation.

[0642] In addition, CpG motifs were introduced into the gene so as to increase its immunogenicity as DNA vaccine. In order to facilitate the manipulation of the synthetic gene, two BamH1 and Xho1 restriction sites were placed on either side of the open reading frame of the S protein, and the BamH1, Xho1, Nhe1, Kpn1, BspE1 and Sal1 restriction sites were avoided in the synthetic gene.

[0643] The sequence of the synthetic gene designed (gene 040530) is given in SEQ ID No: 140.

[0644] An alignment of the synthetic gene 040530 with the sequence of the wild-type gene of the isolate 031589 of SARS-CoV deposited at the C.N.C.M. under the number I-3059 (SEQ ID No: 4, plasmid pSRAS-S) is presented in FIG. 32.

2) Plasmid Constructs

[0645] The synthetic gene SEQ ID No: 140 was assembled from synthetic oligonucleotides and cloned between the Kpn1 and Sac1 sites of the plasmid pUC-Kana in order to give the plasmid 040530pUC-Kana. The nucleotide sequence of the insert of the plasmid 040530pUC-Kana was verified by automated sequencing (Applied).

[0646] A Kpn1-Xho1 fragment containing the synthetic gene 040530 was excised from the plasmid 040530pUC-Kana and subcloned between the Nhe1 and Xho1 sites of the expression plasmic pCI (Promega) in order to obtain the plasmid pCI-SSYNTH, deposited at the CNCM on Dec. 1, 2004, under the number I-3333.

[0647] A synthetic gene encoding the soluble form of the S protein was then obtained by fusing the synthetic sequences encoding the ectodomain of the S protein (amino acids 1 to 1193) with those of the tag (FLAG: DYKDDDDK) via a linker BspE1 encoding the dipeptide SG. Practically, a DNA fragment encoding the ectodomain of the SARS-CoV S was amplified by PCR with the aid of the oligonucleotides 5'-ACTAGCTAGCGGATCCACCATGTTTCATCTT CCTG-3' and 5'-AGTATCCGGAC TTG ATGTACT GCTCG-TACTTGC-3' from the plasmid 04053-0pUC-Kana, digested with Nhe1 and BspE1 and then inserted between the unique Nhe1 and BspE1 sites of the plasmid pCI-Ssol, to give the plasmid pCI-SCUBE, deposited at the CNCM on Dec. 1, 2004, under the number I-3332. The plasmids pCI-Ssol, pCI-Ssol-CTE, and pCI-Ssol-WPRE (deposited at the CNCM, on Nov. 22, 2004, under the number I-3324) had been previously obtained by subcloning the Kpn1-Xho1 fragment excised from the plasmid pcDNA-Ssol (see technical note of DI 2004-106) between the Nhe1 and Xho1 sites of the plasmids pCI, pCI-S-CTE and pCI-S-WPRE respectively.)

[0648] The plasmids pCI-Scube and pCI-Ssol encode the same recombinant Ssol polypeptide.

3) Results

[0649] The capacity of the synthetic gene encoding the S protein to efficiently direct the expression of the SARS-CoV S in mammalian cells was compared with that of the wild-type gene after transient transfection of primate cells (VeroE6) and of human cells (293T).

[0650] In the experiment presented in FIG. 33 and in table XIV, monolayers of 5×10^5 VeroE6 cells or 7×10^5 293T cells in 35 mm Petri dishes were transfected with 2 μ g of plasmids pCI (as control), pCI-S, pCI-S-CTE, pCI-S-WPRE and pCI-S-Synth and 6 μ l of Eugene6 reagent according to the manufacturer's instructions (Roche). After 48 hours of incubation at 37° C. and under 5% CO₂, cell extracts were prepared in loading buffer according to Laemmli, separated on 8% SDS polyacrylamide gel and then transferred onto a PVDF membrane (BioRad). The detection of this immunoblot (Western blot) was carried out with the aid of an anti-S rabbit polyclonal serum (immune serum of the rabbit P11135: cf example 4 above) and of donkey polyclonal antibodies directed against rabbit IgGs and coupled with peroxidase (NA934V, Amersham). The immunoblot was quantitatively visualized by luminescence with the aid of the ECL+ kit (Amersham) and acquisition on a digital imaging device (FluorS, BioRad).

[0651] The analysis of the results obtained with the software QuantityOne v4.2.3 (BioRad) shows that in this experiment, the plasmid pCI-Synth allows the transient expression of the S protein at high levels in the VeroE6 and 293T cells, whereas the plasmid pCI-S does not make it possible to induce expression at sufficient levels to be detected. The expression levels observed are of the order of twice as high as those observed with the plasmid pCI-S-WPRE.

TABLE XIV

Use of a synthetic gene for the expression of the SARS-CoV S. Cell extracts prepared 48 hours after transfection of VeroE6 or 293T cells with the plasmids pCI, pCI-S, pCI-S-CTE, pCI-S-WPRE and pCI-S-Synth were separated on 8% SDS acrylamide gel and analyzed by Western blotting with the aid of an anti-S rabbit polyclonal antibody and an anti-rabbit IgG(H + L) polyclonal antibody coupled with peroxidase (NA934V, Amersham). The Western blot is visualized by luminescence (ECL+, Amersham) and acquisition on a digital imaging device (FluorS, BioRad). The expression levels of the S protein were measured by quantifying the two predominant bands identified on the image (see FIG. 33) and are indicated according to an arbitrary scale where the value 1 represents the level measured after transfection of the plasmid pCI-S-WPRE.		
Plasmid	VeroE6	293T
pCI	0.0	0.0
pCI-S	≤ 0.1	≤ 0.1
pCI-S-CTE	0.5	≤ 0.1
pCI-S-WPRE	1.0	1.0
pCI-S-synth	1.8	1.9

[0652] In a second instance, the capacity of the synthetic gene Scube to efficiently direct the synthesis and the secretion of the Ssol polypeptide by mammalian cells was compared with that of the wild-type gene after transient transfection of hamster cells (BHK-21) and of human cells (293T).

[0653] In the experiment presented in table XV, monolayers of 6×10^5 BHK-21 cells and 7×10^5 293T cells in 35 mm Petri dishes were transfected with 2 μ g of plasmids pCI (as control), pCI-Ssol, pCI-Ssol-CTE, pCI-Ssol-WPRE and pCI-Scube and 6 μ l of Eugene6 reagent according to the manufacturer's instructions (Roche). After 48 hours of incubation at 37° C. and under 5% CO₂, the cellular supernatants were collected and quantitatively analyzed for the secretion of the Ssol polypeptide by a capture ELISA test specific for the Ssol polypeptide.

[0654] Analysis of the results shows that, in this experiment, the plasmid pCI-Scube allows the expression of the Ssol polypeptide at levels 8 times (BHK-21 cells) to 20 times (293T cells) higher than the plasmid pCI-Ssol. The levels of expression observed are of the order of twice (293T cells) to 5 times (BHK-21 cells) as high as those observed with the plasmid pCI-Ssol-WPRE.

TABLE XV

Use of a synthetic gene for the expression of the Ssol polypeptide. The supernatants were harvested 48 hours after transfection of BHK or 293T cells with the plasmids pCI, pCI-Ssol, pCI-Ssol-CTE, pCI-Ssol-WPRE and pCI-Scube and quantitatively analyzed for the secretion of the Ssol polypeptide by an ELISA test specific for the Ssol polypeptide. The transfections were carried out in duplicate and the results are presented in the form of means and standard deviations of the concentrations of Ssol polypeptide (ng/ml) measured in the supernatants.		
Plasmid	BHK	293T
pCI	<20	<20
pCI-Ssol	<20	56 \pm 10
pCI-Ssol-CTE	<20	63 \pm 8
pCI-Ssol-WPRE	28 \pm 1	531 \pm 15
pCI-Scube	152 \pm 6	1140 \pm 20

[0655] In summary, these results show that the expression, in mammalian cells, of the synthetic gene 040530 encoding SARS-CoV S under the control of RNA polymerase II promoter sequences is much more efficient than that of the wild-type gene of the 031589 isolate. This expression is even more efficient than that directed by the wild-type gene in the presence of the WPRE sequences of the woodchuck hepatitis virus.

4) Applications

[0656] The use of the synthetic gene 040530 encoding SARS-CoV S or its Scube variant encoding the polypeptide Ssol is capable of advantageously replacing the wild-type gene in numerous applications where the expression of S is necessary at high levels. In particular in order to:

[0657] improve the efficiency of gene immunization with plasmids of the pCI-Ssynth or even pCI-Ssynth-CTE or pCI-Ssynth-WPRE type

[0658] establish novel cell lines expressing higher quantities of the S protein or of the Ssol polypeptide with the aid of recombinant lentiviral vectors carrying the Ssynth gene or the Scube gene respectively

[0659] improve the immunogenicity of the recombinant lentiviral vectors allowing the expression of the S protein or of the Ssol polypeptide

[0660] improve the immunogenicity of live vectors allowing the expression of the S protein or of the Ssol polypeptide like recombinant vaccinia viruses or recombinant measles viruses (see examples 16 and 17 below)

EXAMPLE 16

Expression of the SARS-associated Coronavirus (SARS-CoV) Spicule (S) Protein with the Aid of Recombinant Vaccinia Viruses

Vaccine Application

Application to the Production of a Soluble Form of the Spicule (S) Protein and Design of a Serological Test for SARS

1) Introduction

[0661] The aim of this example is to evaluate the capacity of recombinant vaccinia viruses (VV) expressing various SARS-associated coronavirus (SARS-CoV) antigens to constitute novel vaccine candidates against SARS and a means of producing recombinant antigens in mammalian cells.

[0662] For that, the inventors focused on the SARS-CoV spicule (S) protein which makes it possible to induce, after gene immunization in animals, antibodies neutralizing the infectivity of SARS-CoV, and a soluble and secreted form of this protein, the Ssol polypeptide, which is composed of the ectodomain (aa 1-1193) of S fused at its C-ter end with a tag FLAG (DYKDDDDK) via a BspEI linker encoding the SG dipeptide. This Ssol polypeptide exhibits an antigenicity similar to that of the S protein and allows, after injection into mice in the form of a purified protein adjuvanted with aluminum hydroxide, the induction of high neutralizing antibody titers against SARS-CoV.

[0663] The various forms of the S gene were placed under the control of the promoter of the 7.5K gene and then introduced into the thymidine kinase (TK) locus of the Copenhagen strain of the vaccinia virus by double homologous recombination in vivo. In order to improve the immunogenicity of the recombinant vaccinia viruses, a synthetic late promoter was chosen in place of the 7.5K promoter, in order to increase the production of S and Ssol during the late phases of the viral cycle.

[0664] After having isolated the recombinant vaccinia viruses and verified their capacity to express the SARS-CoV S antigen, their capacity to induce in mice an immune response against SARS was tested. After having purified the Ssol antigen from the supernatant of infected cells, an ELISA test for serodiagnosis of SARS was designed, and its efficiency was evaluated with the aid of sera from probable cases of SARS.

2) Construction of the Recombinant Viruses

[0665] Recombinant vaccinia viruses directing the expression of the S glycoprotein of the 031589 isolate of SARS-CoV and of a soluble and secreted form of this protein, the Ssol polypeptide, under the control of the 7.5K promoter were obtained. With the aim of increasing the levels of expression of S and Ssol, recombinant viruses in which the cDNAs for S and for Ssol are placed under the control of a late synthetic promoter were also obtained.

[0666] The plasmid pTG186poly is a transfer plasmid for the construction of recombinant vaccinia viruses (Kieny, 1986, *Biotechnology*, 4:790-795). As such, it contains the VV thymidine kinase gene into which the promoter of the 7.5K gene has been inserted followed by a multiple cloning site allowing the insertion of heterologous genes (FIG. 34A). The promoter of the 7.5K gene in fact contains a tandem of two promoter sequences that are respectively active during the early (P_E) and late (P_L) phases of the vaccinia virus replication cycle. The BamH1-Xho1 fragments were excised from the plasmids pTRIP-S and pcDNA-Ssol respectively and inserted between the BamH1 and Sma1 sites of the plasmid pTG186poly in order to give the plasmids pTG-S and pTG-Ssol (FIG. 34A). The plasmids pTG-S and pTG-Ssol were deposited at the CNCM, on Dec. 2, 2004, under the numbers I-3338 and I-3339, respectively.

[0667] The plasmids pTN480, pTN-S and pTN-Ssol were obtained from the plasmids pTG186poly, pTG-S and pTG-Ssol respectively, by substituting the Nde1-Pst1 fragment containing the 7.5K promoter by a DNA fragment containing the synthetic late promoter 480, which was obtained by hybridization of the oligonucleotides 5'-TATGAGCTTT TTTTTTTTTT TTTTTTGGC ATATAAATAG ACTCG-GCGCG CCATCTGCA-3' and 5'-GATGGCGCGC-GCAGTCTATT TATATGCCAA AAAAAAAAAA AAAAAAAGC TCA-3' (FIG. 34B). The insert was sequenced with the aid of a BigDye Terminator v1.1 kit (Applied Biosystems) and an automated sequencer ABI377. The sequence of the late synthetic promoter 480 as cloned into the transfer plasmids of the pTN series is indicated in FIG. 34C. The plasmids pTN-S and pTN-Ssol were deposited at the CNCM, on Dec. 2, 2004, under the numbers I-3340 and I-3341, respectively.

[0668] The recombinant vaccinia viruses were obtained by double homologous recombination in vivo between the TK cassette of the transfer plasmids of the series pTG and pTN and the TK gene of the Copenhagen strain of the vaccinia virus according to a procedure described by Kieny et al. (1984, *Nature*, 312:163-166). Briefly, CV-1 cells are transfected with the aid of DOTAP (Roche) with genomic DNA of the Copenhagen strain of the vaccinia virus and each of the transfer plasmids of the pTG and pTN series described above, and then superinfected with the helper vaccinia virus VV-ts7 for 24 hours at 33° C. The helper virus is counter-selected by incubation at 40° C. for 2 days and then the recombinant viruses (TK- phenotype) selected by two cloning cycles under agar medium on 143Btk- cells in the presence of BuDr (25 µg/ml). The 6 viruses VV-TG, VV-TG-S, VV-TG-Ssol, VV-TN, VV-TN-S, and VV-TN-Ssol are respectively obtained with the aid of the transfer plasmids pTG186poly, pTG-S, pTG-Ssol, pTN480, pTN-S, pTN-Ssol. The viruses VV-TG and VV-TN do not express any heterologous gene and were used as TK- control in the experiments. The preparations of recombinant viruses were performed on monolayers of CV-1 or BHK-21 cells and the titer in plaque forming units (p.f.u) determined on CV-1 cells according to Earl and Moss (1998, *Current Protocols in Molecular Biology*, 16.16.1-16.16.13).

3) Characterization of the Recombinant Viruses

[0669] The expression of the transgenes encoding the S protein and the Ssol polypeptide was assessed by Western blotting.

[0670] Monolayers of CV-1 cells were infected at a multiplicity of 2 with various recombinant vaccinia viruses VV-TG, VV-TG-S, VV-TG-Ssol, VV-TN, VV-TN-S and VV-TN-Ssol. After 18 hours of incubation at 37° C. and under 5% CO₂, cellular extracts were prepared in loading buffer according to Laemmli, separated on 8% SDS polyacrylamide gel and then transferred onto a PVDF membrane (BioRad). The detection of this immunoblot (Western blot) was performed with the aid of an anti-S rabbit polyclonal serum (immune serum from the rabbit P11135: cf. example 4) and donkey polyclonal antibodies directed against rabbit IgGs and coupled with peroxidase (NA934V, Amersham). The bound antibodies were visualized by luminescence with the aid of the ECL+kit (Amersham) and autoradiography films Hyperfilm MP (Amersham).

[0671] As shown in FIG. 35A, the recombinant virus VV-TN-S directs the expression of the S protein at levels which are comparable to those which can be observed 8 h after infection with SARS-CoV but which are much higher than those which can be observed after infection with VV-TG-S. In a second experiment (FIG. 35B), the analysis of variable quantities of cellular extracts shows that the levels of expression observed after infection with viruses of the TN series (VV-TN-S and VV-TN-Ssol) are about 10 times as high as those observed with the viruses of the TG series (VV-TG-S and VV-TG-Ssol, respectively); In addition, the Ssol polypeptide is secreted into the supernatant of CV-1 cells infected with the VV-TN-Ssol virus more efficiently than in the supernatant of cells infected with VV-TG-Ssol (FIG. 36A). In this experiment, the VV-TN-Sflag virus was used as a control because it expresses the membrane form of the S protein fused at its C-ter end with the FLAG tag. The Sflag protein is not detected in the supernatant of cells infected with VV-TN-Sflag, demonstrating that the Ssol polypeptide is indeed actively secreted after infection with VV-TN-Ssol.

[0672] These results demonstrate that the recombinant vaccinia viruses are indeed carriers of the transgenes and allow the expression of the SRAS glycoprotein in its membrane form (S) or in a soluble or secreted form (Ssol). The vaccinia viruses carrying the synthetic promoter 480 allow the expression of S and the secretion of Ssol at levels much higher than the viruses carrying the promoter of the 7.5K gene.

4) Application to the Production of a Soluble Form of SARS-CoV S. Purification of this Recombinant Antigen and Diagnostic Applications

[0673] The BHK-21 line is the cell line which secretes the highest quantities of Ssol polypeptide after infection with the VV-TN-Ssol virus among the lines tested (BHK-21, CV1, 293T and FrhK-4, FIG. 36B); it allows the quantitative production and purification of the recombinant Ssol polypeptide. In a typical experiment where the experimental conditions for infection, production and purification were optimized, the BHK-21 cells are inoculated in standard culture medium (pyruvate-free DMEM containing 4.5 g/l of glucose and supplemented with 5% TPB, 5% FCS, 100 U/ml of penicillin and 100 µg/ml of streptomycin) in the form of a subconfluent monolayer (10 million cells for each 100 cm² in 25 ml of medium). After 24 h of incubation at 37° C. under 5% CO₂, the cells are infected at an M.O.I. of 0.03 and the standard medium replaced with the secretion medium

where the quantity of FCS is reduced to 0.5% and the TPB eliminated. The culture supernatant is removed after 2.5 days of incubation at 35° C. and under 5% CO₂ and the vaccinia virus inactivated by addition of Triton X-100 (0.1%). After filtration on 0.1 µm polyethersulfone (PES) membrane, the recombinant Ssol polypeptide is purified by affinity chromatography on an anti-FLAG matrix with elution with a solution of FLAG peptide (DYKDDDDK) at 100 µg/ml in TBS (50 mM Tris, pH 7.4, 150 mM NaCl).

[0674] The analysis by 8% SDS acrylamide gel stained with silver nitrate identified a predominant polypeptide whose molecular mass is about 180 kD and whose degree of purity is greater than 90% (FIG. 37). The concentration of the purified Ssol recombinant polypeptide was determined by comparison with molecular mass markers and estimated at 24 ng/µl.

[0675] This purified Ssol polypeptide preparation makes it possible to produce a calibration series in order to measure, with the aid of a capture ELISA test, the Ssol concentrations present in the culture supernatants. According to this test, the BHK-21 line secretes about 1 µg/ml of Ssol polypeptide under the production conditions described above. In addition, the purification scheme presented makes it possible to purify of the order of 160 µg of Ssol polypeptide per liter of culture supernatant.

[0676] The ELISA reactivity of the recombinant Ssol polypeptide was analyzed toward sera from patients suffering from SARS.

[0677] The sera of probable cases of SARS tested were chosen on the basis of the results (positive or negative) of analysis of their specific reactivity toward the native antigens of SARS-CoV by immunofluorescence test on VeroE6 cells infected with SARS-CoV and/or by indirect ELISA test using, as antigen, a lysate of VeroE6 cells infected with SARS-CoV. The sera of these patients are identified by a serial number of the National Reference Center for Influenza Viruses and by the patient's initials and the number of days elapsed since the onset of the symptoms. All the sera of probable cases (cf. table XVI) recognize the native antigens of SARS-CoV with the exception of the serum 032552 of the patient VTT, for which infection with SARS-CoV could not be confirmed by RT-PCR performed on respiratory samples of days 3, 8 and 12. A panel of control sera was used as control (TV sera): they are sera collected in France before the SARS epidemic which occurred in 2003.

TABLE XVI

Sera of probable cases of SARS		
Serum	Patient	Sample collection day
033168	JYK	38
033597	JYK	74
032632	NTM	17
032634	THA	15
032541	PHV	10
032542	NIH	17
032552	VTT	8
032633	PTU	16

[0678] Solid phases sensitized with the recombinant Ssol polypeptide were prepared by adsorption of a solution of purified Ssol polypeptide at 4 µg/ml in PBS in the wells of

an ELISA plate. The plates are incubated overnight at 4° C. and then washed with PBS-Tween buffer (PBS, 0.1% Tween 20). After washing with PBS-Tween, the sera to be tested (100 µl) are diluted 1/100 and 1/400 in PBS-skimmed milk-Tween buffer (PBS, 3% skimmed milk, 0.1% Tween) and then added to the wells of the sensitized ELISA plate. The plates are then incubated for 1 h at 37° C. After 3 washings with PBS-Tween buffer, the anti-human IgG conjugate labeled with peroxidase (ref. NA933V, Amersham) diluted 1/4000 in PBS-skimmed milk-Tween buffer is added and then the plates are incubated for one hour at 37° C. After 6 washings with PBS-Tween buffer, the chromogen (TMB) and the substrate (H₂O₂) are added and the plates are incubated for 10 minutes protected from light. The reaction is stopped by adding a 1M solution of H₃PO₄ and then the absorbance is measured at 450 nm with a reference at 620 nm.

[0679] The ELISA tests (FIG. 38) demonstrate that the recombinant Ssol polypeptide is specifically recognized by the serum antibodies of patients suffering from SARS, collected at the middle or late phase of infection (≥10 days after the onset of the symptoms), whereas it is not significantly recognized by the serum antibodies of the control sera of subjects not suffering from SARS.

[0680] In conclusion, these results demonstrate that the recombinant Ssol polypeptide can be purified from the supernatant of mammalian cells infected with the recombinant vaccinia virus VV-TN-Ssol and can be used as antigen for developing an ELISA test for serological diagnosis of infection with SARS-CoV.

5. Vaccine Applications

[0681] The immunogenicity of the recombinant vaccinia viruses was studied in mice.

[0682] For that, groups of 7 BALB/c mice were immunized by the i.v. route twice at 4 weeks' interval with 10⁶ p.f.u. of recombinant vaccinia viruses VV-TG, VV-T-G-S, VV-TG-Ssol, VV-TN, VV-TN-S and VV-TN-Ssol and, as a control, VV-TG-HA which directs the expression of hemagglutinin of the A/PR/8/34 strain of the influenza virus. The immune sera were collected 3 weeks after each of the immunizations (IS1, IS2).

[0683] The immune sera were analyzed per pool for each of the groups by indirect ELISA using a lysate of VeroE6 cells infected with SARS-CoV as antigen and, as control, a lysate of noninfected VeroE6 cells. The anti-SARS-CoV antibody titers (TI) are calculated as the reciprocal of the dilution producing a specific OD of 0.5 after visualization with an anti-mouse IgG(H+L) polyclonal antibody coupled with peroxidase (NA931V, Amersham) and TMB supplemented with H₂O₂ (KPL). This analysis (FIG. 39A) shows that immunization with the virus VV-TG-S and VV-TN-S induces in mice, from the first immunization, antibodies directed against the native form of the SARS-CoV spicule protein present in the lysate of infected VeroE6 cells. The responses induced by the VV-TN-S virus are higher than those induced by the VV-TG-S virus after the first (TI=740 and TI=270 respectively) and the second (TI=3230 and TI=600 respectively) immunization. The VV-TN-Ssol virus induces high anti-SARS-CoV antibody titers after two immunizations (TI=640), whereas the virus VV-TG-Ssol induces a response at the detection limit (TI=40).

[0684] The immune sera were analyzed per pool for each of the groups for their capacity to seroneutralize the infectivity of SARS-CoV. 4 seroneutralization points on FRhK-4 cells (100 TCID₅₀ of SARS-CoV) are produced for each of the 2-fold dilutions tested from 1/20. The seroneutralizing titer is calculated according to the Reed and Munsch method as the reciprocal of the dilution neutralizing the infectivity of 2 wells out of 4. This analysis shows that the antibodies induced in mice by the vaccinia viruses expressing the S protein or the Ssol polypeptide are neutralizing and that the viruses with synthetic promoters are more efficient immunogens than the viruses carrying the 7.5K promoter: the highest titers (640) are observed after 2 immunizations with the virus VV-TN-S (FIG. 39B).

[0685] The protective power of the neutralizing antibodies induced in mice after immunization with the recombinant vaccinia viruses is evaluated with the aid of a challenge infection with SARS-CoV.

6) Other Applications

[0686] Third generation recombinant vaccinia viruses are constructed by substituting the wild-type sequences of the S and Ssol genes by synthetic genes optimized for the expression in mammalian cells, described above. These recombinant vaccinia viruses are capable of expressing larger quantities of S and Ssol antigens and therefore of exhibiting increased immunogenicity.

[0687] The recombinant vaccinia virus VV-TN-Ssol can be used for the quantitative production and purification of the Ssol antigen for diagnostic (serology by ELISA) and vaccine (subunit vaccine) applications.

EXAMPLE 17

Recombinant Measles Virus Expressing the SARS-associated Coronavirus (SARS-CoV) Spicule (S) Protein. Vaccine Applications.

1) Introduction

[0688] The measles vaccine (MV) induces a lasting protective immunity in humans after a single injection (Hilleman, 2002, Vaccine, 20: 651-665). The protection conferred is very robust and is based on the induction of an antibody response and of a CD4 and CD8 cell response. The MV genome is very stable and no reversion of the vaccine strains to virulence has ever been observed. The measles virus belongs to the genus *Morbillivirus* of the Paramyxoviridae family; it is an enveloped virus whose genome is a 16 kb single-stranded RNA of negative polarity (FIG. 40A) and whose exclusively cytoplasmic replication cycle excludes any possibility of integration into the genome of the host. The measles vaccine is thus one of the most effective and one of the safest live vaccines used in the human population. Frederic Tangy's team recently developed an expression vector on the basis of the Schwarz strain of the measles virus, which is the safest attenuated strain and the most widely used in humans as vaccine against measles. This vaccine strain may be isolated from an infectious molecular clone while preserving its immunogenicity in primates and in mice that are sensitive to the infection. It constitutes, after insertion of additional transcription units, a vector for the expression of heterologous sequences (Combretet, 2003, J. Virol. 77: 11546-11554). In addition, a recombinant MV Schwarz expressing the envelope glycoprotein of the West Nile virus (WNV) induces an effective and lasting antibody response which protects mice from a lethal challenge infec-

tion with WNV (Despres et al., 2004, J. Infect. Dis., in press). All these characteristics make the attenuated Schwarz strain of the measles virus an extremely promising candidate vector for the construction of novel recombinant live vaccines.

[0689] The aim of this example is to evaluate the capacity of recombinant measles viruses (MV) expressing various SARS-associated coronavirus (SARS-CoV) antigens to constitute novel candidate vaccines against SARS.

[0690] The inventors focused on the SARS-CoV spicule (S) protein, which makes it possible to induce, after gene immunization in animals, antibodies neutralizing the infectivity of SARS-CoV, and on a soluble and secreted form of this protein, the Ssol polypeptide, which is composed of the ectodomain (aa 1-1193) of S fused at its C-ter end with a FLAG tag (DYKDDDDK) via a BspEI linker encoding the SG dipeptide. This Ssol polypeptide exhibits a similar antigenicity to that of the S protein and allows, after injection into mice in the form of a purified protein adjuvanted with aluminum hydroxide, the induction of high neutralizing antibody titers against SARS-CoV.

[0691] The various forms of the S gene were introduced in the form of an additional transcription unit between the P (phosphoprotein) and M (matrix) genes into the cDNA of the Schwarz strain of MV previously described (Combretet, 2003, J. Virol. 77: 11546-11554; EP application No. 02291551.6 of Jun. 20, 2002, and EP application No. 02291550.8 of Jun. 20, 2002). After having isolated the recombinant viruses MVSchw2-SARS-S and MVSchw2-SARS-Ssol and checked their capacity to express the SARS-CoV S antigen, their capacity to induce a protective immune response against SARS in mice and then in monkeys was tested.

2) Construction of the Recombinant Viruses

[0692] The plasmid pTM-MVSchw-ATU2 (FIG. 40B) contains an infectious cDNA corresponding to the antigenome of the Schwarz vaccine strain of the measles virus (MV) into which an additional transcription unit (ATU) has been introduced between the P (phosphoprotein) and M (matrix) genes (Combretet, 2003, Journal of Virology, 77: 11546-11554). Recombinant genomes MVSchw2-SARS-S and MVSchw2-SARS-Ssol of the measles virus were constructed by inserting ORFs of the S protein and of the Ssol polypeptide into the additional transcription unit of the MVSchw-ATU2 vector.

[0693] For that, a DNA fragment containing the SARS-CoV S cDNA was amplified by PCR with the aid of the oligonucleotides 5'-ATACGTACGA CCATGTTTAT TTTCTTATTA TTTCTTACTC TCACT-3' and 5'-AT-AGCGCGCT CATTATGTGT AATGTAATTT GACAC-CCTTG-3' using the plasmid pcDNA-S as template and then inserted into the plasmid pCR®2.1-TOPO (Invitrogen) in order to obtain the plasmid pTOPO-S-MV. The two oligonucleotides used contain restriction sites BsiWI and BssHII, so as to allow subsequent insertion into the measles vector, and were designed so as to generate a sequence of 3774 nt including the codons for initiation and termination, so as to observe the rule of 6 which stipulates that the length of the genome of a measles virus must be divisible by 6 (Calain & Roux, 1993, J. Virol., 67: 4822-4830; Schneider et al., 1997, Virology, 227: 314-322). The insert was sequenced with the aid of a BigDye Terminator v1.1 kit (Applied Biosystems) and an automated sequencer ABI377.

[0694] To express a soluble and secreted form of SARS-CoV S, a plasmid containing the cDNA of the Ssol polypep-

tide corresponding to the ectodomain (aa 1-1193) of SARS-CoV S fused at its C-ter end with the sequence of a FLAG tag (DYKDDDDK) via a BspE1 linker encoding the SG dipeptide was then obtained. For that, a DNA fragment was amplified with the aid of the oligonucleotides 5'-CCATTTCAAC AATTTGGCCG-3' and 5'-ATAGGATC-CGCGCGCTCAT TTTATCGTC GTCATCTTTA TAATC-3' from the plasmid pcDNA-Ssol and then inserted into the plasmid pTOPO-S-MV between the SalI and BamHI sites in order to obtain the plasmid pTOPO-S-MV-SF. The sequence generated is 3618 nt long between the BsiW1 and BssHII sites and observes the rule of 6. The insert was sequenced as indicated above.

[0695] The BsiW1-BssHII fragments containing the cDNAs for the S protein and the Ssol polypeptide were then excised by digestion of the plasmids pTOPO-S-MV and pTOPO-S-MV-SF and then subcloned between the corresponding sites of the plasmid pTM-MV Schw-ATU2 in order to give the plasmids pTM-MV Schw2-SARS-S and pTM-MV Schw2-SARS-Ssol (FIG. 40B). These two plasmids were deposited at the C.N.C.M. on Dec. 1, 2004, under the numbers I-3326 and I-3327, respectively.

[0696] The recombinant measles viruses corresponding to the plasmids pTM-MV Schw2-SARS-S and pTM-MV Schw2-SARS-Ssol were obtained by reverse genetics according to the system based on the use of a helper cell line, described by Radecke et al. (1995, Embo J., 14: 5773-5784) and modified by Parks et al. (1999, J. Virol., 73: 3560-3566). Briefly, the helper cells 293-3-46 are transfected according to the calcium phosphate method with 5 µg of the plasmids pTM-MV Schw2-SARS-S or pTM-MV Schw2-SARS-Ssol and 0.02 µg of the plasmid pEMC-La directing the expression of the MV L polymerase (gift from M. A. Billeter). After incubating overnight at 37° C., a heat shock is produced for 2 hours at 43° C. and the transfected cells are transferred onto a monolayer of Vero cells. For each of the two plasmids, syncytia appeared after 2 to 3 days of coculture and were transferred successively onto monolayers of Vero cells at 70% confluence in 35 mm Petri dishes and then in 25 and 75 cm² flasks. When the syncytia have reached 80-90% confluence, the cells are recovered with the aid of a scraper and then frozen and thawed once. After low-speed centrifugation, the supernatant containing the virus is stored in aliquots at -80° C. The titers of the recombinant viruses MV Schw2-SARS-S and MV Schw2-SARS-Ssol were determined by limiting dilution on Vero cells and the titer as dose infecting 50% of the wells (TCID₅₀) calculated according to the Kärber method.

3) Characterization of the Recombinant Viruses

[0697] The expression of the transgenes encoding the S protein and the Ssol polypeptide was assessed by Western blotting and immunofluorescence.

[0698] Monolayers of Vero cells in T-25 flasks were infected at a multiplicity of 0.05 by various passages of the two viruses MV Schw2-SARS-S and MV Schw2-SARS-Ssol and the wild-type virus MWSchw as a control. When the syncytia had reached 80 to 90% confluence, cytoplasmic extracts were prepared in an extraction buffer (150 mM NaCl, 50 mM Tris-HCl, pH 7.2, 1% Triton X-100, 0.1% SDS, 1% DOC) and then diluted in loading buffer according to Laemmli, separated on 8% SDS polyacrylamide gel and transferred onto a PVDF membrane (BioRad). The detection

of this immunoblot (Western blot) was carried out with the aid of an anti-S rabbit polyclonal serum (immune serum of the rabbit P11135: cf. example 4 above) and donkey polyclonal antibodies directed against rabbit IgGs and coupled with peroxidase (NA934V, Amersham). The bound antibodies were visualized by luminescence with the aid of the ECL+kit (Amersham) and Hyperfilm MP autoradiography films (Amersham).

[0699] Vero cells in monolayers on glass slides were infected with the two viruses MV Schw2-SARS-S and MV Schw2-SARS-Ssol and the wild-type virus MWSchw as a control at multiplicities of infection of 0.05. When the syncytia had reached 90 to 100% (MV Schw2-SARS-Ssol virus) or 30 to 40% (MV Schw2-SARS-S, MWSchw) confluence, the cells were fixed in a 4% PBS-PFA solution, permeabilized with a PBS solution containing 0.2% Triton and then labeled with rabbit polyclonal antibodies hyperimmunized with purified and inactivated SARS-CoV virions and with an anti-rabbit IgG(H+L) goat antibody conjugate coupled with FITC (Jackson).

[0700] As shown in FIGS. 41 and 42, the recombinant viruses MV Schw2-SARS-S and MV Schw2-SARS-Ssol direct the expression of the S protein and the Ssol polypeptide respectively at levels comparable to those which can be observed 8 h after infection with SARS-CoV. The expression of these polypeptides is stable after 3 passages of the recombinant viruses in cell culture. These results demonstrate that the recombinant measles viruses are indeed carriers of the transgenes and allow the expression of the SARS glycoprotein in its membrane form (S) or in a soluble form (Ssol). The Ssol polypeptide is expected to be secreted by cells infected with the MV Schw2-SARS-Ssol virus as is the case when this same polypeptide is expressed in mammalian cells after transient transfection of the corresponding sequences (cf. example 11 above).

4) Applications

[0701] Having shown that the viruses MV Schw2-SARS-S and MV Schw2-SARS-Ssol allow the expression of the SARS-CoV S, their capacity to induce a protective immune response against SARS-CoV in CD46^{+/+} IFN-αβR^{-/-} mice, which is sensitive to infection by MV, is evaluated. The antibody response of the immunized mice is evaluated by ELISA test against the native antigens of SARS-CoV and for their capacity to neutralize the infectivity of SARS-CoV in vitro, using the methodologies described above. The protective power of the response will be evaluated by measuring the reduction in the pulmonary viral load 2 days after a nonlethal challenge infection with SARS-CoV.

[0702] Second generation recombinant measles viruses are constructed by substituting the wild-type sequences of the S and Sol genes by synthetic genes optimized for expression in mammalian cells, described in example 15 above. These recombinant measles viruses are capable of expressing larger quantities of the S and Ssol antigens and therefore of exhibiting increased immunogenicity.

[0703] Alternatively, the wild-type or synthetic genes encoding the S protein or the Ssol polypeptide may be inserted into the measles vector MV Schw-ATU3 in the form of an additional transcription unit located between the H and L genes, and then the recombinant viruses produced and characterized in a similar manner. This insertion is capable

of generating recombinant viruses possessing different characteristics (multiplication of the virus, level of expression of the transgene) and possibly an improved immunogenicity compared with those obtained after insertion of the transgenes between the P and N genes.

[0704] The recombinant measles virus MVSchw2-SARS-Ssol may be used for the quantitative production and the purification of the Ssol antigen for diagnostic and vaccine applications.

EXAMPLE 18

Other Applications Linked to the S Protein

[0705] a) The lentiviral vectors allowing the expression of S or Ssol (or even of fragments of S) can constitute a

recombinant vaccine against SARS-CoV, to be used in human or veterinary prophylaxis. In order to demonstrate the feasibility of such a vaccine, the immunogenicity of the recombinant lentiviral vectors TRIP-SD/SA-S-WPRE and TRIP-SD/SA-Ssol-WPRE is studied in mice.

[0706] b) Monoclonal antibodies are produced with the aid of the recombinant Ssol polypeptide. According to the results presented in example 14 above, these antibodies or at least the majority of them will recognize the native form of the SARS-CoV S and will be capable of diagnostic and/or prophylactic applications.

[0707] c) A serological test for SARS is developed with the Ssol polypeptide used as antigen and the double epitope methodology.

SEQUENCE LISTING

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<211> LENGTH: 29746

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

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	985				990					995					1000	
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Arg	Ala	Ser	Ala	Asn	Leu	Ala	Ala	Thr	Lys	Met	Ser	Glu	Cys	Val	
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Leu	Gly	Gln	Ser	Lys	Arg	Val	Asp	Phe	Cys	Gly	Lys	Gly	Tyr	His	
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ctt	atg	tcc	ttc	cca	caa	gca	gcc	ccg	cat	ggg	gtt	gtc	ttc	cta	3223
Leu	Met	Ser	Phe	Pro	Gln	Ala	Ala	Pro	His	Gly	Val	Val	Phe	Leu	
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cat	gtc	acg	tat	gtg	cca	tcc	cag	gag	agg	aac	ttc	acc	aca	gcg	3268
His	Val	Thr	Tyr	Val	Pro	Ser	Gln	Glu	Arg	Asn	Phe	Thr	Thr	Ala	
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cca	gca	att	tgt	cat	gaa	ggc	aaa	gca	tac	ttc	cct	cgt	gaa	ggg	3313
Pro	Ala	Ile	Cys	His	Glu	Gly	Lys	Ala	Tyr	Phe	Pro	Arg	Glu	Gly	
				1065					1070					1075	
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Val	Phe	Val	Phe	Asn	Gly	Thr	Ser	Trp	Phe	Ile	Thr	Gln	Arg	Asn	
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ttc	ttt	tct	cca	caa	ata	att	act	aca	gac	aat	aca	ttt	gtc	tca	3403
Phe	Phe	Ser	Pro	Gln	Ile	Ile	Thr	Thr	Asp	Asn	Thr	Phe	Val	Ser	
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Asp	Pro	Leu	Gln	Pro	Glu	Leu	Asp	Ser	Phe	Lys	Glu	Glu	Leu	Asp	
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Lys	Tyr	Phe	Lys	Asn	His	Thr	Ser	Pro	Asp	Val	Asp	Leu	Gly	Asp	
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Ile	Ser	Gly	Ile	Asn	Ala	Ser	Val	Val	Asn	Ile	Gln	Lys	Glu	Ile	
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Met	Val	Thr	Ile	Leu	Leu	Cys	Cys	Met	Thr	Ser	Cys	Cys	Ser	Cys	
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ctc	aag	ggg	gca	tgc	tct	tgt	ggg	tct	tgc	tgc	aag	ttt	gat	gag	3808
Leu	Lys	Gly	Ala	Cys	Ser	Cys	Gly	Ser	Cys	Cys	Lys	Phe	Asp	Glu	
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gat	gac	tct	gag	cca	gtt	ctc	aag	ggg	gtc	aaa	tta	cat	tac	aca	3853
Asp	Asp	Ser	Glu	Pro	Val	Leu	Lys	Gly	Val	Lys	Leu	His	Tyr	Thr	
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<210> SEQ ID NO 3

<211> LENGTH: 1255

<212> TYPE: PRT

<213> ORGANISM: CORONAVIRUS

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<400> SEQUENCE: 3

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His Thr Ser Ser Met Arg Gly Val Tyr Tyr Pro Asp Glu Ile Phe Arg
 35           40           45
Ser Asp Thr Leu Tyr Leu Thr Gln Asp Leu Phe Leu Pro Phe Tyr Ser
 50           55           60
Asn Val Thr Gly Phe His Thr Ile Asn His Thr Phe Gly Asn Pro Val
 65           70           75           80
Ile Pro Phe Lys Asp Gly Ile Tyr Phe Ala Ala Thr Glu Lys Ser Asn
 85           90           95
Val Val Arg Gly Trp Val Phe Gly Ser Thr Met Asn Asn Lys Ser Gln
100          105          110
Ser Val Ile Ile Ile Asn Asn Ser Thr Asn Val Val Ile Arg Ala Cys
115          120          125
Asn Phe Glu Leu Cys Asp Asn Pro Phe Phe Ala Val Ser Lys Pro Met
130          135          140
Gly Thr Gln Thr His Thr Met Ile Phe Asp Asn Ala Phe Asn Cys Thr
145          150          155          160
Phe Glu Tyr Ile Ser Asp Ala Phe Ser Leu Asp Val Ser Glu Lys Ser
165          170          175
Gly Asn Phe Lys His Leu Arg Glu Phe Val Phe Lys Asn Lys Asp Gly
180          185          190
Phe Leu Tyr Val Tyr Lys Gly Tyr Gln Pro Ile Asp Val Val Arg Asp
195          200          205
Leu Pro Ser Gly Phe Asn Thr Leu Lys Pro Ile Phe Lys Leu Pro Leu
210          215          220
Gly Ile Asn Ile Thr Asn Phe Arg Ala Ile Leu Thr Ala Phe Ser Pro
225          230          235          240
Ala Gln Asp Ile Trp Gly Thr Ser Ala Ala Ala Tyr Phe Val Gly Tyr
245          250          255
Leu Lys Pro Thr Thr Phe Met Leu Lys Tyr Asp Glu Asn Gly Thr Ile
260          265          270
Thr Asp Ala Val Asp Cys Ser Gln Asn Pro Leu Ala Glu Leu Lys Cys
275          280          285
Ser Val Lys Ser Phe Glu Ile Asp Lys Gly Ile Tyr Gln Thr Ser Asn
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Phe Arg Val Val Pro Ser Gly Asp Val Val Arg Phe Pro Asn Ile Thr
305          310          315          320
Asn Leu Cys Pro Phe Gly Glu Val Phe Asn Ala Thr Lys Phe Pro Ser
325          330          335
Val Tyr Ala Trp Glu Arg Lys Lys Ile Ser Asn Cys Val Ala Asp Tyr
340          345          350
Ser Val Leu Tyr Asn Ser Thr Phe Phe Ser Thr Phe Lys Cys Tyr Gly
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Val Ser Ala Thr Lys Leu Asn Asp Leu Cys Phe Ser Asn Val Tyr Ala
370          375          380
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Thr Gly Asn Tyr Asn 435	Tyr Lys Tyr Arg 440	Tyr Leu Arg His 445	Gly Lys Leu
Arg Pro Phe Glu Arg 450	Asp Ile Ser Asn 455	Val Pro Phe Ser 460	Pro Asp Gly
Lys Pro Cys Thr Pro 465	Pro Ala Leu Asn 470	Cys Tyr Trp Pro 475	Leu Asn Asp 480
Tyr Gly Phe Tyr Thr 485	Thr Thr Gly Ile 490	Gly Tyr Gln Pro 495	Tyr Arg Val
Val Val Leu Ser Phe 500	Glu Leu Leu Asn 505	Ala Pro Ala Thr 510	Val Cys Gly
Pro Lys Leu Ser Thr 515	Asp Leu Ile Lys 520	Asn Gln Cys Val 525	Asn Phe Asn
Phe Asn Gly Leu Thr 530	Gly Thr Gly Val 535	Leu Thr Pro Ser 540	Ser Ser Lys Arg
Phe Gln Pro Phe Gln 545	Gln Phe Gly Arg 550	Asp Val Ser Asp 555	Phe Thr Asp 560
Ser Val Arg Asp Pro 565	Lys Thr Ser Glu 570	Ile Leu Asp Ile 575	Ser Pro Cys
Ser Phe Gly Gly Val 580	Ser Val Ile Thr 585	Pro Gly Thr Asn 590	Ala Ser Ser
Glu Val Ala Val Leu 595	Tyr Gln Asp Val 600	Asn Cys Thr Asp 605	Val Ser Thr
Ala Ile His Ala Asp 610	Gln Leu Thr Pro 615	Ala Trp Arg Ile 620	Tyr Ser Thr
Gly Asn Asn Val Phe 625	Gln Thr Gln Ala 630	Gly Cys Leu Ile 635	Gly Ala Glu 640
His Val Asp Thr Ser 645	Tyr Glu Cys Asp 650	Ile Pro Ile Gly 655	Ala Gly Ile 655
Cys Ala Ser Tyr His 660	Thr Val Ser Leu 665	Leu Arg Ser Thr 670	Ser Ser Gln Lys
Ser Ile Val Ala Tyr 675	Thr Met Ser Leu 680	Gly Ala Asp Ser 685	Ser Ser Ile Ala
Tyr Ser Asn Asn Thr 690	Ile Ala Ile Pro 695	Thr Asn Phe Ser 700	Ile Ser Ile
Thr Thr Glu Val Met 705	Pro Val Ser Met 710	Ala Lys Thr Ser 715	Val Asp Cys 720
Asn Met Tyr Ile Cys 725	Gly Asp Ser Thr 730	Glu Cys Ala Asn 735	Leu Leu Leu
Gln Tyr Gly Ser Phe 740	Cys Thr Gln Leu 745	Asn Arg Ala Leu 750	Ser Gly Ile
Ala Ala Glu Gln Asp 755	Arg Asn Thr Arg 760	Glu Val Phe Ala 765	Gln Val Lys
Gln Met Tyr Lys Thr 770	Pro Thr Leu Lys 775	Tyr Phe Gly Gly 780	Phe Asn Phe
Ser Gln Ile Leu Pro 785	Asp Pro Leu Lys 790	Pro Thr Lys Arg 795	Ser Phe Ile 800

Glu 805	Asp	Leu	Leu	Phe	Asn	Lys	Val	Thr	Leu	Ala	Asp	Ala	Gly	Phe	Met
Lys 820	Gln	Tyr	Gly	Glu	Cys	Leu	Gly	Asp	Ile	Asn	Ala	Arg	Asp	Leu	Ile
Cys 835	Ala	Gln	Lys	Phe	Asn	Gly	Leu	Thr	Val	Leu	Pro	Pro	Leu	Leu	Thr
Asp 850	Asp	Met	Ile	Ala	Ala	Tyr	Thr	Ala	Ala	Leu	Val	Ser	Gly	Thr	Ala
Thr 865	Ala	Gly	Trp	Thr	Phe	Gly	Ala	Gly	Ala	Ala	Leu	Gln	Ile	Pro	Phe
Ala	Met	Gln	Met	Ala	Tyr	Arg	Phe	Asn	Gly	Ile	Gly	Val	Thr	Gln	Asn
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Gln	Leu	Ile	Arg	Ala	Ala	Glu	Ile	Arg	Ala	Ser	Ala	Asn	Leu	Ala	Ala
Thr 1010	Lys	Met	Ser	Glu	Cys	Val	Leu	Gly	Gln	Ser	Lys	Arg	Val	Asp	
Phe 1025	Cys	Gly	Lys	Gly	Tyr	His	Leu	Met	Ser	Phe	Pro	Gln	Ala	Ala	
Pro 1040	His	Gly	Val	Val	Phe	Leu	His	Val	Thr	Tyr	Val	Pro	Ser	Gln	
Glu 1055	Arg	Asn	Phe	Thr	Thr	Ala	Pro	Ala	Ile	Cys	His	Glu	Gly	Lys	
Ala 1070	Tyr	Phe	Pro	Arg	Glu	Gly	Val	Phe	Val	Phe	Asn	Gly	Thr	Ser	
Trp 1085	Phe	Ile	Thr	Gln	Arg	Asn	Phe	Phe	Ser	Pro	Gln	Ile	Ile	Thr	
Thr 1100	Asp	Asn	Thr	Phe	Val	Ser	Gly	Asn	Cys	Asp	Val	Val	Ile	Gly	
Ile 1115	Ile	Asn	Asn	Thr	Val	Tyr	Asp	Pro	Leu	Gln	Pro	Glu	Leu	Asp	
Ser 1130	Phe	Lys	Glu	Glu	Leu	Asp	Lys	Tyr	Phe	Lys	Asn	His	Thr	Ser	
Pro 1145	Asp	Val	Asp	Leu	Gly	Asp	Ile	Ser	Gly	Ile	Asn	Ala	Ser	Val	
Val 1160	Asn	Ile	Gln	Lys	Glu	Ile	Asp	Arg	Leu	Asn	Glu	Val	Ala	Lys	
Asn 1175	Leu	Asn	Glu	Ser	Leu	Ile	Asp	Leu	Gln	Glu	Leu	Gly	Lys	Tyr	

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1205						1210					1215			
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1220						1225					1230			
Ser	Cys	Cys	Lys	Phe	Asp	Glu	Asp	Asp	Ser	Glu	Pro	Val	Leu	Lys
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<210> SEQ ID NO 4

<211> LENGTH: 3943

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 4

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<210> SEQ ID NO 5

<211> LENGTH: 2049

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 5

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tgccagatga	tttcattggg	tgtgtccttg	cttggaatac	taggaacatt	gatgctactt	1380
caactggtaa	ttataattat	aaatataggt	atcttagaca	tggaagcctt	aggccctttg	1440
agagagacat	atctaattgt	cctttctccc	ctgatggcaa	accttgccac	ccacctgctc	1500
ttaattgtta	ttggccatta	aatgattatg	gtttttacac	cactactggc	attggctacc	1560
aaccttacag	agttgtagta	ctttcttttg	aacttttaaa	tgaccgggcc	acggtttgtg	1620
gacccaaaatt	atccactgac	cttattaaga	accagtgtgt	caattttaat	tttaatggac	1680
tcactggtag	tggtgtgtta	actccttctt	caaagagatt	tcaaccattt	caacaatttg	1740
gccgtgatgt	ctctgatttc	actgattccg	ttcgagatcc	taaaacatct	gaaatattag	1800
acatttcacc	ttgctctttt	gggggtgtaa	gtgtaattac	acctggaaca	aatgcttcac	1860
ctgaagttgc	tggtctatat	caagatgtta	actgcactga	tgtttctaca	gcaatccatg	1920
cagatcaact	cacaccagct	tggcgcatat	attctactgg	aaacaatgta	ttccagactc	1980

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aagcaggctg tcttatagga gctgagcatg tcgacacttc ttatgagtgc gacattccta 2040
ttggagctg 2049

<210> SEQ ID NO 6
<211> LENGTH: 2027
<212> TYPE: DNA
<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 6
catgcagatc aactcacacc agcttggcgc atatattcta ctggaacaa tgtattccag 60
actcaagcag gctgtcttat aggagctgag catgtcgaca cttcttatga gtgcgacatt 120
cctattggag ctggcatttg tgctagttag catacagttt ctttattacg tagtactagc 180
caaaaatcta ttgtggctta tactatgtct ttaggtgctg atagttcaat tgcttactct 240
aataacacca ttgtataacc tactaacttt tcaattagca ttactacaga agtaatgcct 300
gtttctatgg ctaaacctc cgtagattgt aatatgtaca tctgcggaga ttctactgaa 360
tgtgctaatt tgcttctcca atatggtagc ttttgcacac aactaaatcg tgcactctca 420
ggtattgctg ctgaacagga tcgcaacaca cgtgaagtgt tcgctcaagt caaacaaatg 480
tacaaaaccc caactttgaa atattttggt ggttttaatt tttcaciaat attacctgac 540
cctctaaagc caactaagag gtcttttatt gaggacttgc tctttaataa ggtgacactc 600
gctgatgctg gcttcatgaa gcaatatggc gaatgcctag gtgatattaa tgctagagat 660
ctcatttgctg cgcagaagtt caatgggctt acagtgttgc cacctctgct cactgatgat 720
atgattgctg cctacactgc tgctctagtt agtggtactg ccactgctgg atggacattt 780
ggtgctggcg ctgctcttca aatacctttt gctatgcaa tggcatatag gttcaatggc 840
attggagtta cccaaaatgt tctctatgag aacaaaaaac aaatcgccaa ccaatttaac 900
aaggcgatta gtcaaatcca agaatcactt acaacaacat caactgcatt gggcaagctg 960
caagacgttg ttaaccagaa tgctcaagca ttaaacacac ttgttaaaca acttagctct 1020
aattttgggt caatttcaag tgtgctaaat gatacctttt cgcgacttga taaagtcgag 1080
gcgaggttac aaattgacag gttaattaca ggcagacttc aaagccttca aacctatgta 1140
acacaacaac taatcagggc tgctgaaatc agggcttctg ctaatcttgc tgctactaaa 1200
atgtctgagt gtgttcttgg acaatcaaaa agagttgact tttgtgaaa gggctaccac 1260
cttatgtcct tcccacaagc agcccgcgat ggtgttgtct tcctacatgt cacgtatgtg 1320
ccatcccagg agaggaactt caccacagcg ccagcaatth gtcatgaagg caaagcatac 1380
ttccctcgtg aagggtgtttt tgtgtttaat ggcacttctt ggtttattac acagaggaac 1440
ttcttttctc cacaaataat tactacagac aatacatttg tctcaggaaa ttgtgatgtc 1500
gttattggcg tcattaacaa cacagtttat gatcctctgc aacctgagct tgactcattc 1560
aaagaagagc tggacaagta cttcaaaaat catacatcac cagatgttga tcttggcgac 1620
atttcaggca ttaacgcttc tgcgtcaac attcaaaaag aaattgaccg cctcaatgag 1680
gtcgctaaaa atttaaatga atcactcatt gaccttcaag aattgggaaa atatgagcaa 1740
tatattaaat ggccttggtg tgtttggctc ggcttcattg ctggactaat tgccatcgtc 1800
atggttacaa tcttgctttg ttgcatgact agttgttgca gttgcctcaa ggggtcatgc 1860
tcttgtggtt cttgctgcaa gtttgatgag gatgactctg agccagttct caagggtgtc 1920

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aaattacatt acacataaac gaacttatgg atttgtttat gagatTTTTT actcttggat 1980
caattactgc acagccagta aaaattgaca atgcttctcc tgcaagt 2027

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<210> SEQ ID NO 7
<211> LENGTH: 1096
<212> TYPE: DNA
<213> ORGANISM: CORONAVIRUS

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<400> SEQUENCE: 7

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tcttgctttg ttgcatgact agttgttgca gttgcctcaa gggtgcatgc tcttgtgggt 60
cttgtgctga gtttgatgag gatgactctg agccagttct caagggtgtc aaattacatt 120
acacataaac gaacttatgg atttgtttat gagatTTTTT actcttggat caattactgc 180
acagccagta aaaattgaca atgcttctcc tgcaagtact gttcatgcta cagcaacgat 240
accgctacaa gcctcactcc ctttcggatg gcttgttatt ggcgttgcat ttcttgctgt 300
ttttcagagc gctacaaaa taattgcgct caataaaaga tggcagctag ccctttataa 360
gggcttccag ttcatattga atttactgct gctatttggt accatctatt cacatctttt 420
gcttgcgctg gcaggtatgg aggcgcaatt ttgtacctc tatgcctga tatattttct 480
acaatgcctc aacgcatgta gaattattat gagatgttgg ctttgttggg agtgcaaatc 540
caagaaccca ttactttatg atgccaacta ctttgtttgc tggcacacac ataactatga 600
ctactgtata ccatataaca gtgtcacaga tacaattgct gttactgaag gtgacggcat 660
ttcaacacca aaactcaaag aagactacca aattgggtgg tattctgagg ataggcactc 720
aggtgttaaa gactatgtcg ttgtacatgg ctatttcacc gaagtttact accagcttga 780
gtctacacaa attactacag acactggtat tgaaaatgct acattcttca tctttaacaa 840
gcttgttaaa gaccaccga atgtgcaaat acacacaatc gacggctctt caggagtgtc 900
taatccagca atggatccaa tttatgatga gccgacgacg actactagcg tgcctttgta 960
agcacaagaa agtgagtacg aacttatgta ctcatctggt tcggaagaaa caggtagctt 1020
aatagttaat agcgtacttc tttttcttgc tttcgtggta ttcttgctag tcacactagc 1080
catccttact gcgctt 1096

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<210> SEQ ID NO 8
<211> LENGTH: 1135
<212> TYPE: DNA
<213> ORGANISM: CORONAVIRUS

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<400> SEQUENCE: 8

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attgccatcg tcatgggttac aatcttgctt tgttgcatga ctagtgttg cagttgcctc 60
aagggtgcat gctcttggg ttcttgctgc aagtttgatg aggatgactc tgagccagtt 120
ctcaagggtg tcaattata ttacacataa acgaacttat ggatttgttt atgagatttt 180
ttactcttgg atcaattact gcacagccag taaaaattga caatgcttct cctgcaagta 240
ctgttcatgc tacagcaacg ataccgctac aagcctcact ccctttcgga tggcttgtta 300
ttggcggtgc atttcttctg gtttttcaga gcgctaccaa aataattgcg ctcaataaaa 360
gatggcagct agccctttat aagggtctcc agttcatttg caatttactg ctgctatttg 420
ttaccatcta ttcacatctt ttgcttgcg ctgcaggtat ggaggcgcaa tttttgtacc 480
tctatgcctt gatataattt ctacaatgca tcaacgcag tagaattatt atgagatgtt 540

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ggctttgttg gaagtgc aaa tccaagaacc cttactttta tgatgccaac tactttgttt 600
gctggcacac acataactat gactactgta taccatataa cagtgtcaca gatacaattg 660
tcgttactga aggtgacggc atttcaacac caaaactcaa agaagactac caaattggtg 720
gttattctga ggataggcac tcaggtgtta aagactatgt cgtgtgacat ggctatttca 780
ccgaagttta ctaccagctt gagtctacac aaattactac agacactggg attgaaaatg 840
ctacattctt catctttaac aagcttgta aagaccacc gaatgtgcaa atacacacaa 900
tcgacggctc ttcaggagtt gctaattccag caatggatcc aatttatgat gagccgacga 960
cgactactag cgtgcctttg taagcacaag aaagtgaata cgaacttatg tactcattcg 1020
tttcggaaga aacaggtagc ttaatagta atagcgtact tctttttctt gctttcgtgg 1080
tattcttgct agtcacacta gccatcctta ctgcgcttcg attgtgtgcg tactg 1135

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<210> SEQ ID NO 9
<211> LENGTH: 1096
<212> TYPE: DNA
<213> ORGANISM: CORONAVIRUS
<220> FEATURE:
<221> NAME/KEY: CDS
<222> LOCATION: (137)..(958)
<223> OTHER INFORMATION:

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<400> SEQUENCE: 9

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cttgctgcaa gtttgatgag gatgactctg agccagttct caagggtgtc aaattacatt 120
acacataaac gaactt atg gat ttg ttt atg aga ttt ttt act ctt gga tca 172
          Met Asp Leu Phe Met Arg Phe Phe Thr Leu Gly Ser
          1             5             10
att act gca cag cca gta aaa att gac aat gct tct cct gca agt act 220
Ile Thr Ala Gln Pro Val Lys Ile Asp Asn Ala Ser Pro Ala Ser Thr
          15             20             25
gtt cat gct aca gca acg ata ccg cta caa gcc tca ctc cct ttc gga 268
Val His Ala Thr Ala Thr Ile Pro Leu Gln Ala Ser Leu Pro Phe Gly
          30             35             40
tgg ctt gtt att ggc gtt gca ttt ctt gct gtt ttt cag agc gct acc 316
Trp Leu Val Ile Gly Val Ala Phe Leu Ala Val Phe Gln Ser Ala Thr
          45             50             55             60
aaa ata att gcg ctc aat aaa aga tgg cag cta gcc ctt tat aag ggc 364
Lys Ile Ile Ala Leu Asn Lys Arg Trp Gln Leu Ala Leu Tyr Lys Gly
          65             70             75
ttc cag ttc att tgc aat tta ctg ctg cta ttt gtt acc atc tat tca 412
Phe Gln Phe Ile Cys Asn Leu Leu Leu Leu Phe Val Thr Ile Tyr Ser
          80             85             90
cat ctt ttg ctt gtc gct gca ggt atg gag gcg caa ttt ttg tac ctc 460
His Leu Leu Leu Val Ala Ala Gly Met Glu Ala Gln Phe Leu Tyr Leu
          95             100             105
tat gcc ttg ata tat ttt cta caa tgc atc aac gca tgt aga att att 508
Tyr Ala Leu Ile Tyr Phe Leu Gln Cys Ile Asn Ala Cys Arg Ile Ile
          110             115             120
atg aga tgt tgg ctt tgt tgg aag tgc aaa tcc aag aac cca tta ctt 556
Met Arg Cys Trp Leu Cys Trp Lys Cys Lys Ser Lys Asn Pro Leu Leu
          125             130             135             140
tat gat gcc aac tac ttt gtt tgc tgg cac aca cat aac tat gac tac 604
Tyr Asp Ala Asn Tyr Phe Val Cys Trp His Thr His Asn Tyr Asp Tyr
          145             150             155

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tgt ata cca tat aac agt gtc aca gat aca att gtc gtt act gaa ggt Cys Ile Pro Tyr Asn Ser Val Thr Asp Thr Ile Val Val Thr Glu Gly 160 165 170	652
gac ggc att tca aca cca aaa ctc aaa gaa gac tac caa att ggt ggt Asp Gly Ile Ser Thr Pro Lys Leu Lys Glu Asp Tyr Gln Ile Gly Gly 175 180 185	700
tat tct gag gat agg cac tca ggt gtt aaa gac tat gtc gtt gta cat Tyr Ser Glu Asp Arg His Ser Gly Val Lys Asp Tyr Val Val Val His 190 195 200	748
ggc tat ttc acc gaa gtt tac tac cag ctt gag tct aca caa att act Gly Tyr Phe Thr Glu Val Tyr Tyr Gln Leu Glu Ser Thr Gln Ile Thr 205 210 215 220	796
aca gac act ggt att gaa aat gct aca ttc ttc atc ttt aac aag ctt Thr Asp Thr Gly Ile Glu Asn Ala Thr Phe Phe Ile Phe Asn Lys Leu 225 230 235	844
gtt aaa gac cca ccg aat gtg caa ata cac aca atc gac ggc tct tca Val Lys Asp Pro Pro Asn Val Gln Ile His Thr Ile Asp Gly Ser Ser 240 245 250	892
gga gtt gct aat cca gca atg gat cca att tat gat gag ccg acg acg Gly Val Ala Asn Pro Ala Met Asp Pro Ile Tyr Asp Glu Pro Thr Thr 255 260 265	940
act act agc gtg cct ttg taagcacaag aaagtgagta cgaacttatg Thr Thr Ser Val Pro Leu 270	988
tactcattcg tttcgggaaga aacaggtacg ttaatagtta atagegtact tctttttctt	1048
gctttcgtgg tattcttgct agtcacacta gccatcctta ctgcgctt	1096

<210> SEQ ID NO 10

<211> LENGTH: 274

<212> TYPE: PRP

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 10

Met Asp Leu Phe Met Arg Phe Phe Thr Leu Gly Ser Ile Thr Ala Gln 1 5 10 15
Pro Val Lys Ile Asp Asn Ala Ser Pro Ala Ser Thr Val His Ala Thr 20 25 30
Ala Thr Ile Pro Leu Gln Ala Ser Leu Pro Phe Gly Trp Leu Val Ile 35 40 45
Gly Val Ala Phe Leu Ala Val Phe Gln Ser Ala Thr Lys Ile Ile Ala 50 55 60
Leu Asn Lys Arg Trp Gln Leu Ala Leu Tyr Lys Gly Phe Gln Phe Ile 65 70 75 80
Cys Asn Leu Leu Leu Leu Phe Val Thr Ile Tyr Ser His Leu Leu Leu 85 90 95
Val Ala Ala Gly Met Glu Ala Gln Phe Leu Tyr Leu Tyr Ala Leu Ile 100 105 110
Tyr Phe Leu Gln Cys Ile Asn Ala Cys Arg Ile Ile Met Arg Cys Trp 115 120 125
Leu Cys Trp Lys Cys Lys Ser Lys Asn Pro Leu Leu Tyr Asp Ala Asn 130 135 140
Tyr Phe Val Cys Trp His Thr His Asn Tyr Asp Tyr Cys Ile Pro Tyr 145 150 155 160
Asn Ser Val Thr Asp Thr Ile Val Val Thr Glu Gly Asp Gly Ile Ser

	165		170		175
Thr Pro Lys Leu Lys Glu Asp Tyr Gln Ile Gly Gly Tyr Ser Glu Asp 180 185 190					
Arg His Ser Gly Val Lys Asp Tyr Val Val Val His Gly Tyr Phe Thr 195 200 205					
Glu Val Tyr Tyr Gln Leu Glu Ser Thr Gln Ile Thr Thr Asp Thr Gly 210 215 220					
Ile Glu Asn Ala Thr Phe Phe Ile Phe Asn Lys Leu Val Lys Asp Pro 225 230 235 240					
Pro Asn Val Gln Ile His Thr Ile Asp Gly Ser Ser Gly Val Ala Asn 245 250 255					
Pro Ala Met Asp Pro Ile Tyr Asp Glu Pro Thr Thr Thr Thr Ser Val 260 265 270					
Pro Leu					
<210>	SEQ ID NO 11				
<211>	LENGTH: 1096				
<212>	TYPE: DNA				
<213>	ORGANISM: CORONAVIRUS				
<220>	FEATURE:				
<221>	NAME/KEY: CDS				
<222>	LOCATION: (558)..(1019)				
<223>	OTHER INFORMATION:				
<400>	SEQUENCE: 11				
tcttgccttg ttgcatgact agttgttgca gttgcctcaa gggtgcatgc tcttgtggtt	60				
cttgcctgcaa gttgatgag gatgactctg agccagtctc caagggtgtc aaattacatt	120				
acacataaac gaacttatgg atttgtttat gagatttttt actccttgga caattactgc	180				
acagccagta aaaattgaca atgctctctc tgcaagtact gtcatgcta cagcaacgat	240				
accgctacaa gcctcactcc ctctcgatg gcttggtatt gccgttgcat ttcttgctgt	300				
ttttcagagc gctaccaaaa taattgccgt caataaaaga tggcagctag ccctttataa	360				
gggcttccag ttcatttgca atttactgct gctatttggt accatctatt cacatctttt	420				
gcttgctgct gcaggtagg aggcgcaatt tttgtacctc tatgcottga tatattttct	480				
acaatgcatc aacgcatgta gaattattat gagatgttgg ctttgttgga agtgcaaatc	540				
caagaaccce ttactttt atg atg cca act act ttg ttt gct gcc aca cac Met Met Pro Thr Thr Leu Phe Ala Gly Thr His 1 5 10	590				
ata act atg act act gta tac cat ata aca gtg tca cag ata caa ttg Ile Thr Met Thr Thr Val Tyr His Ile Thr Val Ser Gln Ile Gln Leu 15 20 25	638				
tcg tta ctg aag gtg acg gca ttt caa cac caa aac tca aag aag act Ser Leu Leu Lys Val Thr Ala Phe Gln His Gln Asn Ser Lys Lys Thr 30 35 40	686				
acc aaa ttg gtg gtt att ctg agg ata ggc act cag gtg tta aag act Thr Lys Leu Val Val Ile Leu Arg Ile Gly Thr Gln Val Leu Lys Thr 45 50 55	734				
atg tcg ttg tac atg gct att tca ccg aag ttt act acc agc ttg agt Met Ser Leu Tyr Met Ala Ile Ser Pro Lys Phe Thr Thr Ser Leu Ser 60 65 70 75	782				
cta cac aaa tta cta cag aca ctg gta ttg aaa atg cta cat tct tca Leu His Lys Leu Leu Gln Thr Leu Val Leu Lys Met Leu His Ser Ser 80 85 90	830				

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tct tta aca agc ttg tta aag acc cac cga atg tgc aaa tac aca caa	878
Ser Leu Thr Ser Leu Leu Lys Thr His Arg Met Cys Lys Tyr Thr Gln	
95 100 105	
tcg acg gct ctt cag gag ttg cta atc cag caa tgg atc caa ttt atg	926
Ser Thr Ala Leu Gln Glu Leu Ile Gln Gln Trp Ile Gln Phe Met	
110 115 120	
atg agc cga cga cga cta cta gcg tgc ctt tgt aag cac aag aaa gtg	974
Met Ser Arg Arg Arg Leu Leu Ala Cys Leu Cys Lys His Lys Lys Val	
125 130 135	
agt acg aac tta tgt act cat tcg ttt cgg aag aaa cag gta cgt	1019
Ser Thr Asn Leu Cys Thr His Ser Phe Arg Lys Lys Gln Val Arg	
140 145 150	
taatagttaa tagcgtactt ctttttcttg ctttcgtggg attcttgcta gtcacactag	1079
ccatcccttac tgcgcctt	1096

<210> SEQ ID NO 12
 <211> LENGTH: 154
 <212> TYPE: PRT
 <213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 12

Met Met Pro Thr Thr Leu Phe Ala Gly Thr His Ile Thr Met Thr Thr	
1 5 10 15	
Val Tyr His Ile Thr Val Ser Gln Ile Gln Leu Ser Leu Leu Lys Val	
20 25 30	
Thr Ala Phe Gln His Gln Asn Ser Lys Lys Thr Thr Lys Leu Val Val	
35 40 45	
Ile Leu Arg Ile Gly Thr Gln Val Leu Lys Thr Met Ser Leu Tyr Met	
50 55 60	
Ala Ile Ser Pro Lys Phe Thr Thr Ser Leu Ser Leu His Lys Leu Leu	
65 70 75 80	
Gln Thr Leu Val Leu Lys Met Leu His Ser Ser Ser Leu Thr Ser Leu	
85 90 95	
Leu Lys Thr His Arg Met Cys Lys Tyr Thr Gln Ser Thr Ala Leu Gln	
100 105 110	
Glu Leu Leu Ile Gln Gln Trp Ile Gln Phe Met Met Ser Arg Arg Arg	
115 120 125	
Leu Leu Ala Cys Leu Cys Lys His Lys Lys Val Ser Thr Asn Leu Cys	
130 135 140	
Thr His Ser Phe Arg Lys Lys Gln Val Arg	
145 150	

<210> SEQ ID NO 13
 <211> LENGTH: 332
 <212> TYPE: DNA
 <213> ORGANISM: CORONAVIRUS
 <220> FEATURE:
 <221> NAME/KEY: CDS
 <222> LOCATION: (36)..(263)
 <223> OTHER INFORMATION:

<400> SEQUENCE: 13

tgcctttgta agcacaagaa agtgagtacg aactt atg tac tca ttc gtt tcg	53
Met Tyr Ser Phe Val Ser	
1 5	
gaa gaa aca ggt acg tta ata gtt aat agc gta ctt ctt ttt ctt gct	101
Glu Glu Thr Gly Thr Leu Ile Val Asn Ser Val Leu Leu Phe Leu Ala	

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10	15	20	
ttc gtg gta ttc ttg cta gtc aca cta gcc atc ctt act gcg ctt cga			149
Phe Val Val Phe Leu Leu Val Thr Leu Ala Ile Leu Thr Ala Leu Arg			
25	30	35	
ttg tgt gcg tac tgc tgc aat att gtt aac gtg agt tta gta aaa cca			197
Leu Cys Ala Tyr Cys Cys Asn Ile Val Asn Val Ser Leu Val Lys Pro			
40	45	50	
acg gtt tac gtc tac tcg cgt gtt aaa aat ctg aac tct tct gaa gga			245
Thr Val Tyr Val Tyr Ser Arg Val Lys Asn Leu Asn Ser Ser Glu Gly			
55	60	65	70
ggt cct gat ctt ctg gtc taaacgaact aactattatt attattctgt			293
Val Pro Asp Leu Leu Val			
75			
ttggaacttt aacattgctt atcatggcag acaacggta			332

<210> SEQ ID NO 14
 <211> LENGTH: 76
 <212> TYPE: PRT
 <213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 14

Met Tyr Ser Phe Val Ser Glu Glu Thr Gly Thr Leu Ile Val Asn Ser	
1	15
Val Leu Leu Phe Leu Ala Phe Val Val Phe Leu Leu Val Thr Leu Ala	
20	30
Ile Leu Thr Ala Leu Arg Leu Cys Ala Tyr Cys Cys Asn Ile Val Asn	
35	45
Val Ser Leu Val Lys Pro Thr Val Tyr Val Tyr Ser Arg Val Lys Asn	
50	60
Leu Asn Ser Ser Glu Gly Val Pro Asp Leu Leu Val	
65	75

<210> SEQ ID NO 15
 <211> LENGTH: 332
 <212> TYPE: DNA
 <213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 15

tgccctttgta agcacaagaa agtgagtacg aacttatgta ctcattcggt tcggaagaaa	60
caggtagcgtt aatagttaat agcgtacttc tttttcttgc tttcgtggta ttcttgctag	120
tcacactagc catccttact gcgcttcgat tgtgtgcgta ctgctgcaat attgttaacg	180
tgagtttagt aaaaccaacg gtttaagtct actcgcgtgt taaaaatctg aactcttctg	240
aaggagttcc tgatcttctg gtctaaacga actaactatt attattattc tgtttggaac	300
tttaacattg cttatcatgg cagacaacgg ta	332

<210> SEQ ID NO 16
 <211> LENGTH: 708
 <212> TYPE: DNA
 <213> ORGANISM: CORONAVIRUS
 <220> FEATURE:
 <221> NAME/KEY: CDS
 <222> LOCATION: (41)..(703)
 <223> OTHER INFORMATION:

<400> SEQUENCE: 16

tattattatt attctgtttg gaactttaac attgcttata atg gca gac aac ggt	55
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<210> SEQ ID NO 17
<211> LENGTH: 221
<212> TYPE: PRT
<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 17
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Met	Ala	Asp	Asn	Gly	Thr	Ile	Thr	Val	Glu	Glu	Leu	Lys	Gln	Leu	Leu
1				5					10					15	
Glu	Gln	Trp	Asn	Leu	Val	Ile	Gly	Phe	Leu	Phe	Leu	Ala	Trp	Ile	Met
			20					25					30		
Leu	Leu	Gln	Phe	Ala	Tyr	Ser	Asn	Arg	Asn	Arg	Phe	Leu	Tyr	Ile	Ile
		35					40					45			

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Lys Leu Val Phe Leu Trp Leu Leu Trp Pro Val Thr Leu Ala Cys Phe
 50 55 60
 Val Leu Ala Ala Val Tyr Arg Ile Asn Trp Val Thr Gly Gly Ile Ala
 65 70 75 80
 Ile Ala Met Ala Cys Ile Val Gly Leu Met Trp Leu Ser Tyr Phe Val
 85 90 95
 Ala Ser Phe Arg Leu Phe Ala Arg Thr Arg Ser Met Trp Ser Phe Asn
 100 105 110
 Pro Glu Thr Asn Ile Leu Leu Asn Val Pro Leu Arg Gly Thr Ile Val
 115 120 125
 Thr Arg Pro Leu Met Glu Ser Glu Leu Val Ile Gly Ala Val Ile Ile
 130 135 140
 Arg Gly His Leu Arg Met Ala Gly His Ser Leu Gly Arg Cys Asp Ile
 145 150 155 160
 Lys Asp Leu Pro Lys Glu Ile Thr Val Ala Thr Ser Arg Thr Leu Ser
 165 170 175
 Tyr Tyr Lys Leu Gly Ala Ser Gln Arg Val Gly Thr Asp Ser Gly Phe
 180 185 190
 Ala Ala Tyr Asn Arg Tyr Arg Ile Gly Asn Tyr Lys Leu Asn Thr Asp
 195 200 205
 His Ala Gly Ser Asn Asp Asn Ile Ala Leu Leu Val Gln
 210 215 220

<210> SEQ ID NO 18

<211> LENGTH: 769

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 18

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cctgatcttc tggctctaac gaactaacta ttattattat tctgtttgga actttaacat    60
tgcttatcat ggcagacaac ggtactatta ccgttgagga gcttaaacaa ctccctggaac    120
aatggaacct agtaaatggt ttcctattcc tagcctggat tatgttacta caatttgcct    180
attctaatacg gaacagggttt ttgtacataa taaagcttgt tttcctctgg ctcttggtggc    240
cagtaaacact tgcttgcttt gtgcttgctg ctgtctacag aattaattgg gtgactggcg    300
ggattgcgat tgcaatggct tgtattgtag gcttgatgtg gcttagctac ttcgttgctt    360
ccttcaggct gtttgctcgt acccgctcaa tgtggtcatt caaccagaa acaaacattc    420
ttctcaatgt gcctctccgg gggacaattg tgaccagacc gctcatggaa agtgaacttg    480
tcattggtgc tgtgatcatt cgtggtcact tgcgaatggc cggacactcc ctagggcgct    540
gtgacattaa ggacctgcca aaagagatca ctgtggctac atcacgaacg ctttcttatt    600
acaaattagg agcgtgcgag cgtgtaggca ctgattcagg ttttgctgca tacaaccgct    660
accgtattgg aaactataaa ttaaatacag accacgccgg tagcaacgac aatattgctt    720
tgctagtaca gtaagtgaca acagatgttt catcttggtg acttocagg                769

```

<210> SEQ ID NO 19

<211> LENGTH: 1231

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 19

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taccgtattg gaaactataa attaaatata gaccacgccg gtagcaacga caatattgct    60

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ttgctagtac agtaagtac aacagatggt tcactcttggt gacttccagg ttacaatagc 120
agagatattg attatcatta tgaggacttt caggattgct atttggaaac ttgacgttat 180
aataagttca atagtggagac aattatttta gcctctaact aagaagaatt attcggagtt 240
agatgatgaa gaacctatgg agttagatta tccataaaac gaacatgaaa attattctct 300
tcctgacatt gattgtatgt acatcttgcg agctatatca ctatcaggag tgtgttagag 360
gtacgactgt actactaaaa gaaccttgcc catcaggaaac atacgagggc aattcaccat 420
ttcacctctt tgctgacaat aaatttgac taacttgac tagcacacac ttgcttttg 480
ctgtgctga cggtagctga catacctatc agctgctgc aagatcagtt tcacaaaaac 540
ttttcatcag acaaggagg gttcaacaag agctctact gccacttttt ctcatgttg 600
ctgctctagt atttttaata ctttgcttca ccattaagag aaagacagaa tgaatgagct 660
cactttaatt gacttctatt tgtgttttt agcctttctg ctattccttg ttttaataat 720
gcttattata ttttggtttt cactcgaaat ccaggatcta gaagaacctt gtaccaaagt 780
ctaaacgaac atgaaacttc tcattgtttt gacttgattt tctctatgca gttgcatatg 840
cactgtagta cagcgctgtg catctaataa acctcatgtg cttgaagatc cttgtaaggt 900
acaacactag gggaataact tatagcactg cttggctttg tgctctagga aaggttttac 960
cttttcatag atggcacact atggttcaaa catgcacacc taatgttact atcaactgtc 1020
aagatccagc tgggtggtgc cttatagcta ggtgttgta ccttcatgaa ggtcaccaaa 1080
ctgctgcatt tagagacgta cttgttggtt taaataaacg aacaaattaa aatgtctgat 1140
aatggacccc aatcaaacca acgtagtgc cccgcatta catttggttg acccacagat 1200
tcaactgaca ataaccagaa tggaggacgc a 1231

```

<210> SEQ ID NO 20

<211> LENGTH: 1242

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 20

```

gcatacaacc gctaccgtat tggaaactat aaattaaata cagaccacgc cggtagcaac 60
gacaatattg ctttgctagt acagtaagt acaacagatg tttcatcttg ttgacttcca 120
ggttacaata gcagagatat tgattatcat tatgaggact ttcaggattg ctatttgga 180
tcttgacgtt ataataagtt caatagttag acagttatgt aagcctctaa ctaagaagaa 240
ttattcggag ttagatgatg aagaacctat ggagtttagat tatccataaa acgaacatga 300
aaattattct cttcctgaca ttgattgtat ttacatcttg cgagctatat cactatcagg 360
agtgtgttag aggtacgact gtactactaa aagaaccttg cccatcagga acatacagg 420
gcaattcacc atttcaccct cttgctgaca ataaatttgc actaacttgc actagcacac 480
actttgcttt tgcttggtgt gacggtagtc gacataccta tcagctgctg gcaagatcag 540
tttcacaaa acttttcatc agacaagagg aggttcaaca agagctctac tcgccacttt 600
ttctcattgt tgctgctcta gtatttttaa tactttgctt caccattaag agaagacag 660
aatgaatgag ctcaatttaa ttgacttcta tttgtgcttt ttagcctttc tgctattcct 720
tgttttaata atgcttatta tattttggtt ttactcga atccaggatc tagaagaacc 780
ttgtacaaa gtctaaacga acatgaaact tctcattgtt ttgacttgta tttctctatg 840

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cagttgcata tgcactgtag tacagcgctg tgcactaat aaacctcatg tgcttgaaga 900
tccttgtaag gtacaacact aggggtaata cttatagcac tgcttggtt tgtgctctag 960
gaaaggtttt accttttcat agatggcaca ctatggttca aacatgcaca cctaattgta 1020
ctatcaactg tcaagatcca gctggtggtg cgcttatagc taggtgttgg taccttcatg 1080
aaggtcacca aactgctgca tttagagacg tacttggtgt tttaaataaa cgaacgaatt 1140
aaaatgtctg ataattggacc ccaatcaaac caacgtagtg ccccccgcac tacatttggt 1200
ggacccacag attcaactga caataaccag aatggaggac gc 1242

```

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<210> SEQ ID NO 21
<211> LENGTH: 1231
<212> TYPE: DNA
<213> ORGANISM: CORONAVIRUS
<220> FEATURE:
<221> NAME/KEY: CDS
<222> LOCATION: (86)..(274)
<223> OTHER INFORMATION:

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<400> SEQUENCE: 21

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taccgtattg gaaacataaa attaaatata gaccacgccg gtacgaacga caatattgct 60
ttgctagtac agtaagtgc aacag atg ttt cat ctt gtt gac ttc cag gtt 112
                Met Phe His Leu Val Asp Phe Gln Val
                1                5
aca ata gca gag ata ttg att atc att atg agg act ttc agg att gct 160
Thr Ile Ala Glu Ile Leu Ile Ile Ile Met Arg Thr Phe Arg Ile Ala
10                15                20                25
att tgg aat ctt gac gtt ata ata agt tca ata gtg aga caa tta ttt 208
Ile Trp Asn Leu Asp Val Ile Ile Ser Ser Ile Val Arg Gln Leu Phe
                30                35                40
aag cct cta act aag aag aat tat tcg gag tta gat gat gaa gaa cct 256
Lys Pro Leu Thr Lys Lys Asn Tyr Ser Glu Leu Asp Asp Glu Glu Pro
                45                50                55
atg gag tta gat tat cca taaaacgaac atgaaaatta ttctcttcct 304
Met Glu Leu Asp Tyr Pro
60
gacattgatt gtatttacct cttgcgagct atatcactat caggagtgtg ttagaggtac 364
gactgtacta ctaaaagaac cttgcccatc aggaacatac gagggcaatt caccatttca 424
ccctcttgct gacaataaat ttgcactaac ttgcactagc acacactttg cttttgcttg 484
tgctgacggt actcgacata cctatcagct gcgtgcaaga tcagtttcac caaaactttt 544
catcagacaa gaggagggtc aacaagagct ctactcgcca ctttttctca ttgttgctgc 604
tctagtattt ttaatacttt gcttcacat taagagaaag acagaatgaa tgagctcact 664
ttaattgact tctatttggt ctttttagcc tttctgctat tccttggttt aataatgctt 724
attatatttt ggttttctc cgaaatccag gatctagaag aacctgttac caaagtctaa 784
acgaacatga aacttctcat tgttttgact tgtatttctc tatgcagttg catatgcact 844
gtagtacagc gctgtgcac taataaacct catgtgcttg aagatccttg taaggtaaa 904
cactaggggt aatacttata gcactgcttg gctttgtgct ctaggaaaag ttttaccttt 964
tcatagatgg cacactatgg ttcaaactg cacacctaata gttactatca actgtcaaga 1024
tccagctggt ggtgcgctta tagctagggt ttggtacett catgaaggtc accaaactgc 1084
tgcatttaga gacgtacttg ttgtttttaa taaacgaaca aattaaaatg tctgataatg 1144

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gaccccaatc aaaccaacgt agtgccccc gcattacatt tgggtggacc acagattcaa 1204

ctgacaataa ccagaatgga ggacgca 1231

<210> SEQ ID NO 22

<211> LENGTH: 63

<212> TYPE: PRT

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 22

Met Phe His Leu Val Asp Phe Gln Val Thr Ile Ala Glu Ile Leu Ile
1 5 10 15

Ile Ile Met Arg Thr Phe Arg Ile Ala Ile Trp Asn Leu Asp Val Ile
20 25 30

Ile Ser Ser Ile Val Arg Gln Leu Phe Lys Pro Leu Thr Lys Lys Asn
35 40 45

Tyr Ser Glu Leu Asp Asp Glu Pro Met Glu Leu Asp Tyr Pro
50 55 60

<210> SEQ ID NO 23

<211> LENGTH: 1231

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<220> FEATURE:

<221> NAME/KEY: CDS

<222> LOCATION: (285)..(650)

<223> OTHER INFORMATION:

<400> SEQUENCE: 23

taccgtattg gaaactataa attaaatata gaccacgccg gtagcaacga caatattgct 60

ttgctagtag agtaagtgc aacagatgtt tcattctgtt gacttccagg ttacaatagc 120

agagatattg attatcatta tgaggacttt caggattgct atttggaatc ttgacgttat 180

aataagttca atagtgcac aattatttaa gcctctaact aagaagaatt attcggagtt 240

agatgatgaa gaacctatgg agttagatta tccataaaac gaac atg aaa att att
Met Lys Ile Ile
1 296

ctc ttc ctg aca ttg att gta ttt aca tct tgc gag cta tat cac tat
Leu Phe Leu Thr Leu Ile Val Phe Thr Ser Cys Glu Leu Tyr His Tyr
5 10 15 20 344

cag gag tgt gtt aga ggt acg act gta cta cta aaa gaa cct tgc cca
Gln Glu Cys Val Arg Gly Thr Thr Val Leu Leu Lys Glu Pro Cys Pro
25 30 35 392

tca gga aca tac gag ggc aat tca cca ttt cac cct ctt gct gac aat
Ser Gly Thr Tyr Glu Gly Asn Ser Pro Phe His Pro Leu Ala Asp Asn
40 45 50 440

aaa ttt gca cta act tgc act agc aca cac ttt gct ttt gct tgt gct
Lys Phe Ala Leu Thr Cys Thr Ser Thr His Phe Ala Phe Ala Cys Ala
55 60 65 488

gac ggt act cga cat acc tat cag ctg cgt gca aga tca gtt tca cca
Asp Gly Thr Arg His Thr Tyr Gln Leu Arg Ala Arg Ser Val Ser Pro
70 75 80 536

aaa ctt ttc atc aga caa gag gag gtt caa caa gag ctc tac tcg cca
Lys Leu Phe Ile Arg Gln Glu Glu Val Gln Gln Glu Leu Tyr Ser Pro
85 90 95 100 584

ctt ttt ctc att gtt gct gct cta gta ttt tta ata ctt tgc ttc acc
Leu Phe Leu Ile Val Ala Ala Leu Val Phe Leu Ile Leu Cys Phe Thr
105 110 115 632

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att aag aga aag aca gaa tgaatgagct cactttaatt gacttctatt      680
Ile Lys Arg Lys Thr Glu
      120
tgtgcttttt agcctttctg ctattccttg ttttaataat gcttattata ttttggtttt      740
cactcgaaat ccaggatcta gaagaacctt gtaccaaagt ctaaacgaac atgaaacttc      800
tcattgtttt gacttgattt tctctatgca gttgcatatg cactgtagta cagcgctgtg      860
catctaataa acctcatgtg cttgaagatc cttgtaaggt acaacactag gggtaatact      920
tatagcactg cttggctttg tgctctagga aaggttttac cttttcatag atggcacact      980
atggttcaaa catgcacacc taatgttact atcaactgtc aagatccagc tgggtgtgctg      1040
cttatagcta ggtgttggtg ccttcatgaa ggtcaccaa ctgctgcatt tagagacgta      1100
cttggtgttt taaataaacg aacaaattaa aatgtctgat aatggacccc aatcaaacca      1160
acgtagtgcc ccccgcatca catttggttg acccacagat tcaactgaca ataaccagaa      1220
tggaggacgc a                                                    1231

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<210> SEQ ID NO 24
<211> LENGTH: 122
<212> TYPE: PRT
<213> ORGANISM: CORONAVIRUS

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<400> SEQUENCE: 24

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```

Met Lys Ile Ile Leu Phe Leu Thr Leu Ile Val Phe Thr Ser Cys Glu
 1             5             10             15
Leu Tyr His Tyr Gln Glu Cys Val Arg Gly Thr Thr Val Leu Leu Lys
      20             25             30
Glu Pro Cys Pro Ser Gly Thr Tyr Glu Gly Asn Ser Pro Phe His Pro
      35             40             45
Leu Ala Asp Asn Lys Phe Ala Leu Thr Cys Thr Ser Thr His Phe Ala
      50             55             60
Phe Ala Cys Ala Asp Gly Thr Arg His Thr Tyr Gln Leu Arg Ala Arg
      65             70             75             80
Ser Val Ser Pro Lys Leu Phe Ile Arg Gln Glu Glu Val Gln Gln Glu
      85             90             95
Leu Tyr Ser Pro Leu Phe Leu Ile Val Ala Ala Leu Val Phe Leu Ile
      100            105            110
Leu Cys Phe Thr Ile Lys Arg Lys Thr Glu
      115            120

```

```

<210> SEQ ID NO 25
<211> LENGTH: 1231
<212> TYPE: DNA
<213> ORGANISM: CORONAVIRUS
<220> FEATURE:
<221> NAME/KEY: CDS
<222> LOCATION: (650)..(781)
<223> OTHER INFORMATION:

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<400> SEQUENCE: 25

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taccgtattg gaaactataa attaaatata gaccacgccg gtagcaacga caatattgct      60
ttgctagtag agtaagtgtc aacagatgtt tcactctgtt gacttccagg ttacaatagc      120
agagatattg attatcatta tgaggacttt caggattgct atttggaatc ttgacgttat      180
aataagttca atagtgtgac aattatttaa gcctctaact aagaagaatt attcggagtt      240

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agatgatgaa gaacctatgg agttagatta tccataaaac gaacatgaaa attattctct 300
tcctgacatt gattgtatatt acatcttgccg agctatatca ctatcaggag tgtgttagag 360
gtacgactgt actactaaaa gaaccttgcc catcaggaaac atacgagggc aattcaccat 420
ttcacctct tgctgacaat aaatttgac taacttgac tagcacacac ttgcttttg 480
cttgtgctga cggctactga catacctatc agctgcgtgc aagatcagtt tcacaaaaac 540
ttttcatcag acaagaggag gttcaacaag agctctactc gccacttttt ctcatgttg 600
ctgctctagt atttttaata ctttgcttca ccattaagag aaagacaga atg aat gag 658
                                     Met Asn Glu
                                     1
ctc act tta att gac ttc tat ttg tgc ttt tta gcc ttt ctg cta ttc 706
Leu Thr Leu Ile Asp Phe Tyr Leu Cys Phe Leu Ala Phe Leu Leu Phe
   5             10             15

ctt gtt tta ata atg ctt att ata ttt tgg ttt tca ctc gaa atc cag 754
Leu Val Leu Ile Met Leu Ile Ile Phe Trp Phe Ser Leu Glu Ile Gln
  20             25             30             35

gat cta gaa gaa cct tgt acc aaa gtc taaacgaaca tgaaacttct 801
Asp Leu Glu Glu Pro Cys Thr Lys Val
          40

cattgttttg acttgtatatt ctctatgcag ttgcatatgc actgtagtac agcgctgtgc 861
atctaataaa cctcatgtgc ttgaagatcc ttgtaaggta caacactagg ggtaatactt 921
atagcactgc ttggctttgt gctctaggaa aggttttacc ttttcataga tggcacacta 981
tggttcaaac atgcacacct aatgttacta tcaactgtca agatccagct ggtgggtgcgc 1041
ttatagctag gtgttggtac cttcatgaag gtcaccaaac tgctgcattt agagacgtac 1101
ttgttgtttt aaataaacga acaaattaaa atgtctgata atggacccca atcaaaccaa 1161
cgtagtgccc cccgcattac atttggtgga cccacagatt caactgacaa taaccagaat 1221
ggaggacgca 1231

```

<210> SEQ ID NO 26
 <211> LENGTH: 44
 <212> TYPE: PRT
 <213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 26

```

Met Asn Glu Leu Thr Leu Ile Asp Phe Tyr Leu Cys Phe Leu Ala Phe
1             5             10             15

Leu Leu Phe Leu Val Leu Ile Met Leu Ile Ile Phe Trp Phe Ser Leu
20             25             30

Glu Ile Gln Asp Leu Glu Glu Pro Cys Thr Lys Val
35             40

```

<210> SEQ ID NO 27
 <211> LENGTH: 1231
 <212> TYPE: DNA
 <213> ORGANISM: CORONAVIRUS
 <220> FEATURE:
 <221> NAME/KEY: CDS
 <222> LOCATION: (791)..(907)
 <223> OTHER INFORMATION:

<400> SEQUENCE: 27

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taccgtattg gaaactataa attaaatata gaccacgccg gtagcaacga caatattgct 60

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ttgctagtagc agtaagtac aacagatggt tcatcttggt gacttccagg ttacaatagc 120
agagatattg attatcatta tgaggacttt caggattgct atttggaatc ttgacgttat 180
aataagttca atagttagac aattatntaa gcctctaact aagaagaatt attcggagtt 240
agatgatgaa gaacctatgg agttagatta tccataaaac gaacatgaaa attattctct 300
tcctgacatt gattgtatnt acatcttgcg agctatatca ctatcaggag tgtgttagag 360
gtacgactgt actactaaaa gaaccttgcc catcaggaac atacgagggc aattcaccat 420
ttcaccctct tgctgacaat aaatttgac taacttgac tagcacacac ttgcttttg 480
cttgtgctga cggtagctga catacctatc agctgctgac aagatcagtt tcacaaaaac 540
ttttcatcag acaaggagg gttcaacaag agctctactc gccacttttt ctcatgttg 600
ctgctctagt atttttaata ctttgcttca ccattaagag aaagacagaa tgaatgagct 660
cactttaatt gacttctatt tgtgcttttt agcctttctg ctattccttg ttttaataat 720
gcttattata ttttggtttt cactcgaaat ccaggatcta gaagaacctt gtaccaaagt 780
ctaaacgaac atg aaa ctt ctc att gtt ttg act tgt att tct cta tgc 829
      Met Lys Leu Leu Ile Val Leu Thr Cys Ile Ser Leu Cys
      1             5             10

agt tgc ata tgc act gta gta cag cgc tgt gca tct aat aaa cct cat 877
Ser Cys Ile Cys Thr Val Val Gln Arg Cys Ala Ser Asn Lys Pro His
      15             20             25

gtg ctt gaa gat cct tgt aag gta caa cac taggggtaat acttatagca 927
Val Leu Glu Asp Pro Cys Lys Val Gln His
      30             35

ctgcttggtt ttgtgctcta ggaaagggtt taccttttca tagatggcac actatgggtc 987
aaacatgcac acctaagtgt actatcaact gtcaagatcc agctgggtgt gcgcttatag 1047
ctaggtgttg gtaccttcat gaaggtcacc aaactgctgc atttagagac gtacttggtg 1107
ttttaaataa acgaacaaat taaaatgtct gataatggac cccaatcaaa ccaacgtagt 1167
gccccccgca ttacatttgg tggaccacac gattcaactg acaataacca gaatggagga 1227
cgca 1231

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<210> SEQ ID NO 28
 <211> LENGTH: 39
 <212> TYPE: PRT
 <213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 28

```

Met Lys Leu Leu Ile Val Leu Thr Cys Ile Ser Leu Cys Ser Cys Ile
1             5             10             15

```

```

Cys Thr Val Val Gln Arg Cys Ala Ser Asn Lys Pro His Val Leu Glu
      20             25             30

```

```

Asp Pro Cys Lys Val Gln His
      35

```

<210> SEQ ID NO 29
 <211> LENGTH: 1231
 <212> TYPE: DNA
 <213> ORGANISM: CORONAVIRUS
 <220> FEATURE:
 <221> NAME/KEY: CDS
 <222> LOCATION: (876)..(1127)
 <223> OTHER INFORMATION:

<400> SEQUENCE: 29

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taccgtattg gaaactataa attaaataca gaccacgccg gtagcaacga caatattgct    60
ttgctagtag agtaagtac aacagatgtt tcattctgtt gacttccagg ttacaatagc    120
agagatattg attatcatta tgaggacttt caggattgct atttggaatc ttgacgttat    180
aataagttca atagttagac aattatttaa gcctctaact aagaagaatt attcggagtt    240
agatgatgaa gaacctatgg agttagatta tccataaaac gaacatgaaa attattctct    300
tcctgacatt gattgtattt acatcttgcg agctatatca ctatcaggag tgtgttagag    360
gtacgactgt actactaaaa gaaccttgcc catcaggaac atacgagggc aattcaccat    420
ttcacccctct tgctgacaat aaatttgcac taacttgcac tagcacacac ttgcttttg    480
cttgtgctga cggctactga catacctatc agctgcgtgc aagatcagtt tcacaaaaac    540
ttttcatcag acaagaggag gttcaacaag agctctactc gccacttttt ctcatgtttg    600
ctgctctagt atttttaata ctttgcttca ccattaagag aaagacagaa tgaatgagct    660
cactttaatt gacttctatt tgtgcttttt agcctttctg ctattccttg ttttaataat    720
gcttattata ttttggtttt cactcgaaat ccaggatcta gaagaacctt gtaccaaagt    780
ctaaacgaac atgaaacttc tcattgtttt gacttgtatt tctctatgca gttgcatatg    840
cactgtagta cagcgcgtgt catctaataa acctc atg tgc ttg aag atc ctt    893
                               Met Cys Leu Lys Ile Leu
                               1             5

gta agg tac aac act agg ggt aat act tat agc act gct tgg ctt tgt    941
Val Arg Tyr Asn Thr Arg Gly Asn Thr Tyr Ser Thr Ala Trp Leu Cys
          10             15             20

gct cta gga aag gtt tta cct ttt cat aga tgg cac act atg gtt caa    989
Ala Leu Gly Lys Val Leu Pro Phe His Arg Trp His Thr Met Val Gln
          25             30             35

aca tgc aca cct aat gtt act atc aac tgt caa gat cca gct ggt ggt    1037
Thr Cys Thr Pro Asn Val Thr Ile Asn Cys Gln Asp Pro Ala Gly Gly
          40             45             50

gcg ctt ata gct agg tgt tgg tac ctt cat gaa ggt cac caa act gct    1085
Ala Leu Ile Ala Arg Cys Trp Tyr Leu His Glu Gly His Gln Thr Ala
          55             60             65             70

gca ttt aga gac gta ctt gtt gtt tta aat aaa cga aca aat    1127
Ala Phe Arg Asp Val Leu Val Val Leu Asn Lys Arg Thr Asn
          75             80

taaaatgtct gataatggac cccaatcaaa ccaacgtagt gccccccgca ttacatttgg    1187

tggaaccaca gattcaactg acaataacca gaatggagga cgca    1231

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<210> SEQ ID NO 30

<211> LENGTH: 84

<212> TYPE: PRT

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 30

```

Met Cys Leu Lys Ile Leu Val Arg Tyr Asn Thr Arg Gly Asn Thr Tyr
1             5             10             15

```

```

Ser Thr Ala Trp Leu Cys Ala Leu Gly Lys Val Leu Pro Phe His Arg
20             25             30

```

```

Trp His Thr Met Val Gln Thr Cys Thr Pro Asn Val Thr Ile Asn Cys
35             40             45

```

```

Gln Asp Pro Ala Gly Gly Ala Leu Ile Ala Arg Cys Trp Tyr Leu His
50             55             60

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Glu Gly His Gln Thr Ala Ala Phe Arg Asp Val Leu Val Val Leu Asn
65 70 75 80

Lys Arg Thr Asn

<210> SEQ ID NO 31

<211> LENGTH: 21221

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 31

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<210> SEQ ID NO 32
<211> LENGTH: 297
<212> TYPE: DNA
<213> ORGANISM: CORONAVIRUS

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<400> SEQUENCE: 32
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caactgacaa taaccagaat ggaggacgca atggggcaag gccaaaacag cgccgacccc 120
aagggtttacc caataatact gcgtcttgggt tcacagctct cactcagcat ggcaaggagg 180
aacttagatt ccctcgaggc cagggcgctc caatcaacac caatagtggt ccagatgacc 240
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<210> SEQ ID NO 33
<211> LENGTH: 98
<212> TYPE: PRT
<213> ORGANISM: CORONAVIRUS

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<400> SEQUENCE: 33
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Met Asp Pro Asn Gln Thr Asn Val Val Pro Pro Ala Leu His Leu Val
1           5           10          15
Asp Pro Gln Ile Gln Leu Thr Ile Thr Arg Met Glu Asp Ala Met Gly
20          25          30
Gln Gly Gln Asn Ser Ala Asp Pro Lys Val Tyr Pro Ile Ile Leu Arg
35          40          45
Leu Gly Ser Gln Leu Ser Leu Ser Met Ala Arg Arg Asn Leu Asp Ser
50          55          60

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Leu Glu Ala Arg Ala Phe Gln Ser Thr Pro Ile Val Val Gln Met Thr
65 70 75 80

Lys Leu Ala Thr Thr Glu Glu Leu Pro Asp Glu Phe Val Val Val Thr
85 90 95

Ala Lys

<210> SEQ ID NO 34
<211> LENGTH: 213
<212> TYPE: DNA
<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 34

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caagaaattc aactcctggc agcagtaggg gaaattctcc tgctcgaatg gctagcggag 180
gtggtgaaac tgccctcgcg ctattgctgc tag 213

<210> SEQ ID NO 35
<211> LENGTH: 70
<212> TYPE: PRT
<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 35

Met Leu Pro Pro Cys Tyr Asn Phe Leu Lys Glu Gln His Cys Gln Lys
1 5 10 15

Ala Ser Thr Gln Arg Glu Ala Glu Ala Val Lys Pro Leu Leu Ala
20 25 30

Pro His His Val Val Ala Val Ile Gln Glu Ile Gln Leu Leu Ala Ala
35 40 45

Val Gly Glu Ile Leu Leu Leu Glu Trp Leu Ala Glu Val Val Lys Leu
50 55 60

Pro Ser Arg Tyr Cys Cys
65 70

<210> SEQ ID NO 36
<211> LENGTH: 1377
<212> TYPE: DNA
<213> ORGANISM: CORONAVIRUS
<220> FEATURE:
<221> NAME/KEY: CDS
<222> LOCATION: (67)..(1335)
<223> OTHER INFORMATION:

<400> SEQUENCE: 36

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attaaa atg tct gat aat gga ccc caa tca aac caa cgt agt gcc ccc 108
Met Ser Asp Asn Gly Pro Gln Ser Asn Gln Arg Ser Ala Pro
1 5 10
cgc att aca ttt ggt gga ccc aca gat tca act gac aat aac cag aat 156
Arg Ile Thr Phe Gly Gly Pro Thr Asp Ser Thr Asp Asn Asn Gln Asn
15 20 25 30
gga gga cgc aat ggg gca agg cca aaa cag cgc cga ccc caa ggt tta 204
Gly Gly Arg Asn Gly Ala Arg Pro Lys Gln Arg Arg Pro Gln Gly Leu
35 40 45
ccc aat aat act gcg tct tgg ttc aca gct ctc act cag cat ggc aag 252
Pro Asn Asn Thr Ala Ser Trp Phe Thr Ala Leu Thr Gln His Gly Lys
50 55 60

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gtt cgt ggt ggt gac ggc aaa atg aaa gag ctc agc ccc aga tgg tac Val Arg Gly Gly Asp Gly Lys Met Lys Glu Leu Ser Pro Arg Trp Tyr 95 100 105 110	396
ttc tat tac cta gga act ggc cca gaa gct tca ctt ccc tac ggc gct Phe Tyr Tyr Leu Gly Thr Gly Pro Glu Ala Ser Leu Pro Tyr Gly Ala 115 120 125	444
aac aaa gaa ggc atc gta tgg gtt gca act gag gga gcc ttg aat aca Asn Lys Glu Gly Ile Val Trp Val Ala Thr Glu Gly Ala Leu Asn Thr 130 135 140	492
ccc aaa gac cac att ggc acc cgc aat cct aat aac aat gct gcc acc Pro Lys Asp His Ile Gly Thr Arg Asn Pro Asn Asn Ala Ala Thr 145 150 155	540
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agt cgc ggt aat tca aga aat tca act cct ggc agc agt agg gga aat Ser Arg Gly Asn Ser Arg Asn Ser Thr Pro Gly Ser Ser Arg Gly Asn 195 200 205	684
tct cct gct cga atg gct agc gga ggt ggt gaa act gcc ctc gcg cta Ser Pro Ala Arg Met Ala Ser Gly Gly Glu Thr Ala Leu Ala Leu 210 215 220	732
ttg ctg cta gac aga ttg aac cag ctt gag agc aaa gtt tct ggt aaa Leu Leu Leu Asp Arg Leu Asn Gln Leu Glu Ser Lys Val Ser Gly Lys 225 230 235	780
ggc caa caa caa caa ggc caa act gtc act aag aaa tct gct gct gag Gly Gln Gln Gln Gln Gly Gln Thr Val Thr Lys Lys Ser Ala Ala Glu 240 245 250	828
gca tct aaa aag cct cgc caa aaa cgt act gcc aca aaa cag tac aac Ala Ser Lys Lys Pro Arg Gln Lys Arg Thr Ala Thr Lys Gln Tyr Asn 255 260 265 270	876
gtc act caa gca ttt ggg aga cgt ggt cca gaa caa acc caa gga aat Val Thr Gln Ala Phe Gly Arg Arg Gly Pro Glu Gln Thr Gln Gly Asn 275 280 285	924
ttc ggg gac caa gac cta atc aga caa gga act gat tac aaa cat tgg Phe Gly Asp Gln Asp Leu Ile Arg Gln Gly Thr Asp Tyr Lys His Trp 290 295 300	972
ccg caa att gca caa ttt gct cca agt gcc tct gca ttc ttt gga atg Pro Gln Ile Ala Gln Phe Ala Pro Ser Ala Ser Ala Phe Phe Gly Met 305 310 315	1020
tca cgc att ggc atg gaa gtc aca cct tcg gga aca tgg ctg act tat Ser Arg Ile Gly Met Glu Val Thr Pro Ser Gly Thr Trp Leu Thr Tyr 320 325 330	1068
cat gga gcc att aaa ttg gat gac aaa gat cca caa ttc aaa gac aac His Gly Ala Ile Lys Leu Asp Asp Lys Asp Pro Gln Phe Lys Asp Asn 335 340 345 350	1116
gtc ata ctg ctg aac aag cac att gac gca tac aaa aca ttc cca cca Val Ile Leu Leu Asn Lys His Ile Asp Ala Tyr Lys Thr Phe Pro Pro 355 360 365	1164

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aca gag cct aaa aag gac aaa aag aaa aag act gat gaa gct cag cct 1212
Thr Glu Pro Lys Lys Asp Lys Lys Lys Lys Thr Asp Glu Ala Gln Pro
      370                      375                      380

ttg ccg cag aga caa aag aag cag ccc act gtg act ctt ctt cct gcg 1260
Leu Pro Gln Arg Gln Lys Lys Gln Pro Thr Val Thr Leu Leu Pro Ala
      385                      390                      395

gct gac atg gat gat ttc tcc aga caa ctt caa aat tcc atg agt gga 1308
Ala Asp Met Asp Asp Phe Ser Arg Gln Leu Gln Asn Ser Met Ser Gly
      400                      405                      410

gct tct gct gat tca act cag gca taa acactcatga tgaccacaca 1355
Ala Ser Ala Asp Ser Thr Gln Ala
415                      420

aggcagatgg gctatgtaaa cg 1377

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<210> SEQ ID NO 37
<211> LENGTH: 422
<212> TYPE: PRT
<213> ORGANISM: CORONAVIRUS

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<400> SEQUENCE: 37

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Thr Phe Gly Gly Pro Thr Asp Ser Thr Asp Asn Asn Gln Asn Gly Gly
20          25          30

Arg Asn Gly Ala Arg Pro Lys Gln Arg Arg Pro Gln Gly Leu Pro Asn
35          40          45

Asn Thr Ala Ser Trp Phe Thr Ala Leu Thr Gln His Gly Lys Glu Glu
50          55          60

Leu Arg Phe Pro Arg Gly Gln Gly Val Pro Ile Asn Thr Asn Ser Gly
65          70          75          80

Pro Asp Asp Gln Ile Gly Tyr Tyr Arg Arg Ala Thr Arg Arg Val Arg
85          90          95

Gly Gly Asp Gly Lys Met Lys Glu Leu Ser Pro Arg Trp Tyr Phe Tyr
100         105         110

Tyr Leu Gly Thr Gly Pro Glu Ala Ser Leu Pro Tyr Gly Ala Asn Lys
115         120         125

Glu Gly Ile Val Trp Val Ala Thr Glu Gly Ala Leu Asn Thr Pro Lys
130         135         140

Asp His Ile Gly Thr Arg Asn Pro Asn Asn Asn Ala Ala Thr Val Leu
145         150         155         160

Gln Leu Pro Gln Gly Thr Thr Leu Pro Lys Gly Phe Tyr Ala Glu Gly
165         170         175

Ser Arg Gly Gly Ser Gln Ala Ser Ser Arg Ser Ser Ser Arg Ser Arg
180         185         190

Gly Asn Ser Arg Asn Ser Thr Pro Gly Ser Ser Arg Gly Asn Ser Pro
195         200         205

Ala Arg Met Ala Ser Gly Gly Gly Glu Thr Ala Leu Ala Leu Leu Leu
210         215         220

Leu Asp Arg Leu Asn Gln Leu Glu Ser Lys Val Ser Gly Lys Gly Gln
225         230         235         240

Gln Gln Gln Gly Gln Thr Val Thr Lys Lys Ser Ala Ala Glu Ala Ser
245         250         255

Lys Lys Pro Arg Gln Lys Arg Thr Ala Thr Lys Gln Tyr Asn Val Thr

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260	265	270
Gln Ala Phe Gly Arg Arg Gly Pro Glu Gln Thr Gln Gly Asn Phe Gly		
275	280	285
Asp Gln Asp Leu Ile Arg Gln Gly Thr Asp Tyr Lys His Trp Pro Gln		
290	295	300
Ile Ala Gln Phe Ala Pro Ser Ala Ser Ala Phe Phe Gly Met Ser Arg		
305	310	315
Ile Gly Met Glu Val Thr Pro Ser Gly Thr Trp Leu Thr Tyr His Gly		
325	330	335
Ala Ile Lys Leu Asp Asp Lys Asp Pro Gln Phe Lys Asp Asn Val Ile		
340	345	350
Leu Leu Asn Lys His Ile Asp Ala Tyr Lys Thr Phe Pro Pro Thr Glu		
355	360	365
Pro Lys Lys Asp Lys Lys Lys Lys Thr Asp Glu Ala Gln Pro Leu Pro		
370	375	380
Gln Arg Gln Lys Lys Gln Pro Thr Val Thr Leu Leu Pro Ala Ala Asp		
385	390	395
Met Asp Asp Phe Ser Arg Gln Leu Gln Asn Ser Met Ser Gly Ala Ser		
405	410	415
Ala Asp Ser Thr Gln Ala		
420		

<210> SEQ ID NO 38

<211> LENGTH: 1377

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 38

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atgaagggtca ccaaactgct gcatttagag acgtacttgt tgttttaaat aaacgaacaa    60
attaaaaatgt ctgataatgg accccaatca aaccaacgta gtgccccccg cattacattt    120
ggtggaccca cagattcaac tgacaataac cagaatggag gacgcaatgg ggcaaggcca    180
aaacagcgcc gacccaagg ttacccaat aatactgcgt cttggttcac agctctcact    240
cagcatggca aggaggaaact tagattccct cgaggccagg gcgttccaat caacaccaat    300
agtgggtccag atgaccaaact tggctactac cgaagagcta cccgacgagt tcgtgggtgg    360
gacggcaaaa tgaaagagct cagcccaga tggctacttct attacctagg aactggccca    420
gaagcttcac ttccctacgg cgctaacaaa gaaggcatcg tatgggttgc aactgaggga    480
gccttgaata cacccaaaga ccacattggc acccgcaatc ctaataacaa tgctgccacc    540
gtgctacaac ttctcaagg aacaacattg ccaaaaggct tctacgcaga gggaagcaga    600
ggcggcagtc aagcctcttc tcgctctcca tcacgtagtc gcggtaatc aagaaattca    660
actcctggca gcagtagggg aaattctcct gctcgaatgg ctacgaggag tggtgaaact    720
gccctcgcg c tattgtgct agacagattg aaccagcttg agagcaaagt ttctggtaaa    780
ggccaacaac aacaaggcca aactgtcact aagaaatctg ctgctgaggc atctaaaaag    840
cctcgccaaa aacgtactgc cacaaaacag tacaacgtca ctcaagcatt tgggagacgt    900
ggtccagaac aaaccaagg aaatttcggg gaccaagacc taatcagaca aggaactgat    960
tacaacatt gccgcgaaat tgcacaattt gctccaagtg cctctgcatt ctttggaatg    1020
tcacgcattg gcatggaagt cacaccttcg ggaacatggc tgacttatca tggagccatt    1080

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aaattggatg acaaagatcc acaattcaaa gacaacgtca tactgctgaa caagcacatt	1140
gacgcataca aaacattccc accaacagag cctaaaaagg acaaaaagaa aaagactgat	1200
gaagctcagc ctttgccgca gagacaaaag aagcagccca ctgtgactct tcttcctgcg	1260
gctgacatgg atgattttctc cagacaactt caaaattcca tgagtggagc ttctgctgat	1320
tcaactcagg cataaacact catgatgacc acacaaggca gatgggctat gtaaacg	1377

<210> SEQ ID NO 39
 <211> LENGTH: 204
 <212> TYPE: DNA
 <213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 39

atattagggtt ttacctacc caggaaaagc caaccaacct cgatctcttg tagatctgtt	60
ctctaaacga actttaaaat ctgtgtagct gtcgctcggc tgcattgccta gtgcacctac	120
gcagtataaa caataataaa ttttactgtc gttgacaaga aacgagtaac tcgtccctct	180
tctgcagact gcttacggtt tcgt	204

<210> SEQ ID NO 40
 <211> LENGTH: 809
 <212> TYPE: DNA
 <213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 40

actcaagcat ttgggagacg tgggtccagaa caaacccaag gaaatttcgg ggaccaagac	60
ctaatacagc aaggaactga ttacaacatc tggccgcaaa ttgcacaatt tgctccaagt	120
gcctctgcat tctttggaat gtcacgcatt ggcatggaag tcacaccttc gggaacatgg	180
ctgacttatac atggagccat taaattggat gacaaagatc cacaattcaa agacaacgtc	240
atactgctga acaagcacat tgacgcatac aaaacattcc caccaacaga gcctaaaaag	300
gacaaaaaga aaaagactga tgaagctcag cctttgccgc agagacaaaa gaagcagccc	360
actgtgactc ttcttcctgc ggctgacatg gatgatttct ccagacaact tcaaaattcc	420
atgagtggag cttctgctga ttcaactcag gcataaacac tcattgatgac cacacaaggc	480
agatgggcta tgtaaacggt ttcgcaattc cgtttacgat acatagtcta ctcttggtga	540
gaatgaattc tcgtaactaa acagcacaag taggtttagt taactttaat ctcacatagc	600
aatctttaat caatgtgtaa cattagggag gacttgaaag agccaccaca ttttcacga	660
ggccacgcgg agtacgatc aggggtacagt gaataatgct agggagagct gcctatatgg	720
aagagcccta atgtgtataa ttaattttag tagtgctatc cccatgtgat ttaaatagct	780
tcttaggaga atgacaaaaa aaaaaaaaa	809

<210> SEQ ID NO 41
 <211> LENGTH: 448
 <212> TYPE: DNA
 <213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 41

aatgaacaca tagggctgtt caagctgggg cagtacgcct tttccagct ctactagacc	60
acaagtgcc aTTTTGAGGT gttcaagtgc ctccgatagg gcctcttcca cagagtcgcc	120
gaagccacgc actagcacgt ctctaacctg aaggacaggc aaactgagtt ggacgtgtgt	180

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tttctcgttg acaccaagaa caaggctctc catcttacct ttcggtcaca cccggacgaa	240
acctaggtat gctgatgac gactgcaaca cggacgaaac cgtaagcagt ctgcagaaga	300
gggacgagtt actcgtttct tgtcaacgac agtaaaatctt attattgttt atactgcgta	360
ggtgcactag gcatgcagcc gagcgacagc tacacagatt ttaaagttcg tttagagAAC	420
agatctacaa gagatcgagg ttggttgg	448

<210> SEQ ID NO 42

<211> LENGTH: 2033

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 42

atacctagggt ttcgtccggg tgtgaccgaa aggtaagatg gagagccttg ttcttggtgt	60
caacgagaaa acacacgtcc aactcagttt gcctgtcctt caggtagag acgtgctagt	120
gcgtggcttc ggggactctg tggaaagagg cctatcggag gcacgtgaac acctcaaaaa	180
tggcacttgt ggtctagtag agctggaaaa aggcgtactg cccagcttg aacagcccta	240
tgtgttcatt aaacgttctg atgccttaag caccaatcac ggccacaagg tcgttgagct	300
ggttgacgaa atggacggca ttcagtacgg tcgtagcggg ataacactgg gactactcgt	360
gccacatgtg ggcgaaaccc caattgcata ccgcaatgtt cttcttcgta agaacggtaa	420
taagggagcc ggtggtcata gctatggcat cgtactaaag tcttatgact taggtgacga	480
gcttgccact gatcccattg aagattatga acaaaactgg aacactaagc atggcagtgg	540
tgcactccgt gaactcactc gtgagctcaa tggaggtgca gtcactcgt atgtcgacaa	600
caatttctgt ggcccagatg ggtaccctct tgattgcac aaagattttc tcgcacgcgc	660
gggcaagtca atgtgcactc tttccgaaca acttgattac atcgagtcga agagaggtgt	720
ctactgctgc cgtgaccatg agcatgaaat tgcctggttc actgagcgt ctgataagag	780
ctacgagcac cagacaccct tcgaaattaa gaggccaag aaatttgaca ctttcaaagg	840
ggaatgcccc aagtttgtgt ttcctcttaa ctcaaaagtc aaagtcattc aaccacgtgt	900
tgaaaagaaa aagactgagg gtttcatggg gcgtatacgc tctgtgtacc ctgttgcatc	960
tccacaggag tgaacaata tgcacttgtc taccttgatg aaatgtaac attgcgatga	1020
agtttcatgg cagacgtgag actttctgaa agccacttgt gaacattgtg gactgaaaa	1080
tttagttatt gaaggacctc ctacatgtgg gtacctacct actaatgctg tagtgaaaat	1140
gccatgtcct gcctgtcaag acccagagat tggacctgag catagtgttg cagattatca	1200
caaccactca aacattgaaa ctgcactcgg caagggaggt aggactagat gttttggagg	1260
ctgtgtgttt gcctatgttg gctgctataa taagcgtgcc tactgggttc ctctgctag	1320
tgctgatatt ggctcaggcc atactggcat tactgggtgac aatgtggaga ccttgaatga	1380
ggatctcctt gagatactga gtcgtgaacg tgttaacatt aacattgttg gcgattttca	1440
tttgaatgaa gaggttgcca tcattttggc atctttctct gcttctacaa gtgcctttat	1500
tgacactata aagagtcttg attacaagtc tttcaaaacc attgttgagt cctgcggtaa	1560
ctataaagtt accaagggaa agcccgtaaa aggtgcttgg aacattggac aacagagatc	1620
agttttaaca ccactgtgtg gttttccctc acaggctgct ggtgttatca gatcaatttt	1680
tgcgcgacaa cttagtcgag caaacctcct aattcctgat ttgcaaagag cagctgtcac	1740

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catacttgat ggtatttctg aacagtcatt acgtcttgtc gacgccatgg tttatacttc	1800
agacctgctc accaacagtg tcattattat ggcatatgta actggtggtc ttgtacaaca	1860
gacttctcag tggttgtcta atcttttggg cactactggt gaaaaactca ggcctatctt	1920
tgaatggatt gaggcgaaac ttagtgacag agttgaattt ctcaaggatg ctggggagat	1980
tctcaaattt ctcatcacag gtgtttttga catcgtcaag ggtcaaatac agg	2033

<210> SEQ ID NO 43

<211> LENGTH: 2018

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 43

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aatttctcat tacagggtgtt ttgacatcg tcaagggtca aatacagggt gcttcagata	120
acatcaagga ttgtgtaaaa tgcttcattg atgttggtta caaggcactc gaaatgtgca	180
ttgatcaagt cactatcgct ggcgcaaagt tgcgatcact caacttaggt gaagtcttca	240
tcgctcaaa gcaaggactt taccgtcagt gtatcgtgg caaggagcag ctgcaactac	300
tcatgcctct taaggcacca aaagaagtaa cctttcttga aggtgattca catgacacag	360
tacttacctc tgaggaggtt gttctcaaga acggtgaact cgaagcactc gagacgccc	420
ttgatagctt cacaaatgga gctatcgttg gcacaccagt ctgtgtaaat ggcctcatgc	480
tcttagagat taaggacaaa gaacaatact gcgcattgtc tcctggttta ctggctacaa	540
acaatgtctt tcgcttaaaa ggggtgcac caattaaagg tgtaaccttt ggagaagata	600
ctgtttggga agttcaaggt tacaagaatg tgagaatcac atttgagctt gatgaacgtg	660
ttgacaaagt gcttaatgaa aagtgtctg tctacactgt tgaatccggt accgaagtta	720
ctgagtttgc atgtgttgta gcagaggctg ttgtgaagac tttacaacca gtttctgac	780
tccttaccaa catgggtatt gatcttgatg agtgagagtg agctacattc tacttatttg	840
atgatgctg tgaagaaaac ttttcatcac gtatgtattg ttccttttac cctccagatg	900
aggaagaaga ggacgatgca gagtgtgagg aagaagaaat tgatgaaacc tgtgaacatg	960
agtacggtac agaggatgat tatcaaggtc tccctctgga atttggtgcc tcagctgaaa	1020
cagttcgagt tgaggaagaa gaagaggaag actggctgga tgatactact gagcaatcag	1080
agattgagcc agaaccagaa cctacacctg aagaaccagt taatcagttt actggttatt	1140
taaaacttac tgacaatgtt gccattaaat gtgttgacat cgttaaggag gcacaaagt	1200
ctaatactat ggtgattgta aatgtgtgta acatacacct gaaacatggg ggtgggtgag	1260
cagggtgact caacaaggca accaatgtg ccatgcaaaa ggagagtgat gattacatta	1320
agctaaatgg ccctcttaca gtaggagggg cttgtttgct ttctggacat aatcttgcta	1380
agaagtgtct gcatgtgtt ggacctaacc taaatgcagg tgaggacatc cagcttctta	1440
aggcagcata tgaaaatttc aattcacagg acatcttact tgcaccattg ttgtcagcag	1500
gcataatttg tgctaaacca cttcagtcct tacaagtgtg cgtgcagacg gttcgtacac	1560
aggtttatat tgcagtcaat gacaaagctc tttatgagca ggttgcacat gattatcttg	1620
ataacctgaa gcctagagtg gaagcaccta aacaagagga gccaccaaac acagaagatt	1680
ccaaaactga ggagaaatct gtcgtacaga agcctgtcga tgtgaagcca aaaattaagg	1740

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cctgcattga tgaggttacc acaacactgg aagaaactaa gtttcttacc aataagttac	1800
tcttgtttgc tgatatcaat ggtaagcttt accatgattc tcagaacatg cttagagggtg	1860
aagatatgtc tttccttgag aaggatgcac cttacatggt aggtgatggt atcactagt	1920
gtgatatac ttgtgttgta ataccctcca aaaaggctgg tggcactact gagatgctct	1980
caagagcttt gaagaaagtg ccagttgatg agtatata	2018

<210> SEQ ID NO 44
 <211> LENGTH: 1442
 <212> TYPE: DNA
 <213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 44

ttgatgagggt taccacaaca ctggaagaaa ctaagtttct taccaataag ttactcttgt	60
ttgctgatat caatggtaag ctttaccatg attctcagaa catgcttaga ggtgaagata	120
tgtctttcct tgagaaggat gcaccttaca tggtaggtga tgttatcact agtggtgata	180
tcacttgtgt tgtaataccc tccaaaaagg ctggtggcac tactgagatg ctctcaagag	240
ctttgaagaa agtgccaggt gatgagtata taaccacgta ccctggacaa ggatgtgctg	300
gttatacact tgaggaagct aagactgctc ttaagaaatg caaatctgca ttttatgtac	360
taccttcaga agcacctaata gctaagggaag agattctagg aactgtatcc tggaatttga	420
gagaaatgct tgctcatgct gaagagacaa gaaaattaat gcctatatgc atggatgtta	480
gagccataat ggcaaccatc caacgtaagt ataaaggaat taaaattcaa gagggcatcg	540
ttgactatgg tgtccgattc ttcttttata ctagttaaaga gcctgtagct tctattatta	600
cgaagctgaa ctctctaaat gagccgcttg tcacaatgcc aattgggtat gtgacacatg	660
gttttaatat tgaagaggct gcgcgctgta tgcgttctct taaagctcct gccgtagtgt	720
cagtatcatc accagatgct gttactacat ataatggata cctcacttcg tcatcaaaga	780
catctgagga gcactttgta gaaacagttt ctttggctgg ctcttacaga gattggtcct	840
attcaggaca gcgtacagag ttagggttg aatttcttaa gcgtggtgac aaaattgtgt	900
accacactct ggagagcccc gtcgagtttc atcttgacgg tgaggttctt tcacttgaca	960
aactaaagag tctcttatcc ctgcgggagg ttaagactat aaaagtgttc acaactgttg	1020
acaacactaa tctccacaca cagcttggtg atatgtctat gacatatgga cagcagttg	1080
gtccaacata cttggatggt gctgatgtta caaaaattaa acctcatgta aatcatgagg	1140
gtaagacttt ctttgtacta cctagtgtat acacactacg tagtgaagct ttcgagtact	1200
accatactct tgatgagagt tttcttggtg ggtacatgct tgctttaaac cacacaaaga	1260
aatggaaatt tcctcaagtt ggtggtttta cttcaattaa atgggctgat aacaattgtt	1320
atttgtctag tgttttatta gcacttcaac agcttgaagt caaattcaat gcaccagcac	1380
ttcaagaggc ttattataga gcccggtgctg gtgatgctgc taacttttgt gcactcatac	1440
tc	1442

<210> SEQ ID NO 45
 <211> LENGTH: 1050
 <212> TYPE: DNA
 <213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 45

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atatgtctat gacatatgga cagcagtttg gtccaacata cttggatggt gctgatgtta	60
caaaaattaa acctcatgta aatcatgagg gtaagacttt ctttgtaacta cctagtgatg	120
acacactacg tagtgaagct ttcgagtact accatactct tgatgagagt tttcttggtta	180
ggtacatgtc tgctttaaac cacacaaaga aatggaaatt tcctcaagtt ggtgggttaa	240
cttcaattaa atgggctgat aacaattggt atttgtctag tgttttatta gcacttcaac	300
agcttgaagt caaattcaat gcaccagcac ttcaagaggc ttattataga gcccgctgtg	360
gtgatgctgc taacttttgt gcactcatac tcgcttacag taataaaaact gttggcgagc	420
ttggtgatgt cagagaaact atgacccatc ttctacagca tgctaatttg gaatctgcaa	480
agcgagtctt taatgtgggt tgtaaacatt gtggtcagaa aactactacc ttaacgggtg	540
tagaagctgt gatgtatatg ggtactctat cttatgataa tcttaagaca ggtgtttcca	600
ttccatgtgt gtgtggctgt gatgctacac aatatctagt acaacaagag tcttcttttg	660
ttatgatgtc tgcaccacct gctgagtata aattacagca aggtacattc ttatgtgcga	720
atgagtacac tggttaactat cagtgtgggtc attacactca tataactgct aaggagaccc	780
tctatcgtat tgacggagct caccttaca agatgtcaga gtacaaagga ccagtgactg	840
atgttttcta caaggaaaca tcttacacta caaccatcaa gcctgtgtcg tataaactcg	900
atggagttac ttacacagag attgaacaa aattggatgg gtattataaa aaggataatg	960
cttactatac agagcagcct atagaccttg taccaactca accattacca aatgcgagtt	1020
ttgataatth caaactcaca tgttctaaca	1050

<210> SEQ ID NO 46

<211> LENGTH: 1995

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 46

tttgtgcact catactcgct tacagtaata aaactggttg cgagcttggt gatgtcagag	60
aaactatgac ccatcttcta cagcatgcta atttggaatc tgcaaagcga gttcttaatg	120
tggtgtgttaa acattgtggt cagaaaacta ctaccttaac ggggttagaa gctgtgatgt	180
atatgggtac tctatcttat gataatctta agacagggtt ttccattcca tgtgtgtgtg	240
gtcgtgatgc tacacaatat ctagtacaac aagagtcttc tttgtttatg atgtctgcac	300
cacctgctga gtataaatta cagcaaggta cattcttatg tgcgaatgag tacactggta	360
actatcagtg tggtcattac actcatataa ctgctaagga gacctctat cgtattgacg	420
gagctcacct tacaaagatg tcagagtaca aaggaccagt gactgatgtt ttctacaagg	480
aaacatctta cactacaacc atcaagcctg tgtcgtataa actcgatgga gttacttaca	540
cagagattga accaaaattg gatgggtatt ataaaaagga taatgcttac tatacagagc	600
agcctataga cttgtacca actcaacat taccaaatgc gagttttgat aatttcaaac	660
tcacatgttc taacacaaaa tttgtgatg atttaaatca aatgacaggc ttcacaaagc	720
cagcttcacg agagctatct gtcacattct tcccagactt gaatggcgat gtagtggcta	780
ttgactatag acactattca gcgagtttca agaaagggtc taaattactg cataagccaa	840
ttgtttggca cattaaccag gctacaacca agacaacgtt caaaccacac acttggtgtt	900
tacgttgtct ttggagtaca aagccagtag atacttcaaa ttcatttgaa gttctggcag	960

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tagaagacac acaaggaatg gacaatcttg cttgtgaaag tcaacaaccc acctctgaag	1020
aagtagtgga aaatcctacc atacagaagg aagtcataga gtgtgacgtg aaaactaccg	1080
aagttgtagc caatgtcata cttaaaccat cagatgaagg tggtaaagta acacaagagt	1140
taggtcatga ggatcttatg gctgcttatg tggaaaacac aagcattacc attaagaaac	1200
ctaatagtgt ttcactagcc ttaggtttta aaacaattgc cactcatggg attgctgcaa	1260
ttaatagtgt tccttgaggt aaaatcttg cttatgtcaa accattctta ggacaagcag	1320
caattacaac atcaaattgc gctaagagat tagcacaacg tgtgtttaac aattatatgc	1380
cttatgtgtt tacattattg ttccaattgt gtacttttac taaaagtacc aattctagaa	1440
ttagagcttc actacctaca actattgcta aaaatagtgt taagagtgtt gctaaattat	1500
gtttggatgc cggcattaat tatgtgaagt caccctaaatt ttctaaattg ttcacaatcg	1560
ctatgtggct attgttggtta agtatttgct taggttctct aatctgtgta actgctgctt	1620
ttggtgtact cttatctaatt tttggtgctc cttcttattg taatggcggt agagaattgt	1680
atcttaattc gtctaacgtt actactatgg atttctgtga aggttctttt ccttcagca	1740
tttgtttaag tggattagac tcccttgatt cttatccagc tcttgaaacc attcaggtga	1800
cgatttcacg gtacaagcta gacttgacaa ttttaggtct ggccgctgag tgggttttg	1860
catatatgtt gttcacaaaa ttcttttatt tattaggtct ttcagctata atgcaggtgt	1920
tctttggcta ttttgctagt catttcatca gcaattcttg gctcatgtgg tttatcatta	1980
gtattgtaca aatgg	1995

<210> SEQ ID NO 47

<211> LENGTH: 1884

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 47

aattcttggc tcatgtgggt tatcattagt attgtacaaa tggcaccgtt ttctgcaatg	60
gttaggatgt acatcttctt tgcttcttct tactacatat ggaagagcta gttcatatc	120
atggatgggt gcacctcttc gacttgcatg atgtgctata agcgcaatcg tgccacacgc	180
gttgagtgtg caactattgt taatggcatg aagagatctt tctatgtcta tgcaaatgga	240
ggccgtggct tctgcaagac tcacaattgg aattgtctca attgtgacac attttgcact	300
ggtagtacat tcattagtga tgaagttgct cgtgatttgt cactccagtt taaaagacca	360
atcaacccta ctgaccagtc atcgtatatg gttgatatgt ttgctgtgaa aaatggcgcg	420
cttcacctct actttgacaa ggctgggtcaa aagacctatg agagacatcc gctctcccat	480
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atagtttttg atggcaagtc caaatgcgac gagtctgctt ctaagtctgc ttctgtgtac	600
tacagtcagc tgatgtgcca acctattctg ttgcttgacc aagctcttgt atcagacgtt	660
ggagatagta ctgaagtttc cgttaagatg tttgatgctt atgtcgacac cttttcagca	720
acttttagtg ttcctatgga aaaacttaag gcacttggtg ctacagctca cagcgagtta	780
gcaaagggtg tagcttttaga tgggtgtcctt tctacattcg tgtcagctgc ccgacaagg	840
gttgttgata ccgatgttga cacaaggat gttattgaat gtctcaaaact ttcacatcac	900
tctgacttag aagtgcagcg tgacagttgt aacaatttca tgctcaccta taataagggt	960

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gaaaacatga cgcccagaga tcttggcgca tgtattgact gtaatgcaag gcatatcaat	1020
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ttatctgaac agctgcgtaa acaaattcgt agtgctgccca agaagaacaa catacctttt	1140
agactaactt gtgtacaac tagacagggt gtcaatgtca taactactaa aatctcactc	1200
aagggtggta agattgttag tacttgtttt aaacttatgc ttaaggccac attattgtgc	1260
gttcttgctg cattggtttg ttatatcggt atgccagtac atacattgtc aatccatgat	1320
ggttacacaa atgaaatcat tggttacaaa gccattcagg atggtgtcac tcgtgacatc	1380
atttctactg atgattgttt tgcaataaaa catgctgggt ttgacgcatg gtttagccag	1440
cgtggtggtt catacaaaaa tgacaaaagc tgccctgtag tagctgctat cattacaaga	1500
gagattgggt tcatagtgcc tggcttaccg ggtactgtgc tgagagcaat caatggtgac	1560
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aaactcattg agtatagtga ttttgctacc tctgcttgcg ttcttgctgc tgagtgtaca	1680
atttttaagg atgctatggg caaacctgtg ccatattggt atgacactaa ttgctagag	1740
ggttctatct cttatagtga gcttcgtcca gacactcgtt atgtgcttat ggatggttcc	1800
atcatacagt ttcctaacac ttacctggag ggttctgtta gagtagtaac aacttttgat	1860
gctgagtact gtagacatgg taca	1884

<210> SEQ ID NO 48

<211> LENGTH: 2020

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 48

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ttctgttaga gtagtaacaa cttttgatgc tgagtactgt agacatggta catgcgaaag	120
gtcagaagta ggtatttgcc tatctaccag tggtagatgg gttcttaata atgagcatta	180
cagagctcta tcaggagttt tctgtggtgt tgatgcgatg aatctcatag ctaacatctt	240
tactcctctt gtgcaacctg tgggtgcttt agatgtgtct gcttcagtag tggctgggtg	300
tattattgcc atattggtga cttgtgctgc ctactacttt atgaaattca gacgtgtttt	360
tggtagtagc aacctggtt ttgctgctaa tgcacttttg tttttgatgt ctttactat	420
actctgtctg gtaccagctt acagctttct gccgggagtc tactcagtct tttacttgta	480
cttgacattc tatttcacca atgatgttct attcttggt caccttcaat ggtttgccat	540
gttttctcct attgtgcctt ttgggataac agcaatctat gtattctgta tttctctgaa	600
gcaactgccat tgggttctta acaactatct taggaaaaga gtcattgtta atggagttac	660
atttagtacc ttcgaggagg ctgctttgtg tacctttttg ctcaacaagg aaatgtacct	720
aaaattgcgt agcgagacac tgttgccact tacacagtat aacaggtatc ttgctctata	780
taacaagtag aagtatttca gtggagcctt agatactacc agctatcgtg aagcagcttg	840
ctgccactta gcaaaggctc taaatgactt tagcaactca ggtgctgatg ttctctacca	900
accaccacag acatcaatca cttctgctgt tctgcagagt ggttttagga aaatggcatt	960
cccgtaggc aaagtgaag ggtgcatggt acaagtaacc tgtggaacta caactcttaa	1020
tggattgtgg ttggatgaca cagtatactg tocaagacat gtcatttgca cagcagaaga	1080

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catgcttaat cctaactatg aagatctgct cattcgcaaa tccaaccata gctttcttgt	1140
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gcttaaagtt gatacttcta accctaagac acccaagtat aaatttgtcc gtatccaacc	1260
tggtcaaaca ttttcagttc tagcatgcta caatggttca ccatctggtg tttatcagtg	1320
tgccatgaga cctaatacata ccattaaagg ttctttcctt aatggatcat gtggtagtgt	1380
tggttttaac attgattatg attgcgtgtc tttctgctat atgcatcata tggagcttcc	1440
aacaggagta cacgctggta ctgacttaga aggtaaattc tatggtccat ttgttgacag	1500
acaaactgca caggctgcag gtacagacac aaccataaca ttaaatgttt tggcatggct	1560
gtatgctgct gttatcaatg gtgatagggt gtttcttaat agattcacca ctactttgaa	1620
tgactttaac cttgtggcaa tgaagtacaa ctatgaacct ttgacacaag atcatgttga	1680
catattggga cctctttctg ctcaaacagg aattgccgtc ttagatatgt gtgctgcttt	1740
gaaagagctg ctgcagaatg gtatgaatgg tcgtactatc cttggtagca ctattttaga	1800
agatgagttt acaccatttg atgttgtagt acaatgctct ggtgttacct tccaaggtaa	1860
gttcaagaaa attgttaagg gcaactcatc ttggatgctt ttaactttct tgacatcact	1920
attgattcct gttcaaagta cacagtggtc actgttttct tttgtttacg agaatgcttt	1980
cttgccattt actcttggtg ttatggcaat tgctgcatgt	2020

<210> SEQ ID NO 49

<211> LENGTH: 2040

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 49

agcatttcca gcctgaagac gtactgtagc agctaaactg cccagcacca tacctctatt	60
taggttggtt aagcctttga tgaagtacaa gtatttcact ttaggccctt ttggtgtgtc	120
tgtaacaaac ctacaagggt gttccagttc tgtgtaaatt gtacctgtac catcactctt	180
agggatctta gccattttga gatcttgggt gtctgatatg aatgccagca caaacctacc	240
tcccttcgaa ttgttatagt aggcaagtgc attgtcatca gtacaagctg tttgtgtggt	300
accagccgca caggacatct gtcgtagtgc tactggactc agttcattat tctgtagtgt	360
aacagctgag ttggctctta gagctgtaac aataagaggc caagccaaat ttggtgaatt	420
gtccatgtta atttactaa gttgaacaat cttgctatcc gcatcaacaa cttgctggat	480
ttcccagagt gcagatgcat atgtaaagggt gttaccatca caagtgttct ttaggtacc	540
ataatcaggg acaacaacca tgagtttggc tgctgtatgc aatggtagta tgttgagtgg	600
aacacaacca tcacgcgcgt tgttgataat gttgttaagt gcatcattat caagcttcct	660
aagcatagtg aagagcattg tttgcatagc actagttact tttgccctct tgcctcaga	720
tcttgctgtt ttgtacattt gggatcatagc ctgatctgcc atcttttcca acttgctgtg	780
catggcagca tcacgggtcaa actcagattt agccacattc aaagatttct ttaacttttt	840
gagaacgact tcagaatcac cattagctac agcctgctca taggcctcct gggcagtggc	900
ataagcggca tatgatggta aagaactaaa ttctgaagca atagcctgaa gagtagcacg	960
gttatcgagc atttcctcgc acaacctatt aatgtctaca gcaccctgca tggatagcaa	1020
aacagacaaa agagaaacca tcttctcgaa agcttcagtt gtgtcttttg caagaagaat	1080

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atcattgtgg agttgtacac attgtgcccc caatttagaa gatgactcta ctctaagttg	1140
ttgaagaacc gagagcagta ccacagatgt gcactttacg tcagacattt tagactgtac	1200
agtagcaacc ttgatacatg gtttacctcc aatacccaac aacttaatgt taagcttgaa	1260
agcatcaata ctactcttag gagggcaaaag cccctgggag ttcataatacc taaattcttg	1320
tgtagagacc aagtagtcat aaacaccaag agtaagcctg aagtaacggg tgagtaaaca	1380
gaaaaggcca aagtagcagc agcaacaata gcctaagaaa caataaaca gcatgataca	1440
ctgtaagggtg ttgccagtaa taaataaaca tgggtaatac tcaacacaca caaacactat	1500
agctctagct aaaaacatga tagtcgtaac gacaccagaa tagttagagg ttacagaaat	1560
aactaaggcc cacatggaaa tagcttgatc taaagcatta ccatagtaga ctttgtaaac	1620
aagtgtaatg acattcatca gtgtccaaac acgtctagca gcatcatcat aaacagtgcg	1680
agctgtcatg agaataagca aaactaaagc tgaagcatat ataacacaat ccttaagcct	1740
ataaccagac aagctagtgt cagccaattc aagccatgtc atgatacgca tccccagct	1800
agcaggcatg tagaccatat taaagtaagc aactgttgca agagaaggta acagaaacaa	1860
gcacaagaat gcgtgcttat gcttaacaag cagcatagca catgcagcaa ttgccataat	1920
accaagagta aatggcaaga aagcattctc gtaaacaaag aaaacagtg accactgtgt	1980
actttgaaca agaatcaata gtgatgtcaa gaaagttaaa agcatccaat gatgagtgc	2040

<210> SEQ ID NO 50

<211> LENGTH: 2012

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 50

cttgtaggtt tgttacagac acacccaaaag ggcctaaagt gaaatacttg tacttcatca	60
aagggttaaa caacctaat agaggtatgg tgctgggcag tttagctgct acagtacgtc	120
ttcaggctgg aaatgtaca gaagtacctg ccaattcaac tgtgctttcc ttctgtgctt	180
ttgcagtaga ccctgctaaa gcatataagg attacctagc aagtggagga caaccaatca	240
ccaactgtgt gaagatgttg tgtacacaca ctggtacagg acaggcaatt actgtaacac	300
cagaagctaa catggacca gagtcctttg gtggtgcttc atgttgtctg tattgtagat	360
gccacattga ccatccaaat cctaaaggat tctgtgactt gaaaggtaag tacgtccaaa	420
tacctaccac ttgtgctaata gaccagtggt gttttacact tagaaacaca gtctgtaccg	480
tctgcggaat gtggaaagggt tatggctgta gttgtgacca actccgcgaa cccttgatgc	540
agtctgcgga tgcatacaacg tttttaaacg ggtttgcggt gtaagtgcag cccgtcttac	600
accgtgcggc acaggcacta gtactgatgt cgtctacagg gcttttgata ttacaacga	660
aaaagttgct ggttttgcaa agttcctaaa aactaattgc tgcgcttcc aggagaagga	720
tgaggaaggc aattttattag actcttactt tgtagttaag aggcatacta tgtctaacta	780
ccaacatgaa gagactattt ataacttggt taaagattgt ccagcgggtg ctgtccatga	840
ctttttcaag tttagagtag atggtgacat ggtaccacat atatcacgtc agcgtctaac	900
taaatacaca atggctgatt tagtctatgc tctacgtcat tttgatgagg gtaattgtga	960
tacattaaaa gaaatactcg tcacatacaa ttgctgtgat gatgattatt tcaataagaa	1020
ggattgggat gacttcgtag agaatcctga catcttacgc gtatatgcta acttaggtga	1080

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gcgtgtacgc caatcattat taaagactgt acaattctgc gatgctatgc gtgatgcagg	1140
cattgttaggc gtactgacat tagataatca ggatcttaat gggaactggg acgatttcgg	1200
tgatttcgta caagtagcac caggctgcgg agttcctatt gtggattcat attactcatt	1260
gctgatgccc atcctcactt tgactagggc attggctgct gagtcccata tggatgctga	1320
tctcgcaaaa ccacttatta agtgggattt gctgaaatat gattttacgg aagagagact	1380
ttgtctcttc gaccgttatt ttaaattatt ggaccagaca taccatccca attgtattaa	1440
ctgtttggat gatagggtga tccttcattg tgcaaaacttt aatgtgttat tttctactgt	1500
gtttccacct acaagttttg gaccactagt aagaaaaata tttgtagatg gtgttccttt	1560
tgttgtttca actggatacc attttcgtga gttaggagtc gtacataatc aggatgtaaa	1620
cttcatagc tcgctctca gtttcaagga acttttagtg tatgctgctg atccagctat	1680
gcatgcagct tctggcaatt tattgctaga taaacgcact acatgctttt cagtagctgc	1740
actaacaac aatgttgctt ttcaaactgt caaacccggg aattttaata aagactttta	1800
tgactttgct gtgtctaaag gtttctttaa ggaaggaagt tctgttgaac taaaacactt	1860
cttctttgct caggatggca acgctgctat cagtgattat gactattatc gttataatct	1920
gccaacaatg tgtgatatca gacaactcct attcgtagtt gaagtgttg ataaatactt	1980
tgattgttac gatggtggct gtattaatgc ca	2012

<210> SEQ ID NO 51

<211> LENGTH: 1877

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 51

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gcgtttctgc tgcgaaaagc ttgagtctct cagtacaagt gttggcaagt atgtaatcgc	120
cagcattagt ccaatcacat gttgctatcg cattgaagtc agtgacattg tcaactgccta	180
cacatgtggt tttgtataaa ccaaaaacct gaccattagc acataatgga aaactaatgg	240
gaggcttatg tgacttgcaa taatagctca tacctcctag atacagtgtg gtcacatcag	300
tgacatcaca acctggggca ttgcaaacat agggattaac agacaacact aatttgtgtg	360
atgttgaaat gacatgggtca tagcagcact tgcaacatag gaatgggtct ctaatacagg	420
caccgcaacg aagtgaagtc tgtgaattgc acaatacaca agcacctaca gcctgcaaga	480
ctgtatgtgg tgtgtacata gcctcataaa actcaggttc ccagtaccgt gaggtgttat	540
cattagttag cattacggaa tacatgtcca acatgtggcc agtaagctca tcatgtaact	600
ttctaataga ttgtaataac aagtgaagca catcagcata ctcttgatta ggatgttttg	660
taagtgggta agcatcaata gccagtgaac cgaacctttc aatcataagt gtacatctg	720
ttttgacaat atcatcgaca aaacagcctg cgcctaatat tcttgatgga tctgggtaag	780
gcaggtaac gtaatcatct ccttgtttta ctagcattgt atgctgtgag caaaattcgt	840
gaggtccttt agtaagggtc gtctcagtc aacattttgc ctacagacatg aacacattat	900
tttgataata aagaactgcc ttaaagtctt taatgctagc tactaaacct tgagccgcat	960
agttactggt atagcacaca acggcatcat cagaaagaat catcatggag aaatgtttac	1020
gcaggtaagc gtaaaactca tccacgaatt catgatcaac atccctatct ctatagagac	1080

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actcatagag cctgtgttgt agattgcgga catacttgtc agctatctta ttaccatcag	1140
ttgaaagaag tgcatttaca ttggctgtaa cagcttgaca aatgttaaag aactatttag	1200
cataagcagt ttagcatca cggatgatg ttccacctgg tttacatat agtgagccgc	1260
cacacatgac catctcactt aatacttgcg cacactcgtt agctaacctg tagaaacggt	1320
gtgataagtt acagcaagtg ttatgtttgc gagcaagaac aagagaggcc attatcctaa	1380
gcatgttagg catggctctg tcacattttg gataatccca acccataagg tgtggagttt	1440
ctacatcact gtaaacagtt tttacatat tatgccagcc accgtaaaac ttgcttggtc	1500
caattaccac agtagctcct ctagtggcgg ctattgactt caataatttc tgatgaaact	1560
gtctatttgt catagtacta cagatagaga caccagctac ggtgcgagct ctattctttg	1620
cactaatggc atacttaaga ttcatttgag ttatagtagg gatgacatta cgcttagtat	1680
acgcgaaaag tgcactctga tcctcataac tcattgagtc ataataaagt ctagccttac	1740
cccatttatt aaatgggaaa ccagctgatt tatccagatt gttaacgatt acttggttg	1800
cattaatata gccaccatcg taacaatcaa agtatattatc aacaacttca actacgaata	1860
ggagttgtct gatatca	1877

<210> SEQ ID NO 52

<211> LENGTH: 2051

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 52

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acgaacacga ctctgtctga caatccttcc agtgtatcac tgagcatttg tactatctta	120
atacgcacta cattccaggg caagccttta tacatgagtg gtataagatg tttaaactgg	180
tcacctgggt gaggttttgc attaactctg gtgaattctg tgttattttc agtgtcaaca	240
taaccagtcg gtacagctac taagttaaca cctgtagaaa atcctagctg gagaggtagg	300
ttagtaccca cagcatctct agttgcatga cagccctcta catcaaagcc aatccacgca	360
cgaacgtgac gaatagcttc ttgcgggtg ataaacatat tagggtaacc attgacttg	420
taattcattt tgaaccocat catagagatg agtctacggt aggtcatgtc ctttggtatg	480
cctggtatgt caacacataa tccttcagtc ttgaacttta tatcaacgct gaggtgtgta	540
ggtgcctgtg taggatgaag accagtaatg atcttactac agtccttaaa aagtccagtt	600
acattttctg cttgtaattg agccacattg cgacgtggta tttctagact tgtaaattgc	660
agtttgtcat aaagatctct atcagacatt atgcacaaaa tgccaatttt tgcccttggt	720
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atgacatagt catattcaga accctgtgat gaatcaacag tctgcgtagg caatcctaag	840
atttttgaag ctacagcgtt ctgtgaatta taaggtaga taaaaacagc ttttctccaa	900
gcaggattgc gtgtaagaaa ttctcttaca acgcctatct gaggtctgtt gattgcagat	960
gaaacatcat gtgtaataac accctttagt aacattttga agcattgagc tgacttatcc	1020
ttgtgtgctt ttagcttatt gtcataaaact aaagcactca cagtgtcaac aatttcagca	1080
ggacaacggc gacaagtacc aaggaacatg tctggaccta ttgttttcat aagtctgcac	1140
actgaattaa aatattcttg ttctagtgtg ccttagtca gcaatgtgcg gggggctggt	1200

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aattgagcag gatcgccaat atagacgtag tgttttgac gaagtctagc attgacaaca	1260
ctcaagtcac aattagtagc catagagatt tcatcaaaga ctacaatgac agcagttggt	1320
tctggcaatg catttacagt gcagaaaaca tactgttcta gtgttggaatt cactttgaat	1380
ttatcaaaac actctacgag cgcacgcgca ggtatgattc tactacattt atctatgggc	1440
aaatatttta atgccttttc acatagggca tcaacagctg catgagagca tgcggtatac	1500
actatgcgag cagatgggta atagagagca agtccgatgg caaaatgact cttaccagta	1560
ccagggtggc cttggagtgt agagtacttt tgcattgccg ccttttgata atttgcaaca	1620
ttgctagaaa actcatctga gatgttgagt gttgggtaca agccagtaat tctcacatag	1680
tgctcttggt gcactagagt aggtgcacta agtggcatta cagtgtgaga tgtcaacaca	1740
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taaccagtaa agacatagtt tctgttcaat ggtggtctag gttttccaac ctcccatgaa	1920
agatgcaatt ctctgtcaga gagtacttcg cgtacagtgg caataccata tgacagctta	1980
aatgtttcct cagtggcttt gagcgtttct gctgcgaaaa gcttgagtct ctcagtacaa	2040
gtgttggtgca g	2051

<210> SEQ ID NO 53

<211> LENGTH: 2075

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 53

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aatttcagca tagtcaattg taaccttgac cacttttgaa atcactgaca aatcttgtga	120
ctttattatc tcgacaaagt catcaagtaa aagatcaatc acagaacaca cacattttga	180
tgaacctggt tgcgcactctg ttatgaagta atttttcact gtgctgtcca tagggataaa	240
atcctcta at taaagtgggt aatcttgtga gcgcttggt aagcctatca ttaaatgaag	300
accgccaagt tgtccatgac tgaatctcc ataaacgatg tgttcgaagg catagccctc	360
gagcttatat cgctgtatga attcatccat agcgagctcg agaaagtcag tttccatttg	420
tgatctgggc ttaaaatcct ctaagtctct gctctgagta aagtaggttt caggcaactg	480
ttgaataatg ccgtctactt tcttaaagta gttaaactgt gtttttactg attctccaat	540
taatgtgact ccattgacgc tagcttgtgc tgggtccctt gaagggtgta gacctttgac	600
tgaaccttct gttattaaaa caccattacg ggcgtttcta aaaaggctta cctgtccttc	660
cactctacca tcaaacaaga cagtaagtga agaacaagca ctctcagtag gtttcttggc	720
aatgtcagtc attgtgcaga cacctattgt agatacatgt gctggggctt ctcttttgta	780
gtcccagatt acagtattag cagcgatata aacacccaaa ttattgagta tcttaattct	840
tggcactggt ttaattgtac gcttagccca aagctcaaat gcaacattaa cagggaagtgt	900
tgtcttattt tcaaagatct ccacatcaat accatctacc tttgtgtaaa cagcattatt	960
aatgatggaa acagggtgct cgccggcggtg tccatcaaag tgccttttat taacaacatt	1020
ataagccaca ttttctaaac tctgtaacct ggtaaatgta ttccacaggt tataagtatc	1080
aaattgtttg taaatccata ggctaaatcc agcagaaatc atcatattat atgcatccaa	1140

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gtactgtcgg tactcatttg catggtgtct gcaaacagca ccacctaaat tgcacgtgt	1200
aatacacgta gcagatttga gtggaacata atcaatatcc gacactactt gtttgccatg	1260
agactcacia ggactatcag aatagtaaaa gaaaggcaat tgctttaaat tagtaaatgc	1320
acttttatcg aaagctggag tgtggaatgc atgcttattc acatacaaac taccaccatc	1380
acagcctggt aagtccaagt ttgacaagac tcttgtgtca aacctacaca caattgcatt	1440
ggctgggtaa cgatcaacgt tacaattcca aaacaaacaa acaccatcag tgaatttatc	1500
gtgatgtgta gcataagaat agaagagtcc ctctattttg taagctttgt cactacatgg	1560
ctgagcatcg tagaacttcc attctacttc agcctgaggg acacacttga tagcctttgg	1620
atttccaatg tcatgaagaa ctggaaactt atcagcaagc aatgcagact tcacaaccat	1680
gtgtgtgact tttctgcaag cagaattaac cctcagttca tctcctataa taggggtattc	1740
aacagaccaa tcaacgcgct taacaaagca ctcatggact gctaaacatc tagtcatgat	1800
agcatcacia ctagccacat gtgcatttcc atgtacctgg caatgttggg catggttact	1860
ctgaaggtta cccgtaaagc cccactgctg aacatcaatc ataatgggt tatagacata	1920
gtcaaaacc acagaatgat tccagcaggc ataagtatct gatgaagtag aaaagcaagt	1980
tgacgctttg tcacacagac aacacgttct ttcaggtcca atcttgacaa agtacttcat	2040
tgatgtaagc tcaaagccat gcgcccacaa gacga	2075

<210> SEQ ID NO 54

<211> LENGTH: 1891

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 54

aagattcacc acttaaatga gaggatttta tccctatgga cagcacagtg aaaaattact	60
tcataacaga tgcgcaaaca ggttcatcaa aatgtgtgtg ttctgtgatt gatcttttac	120
ttgatgactt tgtcgagata ataaagtcac aagatttgct agtgatttca aaagtgggtca	180
aggttacaat tgactatgct gaaatttcat tcatgctttg gtgtaaggat ggacatgttg	240
aaaccttcta cccaaaacta caagcaagtc aagcgtggca accaggtgtt gcgatgccta	300
acttgtaaca gatgcaaaga atgcttcttg aaaagtgtga ccttcagaat tatggtgaaa	360
atgctgttat accaaaagga ataatgatga atgtcgcaa gtatactcaa ctgtgtcaat	420
acttaaatat acttacttta gctgtaccct acaacatgag agttattcac tttggtgctg	480
gctctgataa aggagttgca ccaggtacag ctgtgctcag acaatgggtg ccaactggca	540
cactacttgt cgattcagat cttaatgact tcgtctccga cgcagattct actttaattg	600
gagactgtgc aacagtacat acggctaata aatgggacct tattattagc gatatgtatg	660
accctaggac caaacatgtg acaaaagaga atgactctaa agaagggttt ttcacttatc	720
tgtgtggatt tataaagcaa aaactagccc tgggtgggtc tatagctgta aagataacag	780
agcattcttg gaatgctgac ctttacaagc ttatggggcca tttctcatgg tggacagctt	840
ttgttacaaa tgtaaatgca tcatcatcgg aagcattttt aattggggct aactatcttg	900
gcaagccgaa ggaacaaatt gatggctata ccatgcatgc taactacatt ttctggagga	960
acacaaatcc tatccagttg tcttcctatt cactctttga catgagcaaa tttcctctta	1020
aattaagagg aactgctgta atgtctctta aggagaatca aatcaatgat atgatttatt	1080

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ctcttctgga aaaaggtagg cttatcatta gagaaaacaa cagagttgtg gtttcaagtg 1140
atattcttgt taacaactaa acgaacatgt ttattttctt attatttctt actctcacta 1200
gtggtagtga ccttgaccgg tgcaccactt ttgatgatgt tcaagctcct aattacactc 1260
aacatacttc atctatgagg ggggtttact atcctgatga aatttttaga tcagacactc 1320
tttatttaac tcaggattta tttcttccat tttattctaa tgttacaggg tttcatacta 1380
ttaatcatac gtttggaac cctgtcatac cttttaagga tggatttat tttgctgcca 1440
cagagaaatc aaatgtgtgc cgtggttggg tttttggttc taccatgaac aacaagtcac 1500
agtcggtgat tattattaac aattctacta atgttggtat acgagcatgt aactttgaat 1560
tgtgtgacaa cctttctttt gctgtttcta aacccatggg tacacagaca catactatga 1620
tattcgataa tgcatttaat tgcactttcg agtacatata tgatgccttt tcgcttgatg 1680
tttcagaaaa gtcaggtaat tttaaacact tacgagagtt tgtgtttaaa aataaagatg 1740
ggtttctcta tgtttataag ggctatcaac ctatagatgt agttcgtgat ctaccttctg 1800
gttttaacac ttgaaacct atttttaagt tgcctcttgg tattaacatt acaaatttta 1860
gagccattct tacagccttt tcacctgctc a 1891

```

```

<210> SEQ ID NO 55
<211> LENGTH: 32
<212> TYPE: DNA
<213> ORGANISM: artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: N sens primer

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<400> SEQUENCE: 55

```

```

cccatatgtc tgataatgga cccaatcaa ac 32

```

```

<210> SEQ ID NO 56
<211> LENGTH: 32
<212> TYPE: DNA
<213> ORGANISM: artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: N antisens primer

```

```

<400> SEQUENCE: 56

```

```

cccccggtg cctgagttga atcagcagaa gc 32

```

```

<210> SEQ ID NO 57
<211> LENGTH: 31
<212> TYPE: DNA
<213> ORGANISM: artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: Sc sens primer

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```

<400> SEQUENCE: 57

```

```

cccatatgag tgaccttgac cggcgacca c 31

```

```

<210> SEQ ID NO 58
<211> LENGTH: 30
<212> TYPE: DNA
<213> ORGANISM: artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: SL sens primer

```

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<400> SEQUENCE: 58

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```

cccatatgaa accttgaccc ccacctgctc 30

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<210> SEQ ID NO 59
<211> LENGTH: 33
<212> TYPE: DNA
<213> ORGANISM: Sc and SL antisens primer

<400> SEQUENCE: 59

cccccggtt taatatattg ctcattttt ccc 33

<210> SEQ ID NO 60
<211> LENGTH: 16
<212> TYPE: DNA
<213> ORGANISM: Sens set 1 primer

<400> SEQUENCE: 60

ggcatcgat ggggtg 16

<210> SEQ ID NO 61
<211> LENGTH: 16
<212> TYPE: DNA
<213> ORGANISM: Antisens set 2 (28774-28759) primer

<400> SEQUENCE: 61

cagtttcacc acctcc 16

<210> SEQ ID NO 62
<211> LENGTH: 16
<212> TYPE: DNA
<213> ORGANISM: Sens set 2 (28375-28390) primer

<400> SEQUENCE: 62

ggctactacc gaagag 16

<210> SEQ ID NO 63
<211> LENGTH: 16
<212> TYPE: DNA
<213> ORGANISM: Antisens set 2 (28702-28687)primer

<400> SEQUENCE: 63

aattaccgag actacg 16

<210> SEQ ID NO 64
<211> LENGTH: 26
<212> TYPE: DNA
<213> ORGANISM: Probe 1/set 1 (28561-28586)

<400> SEQUENCE: 64

ggcaccgca atcctaataa caatgc 26

<210> SEQ ID NO 65
<211> LENGTH: 21
<212> TYPE: DNA
<213> ORGANISM: Probe 2/set 1 (28588-28608)

<400> SEQUENCE: 65

gccaccgtgc tacaacttcc t 21

<210> SEQ ID NO 66
<211> LENGTH: 23
<212> TYPE: DNA
<213> ORGANISM: Probe 1/set 2 /probe N/FL (28541-28563)

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<400> SEQUENCE: 66

atacacccaa agaccacatt ggc 23

<210> SEQ ID NO 67

<211> LENGTH: 25

<212> TYPE: DNA

<213> ORGANISM: Probe 2/set 2/probe SARS/N/LC705 (28565-28589)

<400> SEQUENCE: 67

cccgcaatcc taataacaat gctgc 25

<210> SEQ ID NO 68

<211> LENGTH: 30

<212> TYPE: DNA

<213> ORGANISM: artificial sequence

<220> FEATURE:

<223> OTHER INFORMATION: Anchor primer 14T

<400> SEQUENCE: 68

agatgaattc ggtacctttt tttttttttt 30

<210> SEQ ID NO 69

<211> LENGTH: 13

<212> TYPE: PRT

<213> ORGANISM: artificial sequence

<220> FEATURE:

<223> OTHER INFORMATION: M2-14 peptide

<400> SEQUENCE: 69

Ala Asp Asn Gly Thr Ile Thr Val Glu Glu Leu Lys Gln
1 5 10

<210> SEQ ID NO 70

<211> LENGTH: 12

<212> TYPE: PRT

<213> ORGANISM: artificial sequence

<220> FEATURE:

<223> OTHER INFORMATION: E1-12 peptide

<400> SEQUENCE: 70

Met Tyr Ser Phe Val Ser Glu Glu Thr Gly Thr Leu
1 5 10

<210> SEQ ID NO 71

<211> LENGTH: 24

<212> TYPE: PRT

<213> ORGANISM: artificial sequence

<220> FEATURE:

<223> OTHER INFORMATION: E53-72 peptide

<400> SEQUENCE: 71

Lys Pro Thr Val Tyr Val Tyr Ser Arg Val Lys Asn Leu Asn Ser Ser
1 5 10 15Glu Gly Val Pro Asp Leu Leu Val
20

<210> SEQ ID NO 72

<211> LENGTH: 153

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 72

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gatattaggt ttttacctac ccaggaaaag ccaaccaacc tcgatctctt gtagatctgt      60
tctctaaacg aactttaaaa tctgtgtagc tgtcgctcgg ctgcatgcct agtgcaccta    120
cgcagtataa acaataataa attttactgt cgt                                  153

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<210> SEQ ID NO 73
<211> LENGTH: 410
<212> TYPE: DNA
<213> ORGANISM: CORONAVIRUS

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<400> SEQUENCE: 73

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ttctccagac aacttcaaaa ttccatgagt ggagcttctg ctgattcaac tcaggcataa      60
acactcatga tgaccacaca aggcagatgg gctatgtaaa cgttttcgca attccgttta    120
cgatacatag tctactcttg tgcagaatga attctcgtaa ctaaacagca caagtaggtt    180
tagttaactt taatctcaca tagcaatctt taatcaatgt gtaacattag ggaggacttg    240
aaagagccac cacattttca tcgaggccac gcggagtacg atcgagggta cagtgaataa    300
tgctaggggag agctgcctat atggaagagc cctaattgtgt aaaattaatt ttagtagtgc    360
tatcccatg  tgattttaat agcttcttag gagaatgaca aaaaaaaaaa                410

```

```

<210> SEQ ID NO 74
<211> LENGTH: 4382
<212> TYPE: PRT
<213> ORGANISM: CORONAVIRUS

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```

<400> SEQUENCE: 74

```

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Met Glu Ser Leu Val Leu Gly Val Asn Glu Lys Thr His Val Gln Leu
 1             5             10            15

Ser Leu Pro Val Leu Gln Val Arg Asp Val Leu Val Arg Gly Phe Gly
      20            25            30

Asp Ser Val Glu Glu Ala Leu Ser Glu Ala Arg Glu His Leu Lys Asn
      35            40            45

Gly Thr Cys Gly Leu Val Glu Leu Glu Lys Gly Val Leu Pro Gln Leu
      50            55            60

Glu Gln Pro Tyr Val Phe Ile Lys Arg Ser Asp Ala Leu Ser Thr Asn
      65            70            75            80

His Gly His Lys Val Val Glu Leu Val Ala Glu Met Asp Gly Ile Gln
      85            90            95

Tyr Gly Arg Ser Gly Ile Thr Leu Gly Val Leu Val Pro His Val Gly
      100           105           110

Glu Thr Pro Ile Ala Tyr Arg Asn Val Leu Leu Arg Lys Asn Gly Asn
      115           120           125

Lys Gly Ala Gly Gly His Ser Tyr Gly Ile Asp Leu Lys Ser Tyr Asp
      130           135           140

Leu Gly Asp Glu Leu Gly Thr Asp Pro Ile Glu Asp Tyr Glu Gln Asn
      145           150           155           160

Trp Asn Thr Lys His Gly Ser Gly Ala Leu Arg Glu Leu Thr Arg Glu
      165           170           175

Leu Asn Gly Gly Ala Val Thr Arg Tyr Val Asp Asn Asn Phe Cys Gly
      180           185           190

Pro Asp Gly Tyr Pro Leu Asp Cys Ile Lys Asp Phe Leu Ala Arg Ala
      195           200           205

Gly Lys Ser Met Cys Thr Leu Ser Glu Gln Leu Asp Tyr Ile Glu Ser

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210					215					220					
Lys 225	Arg	Gly	Val	Tyr	Cys 230	Cys	Arg	Asp	His	Glu 235	His	Glu	Ile	Ala	Trp 240
Phe	Thr	Glu	Arg	Ser 245	Asp	Lys	Ser	Tyr	Glu 250	His	Gln	Thr	Pro	Phe 255	Glu
Ile	Lys	Ser	Ala	Lys 260	Lys	Phe	Asp	Thr	Phe 265	Lys	Gly	Glu	Cys 270	Pro	Lys
Phe	Val	Phe	Pro	Leu 275	Asn	Ser	Lys 280	Val	Lys	Val	Ile 285	Gln	Pro	Arg	Val
Glu	Lys 290	Lys	Lys	Thr	Glu 295	Gly	Phe	Met	Gly	Arg	Ile 300	Arg	Ser	Val	Tyr
Pro 305	Val	Ala	Ser	Pro	Gln 310	Glu	Cys	Asn	Asn	Met 315	His	Leu	Ser	Thr	Leu 320
Met	Lys	Cys	Asn	His 325	Cys	Asp	Glu	Val	Ser 330	Trp	Gln	Thr	Cys	Asp 335	Phe
Leu	Lys	Ala	Thr	Cys 340	Glu	His	Cys	Gly 345	Thr	Glu	Asn	Leu	Val 350	Ile	Glu
Gly	Pro	Thr	Thr	Cys 355	Gly	Tyr	Leu 360	Pro	Thr	Asn	Ala 365	Val	Val	Lys	Met
Pro 370	Cys	Pro	Ala	Cys	Gln 375	Asp	Pro	Glu	Ile	Gly 380	Pro	Glu	His	Ser	Val
Ala 385	Asp	Tyr	His	Asn 390	His	Ser	Asn	Ile	Glu	Thr 395	Arg	Leu	Arg	Lys	Gly 400
Gly	Arg	Thr	Arg	Cys 405	Phe	Gly	Gly	Cys	Val 410	Phe	Ala	Tyr	Val	Gly 415	Cys
Tyr	Asn	Lys	Arg	Ala 420	Tyr	Trp	Val	Pro 425	Arg	Ala	Ser	Ala	Asp 430	Ile	Gly
Ser	Gly	His	Thr	Gly 435	Ile	Thr	Gly 440	Asp	Asn	Val	Glu 445	Thr	Leu	Asn	Glu
Asp 450	Leu	Leu	Glu	Ile 455	Leu	Ser	Arg	Glu	Arg	Val 460	Asn	Ile	Asn	Ile	Val
Gly 465	Asp	Phe	His	Leu 470	Asn	Glu	Glu	Val	Ala 475	Ile	Ile	Leu	Ala	Ser	Phe 480
Ser	Ala	Ser	Thr	Ser 485	Ala	Phe	Ile	Asp 490	Thr	Ile	Lys	Ser	Leu	Asp 495	Tyr
Lys	Ser	Phe	Lys	Thr 500	Ile	Val	Glu	Ser 505	Cys	Gly	Asn	Tyr	Lys 510	Val	Thr
Lys	Gly	Lys	Pro	Val 515	Lys	Gly	Ala 520	Trp	Asn	Ile	Gly	Gln 525	Gln	Arg	Ser
Val 530	Leu	Thr	Pro	Leu 535	Cys	Gly	Phe	Pro	Ser	Gln 540	Ala	Ala	Gly	Val	Ile
Arg 545	Ser	Ile	Phe	Ala 550	Arg	Thr	Leu	Asp	Ala 555	Ala	Asn	His	Ser	Ile	Pro 560
Asp	Leu	Gln	Arg	Ala 565	Ala	Val	Thr	Ile 570	Leu	Asp	Gly	Ile	Ser	Glu	Gln
Ser	Leu	Arg	Leu	Val 580	Asp	Ala	Met	Val 585	Tyr	Thr	Ser	Asp	Leu 590	Leu	Thr
Asn	Ser	Val	Ile	Ile 595	Met	Ala	Tyr 600	Val	Thr	Gly	Gly 605	Leu	Val	Gln	Gln
Thr 610	Ser	Gln	Trp	Leu 615	Ser	Asn	Leu 620	Leu	Gly	Thr	Thr 625	Val	Glu	Lys	Leu

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Arg	Pro	Ile	Phe	Glu	Trp	Ile	Glu	Ala	Lys	Leu	Ser	Ala	Gly	Val	Glu	625	630	635	640
Phe	Leu	Lys	Asp	Ala	Trp	Glu	Ile	Leu	Lys	Phe	Leu	Ile	Thr	Gly	Val	645	650	655	
Phe	Asp	Ile	Val	Lys	Gly	Gln	Ile	Gln	Val	Ala	Ser	Asp	Asn	Ile	Lys	660	665	670	
Asp	Cys	Val	Lys	Cys	Phe	Ile	Asp	Val	Val	Asn	Lys	Ala	Leu	Glu	Met	675	680	685	
Cys	Ile	Asp	Gln	Val	Thr	Ile	Ala	Gly	Ala	Lys	Leu	Arg	Ser	Leu	Asn	690	695	700	
Leu	Gly	Glu	Val	Phe	Ile	Ala	Gln	Ser	Lys	Gly	Leu	Tyr	Arg	Gln	Cys	705	710	715	720
Ile	Arg	Gly	Lys	Glu	Gln	Leu	Gln	Leu	Leu	Met	Pro	Leu	Lys	Ala	Pro	725	730	735	
Lys	Glu	Val	Thr	Phe	Leu	Glu	Gly	Asp	Ser	His	Asp	Thr	Val	Leu	Thr	740	745	750	
Ser	Glu	Glu	Val	Val	Leu	Lys	Asn	Gly	Glu	Leu	Glu	Ala	Leu	Glu	Thr	755	760	765	
Pro	Val	Asp	Ser	Phe	Thr	Asn	Gly	Ala	Ile	Val	Gly	Thr	Pro	Val	Cys	770	775	780	
Val	Asn	Gly	Leu	Met	Leu	Leu	Glu	Ile	Lys	Asp	Lys	Glu	Gln	Tyr	Cys	785	790	795	800
Ala	Leu	Ser	Pro	Gly	Leu	Leu	Ala	Thr	Asn	Asn	Val	Phe	Arg	Leu	Lys	805	810	815	
Gly	Gly	Ala	Pro	Ile	Lys	Gly	Val	Thr	Phe	Gly	Glu	Asp	Thr	Val	Trp	820	825	830	
Glu	Val	Gln	Gly	Tyr	Lys	Asn	Val	Arg	Ile	Thr	Phe	Glu	Leu	Asp	Glu	835	840	845	
Arg	Val	Asp	Lys	Val	Leu	Asn	Glu	Lys	Cys	Ser	Val	Tyr	Thr	Val	Glu	850	855	860	
Ser	Gly	Thr	Glu	Val	Thr	Glu	Phe	Ala	Cys	Val	Val	Ala	Glu	Ala	Val	865	870	875	880
Val	Lys	Thr	Leu	Gln	Pro	Val	Ser	Asp	Leu	Leu	Thr	Asn	Met	Gly	Ile	885	890	895	
Asp	Leu	Asp	Glu	Trp	Ser	Val	Ala	Thr	Phe	Tyr	Leu	Phe	Asp	Asp	Ala	900	905	910	
Gly	Glu	Glu	Asn	Phe	Ser	Ser	Arg	Met	Tyr	Cys	Ser	Phe	Tyr	Pro	Pro	915	920	925	
Asp	Glu	Glu	Glu	Glu	Asp	Asp	Ala	Glu	Cys	Glu	Glu	Glu	Glu	Ile	Asp	930	935	940	
Glu	Thr	Cys	Glu	His	Glu	Tyr	Gly	Thr	Glu	Asp	Asp	Tyr	Gln	Gly	Leu	945	950	955	960
Pro	Leu	Glu	Phe	Gly	Ala	Ser	Ala	Glu	Thr	Val	Arg	Val	Glu	Glu	Glu	965	970	975	
Glu	Glu	Glu	Asp	Trp	Leu	Asp	Asp	Thr	Thr	Glu	Gln	Ser	Glu	Ile	Glu	980	985	990	
Pro	Glu	Pro	Glu	Pro	Thr	Pro	Glu	Glu	Pro	Val	Asn	Gln	Phe	Thr	Gly	995	1000	1005	
Tyr	Leu	Lys	Leu	Thr	Asp	Asn	Val	Ala	Ile	Lys	Cys	Val	Asp	Ile		1010	1015	1020	

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Val	Lys	Glu	Ala	Gln	Ser	Ala	Asn	Pro	Met	Val	Ile	Val	Asn	Ala
1025						1030					1035			
Ala	Asn	Ile	His	Leu	Lys	His	Gly	Gly	Gly	Val	Ala	Gly	Ala	Leu
1040						1045					1050			
Asn	Lys	Ala	Thr	Asn	Gly	Ala	Met	Gln	Lys	Glu	Ser	Asp	Asp	Tyr
1055						1060					1065			
Ile	Lys	Leu	Asn	Gly	Pro	Leu	Thr	Val	Gly	Gly	Ser	Cys	Leu	Leu
1070						1075					1080			
Ser	Gly	His	Asn	Leu	Ala	Lys	Lys	Cys	Leu	His	Val	Val	Gly	Pro
1085						1090					1095			
Asn	Leu	Asn	Ala	Gly	Glu	Asp	Ile	Gln	Leu	Leu	Lys	Ala	Ala	Tyr
1100						1105					1110			
Glu	Asn	Phe	Asn	Ser	Gln	Asp	Ile	Leu	Leu	Ala	Pro	Leu	Leu	Ser
1115						1120					1125			
Ala	Gly	Ile	Phe	Gly	Ala	Lys	Pro	Leu	Gln	Ser	Leu	Gln	Val	Cys
1130						1135					1140			
Val	Gln	Thr	Val	Arg	Thr	Gln	Val	Tyr	Ile	Ala	Val	Asn	Asp	Lys
1145						1150					1155			
Ala	Leu	Tyr	Glu	Gln	Val	Val	Met	Asp	Tyr	Leu	Asp	Asn	Leu	Lys
1160						1165					1170			
Pro	Arg	Val	Glu	Ala	Pro	Lys	Gln	Glu	Glu	Pro	Pro	Asn	Thr	Glu
1175						1180					1185			
Asp	Ser	Lys	Thr	Glu	Glu	Lys	Ser	Val	Val	Gln	Lys	Pro	Val	Asp
1190						1195					1200			
Val	Lys	Pro	Lys	Ile	Lys	Ala	Cys	Ile	Asp	Glu	Val	Thr	Thr	Thr
1205						1210					1215			
Leu	Glu	Glu	Thr	Lys	Phe	Leu	Thr	Asn	Lys	Leu	Leu	Leu	Phe	Ala
1220						1225					1230			
Asp	Ile	Asn	Gly	Lys	Leu	Tyr	His	Asp	Ser	Gln	Asn	Met	Leu	Arg
1235						1240					1245			
Gly	Glu	Asp	Met	Ser	Phe	Leu	Glu	Lys	Asp	Ala	Pro	Tyr	Met	Val
1250						1255					1260			
Gly	Asp	Val	Ile	Thr	Ser	Gly	Asp	Ile	Thr	Cys	Val	Val	Ile	Pro
1265						1270					1275			
Ser	Lys	Lys	Ala	Gly	Gly	Thr	Thr	Glu	Met	Leu	Ser	Arg	Ala	Leu
1280						1285					1290			
Lys	Lys	Val	Pro	Val	Asp	Glu	Tyr	Ile	Thr	Thr	Tyr	Pro	Gly	Gln
1295						1300					1305			
Gly	Cys	Ala	Gly	Tyr	Thr	Leu	Glu	Glu	Ala	Lys	Thr	Ala	Leu	Lys
1310						1315					1320			
Lys	Cys	Lys	Ser	Ala	Phe	Tyr	Val	Leu	Pro	Ser	Glu	Ala	Pro	Asn
1325						1330					1335			
Ala	Lys	Glu	Glu	Ile	Leu	Gly	Thr	Val	Ser	Trp	Asn	Leu	Arg	Glu
1340						1345					1350			
Met	Leu	Ala	His	Ala	Glu	Glu	Thr	Arg	Lys	Leu	Met	Pro	Ile	Cys
1355						1360					1365			
Met	Asp	Val	Arg	Ala	Ile	Met	Ala	Thr	Ile	Gln	Arg	Lys	Tyr	Lys
1370						1375					1380			
Gly	Ile	Lys	Ile	Gln	Glu	Gly	Ile	Val	Asp	Tyr	Gly	Val	Arg	Phe
1385						1390					1395			
Phe	Phe	Tyr	Thr	Ser	Lys	Glu	Pro	Val	Ala	Ser	Ile	Ile	Thr	Lys

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1400	1405	1410
Leu Asn Ser Leu Asn Glu Pro	Leu Val Thr Met Pro	Ile Gly Tyr
1415	1420	1425
Val Thr His Gly Phe Asn Leu	Glu Glu Ala Ala Arg	Cys Met Arg
1430	1435	1440
Ser Leu Lys Ala Pro Ala Val	Val Ser Val Ser Ser	Pro Asp Ala
1445	1450	1455
Val Thr Thr Tyr Asn Gly Tyr	Leu Thr Ser Ser Ser	Lys Thr Ser
1460	1465	1470
Glu Glu His Phe Val Glu Thr	Val Ser Leu Ala Gly	Ser Tyr Arg
1475	1480	1485
Asp Trp Ser Tyr Ser Gly Gln	Arg Thr Glu Leu Gly	Val Glu Phe
1490	1495	1500
Leu Lys Arg Gly Asp Lys Ile	Val Tyr His Thr Leu	Glu Ser Pro
1505	1510	1515
Val Glu Phe His Leu Asp Gly	Glu Val Leu Ser Leu	Asp Lys Leu
1520	1525	1530
Lys Ser Leu Leu Ser Leu Arg	Glu Val Lys Thr Ile	Lys Val Phe
1535	1540	1545
Thr Thr Val Asp Asn Thr Asn	Leu His Thr Gln Leu	Val Asp Met
1550	1555	1560
Ser Met Thr Tyr Gly Gln Gln	Phe Gly Pro Thr Tyr	Leu Asp Gly
1565	1570	1575
Ala Asp Val Thr Lys Ile Lys	Pro His Val Asn His	Glu Gly Lys
1580	1585	1590
Thr Phe Phe Val Leu Pro Ser	Asp Asp Thr Leu Arg	Ser Glu Ala
1595	1600	1605
Phe Glu Tyr Tyr His Thr Leu	Asp Glu Ser Phe Leu	Gly Arg Tyr
1610	1615	1620
Met Ser Ala Leu Asn His Thr	Lys Lys Trp Lys Phe	Pro Gln Val
1625	1630	1635
Gly Gly Leu Thr Ser Ile Lys	Trp Ala Asp Asn Asn	Cys Tyr Leu
1640	1645	1650
Ser Ser Val Leu Leu Ala Leu	Gln Gln Leu Glu Val	Lys Phe Asn
1655	1660	1665
Ala Pro Ala Leu Gln Glu Ala	Tyr Tyr Arg Ala Arg	Ala Gly Asp
1670	1675	1680
Ala Ala Asn Phe Cys Ala Leu	Ile Leu Ala Tyr Ser	Asn Lys Thr
1685	1690	1695
Val Gly Glu Leu Gly Asp Val	Arg Glu Thr Met Thr	His Leu Leu
1700	1705	1710
Gln His Ala Asn Leu Glu Ser	Ala Lys Arg Val Leu	Asn Val Val
1715	1720	1725
Cys Lys His Cys Gly Gln Lys	Thr Thr Thr Leu Thr	Gly Val Glu
1730	1735	1740
Ala Val Met Tyr Met Gly Thr	Leu Ser Tyr Asp Asn	Leu Lys Thr
1745	1750	1755
Gly Val Ser Ile Pro Cys Val	Cys Gly Arg Asp Ala	Thr Gln Tyr
1760	1765	1770
Leu Val Gln Gln Glu Ser Ser	Phe Val Met Met Ser	Ala Pro Pro
1775	1780	1785

Ala	Glu	Tyr	Lys	Leu	Gln	Gln	Gly	Thr	Phe	Leu	Cys	Ala	Asn	Glu
	1790					1795					1800			
Tyr	Thr	Gly	Asn	Tyr	Gln	Cys	Gly	His	Tyr	Thr	His	Ile	Thr	Ala
	1805					1810					1815			
Lys	Glu	Thr	Leu	Tyr	Arg	Ile	Asp	Gly	Ala	His	Leu	Thr	Lys	Met
	1820					1825					1830			
Ser	Glu	Tyr	Lys	Gly	Pro	Val	Thr	Asp	Val	Phe	Tyr	Lys	Glu	Thr
	1835					1840					1845			
Ser	Tyr	Thr	Thr	Thr	Ile	Lys	Pro	Val	Ser	Tyr	Lys	Leu	Asp	Gly
	1850					1855					1860			
Val	Thr	Tyr	Thr	Glu	Ile	Glu	Pro	Lys	Leu	Asp	Gly	Tyr	Tyr	Lys
	1865					1870					1875			
Lys	Asp	Asn	Ala	Tyr	Tyr	Thr	Glu	Gln	Pro	Ile	Asp	Leu	Val	Pro
	1880					1885					1890			
Thr	Gln	Pro	Leu	Pro	Asn	Ala	Ser	Phe	Asp	Asn	Phe	Lys	Leu	Thr
	1895					1900					1905			
Cys	Ser	Asn	Thr	Lys	Phe	Ala	Asp	Asp	Leu	Asn	Gln	Met	Thr	Gly
	1910					1915					1920			
Phe	Thr	Lys	Pro	Ala	Ser	Arg	Glu	Leu	Ser	Val	Thr	Phe	Phe	Pro
	1925					1930					1935			
Asp	Leu	Asn	Gly	Asp	Val	Val	Ala	Ile	Asp	Tyr	Arg	His	Tyr	Ser
	1940					1945					1950			
Ala	Ser	Phe	Lys	Lys	Gly	Ala	Lys	Leu	Leu	His	Lys	Pro	Ile	Val
	1955					1960					1965			
Trp	His	Ile	Asn	Gln	Ala	Thr	Thr	Lys	Thr	Thr	Phe	Lys	Pro	Asn
	1970					1975					1980			
Thr	Trp	Cys	Leu	Arg	Cys	Leu	Trp	Ser	Thr	Lys	Pro	Val	Asp	Thr
	1985					1990					1995			
Ser	Asn	Ser	Phe	Glu	Val	Leu	Ala	Val	Glu	Asp	Thr	Gln	Gly	Met
	2000					2005					2010			
Asp	Asn	Leu	Ala	Cys	Glu	Ser	Gln	Gln	Pro	Thr	Ser	Glu	Glu	Val
	2015					2020					2025			
Val	Glu	Asn	Pro	Thr	Ile	Gln	Lys	Glu	Val	Ile	Glu	Cys	Asp	Val
	2030					2035					2040			
Lys	Thr	Thr	Glu	Val	Val	Gly	Asn	Val	Ile	Leu	Lys	Pro	Ser	Asp
	2045					2050					2055			
Glu	Gly	Val	Lys	Val	Thr	Gln	Glu	Leu	Gly	His	Glu	Asp	Leu	Met
	2060					2065					2070			
Ala	Ala	Tyr	Val	Glu	Asn	Thr	Ser	Ile	Thr	Ile	Lys	Lys	Pro	Asn
	2075					2080					2085			
Glu	Leu	Ser	Leu	Ala	Leu	Gly	Leu	Lys	Thr	Ile	Ala	Thr	His	Gly
	2090					2095					2100			
Ile	Ala	Ala	Ile	Asn	Ser	Val	Pro	Trp	Ser	Lys	Ile	Leu	Ala	Tyr
	2105					2110					2115			
Val	Lys	Pro	Phe	Leu	Gly	Gln	Ala	Ala	Ile	Thr	Thr	Ser	Asn	Cys
	2120					2125					2130			
Ala	Lys	Arg	Leu											

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Asn Ser	Arg Ile Arg Ala	Ser	Leu Pro Thr Thr	Ile	Ala Lys Asn
2165		2170		2175	
Ser Val	Lys Ser Val Ala	Lys	Leu Cys Leu Asp	Ala	Gly Ile Asn
2180		2185		2190	
Tyr Val	Lys Ser Pro Lys	Phe	Ser Lys Leu Phe	Thr	Ile Ala Met
2195		2200		2205	
Trp Leu	Leu Leu Leu Ser	Ile	Cys Leu Gly Ser	Leu	Ile Cys Val
2210		2215		2220	
Thr Ala	Ala Phe Gly Val	Leu	Leu Ser Asn Phe	Gly	Ala Pro Ser
2225		2230		2235	
Tyr Cys	Asn Gly Val Arg	Glu	Leu Tyr Leu Asn	Ser	Ser Asn Val
2240		2245		2250	
Thr Thr	Met Asp Phe Cys	Glu	Gly Ser Phe Pro	Cys	Ser Ile Cys
2255		2260		2265	
Leu Ser	Gly Leu Asp Ser	Leu	Asp Ser Tyr Pro	Ala	Leu Glu Thr
2270		2275		2280	
Ile Gln	Val Thr Ile Ser	Ser	Tyr Lys Leu Asp	Leu	Thr Ile Leu
2285		2290		2295	
Gly Leu	Ala Ala Glu Trp	Val	Leu Ala Tyr Met	Leu	Phe Thr Lys
2300		2305		2310	
Phe Phe	Tyr Leu Leu Gly	Leu	Ser Ala Ile Met	Gln	Val Phe Phe
2315		2320		2325	
Gly Tyr	Phe Ala Ser His	Phe	Ile Ser Asn Ser	Trp	Leu Met Trp
2330		2335		2340	
Phe Ile	Ile Ser Ile Val	Gln	Met Ala Pro Val	Ser	Ala Met Val
2345		2350		2355	
Arg Met	Tyr Ile Phe Phe	Ala	Ser Phe Tyr Tyr	Ile	Trp Lys Ser
2360		2365		2370	
Tyr Val	His Ile Met Asp	Gly	Cys Thr Ser Ser	Thr	Cys Met Met
2375		2380		2385	
Cys Tyr	Lys Arg Asn Arg	Ala	Thr Arg Val Glu	Cys	Thr Thr Ile
2390		2395		2400	
Val Asn	Gly Met Lys Arg	Ser	Phe Tyr Val Tyr	Ala	Asn Gly Gly
2405		2410		2415	
Arg Gly	Phe Cys Lys Thr	His	Asn Trp Asn Cys	Leu	Asn Cys Asp
2420		2425		2430	
Thr Phe	Cys Thr Gly Ser	Thr	Phe Ile Ser Asp	Glu	Val Ala Arg
2435		2440		2445	
Asp Leu	Ser Leu Gln Phe	Lys	Arg Pro Ile Asn	Pro	Thr Asp Gln
2450		2455		2460	
Ser Ser	Tyr Ile Val Asp	Ser	Val Ala Val Lys	Asn	Gly Ala Leu
2465		2470		2475	
His Leu	Tyr Phe Asp Lys	Ala	Gly Gln Lys Thr	Tyr	Glu Arg His
2480		2485		2490	
Pro Leu	Ser His Phe Val	Asn	Leu Asp Asn Leu	Arg	Ala Asn Asn
2495		2500		2505	
Thr Lys	Gly Ser Leu Pro	Ile	Asn Val Ile Val	Phe	Asp Gly Lys
2510		2515		2520	
Ser Lys	Cys Asp Glu Ser	Ala	Ser Lys Ser Ala	Ser	Val Tyr Tyr
2525		2530		2535	
Ser Gln	Leu Met Cys Gln	Pro	Ile Leu Leu Leu	Asp	Gln Ala Leu

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2540	2545	2550
Val Ser Asp Val Gly Asp Ser Thr Glu Val Ser Val Lys Met Phe 2555 2560 2565		
Asp Ala Tyr Val Asp Thr Phe Ser Ala Thr Phe Ser Val Pro Met 2570 2575 2580		
Glu Lys Leu Lys Ala Leu Val Ala Thr Ala His Ser Glu Leu Ala 2585 2590 2595		
Lys Gly Val Ala Leu Asp Gly Val Leu Ser Thr Phe Val Ser Ala 2600 2605 2610		
Ala Arg Gln Gly Val Val Asp Thr Asp Val Asp Thr Lys Asp Val 2615 2620 2625		
Ile Glu Cys Leu Lys Leu Ser His His Ser Asp Leu Glu Val Thr 2630 2635 2640		
Gly Asp Ser Cys Asn Asn Phe Met Leu Thr Tyr Asn Lys Val Glu 2645 2650 2655		
Asn Met Thr Pro Arg Asp Leu Gly Ala Cys Ile Asp Cys Asn Ala 2660 2665 2670		
Arg His Ile Asn Ala Gln Val Ala Lys Ser His Asn Val Ser Leu 2675 2680 2685		
Ile Trp Asn Val Lys Asp Tyr Met Ser Leu Ser Glu Gln Leu Arg 2690 2695 2700		
Lys Gln Ile Arg Ser Ala Ala Lys Lys Asn Asn Ile Pro Phe Arg 2705 2710 2715		
Leu Thr Cys Ala Thr Thr Arg Gln Val Val Asn Val Ile Thr Thr 2720 2725 2730		
Lys Ile Ser Leu Lys Gly Gly Lys Ile Val Ser Thr Cys Phe Lys 2735 2740 2745		
Leu Met Leu Lys Ala Thr Leu Leu Cys Val Leu Ala Ala Leu Val 2750 2755 2760		
Cys Tyr Ile Val Met Pro Val His Thr Leu Ser Ile His Asp Gly 2765 2770 2775		
Tyr Thr Asn Glu Ile Ile Gly Tyr Lys Ala Ile Gln Asp Gly Val 2780 2785 2790		
Thr Arg Asp Ile Ile Ser Thr Asp Asp Cys Phe Ala Asn Lys His 2795 2800 2805		
Ala Gly Phe Asp Ala Trp Phe Ser Gln Arg Gly Gly Ser Tyr Lys 2810 2815 2820		
Asn Asp Lys Ser Cys Pro Val Val Ala Ala Ile Ile Thr Arg Glu 2825 2830 2835		
Ile Gly Phe Ile Val Pro Gly Leu Pro Gly Thr Val Leu Arg Ala 2840 2845 2850		
Ile Asn Gly Asp Phe Leu His Phe Leu Pro Arg Val Phe Ser Ala 2855 2860 2865		
Val Gly Asn Ile Cys Tyr Thr Pro Ser Lys Leu Ile Glu Tyr Ser 2870 2875 2880		
Asp Phe Ala Thr Ser Ala Cys Val Leu Ala Ala Glu Cys Thr Ile 2885 2890 2895		
Phe Lys Asp Ala Met Gly Lys Pro Val Pro Tyr Cys Tyr Asp Thr 2900 2905 2910		
Asn Leu Leu Glu Gly Ser Ile Ser Tyr Ser Glu Leu Arg Pro Asp 2915 2920 2925		

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Thr Arg	Tyr Val	Leu Met	Asp	Gly Ser	Ile Ile	Gln	Phe Pro	Asn	
2930			2935			2940			
Thr Tyr	Leu Glu	Gly Ser	Val	Arg Val	Val Thr	Thr	Phe Asp	Ala	
2945			2950			2955			
Glu Tyr	Cys Arg	His Gly	Thr	Cys Glu	Arg Ser	Glu	Val Gly	Ile	
2960			2965			2970			
Cys Leu	Ser Thr	Ser Gly	Arg	Trp Val	Leu Asn	Asn	Glu His	Tyr	
2975			2980			2985			
Arg Ala	Leu Ser	Gly Val	Phe	Cys Gly	Val Asp	Ala	Met Asn	Leu	
2990			2995			3000			
Ile Ala	Asn Ile	Phe Thr	Pro	Leu Val	Gln Pro	Val	Gly Ala	Leu	
3005			3010			3015			
Asp Val	Ser Ala	Ser Val	Val	Ala Gly	Gly Ile	Ile	Ala Ile	Leu	
3020			3025			3030			
Val Thr	Cys Ala	Ala Tyr	Tyr	Phe Met	Lys Phe	Arg	Arg Val	Phe	
3035			3040			3045			
Gly Glu	Tyr Asn	His Val	Val	Ala Ala	Asn Ala	Leu	Leu Phe	Leu	
3050			3055			3060			
Met Ser	Phe Thr	Ile Leu	Cys	Leu Val	Pro Ala	Tyr	Ser Phe	Leu	
3065			3070			3075			
Pro Gly	Val Tyr	Ser Val	Phe	Tyr Leu	Tyr Leu	Thr	Phe Tyr	Phe	
3080			3085			3090			
Thr Asn	Asp Val	Ser Phe	Leu	Ala His	Leu Gln	Trp	Phe Ala	Met	
3095			3100			3105			
Phe Ser	Pro Ile	Val Pro	Phe	Trp Ile	Thr Ala	Ile	Tyr Val	Phe	
3110			3115			3120			
Cys Ile	Ser Leu	Lys His	Cys	His Trp	Phe Phe	Asn	Asn Tyr	Leu	
3125			3130			3135			
Arg Lys	Arg Val	Met Phe	Asn	Gly Val	Thr Phe	Ser	Thr Phe	Glu	
3140			3145			3150			
Glu Ala	Ala Leu	Cys Thr	Phe	Leu Leu	Asn Lys	Glu	Met Tyr	Leu	
3155			3160			3165			
Lys Leu	Arg Ser	Glu Thr	Leu	Leu Pro	Leu Thr	Gln	Tyr Asn	Arg	
3170			3175			3180			
Tyr Leu	Ala Leu	Tyr Asn	Lys	Tyr Lys	Tyr Phe	Ser	Gly Ala	Leu	
3185			3190			3195			
Asp Thr	Thr Ser	Tyr Arg	Glu	Ala Ala	Cys Cys	His	Leu Ala	Lys	
3200			3205			3210			
Ala Leu	Asn Asp	Phe Ser	Asn	Ser Gly	Ala Asp	Val	Leu Tyr	Gln	
3215			3220			3225			
Pro Pro	Gln Thr	Ser Ile	Thr	Ser Ala	Val Leu	Gln	Ser Gly	Phe	
3230			3235			3240			
Arg Lys	Met Ala	Phe Pro	Ser	Gly Lys	Val Glu	Gly	Cys Met	Val	
3245			3250			3255			
Gln Val	Thr Cys	Gly Thr	Thr	Thr Leu	Asn Gly	Leu	Trp Leu	Asp	
3260			3265			3270			
Asp Thr	Val Tyr	Cys Pro	Arg	His Val	Ile Cys	Thr	Ala Glu	Asp	
3275			3280			3285			
Met Leu	Asn Pro	Asn Tyr	Glu	Asp Leu	Leu Ile	Arg	Lys Ser	Asn	
3290			3295			3300			

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His Ser	Phe Leu Val Gln	Ala Gly	Asn Val Gln	Leu Arg	Val Ile
3305		3310		3315	
Gly His	Ser Met Gln Asn Cys	Leu Leu Arg	Leu Lys	Val Asp	Thr
3320		3325		3330	
Ser Asn	Pro Lys Thr Pro	Lys Tyr Lys	Phe Val Arg	Ile Gln	Pro
3335		3340		3345	
Gly Gln	Thr Phe Ser Val	Leu Ala Cys	Tyr Asn Gly	Ser Pro	Ser
3350		3355		3360	
Gly Val	Tyr Gln Cys Ala Met	Arg Pro Asn His	Thr Ile	Lys Gly	
3365		3370		3375	
Ser Phe	Leu Asn Gly Ser	Cys Gly Ser	Val Gly Phe	Asn Ile	Asp
3380		3385		3390	
Tyr Asp	Cys Val Ser Phe	Cys Tyr Met His	His Met	Glu Leu	Pro
3395		3400		3405	
Thr Gly	Val His Ala Gly	Thr Asp Leu	Glu Gly Lys	Phe Tyr	Gly
3410		3415		3420	
Pro Phe	Val Asp Arg Gln	Thr Ala Gln	Ala Ala Gly	Thr Asp	Thr
3425		3430		3435	
Thr Ile	Thr Leu Asn Val	Leu Ala Trp	Leu Tyr Ala	Ala Val	Ile
3440		3445		3450	
Asn Gly	Asp Arg Trp Phe	Leu Asn Arg	Phe Thr Thr	Thr Leu	Asn
3455		3460		3465	
Asp Phe	Asn Leu Val Ala	Met Lys Tyr	Asn Tyr Glu	Pro Leu	Thr
3470		3475		3480	
Gln Asp	His Val Asp Ile	Leu Gly Pro	Leu Ser Ala	Gln Thr	Gly
3485		3490		3495	
Ile Ala	Val Leu Asp Met	Cys Ala Ala	Leu Lys Glu	Leu Leu	Gln
3500		3505		3510	
Asn Gly	Met Asn Gly Arg	Thr Ile Leu	Gly Ser Thr	Ile Leu	Glu
3515		3520		3525	
Asp Glu	Phe Thr Pro Phe	Asp Val Val	Arg Gln Cys	Ser Gly	Val
3530		3535		3540	
Thr Phe	Gln Gly Lys Phe	Lys Lys Ile	Val Lys Gly	Thr His	His
3545		3550		3555	
Trp Met	Leu Leu Thr Phe	Leu Thr Ser	Leu Leu Ile	Leu Val	Gln
3560		3565		3570	
Ser Thr	Gln Trp Ser Leu	Phe Phe Phe	Val Tyr Glu	Asn Ala	Phe
3575		3580		3585	
Leu Pro	Phe Thr Leu Gly	Ile Met Ala	Ile Ala Ala	Cys Ala	Met
3590		3595		3600	
Leu Leu	Val Lys His Lys	His Ala Phe	Leu Cys Leu	Phe Leu	Leu
3605		3610		3615	
Pro Ser	Leu Ala Thr Val	Ala Tyr Phe	Asn Met Val	Tyr Met	Pro
3620		3625		3630	
Ala Ser	Trp Val Met Arg	Ile Met Thr	Trp Leu Glu	Leu Ala	Asp
3635		3640		3645	
Thr Ser	Leu Ser Gly Tyr	Arg Leu Lys	Asp Cys Val	Met Tyr	Ala
3650		3655		3660	
Ser Ala	Leu Val Leu Leu	Ile Leu Met	Thr Ala Arg	Thr Val	Tyr
3665		3670		3675	
Asp Asp	Ala Ala Arg Arg	Val Trp Thr	Leu Met Asn	Val Ile	Thr

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3680	3685	3690
Leu Val Tyr Lys Val Tyr	Tyr Gly Asn Ala Leu	Asp Gln Ala Ile
3695	3700	3705
Ser Met Trp Ala Leu Val	Ile Ser Val Thr Ser	Asn Tyr Ser Gly
3710	3715	3720
Val Val Thr Thr Ile Met	Phe Leu Ala Arg Ala	Ile Val Phe Val
3725	3730	3735
Cys Val Glu Tyr Tyr Pro	Leu Leu Phe Ile Thr	Gly Asn Thr Leu
3740	3745	3750
Gln Cys Ile Met Leu Val	Tyr Cys Phe Leu Gly	Tyr Cys Cys Cys
3755	3760	3765
Cys Tyr Phe Gly Leu Phe	Cys Leu Leu Asn Arg	Tyr Phe Arg Leu
3770	3775	3780
Thr Leu Gly Val Tyr Asp	Tyr Leu Val Ser Thr	Gln Glu Phe Arg
3785	3790	3795
Tyr Met Asn Ser Gln Gly	Leu Leu Pro Pro Lys	Ser Ser Ile Asp
3800	3805	3810
Ala Phe Lys Leu Asn Ile	Lys Leu Leu Gly Ile	Gly Gly Lys Pro
3815	3820	3825
Cys Ile Lys Val Ala Thr	Val Gln Ser Lys Met	Ser Asp Val Lys
3830	3835	3840
Cys Thr Ser Val Val Leu	Leu Ser Val Leu Gln	Gln Leu Arg Val
3845	3850	3855
Glu Ser Ser Ser Lys Leu	Trp Ala Gln Cys Val	Gln Leu His Asn
3860	3865	3870
Asp Ile Leu Leu Ala Lys	Asp Thr Thr Glu Ala	Phe Glu Lys Met
3875	3880	3885
Val Ser Leu Leu Ser Val	Leu Leu Ser Met Gln	Gly Ala Val Asp
3890	3895	3900
Ile Asn Arg Leu Cys Glu	Glu Met Leu Asp Asn	Arg Ala Thr Leu
3905	3910	3915
Gln Ala Ile Ala Ser Glu	Phe Ser Ser Leu Pro	Ser Tyr Ala Ala
3920	3925	3930
Tyr Ala Thr Ala Gln Glu	Ala Tyr Glu Gln Ala	Val Ala Asn Gly
3935	3940	3945
Asp Ser Glu Val Val Leu	Lys Lys Leu Lys Lys	Ser Leu Asn Val
3950	3955	3960
Ala Lys Ser Glu Phe Asp	Arg Asp Ala Ala Met	Gln Arg Lys Leu
3965	3970	3975
Glu Lys Met Ala Asp Gln	Ala Met Thr Gln Met	Tyr Lys Gln Ala
3980	3985	3990
Arg Ser Glu Asp Lys Arg	Ala Lys Val Thr Ser	Ala Met Gln Thr
3995	4000	4005
Met Leu Phe Thr Met Leu	Arg Lys Leu Asp Asn	Asp Ala Leu Asn
4010	4015	4020
Asn Ile Ile Asn Asn Ala	Arg Asp Gly Cys Val	Pro Leu Asn Ile
4025	4030	4035
Ile Pro Leu Thr Thr Ala	Ala Lys Leu Met Val	Val Val Pro Asp
4040	4045	4050
Tyr Gly Thr Tyr Lys Asn	Thr Cys Asp Gly Asn	Thr Phe Thr Tyr
4055	4060	4065

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Ala Ser	Ala Leu Trp Glu Ile	Gln Gln Val Val Asp	Ala Asp Ser
4070	4075	4080	
Lys Ile	Val Gln Leu Ser Glu Ile	Asn Met Asp Asn	Ser Pro Asn
4085	4090	4095	
Leu Ala	Trp Pro Leu Ile Val	Thr Ala Leu Arg Ala	Asn Ser Ala
4100	4105	4110	
Val Lys	Leu Gln Asn Asn Glu	Leu Ser Pro Val Ala	Leu Arg Gln
4115	4120	4125	
Met Ser	Cys Ala Ala Gly Thr	Thr Gln Thr Ala Cys	Thr Asp Asp
4130	4135	4140	
Asn Ala	Leu Ala Tyr Tyr Asn	Asn Ser Lys Gly Gly	Arg Phe Val
	4145	4150	4155
Leu Ala	Leu Leu Ser Asp His	Gln Asp Leu Lys Trp	Ala Arg Phe
4160	4165	4170	
Pro Lys	Ser Asp Gly Thr Gly	Thr Ile Tyr Thr Glu	Leu Glu Pro
4175	4180	4185	
Pro Cys	Arg Phe Val Thr Asp	Thr Pro Lys Gly Pro	Lys Val Lys
4190	4195	4200	
Tyr Leu	Tyr Phe Ile Lys Gly	Leu Asn Asn Leu Asn	Arg Gly Met
4205	4210	4215	
Val Leu	Gly Ser Leu Ala Ala	Thr Val Arg Leu Gln	Ala Gly Asn
4220	4225	4230	
Ala Thr	Glu Val Pro Ala Asn	Ser Thr Val Leu Ser	Phe Cys Ala
4235	4240	4245	
Phe Ala	Val Asp Pro Ala Lys	Ala Tyr Lys Asp Tyr	Leu Ala Ser
4250	4255	4260	
Gly Gly	Gln Pro Ile Thr Asn	Cys Val Lys Met Leu	Cys Thr His
4265	4270	4275	
Thr Gly	Thr Gly Gln Ala Ile	Thr Val Thr Pro Glu	Ala Asn Met
4280	4285	4290	
Asp Gln	Glu Ser Phe Gly Gly	Ala Ser Cys Cys Leu	Tyr Cys Arg
4295	4300	4305	
Cys His	Ile Asp His Pro Asn	Pro Lys Gly Phe Cys	Asp Leu Lys
4310	4315	4320	
Gly Lys	Tyr Val Gln Ile Pro	Thr Thr Cys Ala Asn	Asp Pro Val
4325	4330	4335	
Gly Phe	Thr Leu Arg Asn Thr	Val Cys Thr Val Cys	Gly Met Trp
4340	4345	4350	
Lys Gly	Tyr Gly Cys Ser Cys	Asp Gln Leu Arg Glu	Pro Leu Met
4355	4360	4365	
Gln Ser	Ala Asp Ala Ser Thr	Phe Leu Asn Gly Phe	Ala Val
4370	4375	4380	

<210> SEQ ID NO 75

<211> LENGTH: 2695

<212> TYPE: PRT

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 75

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1 5 10 15

Thr Ser Thr Asp Val Val Tyr Arg Ala Phe Asp Ile Tyr Asn Glu Lys

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Val	Ala	Gly	Phe	Ala	Lys	Phe	Leu	Lys	Thr	Asn	Cys	Cys	Arg	Phe	Gln
		35					40			45					
Glu	Lys	Asp	Glu	Glu	Gly	Asn	Leu	Leu	Asp	Ser	Tyr	Phe	Val	Val	Lys
		50			55				60						
Arg	His	Thr	Met	Ser	Asn	Tyr	Gln	His	Glu	Glu	Thr	Ile	Tyr	Asn	Leu
65				70				75						80	
Val	Lys	Asp	Cys	Pro	Ala	Val	Ala	Val	His	Asp	Phe	Phe	Lys	Phe	Arg
				85				90						95	
Val	Asp	Gly	Asp	Met	Val	Pro	His	Ile	Ser	Arg	Gln	Arg	Leu	Thr	Lys
		100						105				110			
Tyr	Thr	Met	Ala	Asp	Leu	Val	Tyr	Ala	Leu	Arg	His	Phe	Asp	Glu	Gly
		115				120						125			
Asn	Cys	Asp	Thr	Leu	Lys	Glu	Ile	Leu	Val	Thr	Tyr	Asn	Cys	Cys	Asp
		130				135				140					
Asp	Asp	Tyr	Phe	Asn	Lys	Lys	Asp	Trp	Tyr	Asp	Phe	Val	Glu	Asn	Pro
145				150				155						160	
Asp	Ile	Leu	Arg	Val	Tyr	Ala	Asn	Leu	Gly	Glu	Arg	Val	Arg	Gln	Ser
				165				170						175	
Leu	Leu	Lys	Thr	Val	Gln	Phe	Cys	Asp	Ala	Met	Arg	Asp	Ala	Gly	Ile
		180						185				190			
Val	Gly	Val	Leu	Thr	Leu	Asp	Asn	Gln	Asp	Leu	Asn	Gly	Asn	Trp	Tyr
		195				200						205			
Asp	Phe	Gly	Asp	Phe	Val	Gln	Val	Ala	Pro	Gly	Cys	Gly	Val	Pro	Ile
		210				215				220					
Val	Asp	Ser	Tyr	Tyr	Ser	Leu	Leu	Met	Pro	Ile	Leu	Thr	Leu	Thr	Arg
225				230				235						240	
Ala	Leu	Ala	Ala	Glu	Ser	His	Met	Asp	Ala	Asp	Leu	Ala	Lys	Pro	Leu
				245				250						255	
Ile	Lys	Trp	Asp	Leu	Leu	Lys	Tyr	Asp	Phe	Thr	Glu	Glu	Arg	Leu	Cys
		260						265				270			
Leu	Phe	Asp	Arg	Tyr	Phe	Lys	Tyr	Trp	Asp	Gln	Thr	Tyr	His	Pro	Asn
		275				280						285			
Cys	Ile	Asn	Cys	Leu	Asp	Asp	Arg	Cys	Ile	Leu	His	Cys	Ala	Asn	Phe
		290				295				300					
Asn	Val	Leu	Phe	Ser	Thr	Val	Phe	Pro	Pro	Thr	Ser	Phe	Gly	Pro	Leu
305				310				315						320	
Val	Arg	Lys	Ile	Phe	Val	Asp	Gly	Val	Pro	Phe	Val	Val	Ser	Thr	Gly
				325				330						335	
Tyr	His	Phe	Arg	Glu	Leu	Gly	Val	Val	His	Asn	Gln	Asp	Val	Asn	Leu
		340						345				350			
His	Ser	Ser	Arg	Leu	Ser	Phe	Lys	Glu	Leu	Leu	Val	Tyr	Ala	Ala	Asp
		355				360						365			
Pro	Ala	Met	His	Ala	Ala	Ser	Gly	Asn	Leu	Leu	Leu	Asp	Lys	Arg	Thr
		370				375				380					
Thr	Cys	Phe	Ser	Val	Ala	Ala	Leu	Thr	Asn	Asn	Val	Ala	Phe	Gln	Thr
385				390				395						400	
Val	Lys	Pro	Gly	Asn	Phe	Asn	Lys	Asp	Phe	Tyr	Asp	Phe	Ala	Val	Ser
				405				410						415	
Lys	Gly	Phe	Phe	Lys	Glu	Gly	Ser	Ser	Val	Glu	Leu	Lys	His	Phe	Phe
		420						425				430			

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Phe	Ala	Gln	Asp	Gly	Asn	Ala	Ala	Ile	Ser	Asp	Tyr	Asp	Tyr	Tyr	Arg
	435						440					445			
Tyr	Asn	Leu	Pro	Thr	Met	Cys	Asp	Ile	Arg	Gln	Leu	Leu	Phe	Val	Val
	450						455				460				
Glu	Val	Val	Asp	Lys	Tyr	Phe	Asp	Cys	Tyr	Asp	Gly	Gly	Cys	Ile	Asn
465					470					475					480
Ala	Asn	Gln	Val	Ile	Val	Asn	Asn	Leu	Asp	Lys	Ser	Ala	Gly	Phe	Pro
				485					490					495	
Phe	Asn	Lys	Trp	Gly	Lys	Ala	Arg	Leu	Tyr	Tyr	Asp	Ser	Met	Ser	Tyr
		500						505					510		
Glu	Asp	Gln	Asp	Ala	Leu	Phe	Ala	Tyr	Thr	Lys	Arg	Asn	Val	Ile	Pro
	515						520					525			
Thr	Ile	Thr	Gln	Met	Asn	Leu	Lys	Tyr	Ala	Ile	Ser	Ala	Lys	Asn	Arg
	530					535					540				
Ala	Arg	Thr	Val	Ala	Gly	Val	Ser	Ile	Cys	Ser	Thr	Met	Thr	Asn	Arg
545					550					555					560
Gln	Phe	His	Gln	Lys	Leu	Leu	Lys	Ser	Ile	Ala	Ala	Thr	Arg	Gly	Ala
				565					570					575	
Thr	Val	Val	Ile	Gly	Thr	Ser	Lys	Phe	Tyr	Gly	Gly	Trp	His	Asn	Met
			580					585					590		
Leu	Lys	Thr	Val	Tyr	Ser	Asp	Val	Glu	Thr	Pro	His	Leu	Met	Gly	Trp
	595						600					605			
Asp	Tyr	Pro	Lys	Cys	Asp	Arg	Ala	Met	Pro	Asn	Met	Leu	Arg	Ile	Met
	610					615					620				
Ala	Ser	Leu	Val	Leu	Ala	Arg	Lys	His	Asn	Thr	Cys	Cys	Asn	Leu	Ser
625					630					635					640
His	Arg	Phe	Tyr	Arg	Leu	Ala	Asn	Glu	Cys	Ala	Gln	Val	Leu	Ser	Glu
				645					650					655	
Met	Val	Met	Cys	Gly	Gly	Ser	Leu	Tyr	Val	Lys	Pro	Gly	Gly	Thr	Ser
			660					665					670		
Ser	Gly	Asp	Ala	Thr	Thr	Ala	Tyr	Ala	Asn	Ser	Val	Phe	Asn	Ile	Cys
	675					680						685			
Gln	Ala	Val	Thr	Ala	Asn	Val	Asn	Ala	Leu	Leu	Ser	Thr	Asp	Gly	Asn
	690					695					700				
Lys	Ile	Ala	Asp	Lys	Tyr	Val	Arg	Asn	Leu	Gln	His	Arg	Leu	Tyr	Glu
705					710					715					720
Cys	Leu	Tyr	Arg	Asn	Arg	Asp	Val	Asp	His	Glu	Phe	Val	Asp	Glu	Phe
				725					730					735	
Tyr	Ala	Tyr	Leu	Arg	Lys	His	Phe	Ser	Met	Met	Ile	Leu	Ser	Asp	Asp
		740						745					750		
Ala	Val	Val	Cys	Tyr	Asn	Ser	Asn	Tyr	Ala	Ala	Gln	Gly	Leu	Val	Ala
		755					760					765			
Ser	Ile	Lys	Asn	Phe	Lys	Ala	Val	Leu	Tyr	Tyr	Gln	Asn	Asn	Val	Phe
	770					775					780				
Met	Ser	Glu	Ala	Lys	Cys	Trp	Thr	Glu	Thr	Asp	Leu	Thr	Lys	Gly	Pro
785					790					795					800
His	Glu	Phe	Cys	Ser	Gln	His	Thr	Met	Leu	Val	Lys	Gln	Gly	Asp	Asp
				805					810					815	
Tyr	Val	Tyr	Leu	Pro	Tyr	Pro	Asp	Pro	Ser	Arg	Ile	Leu	Gly	Ala	Gly
			820					825					830		

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Cys	Phe	Val	Asp	Asp	Ile	Val	Lys	Thr	Asp	Gly	Thr	Leu	Met	Ile	Glu
	835						840					845			
Arg	Phe	Val	Ser	Leu	Ala	Ile	Asp	Ala	Tyr	Pro	Leu	Thr	Lys	His	Pro
	850					855					860				
Asn	Gln	Glu	Tyr	Ala	Asp	Val	Phe	His	Leu	Tyr	Leu	Gln	Tyr	Ile	Arg
865					870					875				880	
Lys	Leu	His	Asp	Glu	Leu	Thr	Gly	His	Met	Leu	Asp	Met	Tyr	Ser	Val
			885						890					895	
Met	Leu	Thr	Asn	Asp	Asn	Thr	Ser	Arg	Tyr	Trp	Glu	Pro	Glu	Phe	Tyr
			900					905					910		
Glu	Ala	Met	Tyr	Thr	Pro	His	Thr	Val	Leu	Gln	Ala	Val	Gly	Ala	Cys
		915					920					925			
Val	Leu	Cys	Asn	Ser	Gln	Thr	Ser	Leu	Arg	Cys	Gly	Ala	Cys	Ile	Arg
	930					935					940				
Arg	Pro	Phe	Leu	Cys	Cys	Lys	Cys	Cys	Tyr	Asp	His	Val	Ile	Ser	Thr
945					950					955					960
Ser	His	Lys	Leu	Val	Leu	Ser	Val	Asn	Pro	Tyr	Val	Cys	Asn	Ala	Pro
			965					970						975	
Gly	Cys	Asp	Val	Thr	Asp	Val	Thr	Gln	Leu	Tyr	Leu	Gly	Gly	Met	Ser
		980					985						990		
Tyr	Tyr	Cys	Lys	Ser	His	Lys	Pro	Pro	Ile	Ser	Phe	Pro	Leu	Cys	Ala
		995					1000					1005			
Asn	Gly	Gln	Val	Phe	Gly	Leu	Tyr	Lys	Asn	Thr	Cys	Val	Gly	Ser	
	1010					1015						1020			
Asp	Asn	Val	Thr	Asp	Phe	Asn	Ala	Ile	Ala	Thr	Cys	Asp	Trp	Thr	
	1025					1030						1035			
Asn	Ala	Gly	Asp	Tyr	Ile	Leu	Ala	Asn	Thr	Cys	Thr	Glu	Arg	Leu	
	1040					1045						1050			
Lys	Leu	Phe	Ala	Ala	Glu	Thr	Leu	Lys	Ala	Thr	Glu	Glu	Thr	Phe	
	1055					1060						1065			
Lys	Leu	Ser	Tyr	Gly	Ile	Ala	Thr	Val	Arg	Glu	Val	Leu	Ser	Asp	
	1070					1075						1080			
Arg	Glu	Leu	His	Leu	Ser	Trp	Glu	Val	Gly	Lys	Pro	Arg	Pro	Pro	
	1085					1090						1095			
Leu	Asn	Arg	Asn	Tyr	Val	Phe	Thr	Gly	Tyr	Arg	Val	Thr	Lys	Asn	
	1100					1105						1110			
Ser	Lys	Val	Gln	Ile	Gly	Glu	Tyr	Thr	Phe	Glu	Lys	Gly	Asp	Tyr	
	1115					1120						1125			
Gly	Asp	Ala	Val	Val	Tyr	Arg	Gly	Thr	Thr	Thr	Tyr	Lys	Leu	Asn	
	1130					1135						1140			
Val	Gly	Asp	Tyr	Phe	Val	Leu	Thr	Ser	His	Thr	Val	Met	Pro	Leu	
	1145					1150						1155			
Ser	Ala	Pro	Thr	Leu	Val	Pro	Gln	Glu	His	Tyr	Val	Arg	Ile	Thr	
	1160					1165						1170			
Gly	Leu	Tyr	Pro	Thr	Leu	Asn	Ile	Ser	Asp	Glu	Phe	Ser	Ser	Asn	
	1175					1180						1185			
Val	Ala	Asn	Tyr	Gln	Lys	Val	Gly	Met	Gln	Lys	Tyr	Ser	Thr	Leu	
	1190					1195						1200			
Gln	Gly	Pro	Pro	Gly	Thr	Gly	Lys	Ser	His	Phe	Ala	Ile	Gly	Leu	
	1205					1210						1215			
Ala	Leu	Tyr	Tyr	Pro	Ser	Ala	Arg	Ile	Val	Tyr	Thr	Ala	Cys	Ser	

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1220	1225	1230
His Ala Ala Val Asp Ala Leu Cys Glu Lys Ala Leu Lys Tyr Leu 1235 1240 1245		
Pro Ile Asp Lys Cys Ser Arg Ile Ile Pro Ala Arg Ala Arg Val 1250 1255 1260		
Glu Cys Phe Asp Lys Phe Lys Val Asn Ser Thr Leu Glu Gln Tyr 1265 1270 1275		
Val Phe Cys Thr Val Asn Ala Leu Pro Glu Thr Thr Ala Asp Ile 1280 1285 1290		
Val Val Phe Asp Glu Ile Ser Met Ala Thr Asn Tyr Asp Leu Ser 1295 1300 1305		
Val Val Asn Ala Arg Leu Arg Ala Lys His Tyr Val Tyr Ile Gly 1310 1315 1320		
Asp Pro Ala Gln Leu Pro Ala Pro Arg Thr Leu Leu Thr Lys Gly 1325 1330 1335		
Thr Leu Glu Pro Glu Tyr Phe Asn Ser Val Cys Arg Leu Met Lys 1340 1345 1350		
Thr Ile Gly Pro Asp Met Phe Leu Gly Thr Cys Arg Arg Cys Pro 1355 1360 1365		
Ala Glu Ile Val Asp Thr Val Ser Ala Leu Val Tyr Asp Asn Lys 1370 1375 1380		
Leu Lys Ala His Lys Asp Lys Ser Ala Gln Cys Phe Lys Met Phe 1385 1390 1395		
Tyr Lys Gly Val Ile Thr His Asp Val Ser Ser Ala Ile Asn Arg 1400 1405 1410		
Pro Gln Ile Gly Val Val Arg Glu Phe Leu Thr Arg Asn Pro Ala 1415 1420 1425		
Trp Arg Lys Ala Val Phe Ile Ser Pro Tyr Asn Ser Gln Asn Ala 1430 1435 1440		
Val Ala Ser Lys Ile Leu Gly Leu Pro Thr Gln Thr Val Asp Ser 1445 1450 1455		
Ser Gln Gly Ser Glu Tyr Asp Tyr Val Ile Phe Thr Gln Thr Thr 1460 1465 1470		
Glu Thr Ala His Ser Cys Asn Val Asn Arg Phe Asn Val Ala Ile 1475 1480 1485		
Thr Arg Ala Lys Ile Gly Ile Leu Cys Ile Met Ser Asp Arg Asp 1490 1495 1500		
Leu Tyr Asp Lys Leu Gln Phe Thr Ser Leu Glu Ile Pro Arg Arg 1505 1510 1515		
Asn Val Ala Thr Leu Gln Ala Glu Asn Val Thr Gly Leu Phe Lys 1520 1525 1530		
Asp Cys Ser Lys Ile Ile Thr Gly Leu His Pro Thr Gln Ala Pro 1535 1540 1545		
Thr His Leu Ser Val Asp Ile Lys Phe Lys Thr Glu Gly Leu Cys 1550 1555 1560		
Val Asp Ile Pro Gly Ile Pro Lys Asp Met Thr Tyr Arg Arg Leu 1565 1570 1575		
Ile Ser Met Met Gly Phe Lys Met Asn Tyr Gln Val Asn Gly Tyr 1580 1585 1590		
Pro Asn Met Phe Ile Thr Arg Glu Glu Ala Ile Arg His Val Arg 1595 1600 1605		

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Ala Trp	Ile Gly Phe Asp	Val	Glu Gly Cys His	Ala	Thr Arg Asp
1610		1615		1620	
Ala Val	Gly Thr Asn Leu	Pro	Leu Gln Leu Gly	Phe	Ser Thr Gly
1625		1630		1635	
Val Asn	Leu Val Ala Val	Pro	Thr Gly Tyr Val	Asp	Thr Glu Asn
1640		1645		1650	
Asn Thr	Glu Phe Thr Arg	Val	Asn Ala Lys Pro	Pro	Pro Gly Asp
1655		1660		1665	
Gln Phe	Lys His Leu Ile	Pro	Leu Met Tyr Lys	Gly	Leu Pro Trp
1670		1675		1680	
Asn Val	Val Arg Ile Lys	Ile	Val Gln Met Leu	Ser	Asp Thr Leu
1685		1690		1695	
Lys Gly	Leu Ser Asp Arg	Val	Val Phe Val Leu	Trp	Ala His Gly
1700		1705		1710	
Phe Glu	Leu Thr Ser Met	Lys	Tyr Phe Val Lys	Ile	Gly Pro Glu
1715		1720		1725	
Arg Thr	Cys Cys Leu Cys	Asp	Lys Arg Ala Thr	Cys	Phe Ser Thr
1730		1735		1740	
Ser Ser	Asp Thr Tyr Ala	Cys	Trp Asn His Ser	Val	Gly Phe Asp
1745		1750		1755	
Tyr Val	Tyr Asn Pro Phe	Met	Ile Asp Val Gln	Gln	Trp Gly Phe
1760		1765		1770	
Thr Gly	Asn Leu Gln Ser	Asn	His Asp Gln His	Cys	Gln Val His
1775		1780		1785	
Gly Asn	Ala His Val Ala	Ser	Cys Asp Ala Ile	Met	Thr Arg Cys
1790		1795		1800	
Leu Ala	Val His Glu Cys	Phe	Val Lys Arg Val	Asp	Trp Ser Val
1805		1810		1815	
Glu Tyr	Pro Ile Ile Gly	Asp	Glu Leu Arg Val	Asn	Ser Ala Cys
1820		1825		1830	
Arg Lys	Val Gln His Met	Val	Val Lys Ser Ala	Leu	Leu Ala Asp
1835		1840		1845	
Lys Phe	Pro Val Leu His	Asp	Ile Gly Asn Pro	Lys	Ala Ile Lys
1850		1855		1860	
Cys Val	Pro Gln Ala Glu	Val	Glu Trp Lys Phe	Tyr	Asp Ala Gln
1865		1870		1875	
Pro Cys	Ser Asp Lys Ala	Tyr	Lys Ile Glu Glu	Leu	Phe Tyr Ser
1880		1885		1890	
Tyr Ala	Thr His His Asp	Lys	Phe Thr Asp Gly	Val	Cys Leu Phe
1895		1900		1905	
Trp Asn	Cys Asn Val Asp	Arg	Tyr Pro Ala Asn	Ala	Ile Val Cys
1910		1915		1920	
Arg Phe	Asp Thr Arg Val	Leu	Ser Asn Leu Asn	Leu	Pro Gly Cys
1925		1930		1935	
Asp Gly	Gly Ser Leu Tyr	Val	Asn Lys His Ala	Phe	His Thr Pro
1940		1945		1950	
Ala Phe	Asp Lys Ser Ala	Phe	Thr Asn Leu Lys	Gln	Leu Pro Phe
1955		1960		1965	
Phe Tyr	Tyr Ser Asp Ser	Pro	Cys Glu Ser His	Gly	Lys Gln Val
1970		1975		1980	

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Val Ser 1985	Asp Ile Asp Tyr 1990	Val Pro Leu Lys Ser 1995	Ala Thr Cys Ile 1995
Thr Arg 2000	Cys Asn Leu Gly Gly 2005	Ala Val Cys Arg His 2010	His Ala Asn 2010
Glu Tyr 2015	Arg Gln Tyr Leu Asp 2020	Ala Tyr Asn Met Met 2025	Ile Ser Ala 2025
Gly Phe 2030	Ser Leu Trp Ile Tyr 2035	Lys Gln Phe Asp Thr 2040	Tyr Asn Leu 2040
Trp Asn 2045	Thr Phe Thr Arg Leu 2050	Gln Ser Leu Glu Asn 2055	Val Ala Tyr 2055
Asn Val 2060	Val Asn Lys Gly His 2065	Phe Asp Gly His Ala 2070	Gly Glu Ala 2070
Pro Val 2075	Ser Ile Ile Asn Asn 2080	Ala Val Tyr Thr Lys 2085	Val Asp Gly 2085
Ile Asp 2090	Val Glu Ile Phe Glu 2095	Asn Lys Thr Thr Leu 2100	Pro Val Asn 2100
Val Ala 2105	Phe Glu Leu Trp Ala 2110	Lys Arg Asn Ile Lys 2115	Pro Val Pro 2115
Glu Ile 2120	Lys Ile Leu Asn Asn 2125	Leu Gly Val Asp Ile 2130	Ala Ala Asn 2130
Thr Val 2135	Ile Trp Asp Tyr Lys 2140	Arg Glu Ala Pro Ala 2145	His Val Ser 2145
Thr Ile 2150	Gly Val Cys Thr Met 2155	Thr Asp Ile Ala Lys 2160	Lys Pro Thr 2160
Glu Ser 2165	Ala Cys Ser Ser Leu 2170	Thr Val Leu Phe Asp 2175	Gly Arg Val 2175
Glu Gly 2180	Gln Val Asp Leu Phe 2185	Arg Asn Ala Arg Asn 2190	Gly Val Leu 2190
Ile Thr 2195	Glu Gly Ser Val Lys 2200	Gly Leu Thr Pro Ser 2205	Lys Gly Pro 2205
Ala Gln 2210	Ala Ser Val Asn Gly 2215	Val Thr Leu Ile Gly 2220	Glu Ser Val 2220
Lys Thr 2225	Gln Phe Asn Tyr Phe 2230	Lys Lys Val Asp Gly 2235	Ile Ile Gln 2235
Gln Leu 2240	Pro Glu Thr Tyr Phe 2245	Thr Gln Ser Arg Asp 2250	Leu Glu Asp 2250
Phe Lys 2255	Pro Arg Ser Gln Met 2260	Glu Thr Asp Phe Leu 2265	Glu Leu Ala 2265
Met Asp 2270	Glu Phe Ile Gln Arg 2275	Tyr Lys Leu Glu Gly 2280	Tyr Ala Phe 2280
Glu His 2285	Ile Val Tyr Gly Asp 2290	Phe Ser His Gly Gln 2295	Leu Gly Gly 2295
Leu His 2300	Leu Met Ile Gly Leu 2305	Ala Lys Arg Ser Gln 2310	Asp Ser Pro 2310
Leu Lys 2315	Leu Glu Asp Phe Ile 2320	Pro Met Asp Ser Thr 2325	Val Lys Asn 2325
Tyr Phe 2330	Ile Thr Asp Ala Gln 2335	Thr Gly Ser Ser Lys 2340	Cys Val Cys 2340
Ser Val 2345	Ile Asp Leu Leu Leu 2350	Asp Asp Phe Val Glu 2355	Ile Ile Lys 2355
Ser Gln	Asp Leu Ser Val Ile	Ser Lys Val Val Lys	Val Thr Ile

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2360	2365	2370
Asp Tyr Ala Glu Ile Ser Phe Met Leu Trp Cys Lys Asp Gly His	2380	2385
2375		
Val Glu Thr Phe Tyr Pro Lys Leu Gln Ala Ser Gln Ala Trp Gln	2395	2400
2390		
Pro Gly Val Ala Met Pro Asn Leu Tyr Lys Met Gln Arg Met Leu	2410	2415
2405		
Leu Glu Lys Cys Asp Leu Gln Asn Tyr Gly Glu Asn Ala Val Ile	2425	2430
2420		
Pro Lys Gly Ile Met Met Asn Val Ala Lys Tyr Thr Gln Leu Cys	2440	2445
2435		
Gln Tyr Leu Asn Thr Leu Thr Leu Ala Val Pro Tyr Asn Met Arg	2455	2460
2450		
Val Ile His Phe Gly Ala Gly Ser Asp Lys Gly Val Ala Pro Gly	2470	2475
2465		
Thr Ala Val Leu Arg Gln Trp Leu Pro Thr Gly Thr Leu Leu Val	2485	2490
2480		
Asp Ser Asp Leu Asn Asp Phe Val Ser Asp Ala Asp Ser Thr Leu	2500	2505
2495		
Ile Gly Asp Cys Ala Thr Val His Thr Ala Asn Lys Trp Asp Leu	2515	2520
2510		
Ile Ile Ser Asp Met Tyr Asp Pro Arg Thr Lys His Val Thr Lys	2530	2535
2525		
Glu Asn Asp Ser Lys Glu Gly Phe Phe Thr Tyr Leu Cys Gly Phe	2545	2550
2540		
Ile Lys Gln Lys Leu Ala Leu Gly Gly Ser Ile Ala Val Lys Ile	2560	2565
2555		
Thr Glu His Ser Trp Asn Ala Asp Leu Tyr Lys Leu Met Gly His	2575	2580
2570		
Phe Ser Trp Trp Thr Ala Phe Val Thr Asn Val Asn Ala Ser Ser	2590	2595
2585		
Ser Glu Ala Phe Leu Ile Gly Ala Asn Tyr Leu Gly Lys Pro Lys	2605	2610
2600		
Glu Gln Ile Asp Gly Tyr Thr Met His Ala Asn Tyr Ile Phe Trp	2620	2625
2615		
Arg Asn Thr Asn Pro Ile Gln Leu Ser Ser Tyr Ser Leu Phe Asp	2635	2640
2630		
Met Ser Lys Phe Pro Leu Lys Leu Arg Gly Thr Ala Val Met Ser	2650	2655
2645		
Leu Lys Glu Asn Gln Ile Asn Asp Met Ile Tyr Ser Leu Leu Glu	2665	2670
2660		
Lys Gly Arg Leu Ile Ile Arg Glu Asn Asn Arg Val Val Val Ser	2680	2685
2675		
Ser Asp Ile Leu Val Asn Asn	2690	2695
2690		

<210> SEQ ID NO 76

<211> LENGTH: 20

<212> TYPE: DNA

<213> ORGANISM: Artificial sequence

<220> FEATURE:

<223> OTHER INFORMATION: S/L3/+/4932 primer

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<400> SEQUENCE: 76

ccacacacag cttgtggata

20

<210> SEQ ID NO 77

<211> LENGTH: 20

<212> TYPE: DNA

<213> ORGANISM: Artificial sequence

<220> FEATURE:

<223> OTHER INFORMATION: S/L4/+ /6401 primer

<400> SEQUENCE: 77

ccgaagtgtg aggcaatgtc

20

<210> SEQ ID NO 78

<211> LENGTH: 20

<212> TYPE: DNA

<213> ORGANISM: Artificial sequence

<220> FEATURE:

<223> OTHER INFORMATION: S/L4/+ /6964 primer

<400> SEQUENCE: 78

tttggtgctc cttcttattg

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<210> SEQ ID NO 79

<211> LENGTH: 20

<212> TYPE: DNA

<213> ORGANISM: Artificial sequence

<220> FEATURE:

<223> OTHER INFORMATION: S/L4/- /6817 primer

<400> SEQUENCE: 79

ccggcatcca aacataattt

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<210> SEQ ID NO 80

<211> LENGTH: 20

<212> TYPE: DNA

<213> ORGANISM: Artificial sequence

<220> FEATURE:

<223> OTHER INFORMATION: S/L5/- /7633 primer

<400> SEQUENCE: 80

tggtcagtag ggttgattgg

20

<210> SEQ ID NO 81

<211> LENGTH: 20

<212> TYPE: DNA

<213> ORGANISM: Artificial sequence

<220> FEATURE:

<223> OTHER INFORMATION: S/L5/- /8127 primer

<400> SEQUENCE: 81

catcctttgt gtcaacatcg

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<210> SEQ ID NO 82

<211> LENGTH: 20

<212> TYPE: DNA

<213> ORGANISM: Artificial sequence

<220> FEATURE:

<223> OTHER INFORMATION: S/L5/- /8633 primer

<400> SEQUENCE: 82

gtcacgagtg acaccatcct

20

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<210> SEQ ID NO 83
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L5/+7839 primer

<400> SEQUENCE: 83

atgcgacgag tctgcttcta 20

<210> SEQ ID NO 84
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L5/+8785 primer

<400> SEQUENCE: 84

ttcatagtgc ctggcttacc 20

<210> SEQ ID NO 85
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L5/+8255 primer

<400> SEQUENCE: 85

atcttggcgc atgtattgac 20

<210> SEQ ID NO 86
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L6/-9422 primer

<400> SEQUENCE: 86

tgcattagca gcaacaacat 20

<210> SEQ ID NO 87
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L6/-9966 primer

<400> SEQUENCE: 87

tctgcagaac agcagaagtg 20

<210> SEQ ID NO 88
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L6/-10542 primer

<400> SEQUENCE: 88

cctgtgcagt ttgtctgtca 20

<210> SEQ ID NO 89
<211> LENGTH: 20
<212> TYPE: DNA

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<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L6/+/10677 primer

<400> SEQUENCE: 89
ccttgtggca atgaagtaca 20

<210> SEQ ID NO 90
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L6/+/10106 primer

<400> SEQUENCE: 90
atgtcatttg cacagcagaa 20

<210> SEQ ID NO 91
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L6/+/9571 primer

<400> SEQUENCE: 91
cttcaatggt ttgccatgtt 20

<210> SEQ ID NO 92
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L7/-/11271 primer

<400> SEQUENCE: 92
tgcgagctgt catgagaata 20

<210> SEQ ID NO 93
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L7/-/11801 primer

<400> SEQUENCE: 93
aaccgagagc agtaccacag 20

<210> SEQ ID NO 94
<211> LENGTH: 20
<212> TYPE: DNA
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<210> SEQ ID NO 95
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<212> TYPE: DNA
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<223> OTHER INFORMATION: S/L7/+/12640 primer

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<210> SEQ ID NO 96

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<220> FEATURE:

<223> OTHER INFORMATION: S/L7/+12088 primer

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<210> SEQ ID NO 97

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<212> TYPE: DNA

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<220> FEATURE:

<223> OTHER INFORMATION: S/L7/+11551 primer

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<210> SEQ ID NO 98

<211> LENGTH: 20

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<212> TYPE: DNA

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<210> SEQ ID NO 123
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<223> OTHER INFORMATION: SARS/L3/F5/+ /5822 primer

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<213> ORGANISM: Artificial sequence

<220> FEATURE:

<223> OTHER INFORMATION: SARS/L3/R4/- /4988 primer

<400> SEQUENCE: 138

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<223> OTHER INFORMATION: SARS/L3/R5/- /4437 primer

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tcggtcgccg catacactat tctcagaatg acttggttga gtactacca gtcacagaaa	6420
agcatcttac ggatggcatg acagtaagag aattatgcag tgctgccata accatgagtg	6480
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ttttgcacaa catgggggat catgtaactc gccttgatcg ttgggaaccg gagctgaatg 6600
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<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
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<210> SEQ ID NO 142
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<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
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<223> OTHER INFORMATION: SNE-AS1 primer

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<400> SEQUENCE: 145

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 <212> TYPE: DNA
 <213> ORGANISM: Artificial sequence
 <220> FEATURE:
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<400> SEQUENCE: 146

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<210> SEQ ID NO 147
 <211> LENGTH: 45
 <212> TYPE: DNA
 <213> ORGANISM: Artificial sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: PCR primer

<400> SEQUENCE: 147

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<210> SEQ ID NO 148
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 <212> TYPE: DNA
 <213> ORGANISM: Artificial sequence
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<400> SEQUENCE: 148

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<210> SEQ ID NO 149
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 <212> TYPE: PRT
 <213> ORGANISM: Artificial sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: N-terminal end of SRAS-CoV S protein (amino-acids 1 to 13)

<400> SEQUENCE: 149

Met Phe Ile Phe Leu Leu Phe Leu Thr Leu Thr Ser Gly
 1 5 10

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<210> SEQ ID NO 150
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<212> TYPE: PRT
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<220> FEATURE:
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<400> SEQUENCE: 150

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1 5 10

<210> SEQ ID NO 151
<211> LENGTH: 34
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<220> FEATURE:
<223> OTHER INFORMATION: PCR primer

<400> SEQUENCE: 151

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<210> SEQ ID NO 152
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<220> FEATURE:
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<400> SEQUENCE: 152

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<400> SEQUENCE: 155

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<210> SEQ ID NO 156
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<212> TYPE: DNA
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<220> FEATURE:
<223> OTHER INFORMATION: PCR primer
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<400> SEQUENCE: 156
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40

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<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
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<400> SEQUENCE: 157
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ccatttcaac aatttgcccg
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20

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<210> SEQ ID NO 158
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<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
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<223> OTHER INFORMATION: PCR primer
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<400> SEQUENCE: 158
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1. An isolated or purified strain of severe acute respiratory syndrome-associated human coronavirus, characterized in that its genome has, in the form of complementary DNA, a serine codon at position 23220-23222 of the gene for the S protein or a glycine codon at position 25298-25300 of the gene for ORF3, and an alanine codon at position 7918-7920 of ORF1a or a serine codon at position 26857-26859 of the gene for the M protein, said positions being indicated in terms of reference to the Genbank sequence AY274119.3.

2. The isolated or purified coronavirus strain as claimed in claim 1, characterized in that the DNA equivalent of its genome has a sequence corresponding to the sequence SEQ ID NO: 1.

3. An isolated or purified polynucleotide, characterized in that its sequence is that of the genome of the isolated coronavirus strain as claimed in claim 1 or claim 2.

4. The isolated or purified polynucleotide as claimed in claim 3, characterized in that its sequence is SEQ ID NO: 1.

5. A pair of primers capable of amplifying a fragment of the sequence of the genome of a SARS-associated coronavirus or of its DNA equivalent, characterized in that it is selected from the group consisting of:

the pair of primers No. 1 corresponding respectively to positions 28507 to 28522 (sense primer, SEQ ID NO: 60) and 28774 to 28759 (antisense primer, SEQ ID NO: 61) of the sequence of the polynucleotide as claimed in claim 3 or claim 4,

the pair of primers No. 2 corresponding respectively to positions 28375 to 28390 (sense primer, SEQ ID NO: 62) and 28702 to 28687 (antisense primer, SEQ ID NO: 63) of the sequence of the polynucleotide as claimed in claim 3 or claim 4, and

the pair of primers consisting of the primers SEQ ID Nos: 55 and 56.

6. A probe capable of detecting the presence of the genome of a SARS-associated coronavirus or of a fragment thereof, characterized in that it is selected from the group consisting of the fragments corresponding to the following positions of the polynucleotide sequence as claimed in claim 3 or claim 4: 28561 to 28586, 28588 to 28608, 28541 to 28563 and 28565 to 28589 (SEQ ID NO: 64 to 67).

7. A recombinant cloning and/or expression vector, characterized in that it comprises an insert having the sequence SEQ ID NO: 38 and it is contained in a bacterial strain and it was deposited under the No. I-3048, on Jun. 5, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15.

8. A recombinant cloning and/or expression vector, characterized in that it contains a cDNA fragment selected from the group consisting of:

a cDNA fragment encoding a C-terminal fusion of the N protein (SEQ ID NO: 37) with a polyhistidine tag, and

a cDNA fragment encoding an N-terminal fusion of the N protein (SEQ ID NO: 37) with a polyhistidine tag.

9. The recombinant expression vector as claimed in claim 8, characterized in that it is contained in a bacterial strain which was deposited under the No. I-3117, on Oct. 23, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15.

10. A cell modified with a vector as claimed in any one of claims 7 to 9.

11. A hybridoma producing a monoclonal antibody against the N protein, characterized in that it is chosen from the following hybridomas:

the hybridoma producing the monoclonal antibody 87, deposited at the CNCM on Dec. 1, 2004 under the number I-3328,

the hybridoma producing the monoclonal antibody 86, deposited at the CNCM on Dec. 1, 2004 under the number I-3329,

the hybridoma producing the monoclonal antibody 57, deposited at the CNCM on Dec. 1, 2004 under the number I-3330, and

the hybridoma producing the monoclonal antibody 156, deposited at the CNCM on Dec. 1, 2004 under the number I-3331.

12. A polyclonal or monoclonal antibody or antibody fragment directed against the N protein, characterized in that it is produced by a hybridoma as claimed in claim 11.

13. A chip or filter, characterized in that it comprises an antibody or an antibody fragment as claimed in claim 12.

14. An immunocapture test intended to detect a SARS-associated coronavirus infection, characterized in that it uses a monoclonal antibody specific for the native viral nucleoprotein (N protein).

15. The immunocapture test as claimed in claim 14, characterized in that the antibody used for the capture of the native viral nucleoprotein is a monoclonal antibody specific for the central region and/or for a conformational epitope.

16. The immunocapture test as claimed in claim 14 or 15, characterized in that the antibody used for the capture of the N protein is the monoclonal antibody mAb87, produced by the hybridoma deposited at the CNCM on Dec. 1, 2004 under the number I-3328.

17. The immunocapture test as claimed in claim 14 or 15, characterized in that the antibody used for the capture of the N protein is the monoclonal antibody mAb86, produced by the hybridoma deposited at the CNCM on Dec. 1, 2004 under the number I-3329.

18. The immunocapture test as claimed in claim 14 or 15, characterized in that the monoclonal antibodies mAb86 and mAb87 are used for the capture of the N protein.

19. The immunocapture test as claimed in any one of claims 14 to 18, characterized in that the antibody used for the visualization of the N protein is the monoclonal antibody mAb57, produced by the hybridoma deposited at the CNCM on Dec. 1, 2004 under the number I-3330, said antibody being conjugated with a visualizing molecule or particle.

20. The immunocapture test as claimed in any one of claims 14 to 18, characterized in that a combination of the mAb57 and mAb87 antibodies, conjugated with a visualizing molecule or particle, is used for the visualization of the N protein.

21. A reagent for the detection of a SARS-associated coronavirus, characterized in that it is selected from the group consisting of:

- (a) a pair of primers as claimed in claim 5, or a probe as claimed in claim 6,
- (b) a recombinant vector as claimed in any one of claims 7 to 9 or a modified cell as claimed in claim 10,
- (c) an isolated coronavirus strain as claimed in claim 1 or claim 2 or a polynucleotide as claimed in either of claims 3 and 4,
- (d) an antibody or an antibody fragment as claimed in claim 12,

(e) a combination of antibodies comprising the monoclonal antibodies mAb86 and/or mAb87, and the monoclonal antibody mAb57;

(f) a chip or a filter as claimed in claim 13.

22. The use of a product selected from the group consisting of: a pair of primers as claimed in claim 5, a probe as claimed in claim 6, a recombinant vector as claimed in any one of claims 7 to 9, a modified cell as claimed in claim 10, an isolated coronavirus strain as claimed in claim 1 or claim 2, a polynucleotide as claimed in claim 3 or claim 4, for the preparation of a reagent for the detection and optionally genotyping of a SARS-associated coronavirus.

23. A method for the detection of a SARS-associated coronavirus, from a biological sample, which method is characterized in that it comprises at least:

- (a) the extraction of nucleic acids present in said biological sample,
- (b) the amplification of a fragment of ORF-N by RT-PCR with the aid of a pair of primers as claimed in claim 5, and
- (c) the detection, by any appropriate means, of the amplification products obtained in (b).

24. The method as claimed in claim 23, characterized in that step (b) of detection is carried out with the aid of at least one probe corresponding to positions 28561 to 28586, 28588 to 28608, 28541 to 28563 and 28565 to 28589 of the sequence of the polynucleotide as claimed in claim 3 or claim 4.

25. A method for the detection of a SARS-associated coronavirus infection, from a biological sample, by indirect IgG ELISA using the N protein, which method is characterized in that the plates are sensitized with an N protein solution at a concentration of between 0.5 and 4 µg/ml, preferably 2 µg/ml, in a 10 mM PBS buffer, pH 7.2, phenol red at 0.25 ml/l.

26. A method for the detection of a SARS-associated coronavirus infection, from a biological sample, by double epitope ELISA, characterized in that the serum to be tested is mixed with the visualizing antigen, said mixture then being brought into contact with the antigen attached to a solid support.

27. An immune complex formed of a polyclonal or monoclonal antibody or antibody fragment as claimed in claim 11, and of a SARS-associated coronavirus protein or peptide.

28. A SARS-associated coronavirus detection kit or box, characterized in that it comprises at least one reagent selected from the group consisting of: a pair of primers as claimed in claim 5, a probe as claimed in claim 6, a recombinant vector as claimed in any one of claims 7 to 9, a modified cell as claimed in claim 10, an isolated coronavirus strain as claimed in claim 1 or claim 2 and a polynucleotide as claimed in claim 3 or claim 4.

29. A fragment of the polynucleotide as claimed in claim 3, characterized in that it includes at least one pair of bases or pairs of bases corresponding to the following positions: 7919 and 23220, 7919 and 25298, 16622 and 23220, 19064 and 23220, 16622 and 25298, 19064 and 25298, 23220 and 24872, 23220 and 26857, 24872 and 25298, 25298 and 26857.