



US008343718B2

(12) **United States Patent**
Van Der Werf et al.

(10) **Patent No.:** **US 8,343,718 B2**
(45) **Date of Patent:** **Jan. 1, 2013**

(54) **STRAIN OF SARS-ASSOCIATED
CORONAVIRUS AND APPLICATIONS
THEREOF**

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(*) Notice: Subject to any disclaimer, the term of this patent is extended or adjusted under 35 U.S.C. 154(b) by 0 days.

(21) Appl. No.: **12/754,908**

(22) Filed: **Apr. 6, 2010**

(65) **Prior Publication Data**

US 2011/0065089 A1 Mar. 17, 2011

Related U.S. Application Data

(60) Division of application No. 10/581,356, filed on Feb. 8, 2007, now Pat. No. 7,736,850, which is a continuation of application No. PCT/FR2004/003106, filed on Dec. 2, 2004.

(30) **Foreign Application Priority Data**

Dec. 2, 2003 (FR) 03 14151
Dec. 2, 2003 (FR) 03 14152

(51) **Int. Cl.**

C12Q 1/70 (2006.01)
G01N 33/53 (2006.01)
G01N 33/542 (2006.01)
G01N 33/00 (2006.01)

(52) **U.S. Cl.** **435/5; 435/7.1; 435/7.9; 435/7.92; 435/7.94; 435/7.95**

(58) **Field of Classification Search** None
See application file for complete search history.

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Primary Examiner — Louise Humphrey

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(57) **ABSTRACT**

The invention relates to a novel strain of severe acute respiratory syndrome (SARS)-associated coronavirus, resulting from a sample collected in Hanoi (Vietnam), reference number 031589, nucleic acid molecules originating from the genome of same, proteins and peptides coded by said nucleic acid molecules and, more specifically, protein N and the applications thereof, for example, as diagnostic reagents and/or as a vaccine.

8 Claims, 116 Drawing Sheets

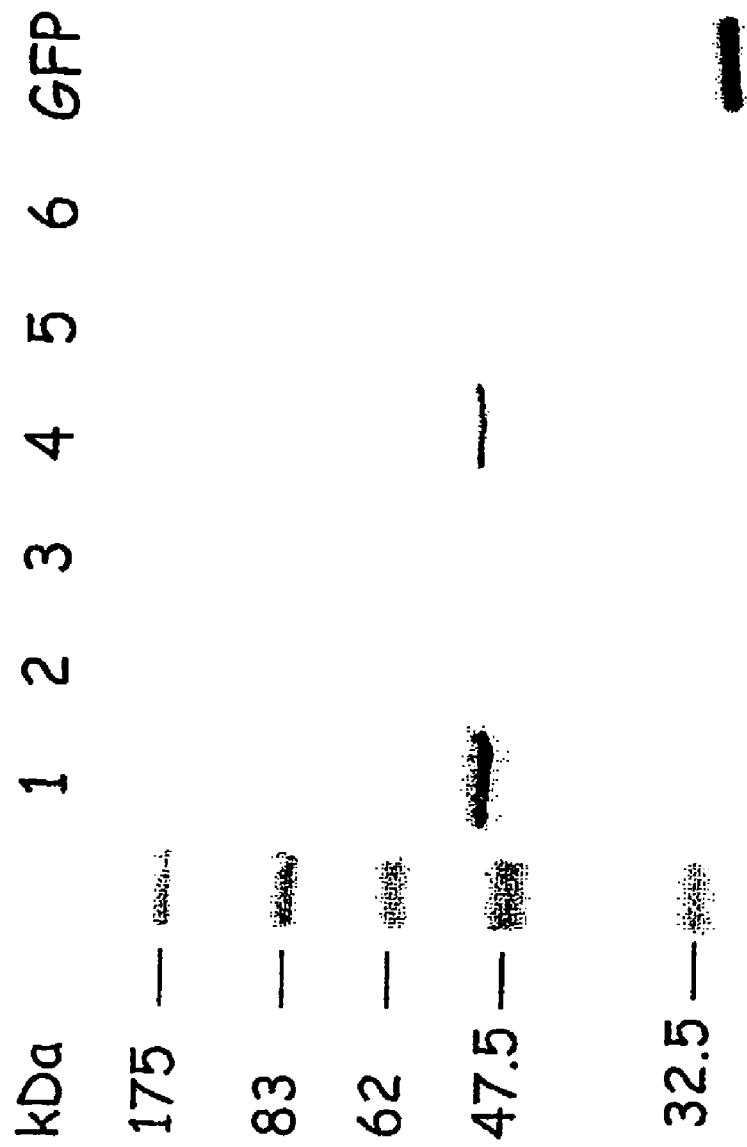


FIGURE 1

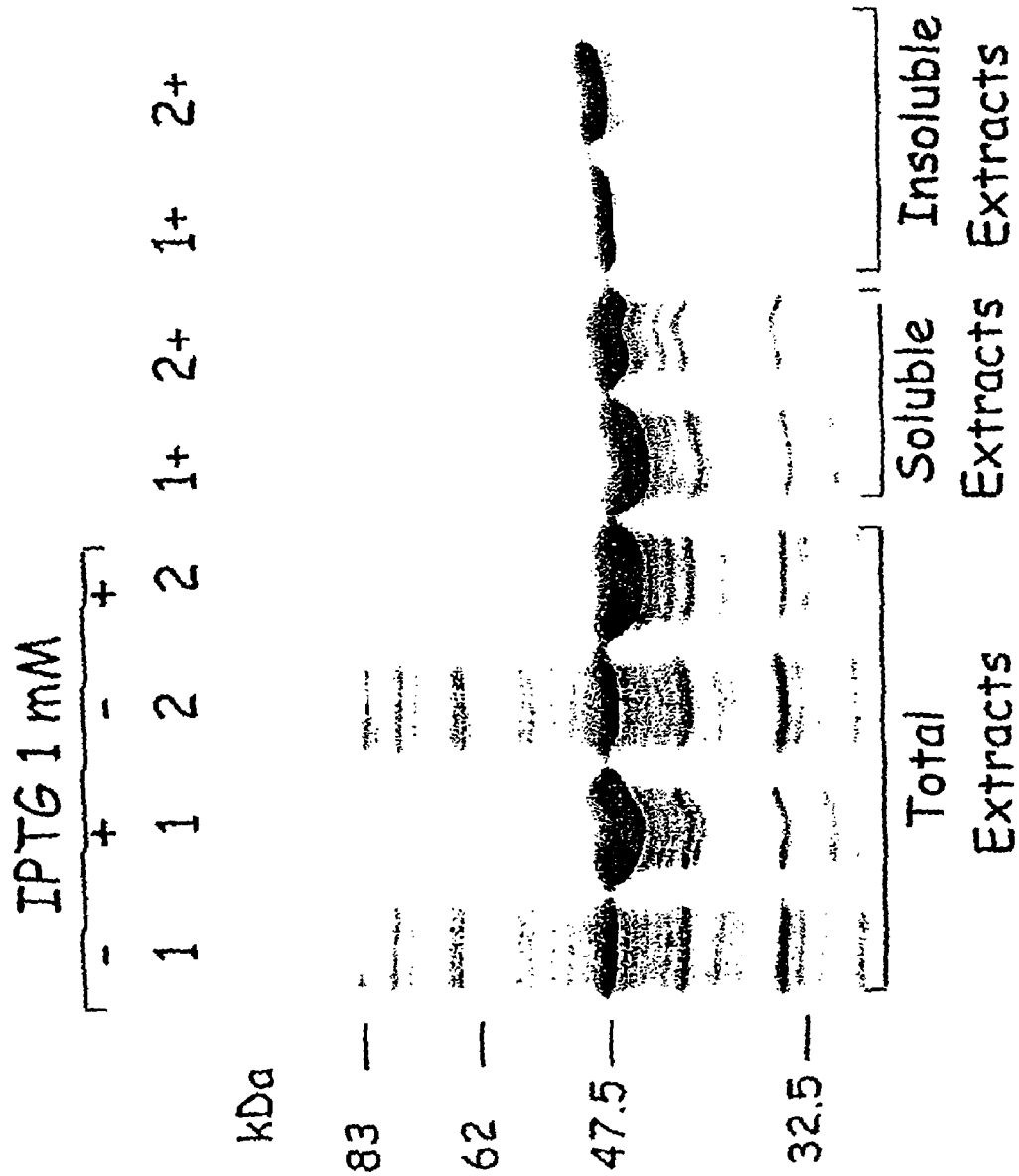


FIGURE 2

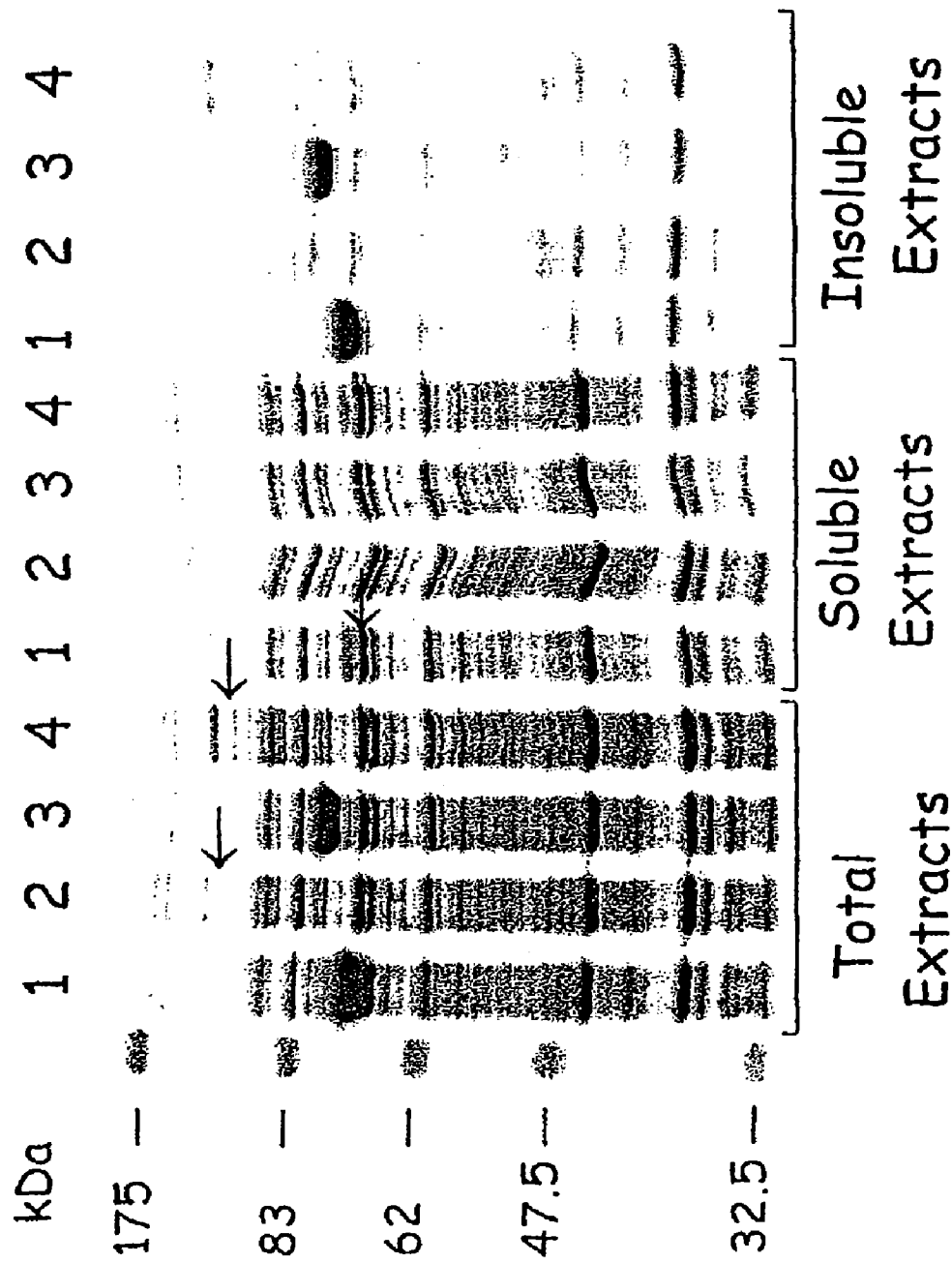


FIGURE 3

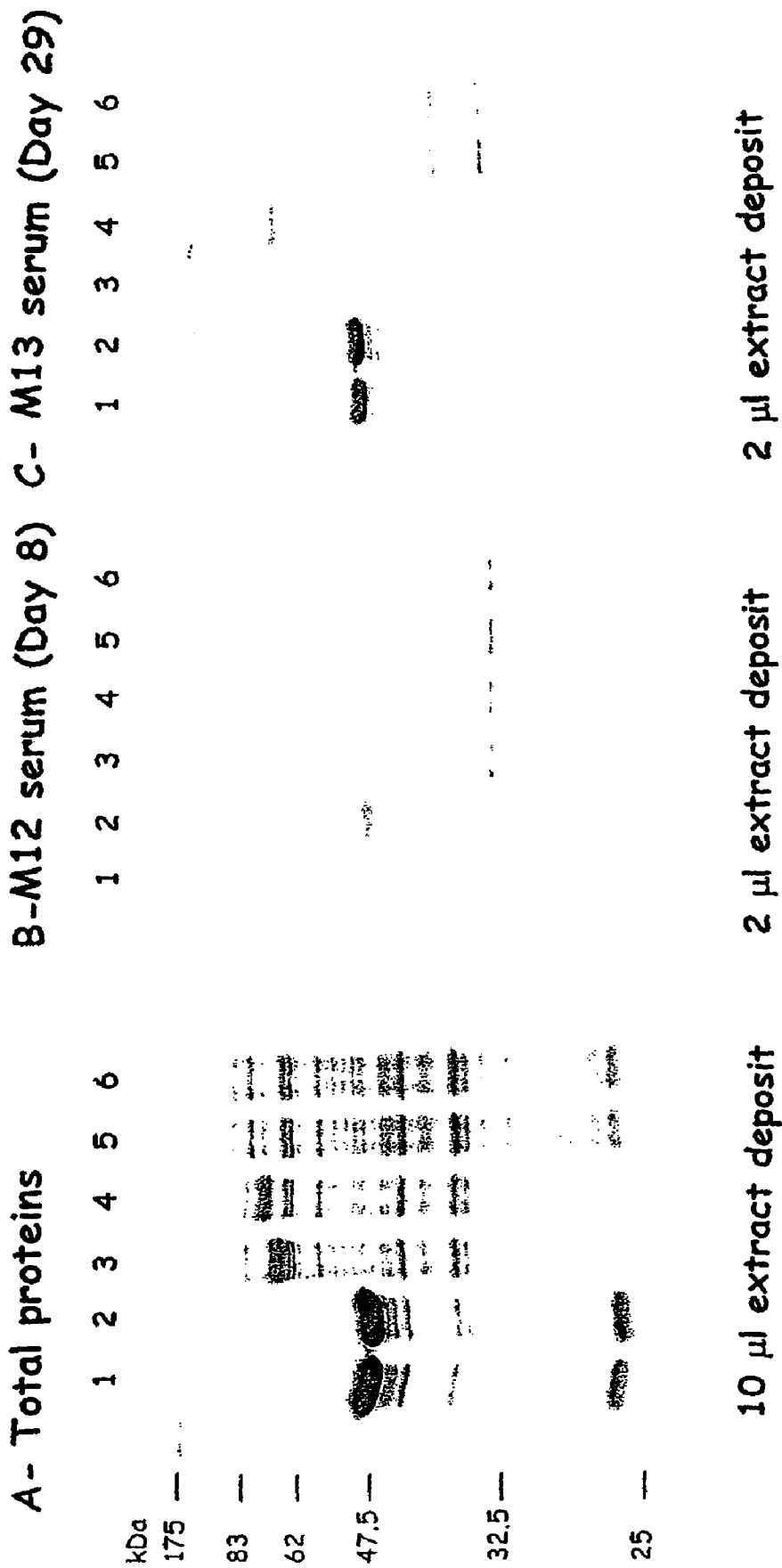


FIGURE 4

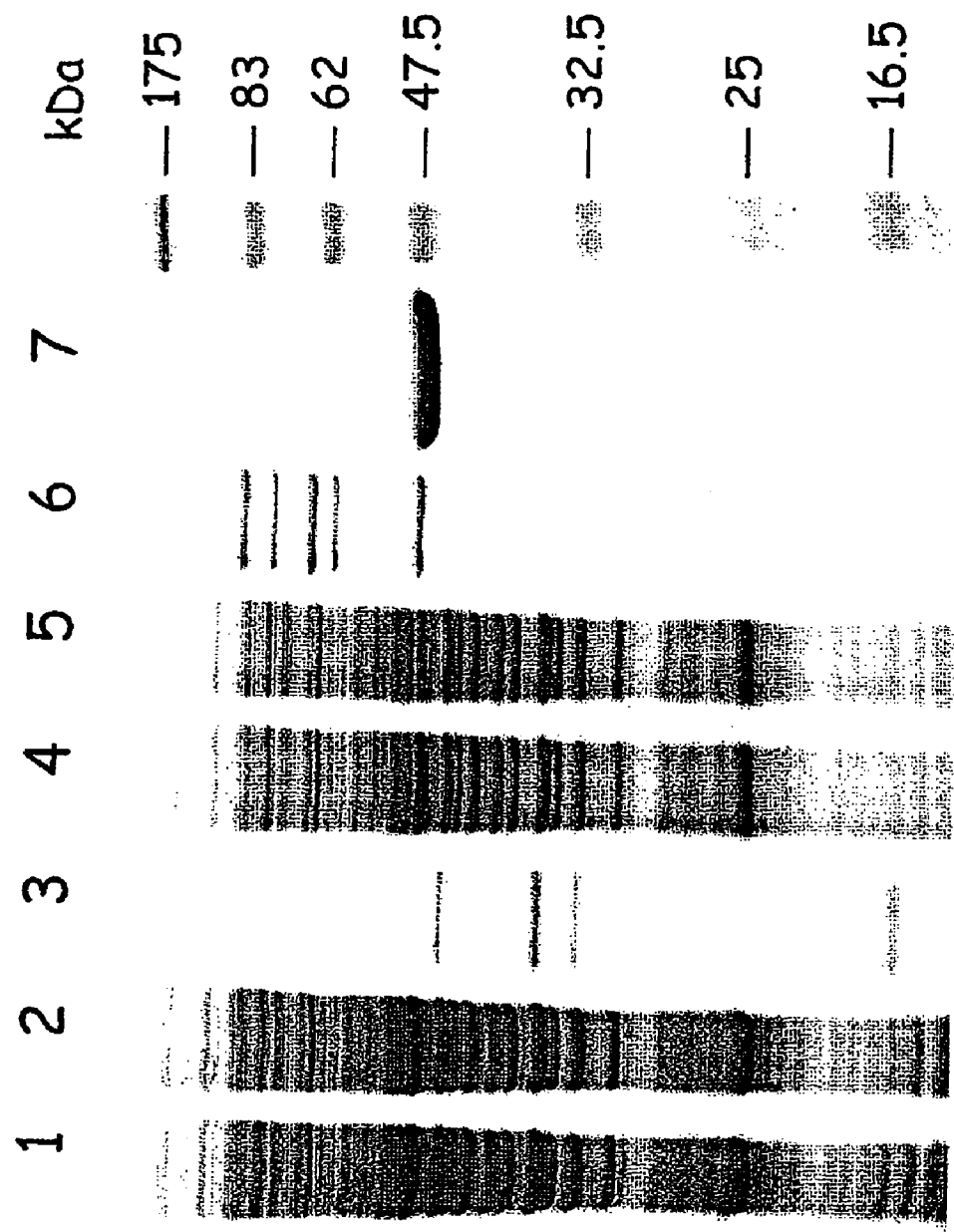


FIGURE 5

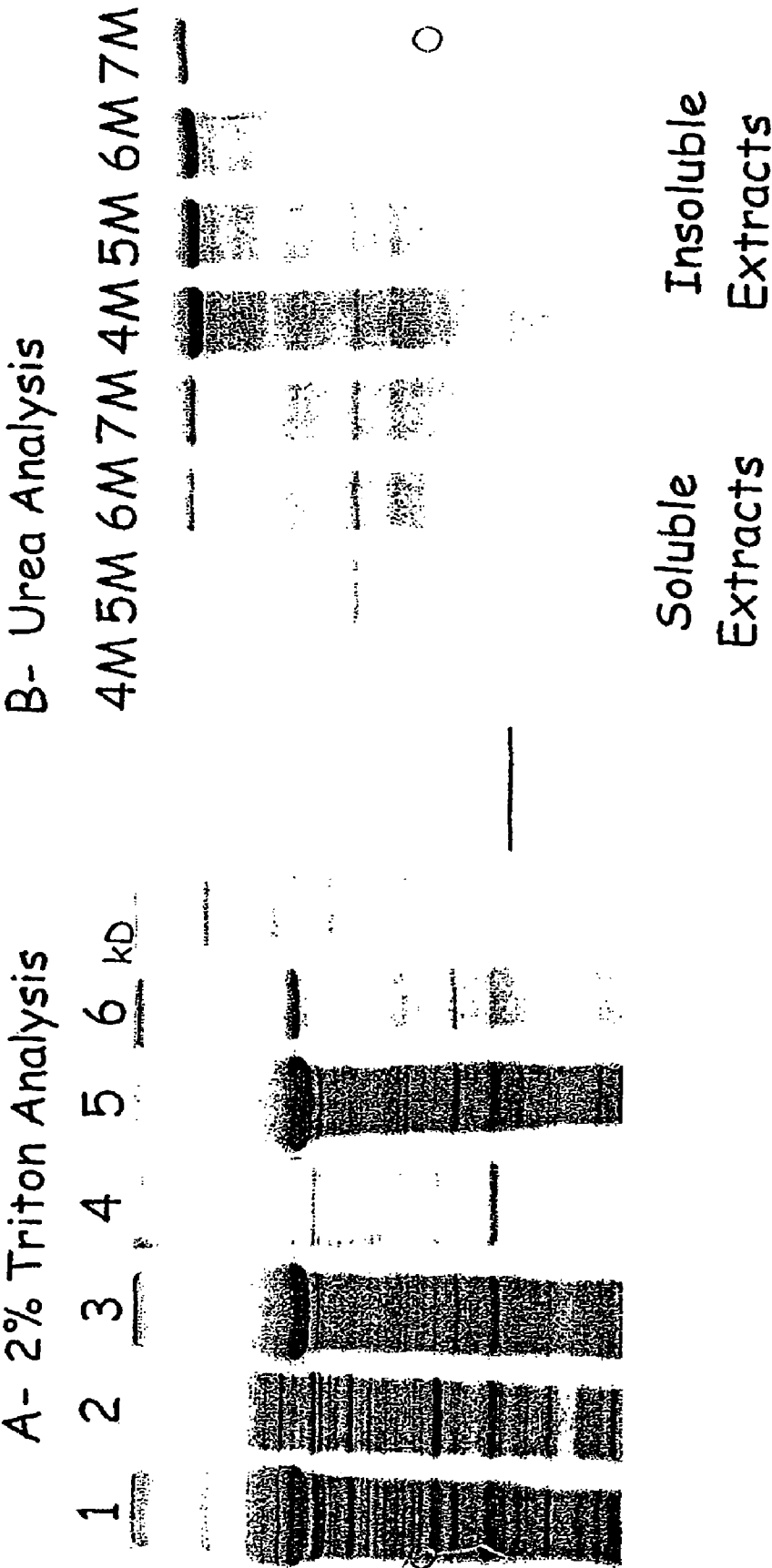


FIGURE 6

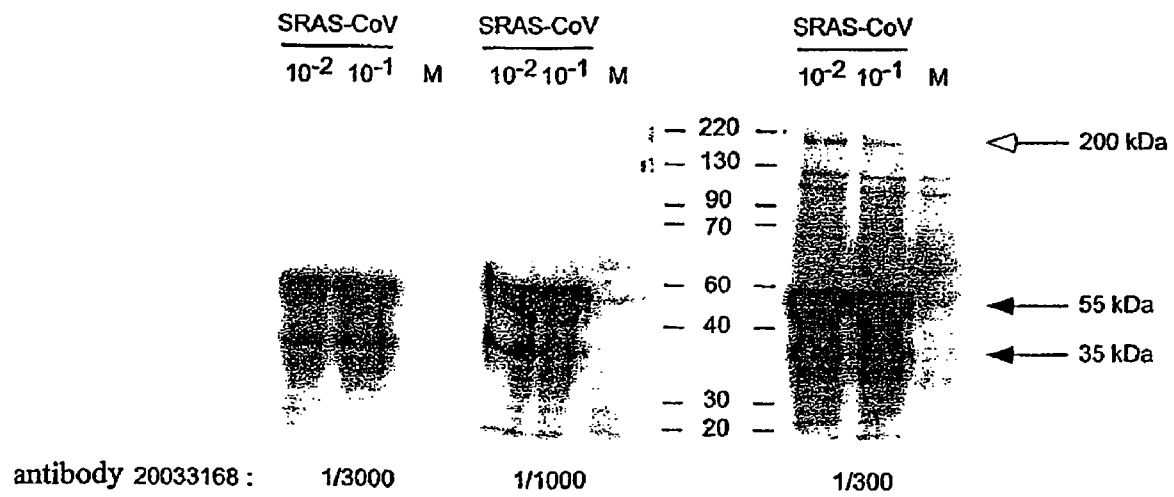


FIGURE 7

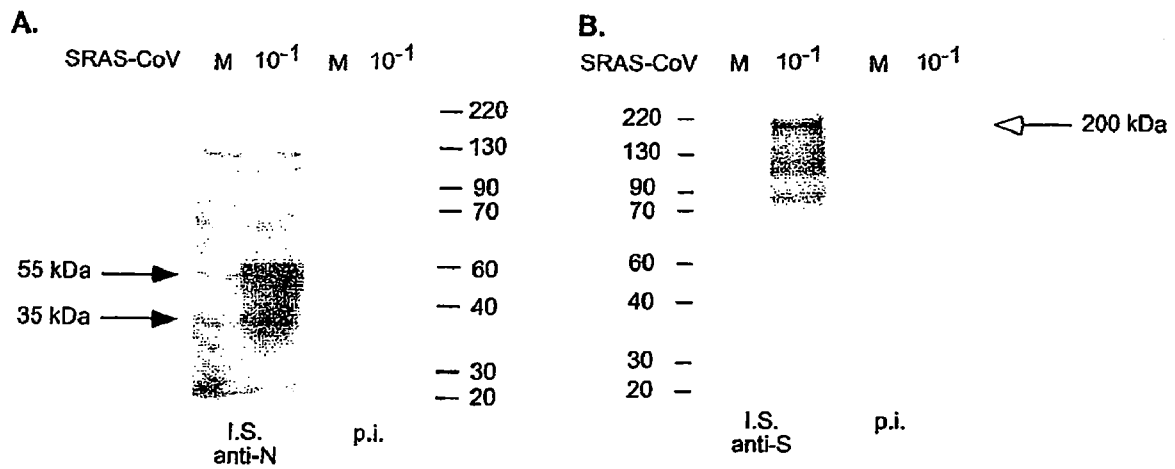
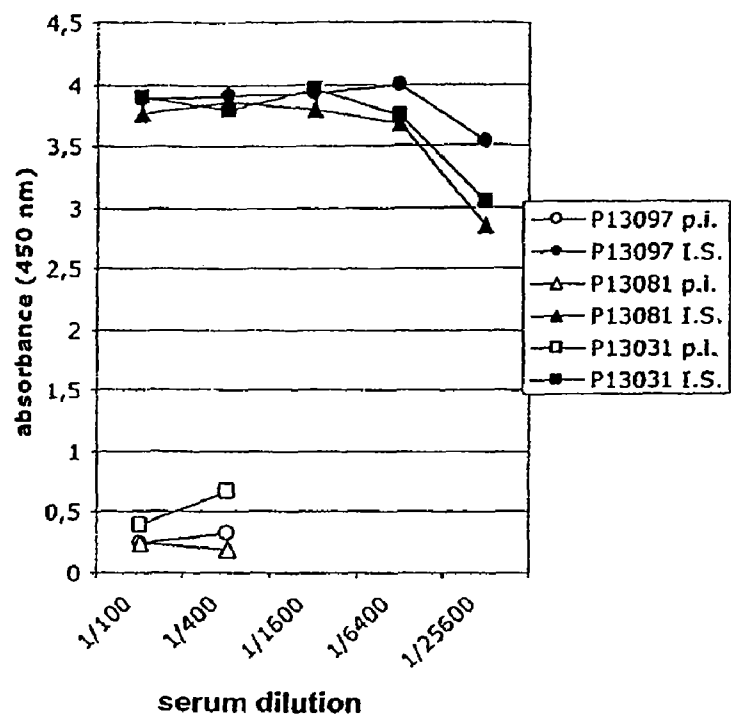


FIGURE 8

A



B

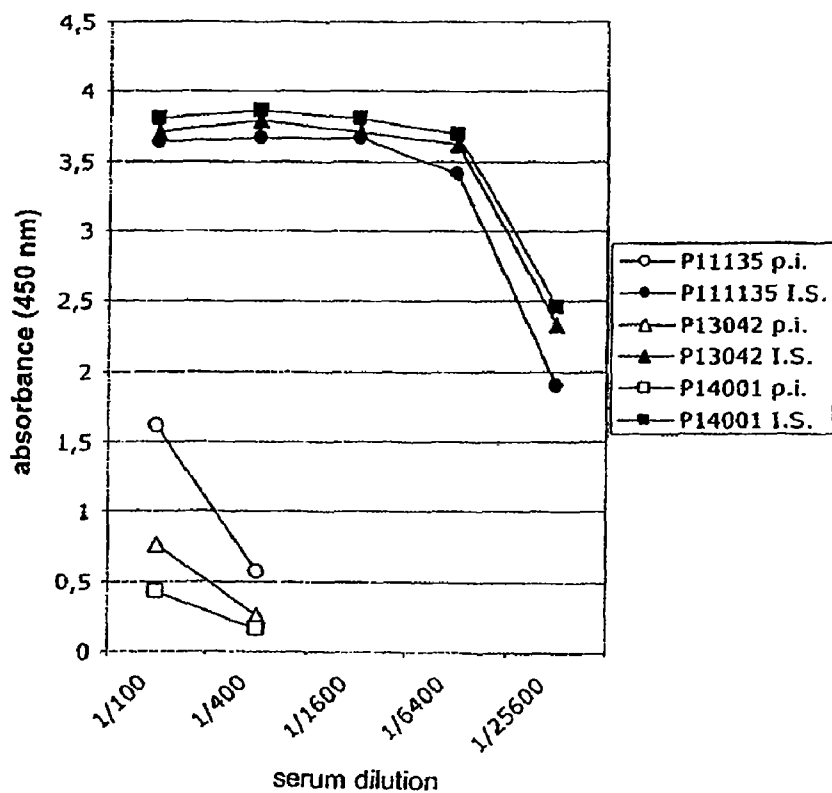


FIGURE 9

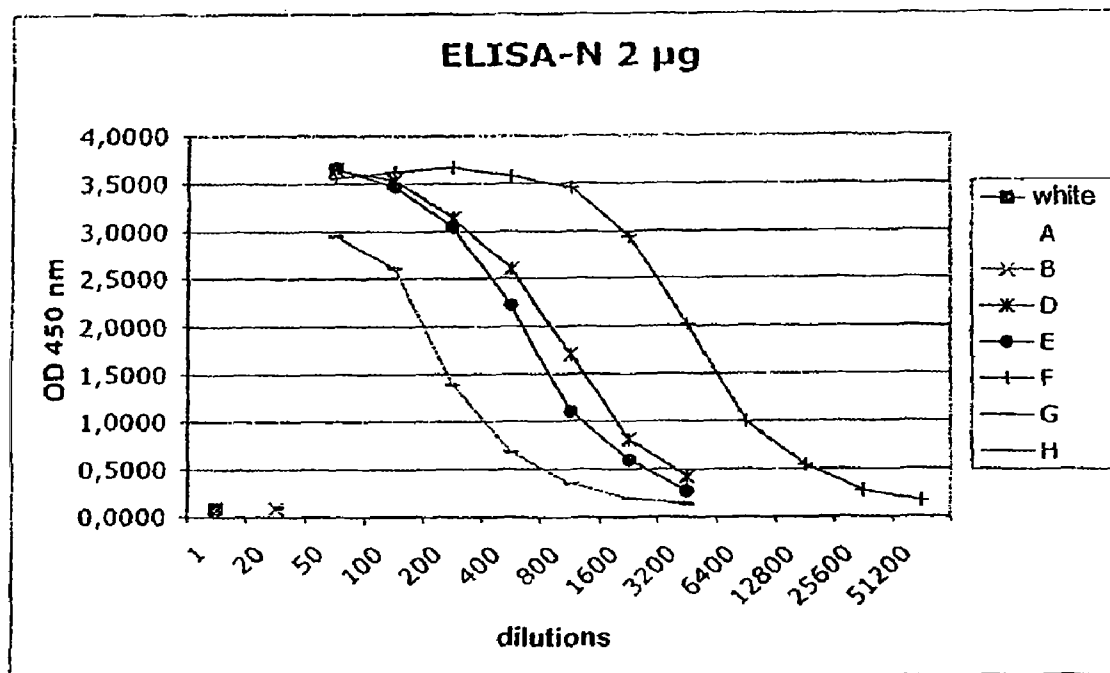
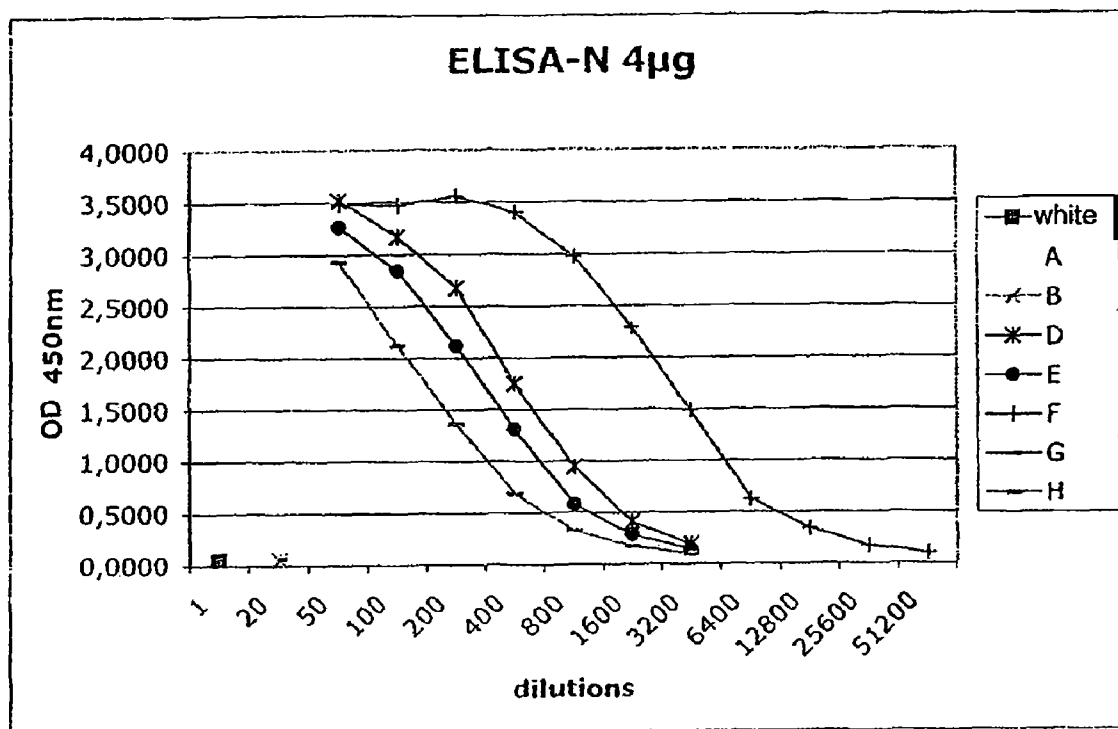


FIGURE 10a

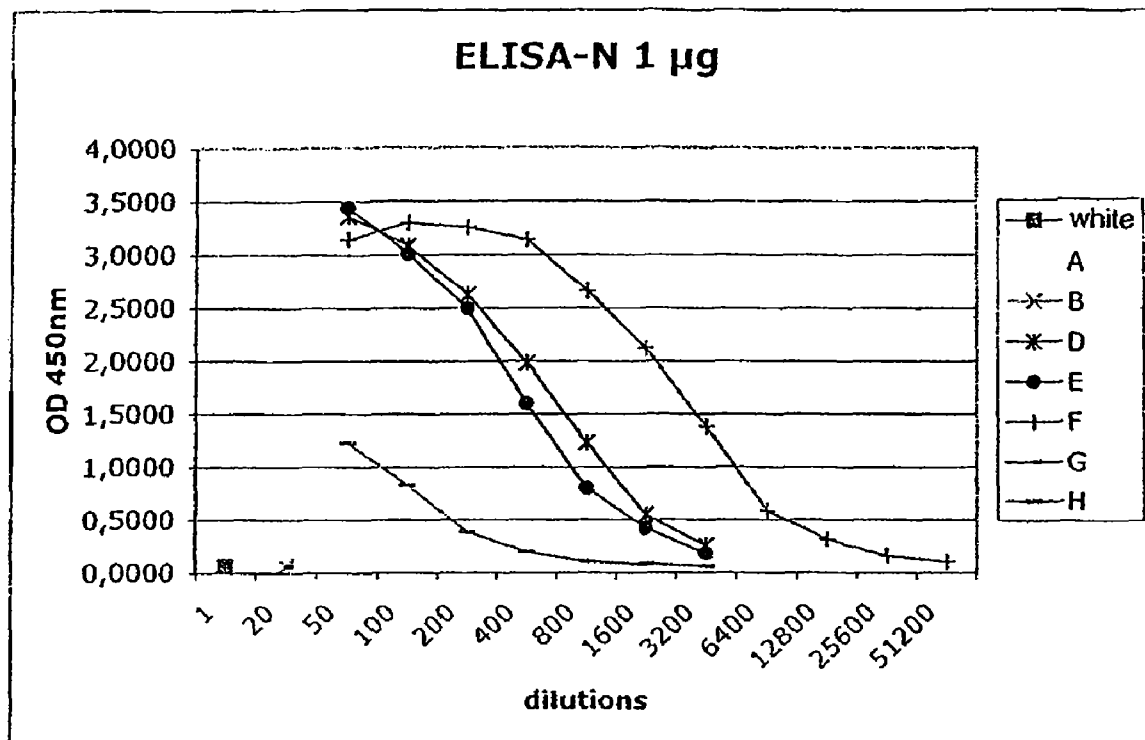


FIGURE 10b

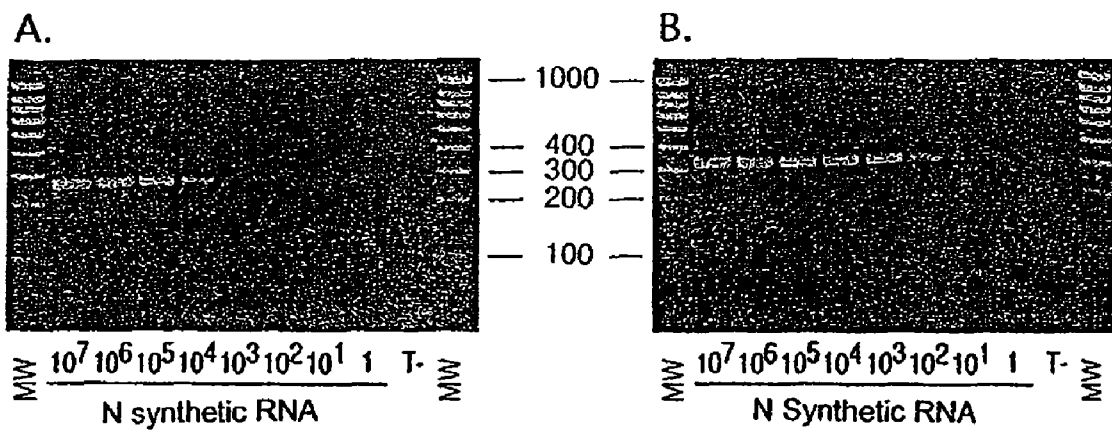


FIGURE 11

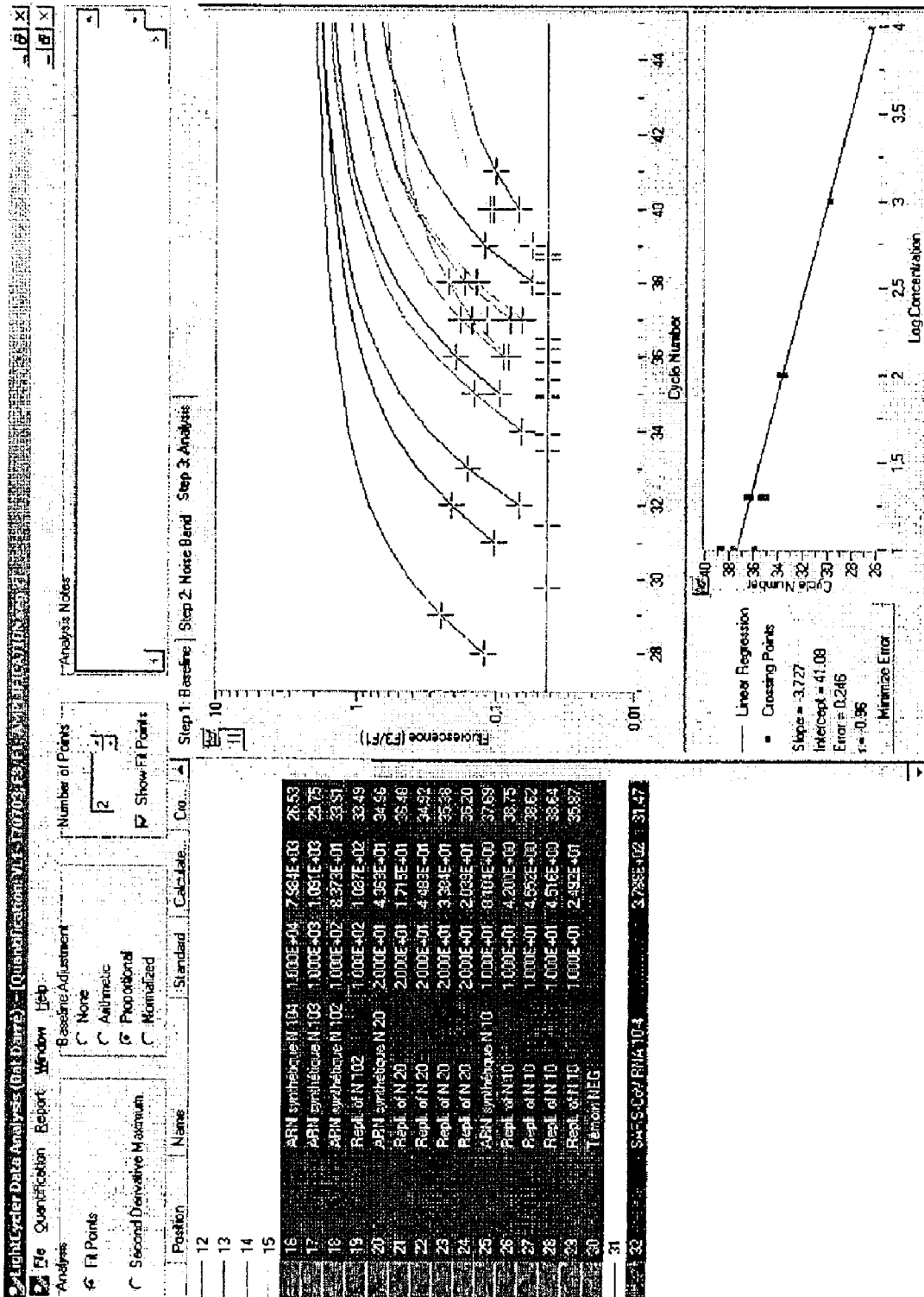


FIGURE 12

FIGURE 13.1

```

                >< Sau96I
                >< PssI
                >< Pali
                >< NspIV
                >< MnlI
                >< HaeIII
                >< EcoO109I
                >< DraII>< MboII >< PmlI
                >< MnlI >< CfrI3I >< PmaCI
                >< Ksp632I >< BsuRI >< MaeII
                >< HinfI >< BsiZI>< EcoNI >< Eco72I
                >< EarI >< BshI >< BslI >< BsaAI
                >< PleI >< Eam1104I>< AsuI >< BsiYI>< BbrPI >< MnlI
TGGCTTCGGG GACTCTGTGG AAGAGGCCCT ATCGGAGGCA CGTGAACACC TCAAAAATGG CACTTGTGGT
    360          370          380          390          400          410          420

                >< Tru9I
                >< SfaNI
                >< RsaI
                >< Csp6I >< BspWI >< MseI
                >< RmaI >< AluI >< AfaI >< AluI >< MaeII
CTAGTAGAGC TGGAAAAAGG CGTACTGCCC CAGCTTGAAC AGCCCTATGT GTTCATTAAA CGTTCTGATG
    430          440          450          460          470          480          490

                >< Pali
                >< HaeIII
                >< Tru9I >< GdiII >< RsaI
                >< MseI >< EaeI >< McrI ><
                >< Esp4I >< BsuRI >< BsmI BsiEI ><
                >< AflII >< BshI >< AluI >< BscCI >< AfaI
CCTTAAGCAC CAATCACGGC CACAAGGTCG TTGAGCTGGT TGCAGAAATG GACGGCATTG AGTACGGTCG
    500          510          520          530          540          550          560

                >< NspI
                >< ScaI >< NspHI
                >< RsaI >< NlaIII
                >< Csp6I >< BslI
                >< BsrI >< BsiYI >< MboII
                >< MboII
                >< AciI >< AfaI >< AflIII >< MunI >< AciI
TAGCGGTATA ACGGTAAATA TACTCGTGCC ACATGTGGGC GAAACCCCAA TTGCATACCG CAATGTTCTT
    570          580          590          600          610          620          630

                >< TthHB8I
                >< TaqI
                >< Sau3AI
                >< NdeII
                >< MboI
                >< DpnII
                >< DpnI
                >< ClaI
                >< Bsu15I
                >< BspDI
                >< BspAI
                >< NlaIV
                >< MspI
                >< HpaII
                >< HapII
                >< CfrI10I
                >< BscBI >< AluI >< BanIII >< BfrI ><
                >< Bsp143I
                >< Bsp106I
                >< BsiXI >< MaeIII >
                >< BscI>< SfaNI DdeI ><
                >< BscBI >< AluI >< BanIII >< BfrI ><
CTTCGTAAGA ACGGTAATAA GGGAGCCGGT GGTTCATAGCT ATGGCATCGA TCTAAAGTCT TATGACTTAG
    640          650          660          670          680          690          700

```

FIGURE 13.2

```

    >> Sau3AI
    >> NdeII
    >> MboI
    >> HphI
    >> DpnII
    >> BspAI
    >> AlwI>> DpnI
    >> AluI    >> Bsp143I
    >> MboII  >> BsrI
    >> DdeI   >> ApaLI
    >> Alw44I
    GTGACGAGCT TGGCACTGAT CCCATTGAAG ATTATGAACA AACTTGGAAC ACTAAGCATG GCAGTGGTGC
    710         720         730         740         750         760         770

    >> SstI
    >> SduI
    >> SacI
    >> NspII
    >> MnlI
    >> HgiAI
    >> Eco24I
    >> TthHB8I
    >> PalI
    >> SduI
    >> NspII
    >> HgiAI
    >> Eco24I
    >> Ecl136II
    >> Bsp1286I
    >> BmyI
    >> BanII
    >> Alw21I
    >> AluI
    >> MaeIII
    >> AccI
    >> Sau96I
    >> PalI
    >> NspIV
    >> HaeIII
    >> RtrI
    >> Cfr13I
    >> HindII
    >> BsuRI
    >> HincII
    >> Bsi2I
    >> BsgI
    >> BshI
    >> AsuI
    ACTCCGTGAA CTCACCTCGTG AGCTCAATGG AGGTGCAGTC ACTCGCTATG TCGACAACAA TTTCTGTGGC
    780         790         800         810         820         830         840

    >> ThaI
    >> ThaI
    >> MvnI
    >> MvnI
    >> HinPII
    >> Hin6I
    >> HhaI
    >> CfoI
    >> BstUI
    >> BstUI
    >> Bsp50I
    >> Bsp50I
    >> AciI
    >> AccII
    >> AccII
    >> MnlI
    >> SfaNI
    >> AccII
    >> Alw21I
    >> VneI
    >> SnoI
    >> SduI
    >> NspII
    >> HgiAI
    >> Bsp1286I
    >> BmyI
    >> ApaLI
    >> Alw44I
    CCAGATGGGT ACCCTCTTGA TTGCATCAAA GATTTTCTCG CACGCGCGGG CAAGTCAATG TGCACCTTTT
    850         860         870         880         890         900         910

    >> TthHB8I
    >> TthHB8I
    >> TaqI
    >> TaqI
    >> MnlI
    >> Ksp632I
    >> HinfI>> PleI
    >> Eam1104I
    >> EarI
    >> BbvI>> AccI
    >> Fnu4HI
    >> NlaIII
    >> NlaIII
    >> EcoRII
    >> DsaV
    CCGAACAAC T GATTACATC GAGTCGAAGA GAGGTGTCTA CTGCTGCCGT GACCATGAGC ATGAAATTGC
    920         930         940         950         960         970         980

    >> TthHB8I
    >> TaqI
    >> SfuI
    >> NspV>> Tru9I
    >> LspI>> MseI
    >> ScrFI
    >> HinPII

```

FIGURE 13.3

```

>< MvaI      >< Hin6I      >< SduI      >< Csp45I
>< Ecl136I   >< HhaI      >< NspII     >< BstBI
>< BstOI     >< HaeII     >< HgiAI     >< Bsp119I
>< BstNI     >< Eco47III   >< Bsp1286I  >< BsiCI
>< BsiLI     >< CfoI      >< BmyI      >< Bpu14I
>< ApyI >< DdeI >< Bsp143II >< AluI   >< Alw21I   >< AsuII
CTGGTTCAC T GAGCGCTCTG ATAAGAGCTA CGAGCACCAG ACACCCTTCG AAATTAAGAG TGCCAAGAAA
    990         1000         1010         1020         1030         1040         1050

                                >< Tru9I
                                >< BsmI
                                >< MseI
                                >< BscCI
                                >< MnlI
TTTGACACTT TCAAAGGGGA ATGCCCAAAG TTTGTGTTTC CTCTTAAGTC AAAAGTCAAA GTCATTCAAC
    1060         1070         1080         1090         1100         1110         1120

>< PmlI
>< PmaCI
>< MaeII
>< Eco72I
>< BsaAI
>< BbrPI
>< AflIII   >< MnlI>< DdeI
CACGTGTTGA AAAGAAAAAG ACTGAGGGTT TCATGGGGCG TATACGCTCT GTGTACCCTG TTGCATCTCC
    1130         1140         1150         1160         1170         1180         1190

>< SfaNI
>< MaeIII   >< AccI
ACAGGAGTGT AACAATATGC ACTTGTCTAC CTTGATGAAA TGTAATCATT GCGATGAAGT TTCATGGCAG
    1200         1210         1220         1230         1240         1250         1260

                                >< SinI
                                >< Sau96I
                                PssI ><
                                >< Psp5II
                                >< PpuMI
                                >< NspIV
                                >< NspHII
                                >< Eco47I
                                >< DraII
                                >< Cfr13I
                                >< BsiZI
                                >< Bmel8I
                                >< AvaII
                                >< AsuI
>< MaeII
                                EcoO109I ><AflIII >
ACGTGCGACT TTCTGAAAGC CACTTGTGAA CATTGTGGCA CTGAAAATTT AGTTATTGAA GGACCTACTA
    1270         1280         1290         1300         1310         1320         1330

                                Van91I ><
                                SinI ><
                                Sau96I ><
                                PflMI ><
                                NspIV ><
                                NspHII >
                                Eco47I ><
                                Cfr13I ><
                                BsiI ><
                                BsiZI ><
                                BsiYI ><
                                Bmel8I ><
                                AvaII ><
                                AsuI ><

>< RsaI
>< NspI
>< NlaIV
>< NlaIII
>< NspHI>< KpnI
>< Eco64I
>< Csp6I
>< BscBI
>< BanI
>< Asp718
>< AfaI
>< AccB1I

```

FIGURE 13. 4

```

    >< Acc65I          >< SfcI          >< NlaIII         AccB7I ><
CATGTGGGTA CCTACCTACT AATGCTGTAG TGAAAATGCC ATGTCCTGCC TGTCAAGACC CAGAGATTGG
    1340          1350          1360          1370          1380          1390          1400

                                >< TthHB8I
                                >< TaqI>< MnlI
                                >< HinfI

    >< DdeI
ACCTGAGCAT AGTGTTCAG ATTATCACAA CCACTCAAAC ATTGAAACTC GACTCCGCAA GGGAGGTAGG
    1410          1420          1430          1440          1450          1460          1470

                                >< PleI          >< AciI

    >< RmaI
    >< MnlI
    >< MaeI          >< BbvI          >< Fnu4HI          BscBI ><
ACTAGATGTT TTGGAGGCTG TGTGTTTGCC TATGTTGGCT GCTATAATAA GCGTGCCTAC TGGGTTCCCTC
    1480          1490          1500          1510          1520          1530          1540

                                XhoII ><
                                Sau3AI ><
                                NdeII ><
                                MflI ><
                                MboI ><
                                >< MaeIII
                                >< Eco31I          DpnII ><
                                >< PalI          >< HaeIII          >< BsrI          >< MnlI DpnI >
    >< RmaI          >< BsuRI          >< BsrI          >< BsmAI          BstYI ><
    >< MnlI          > < DdeI          >< BspWI          >< BsaI>< HphI          BspAI ><
    >< MaeI          >< BshI>< BglI          >< Alw26I          BspI43I >
GTGCTAGTGC TGATATTGGC TCAGGCCATA CTGGCATTAC TGGTGACAAAT GTGGAGACCT TGAATGAGGA
    1550          1560          1570          1580          1590          1600          1610

                                > < Tru9I
                                > < MseI
                                >< MaeII          >< Tru9I
                                >< HpaI
                                >< HindII
                                >< HinfI >< PleI >< HincII
    >< AlwI          >< DdeI          >< AflIII          >< MseI
TCTCCTTGAG ATACTGAGTC GTGAACGTGT TAACATTAAC ATTGTTGGCG ATTTTCATTT GAATGAAGAG
    1620          1630          1640          1650          1660          1670          1680

    >< MboII
    >< BstXI          >< SfaNI
GTTGCCATCA TTTTGGCATC TTTCTCTGCT TCTACAAGTG CCTTTATTGA CACTATAAAG AGTCTTGATT
    1690          1700          1710          1720          1730          1740          1750

                                >< StyI
                                >< MaeIII
                                >< EcoT14I
                                >< Eco130I
                                >< PleI
                                >< MaeIII
                                >< HinfI>< AciI
                                >< BssT1I          BslI ><
                                >< BsaJI          BsiYI ><
ACAAGTCTTT CAAACCATT GTTGAGTCCT GCGGTAAC TAAGTTACC AAGGGAAAGC CCGTAAAGG
    1760          1770          1780          1790          1800          1810          1820

    >< Sau3AI
    >< NdeII
    >< MboI
    >< DpnII
    >< DpnI >< Tru9I
    >< BspAI >< MseI
    >< BspI43I

    >< Van91I
    >< PflMI
    >< DraIII
    >< BslI
    >< BsiYI
    >< BbvI
    >< AccB7I          Fnu4HI ><
    >< MnlI

```

FIGURE 135

```

TGCTTGAAC ATTGGACAAC AGAGATCACT TTTAACACCA CTGTGTGGTT TTCCCTCACA GGCTGCTGGT
1830      1840      1850      1860      1870      1880      1890

      >< ThaI
      >< SfaNI
      >< MvnI
      >< HinPII
>< HinPII
>< Hin6I
>< Hin6I
      >< HhaI
>< Sau3AI      >< HhaI
>< NdeII      >< CfoI
>< MboI      >< CfoI
>< DpnII      >< BstUI
      >< DpnI      >< BssHII
>< BspAI      >< Bsp50I
      >< Bsp143I      >< AccII
GTTATCAGAT CAATTTTTCG GCGCACACTT GATGCAGCAA ACCACTCAAT TCCTGATTTG CAAAGAGCAG
1900      1910      1920      1930      1940      1950      1960

      >< TthHB8I
      >< StyI
      >< NcoI
      >< HindII
      >< HincII
      >< HinII
      >< EcoT14I
      >< Eco57I
      >< TaqI>< Eco130I
>< SalI >< DsaI
>< RtrI >< BssTII
      >< BsaHI
      >< BbiII>< NlaIII
      >< AclI >< HgaI
>< MaeIII
      >< BbvI
      >< MaeII >< AccI>< BsaJI      HphI ><
CTGTCAACAT ACTTGATGGT ATTTCTGAAC AGTCATTACG TCTTGTGCGAC GCCATGGTTT ATACTTCAGA
1970      1980      1990      2000      2010      2020      2030

      >< RsaI
      >< NdeI
      >< Csp6I
      >< BspMI
      >< MaeIII >< BsrI >< AfaI      >< DdeI
CCTGCTCACC AACAGTGTC AATTATATGGC ATATGTAAC TGTGGTCTTG TACAACAGAC TTCTCAGTGG
2040      2050      2060      2070      2080      2090      2100

      >< StuI
      >< Pali
      >< HaeIII
      >< Eco147I
      >< SduI
      >< NspII
      >< Bsp1286I
      >< BmyI
      >< DdeI
      >< BsuRI
      >< BshI
      >< AatI
      >< MnlI
      >< DdeI ><
      >< BfrI ><
TTGTCTAATC TTTTGGGCAC TACTGTTGAA AAACCTCAGGC CTATCTTTGA ATGGATTGAG GCGAAACTTA
2110      2120      2130      2140      2150      2160      2170

      >< TfiI
      >< HinfI
      >< Tth111I ><
      >< SfaNI >< BsgI
      >< FokI
      >< AspI ><
GTGCAGGAGT TGAATTTCTC AAGGATGCTT GGGAGATTCT CAAATTTCTC ATTACAGGTG TTTTGCAT
2180      2190      2200      2210      2220      2230      2240

```

FIGURE 13.6


```

Tru9I ><
MseI ><
HpaI >
HindII >
HincII >
>< Eco57I
CGTCAAGGGT CAAATACAGG TTGCTTCAGA TAACATCAAG GATTGTGTAA AATGCTTCAT TGATGTTGTT
2250      2260      2270      2280      2290      2300      2310

>< Sau3AI
>< NdeII
>< MboI
> < MaeIII
>< FbaI
>< DpnII
>< DpnI
>< BspAI
>< Bsp143I
>< TthHB8I
>< TaqI
AACAAGGCAC TCGAAATGTG CATTGATCAA GTCACATATCG CTGGCGCAAA GTTGCGATCA CTCAACTTAG
2320      2330      2340      2350      2360      2370      2380

>< PvuII
>< MaeII
>< Bst1107I
>< BsaAI
>< BbvI
>< HphI
>< DrdI
>< AccI
GTGAAGTCTT CATCGCTCAA AGCAAGGGAC TTTACCGTCA GTGTATACGT GGCAAGGAGC AGCTGCAACT
2390      2400      2410      2420      2430      2440      2450

>< Tru9I
>< NlaIV
>< MseI
>< MnlI
>< Esp4I
>< Eco64I
>< BscBI
>< NlaIII >< BanI
>< AflII
>< BbvI
>< AccBII
>< MaeIII
ACTCATGCCT CTTAAGGCAC CAAAAGAAGT AACCTTTCCT GAAGGTGATT CACATGACAC AGTACTTACC
2460      2470      2480      2490      2500      2510      2520

> < XhoI
>< TthHB8I
>< TthHB8I>< TaqI
> < SlaI
> < PaeR7I
> < NspIII
>< HphI >< HinII
> < Eco88I
> < CcrI
>< Esp3I >< BsaHI
> < BcoI
>< BsmAI >< BbiII
> < AvaI >< HgaI
>< TaqI > < Ama87I>< BsmBI
>< DdeI>< MnlI
>< Alw26I >< AcyI >< AluI
TCTGAGGAGG TTGTTCTCAA GAACGGTGAA CTCGAAGCAC TCGAGACGCC CGTTGATAGC TTCACAAATG
2530      2540      2550      2560      2570      2580      2590

```

FIGURE 13.7

```

                                >> PstI >> NlaIII
                                >> HaeIII >> MnlI
                                >> BsuRI >> DdeI >> Tru9I
                                >> BshI >> BfrI >> MseI
>> AluI >> BsrI
GAGCTATCGT TGGCACACCA GTCTGTGTAA ATGGCCTCAT GCTCTTAGAG ATTAAGGACA AAGAACAATA
2600 2610 2620 2630 2640 2650 2660

                                >> VneI
                                Tru9I >>
                                >> SmaI
                                >> SduI
                                >> NspII
                                MseI >>
                                >> HgiAI
                                Bsp1286I >> BslI >>
                                BsiYI >>
                                >> BmyI
                                >> ApaLI
                                >> Tru9I >> Alw44I
                                >> MseI >> Alw21I
CTGCGCATTG TCTCCTGGTT TACTGGCTAC AAACAATGTC TTTCGCTTAA AAGGGGGTGC ACCAATTAA
2670 2680 2690 2700 2710 2720 2730

                                >> TfiI
>> MaeIII >> MboII >> MaeIII >> HinfI AluI >>
GGTGTAACTT TTGGAGAAGA TACTGTTTGG GAAGTTCAAG GTTACAAGAA TGTGAGAATC ACATTTGAGC
2740 2750 2760 2770 2780 2790 2800

                                >> RsaI
                                >> NlaIV
                                MaeIII >>
                                >> MspI >> KpnI
                                >> HpaII
                                >> HapII
                                >> Eco64I
                                >> SduI
                                >> NspII >> TfiI >> BscBI
                                >> HgiAI >> BanI
                                >> Bsp1286I >> Asp718
                                >> BmyI >> HinfI >> AfaI
                                >> Alw21I >> AccB1I
                                >> AccI >> Acc65I
TTGATGAACG TGTGACAAA GTGCTTAATG AAAAGTGCTC TGTCTACACT GTTGAATCCG GTACCGAAGT
2810 2820 2830 2840 2850 2860 2870

                                >> Sau3AI
                                >> NdeII
                                >> MboI
                                >> DpnII
                                >> DpnI
                                >> MboII >> BspAI
                                >> BsrI >> Bsp143I
>> DdeI >> MnlI >> AlwNI >> BbsI >> AlwNI
TACTGAGTTT GCATGTGTTG TAGCAGAGGC TGTGTGAAG ACTTTACAAC CAGTTTCTGA TCTCCTTACC
2880 2890 2900 2910 2920 2930 2940

>> Sau3AI
>> NdeII
>> MboI
>> DpnII
>> DpnI
>> BspAI

```

FIGURE 13.8

```

    >< NlaIII>< Bsp143I          >< AluI          >< SfaNI
AACATGGGTA TTGATCTTGA TGAGTGGAGT GTAGCTACAT TCTACTTATT TGATGATGCT GGTGAAGAAA
    2950          2960          2970          2980          2990          3000          3010

                                >< SfaNI
                                >< MnlI
                                >< Ksp632I          >< MnlI
                                >< EarI          >< MboII
    >< MboII          >< GsuI          >< MnlI          >< Eam1104I          >< MboII
        >< BsaAI
    >< HphI >< MaeII>< BpmI          >< MnlI          >< Eam1104I          >< MboII
ACTTTTCATC ACGTATGTAT TGTTCCTTTT ACCCTCCAGA TGAGGAAGAA GAGGACGATG CAGAGTGTGA
    3020          3030          3040          3050          3060          3070          3080

                                >< RsaI
                                >< RsaI
                                >< NlaIII
                                >< MnlI          >< FokI
                                >< Csp6I          Eco31I ><
                                >< Csp6I          >< MamI BsmAI ><
        >< MboII          >< AfaI          >< BsiBI BsaI ><
        >< MboII          >< AfaI          >< BsaB1Alw26I ><
GGAAGAAGAA ATTGATGAAA CCTGTGAACA TGAGTACGGT ACAGAGGATG ATTATCAAGG TCTCCCTCTG
    3090          3100          3110          3120          3130          3140          3150

    >< NlaIV>< PvuII>< XmnI
    >< Eco64I >< Psp5I          >< TthHB8I
    >< MnlI >< DdeI          >< TaqI          >< MnlI          >< MboII
        >< BscBI>< NspBII >< MnlI          >< Ksp632I          >< MboII >< MboII
    >< BanI          >< MnlI          >< EarI          >< BsrI
    >< AccBII >< AluI >< Asp700I          >< Eam1104I >< MboII>< BbsI
GAATTTGGTG CCTCAGCTGA AACAGTTCGA GTTGAGGAAG AAGAAGAGGA AGACTGGCTG GATGATACTA
    3160          3170          3180          3190          3200          3210          3220

                                >< Tru9I
                                >< MseI          >< Eco57I
    >< FokI          >< DdeI          >< BsrI>< MboII BsrI ><
CTGAGCAATC AGAGATTGAG CCAGAACCAG AACCTACACC TGAAGAACCA GTTAATCAGT TTACTGGTTA
    3230          3240          3250          3260          3270          3280          3290

    >< Tru9I          >< MnlI
    >< MseI          >< Tru9I >< HindII>< Tru9I          >< DraIII
    >< DraI          >< MseI >< HincII>< MseI          >< BspWI
TTTAAAACTT ACTGACAATG TTGCCATTAA ATGTGTTGAC ATCGTTAAGG AGGCACAAAG TGCTAATCCT
    3300          3310          3320          3330          3340          3350          3360

                                >< VneI
                                >< SnaI
                                >< SduI
                                >< NspII
                                >< HgiAI
                                >< Bsp1286I
                                >< BmyI
                                >< ApaLI
                                >< Alw44I
    >< BbvI          >< HphI          >< NlaIII          >< Alw21I
        >< Fnu4HI          >< BspMI
ATGGTGATTG TAAATGCTGC TAACATACAC CTGAAACATG GTGGTGGTGT AGCAGGTGCA CTCAACAAGG
    3370          3380          3390          3400          3410          3420          3430

                                >< Sau96I
                                >< Pali
                                >< NspIV
                                >< HaeIII
    >< NlaIV          >< Cfr13I

```

FIGURE 13.9

```

    >< Eco64I
    >< BscBI
    >< BstI
    >< AccB1I>< NlaIII
CAACCAATGG TGCCATGCAA AAGGAGAGTG ATGATTACAT TAAGCTAAAT GGCCCTCTTA CAGTAGGAGG
3440      3450      3460      3470      3480      3490      3500

    >< BsuRI
    >< Tru9I
    >< MseI
    >< AluI >< AsuI >< MnlI
    >< SinI
    >< Sau96I
    >< NspIV
    >< NspHI>< NspHII
    >< Eco47I
    >< Cfr13I
    >< NlaIII >< BspMI
    >< BsiZI
    >< Bme18I
    >< AvaII MnlI ><
    >< DdeI
    >< NspI>< AsuI FokI ><
GTCTTGTTTG CTTTCTGGAC ATAATCTTGC TAAGAAGTGT CTGCATGTTG TTGGACCTAA CCTAAATGCA
3510      3520      3530      3540      3550      3560      3570

    >< Tru9I
    >< HphI> < MseI
    >< Esp4I
    >< AluI >< NdeI
    >< AflIII>< Fnu4HI >< BbvI
GGTGAGGACA TCCAGCTTCT TAAGGCAGCA TATGAAAATT TCAATTCACA GGACATCTTA CTTGCACCAT
3580      3590      3600      3610      3620      3630      3640

    RsaI ><
    Csp6I ><
    >< Eco57I
    >< BcgI
    >< AfaI ><
TGTTGTCAGC AGGCATATTT GGTGCTAAAC CACTTCAGTC TTTACAAGTG TGCCTGCAGA CGGTTTCGTAC
3650      3660      3670      3680      3690      3700      3710

    >< BsgI
    >< BcgI/a
    >< BspMI
    >< AluI
    >< NlaIII
ACAGGTTTAT ATTGCAGTCA ATGACAAAGC TCTTTATGAG CAGGTTGTCA TGGATTATCT TGATAACCTG
3720      3730      3740      3750      3760      3770      3780

    >< MnlI
    >< RmaI
    >< MaeI
    >< MnlI
    >< Eco57I
    >< BscBI
    >< NlaIV
    >< TfiI >< MboII
    >< HinfI >< DdeI
AAGCCTAGAG TGGAAGCACC TAAACAAGAG GAGCCACCAA ACACAGAAGA TTCCAAAACCT GAGGAGAAAT
3790      3800      3810      3820      3830      3840      3850

    >< Tru9I
    >< StuI
    >< PstI
    >< MseI >< MnlI
    >< MaeIII
    >< HaeIII
    >< Eco147I
    >< Eco91I
    >< BsuRI
    >< BstXI ><
    >< Csp6I
    >< TthHB8I
    >< BshI
    >< BstPI
    >< AfaI
    >< TaqI
    >< AatI
    >< BstEII
CTGTCGTACA GAAGCCTGTC GATGTGAAGC CAAAAATTAA GGCCTGCATT GATGAGGTTA CCACAACACT
3860      3870      3880      3890      3900      3910      3920

    TfiI ><
    NlaIII ><
    HinfI ><
    >< DdeI
    >< EcoRV
    >< HindIII

```

FIGURE 13.10

```

>< BsrI      >< MboII      >< MaeIII      >< Eco32I      >< AluI
GGAAGAACT  AAGTTTCTTA  CCAATAAGTT  ACTCTTGTTT  GCTGATATCA  ATGGTAAGCT  TTACCATGAT
3930      3940      3950      3960      3970      3980      3990

      >< NspI
      >< NspHI
      >< NlaIII
>< MnlI      >< SfaNI
      >< EcoNI
      >< MboII >< BslI      >< NlaIII
      >< DdeI      >< BfrI      >< HphI      >< BsiYI      >< FokI
TCTCAGAACA  TGCTTAGAGG  TGAAGATATG  TCTTTCCTTG  AGAAGGATGC  ACCTTACATG  GTAGGTGATG
4000      4010      4020      4030      4040      4050      4060

>< SpeI
>< RmaI
>< MaeI      >< EcoRV>< HphI      >< SfaNI
>< HphI      >< Eco32I      >< MnlI      >< DdeI
TTATCACTAG  TGGTGATATC  ACTTGTTGTTG  TAATACCCTC  CAAAAGGCT  GGTGGCACTA  CTGAGATGCT
4070      4080      4090      4100      4110      4120      4130

      >< ScrFI
      >< RsaI
      >< MvaI
      >< EcoRII
      >< Ecl136I
      >< DsaV
      >< Csp6I >< EcoNI
      >< BstOI
      >< BstNI
      >< BsiLI
      >< BsaJI
      >< BsaAI      >< BslI
      >< MaeII>< ApyI
      >< AfaI      >< BsiYI
CTCAAGAGCT  TTGAAGAAAG  TGCCAGTTGA  TGAGTATATA  ACCACGTACC  CTGGACAAGG  ATGTGCTGGT
4140      4150      4160      4170      4180      4190      4200

      >< Tru9I
      >< MseI
      >< DdeI      >< Esp4I      >< RsaI
>< MnlI      >< BspWI      >< Csp6I
>< FokI      >< AluI      >< AflII      >< Eco57I >< AfaI
TATACACTTG  AGGAAGCTAA  GACTGCTCTT  AAGAAATGCA  AATCTGCATT  TTATGTACTA  CCTTCAGAAG
4210      4220      4230      4240      4250      4260      4270

      >< ScrFI
      >< MvaI
      >< EcoRII
      >< Ecl136I
      >< XmnI
      >< Ksp632I      >< RmaI      >< DsaV      NlaIII ><
      >< EarI      >< TfiI>< MboII      >< BstOI      Ksp632I ><
      >< Eam1104I      >< MaeI      >< BstNI      Eam1104I ><
      >< DdeI      >< HinfI      >< BsiLI      BsmAI ><
      >< BspWI      >< Asp700I      >< ApyI      Alw26I ><
CACCTAATGC  TAAGGAAGAG  ATTCTAGGAA  CTGTATCCTG  GAATTTGAGA  GAAATGCTTG  CTCATGCTGA
4280      4290      4300      4310      4320      4330      4340

      >< VspI      >< Zsp2I
      >< Tru9I      >< Ppu10I
      >< MseI      >< NsiI
      >< MboII      >< NlaIII      >< FokI
      >< Eco57I      >< Mph1103I      >< FokI

```

FIGURE 13. 11

```

                >< AsnI          >< EcoT22I          >< BspWI
                >< AseI          >< AvaIII          >< BglI          >< MaeII
AGAGACAAGA AAATTAATGC CTATATGCAT GGATGTTAGA GCCATAATGG CAACCATCCA ACGTAAGTAT
4350          4360          4370          4380          4390          4400          4410

                >< SfaNI
                >< Tru9I          > < HindII          >< TfiI          >< SpeI
                >< MseI          > < HincII> < MboII          >< RmaI
                >< MnlI          >< DrdI >< HinfI          >< MaeI
AAAGGAATTA AAATTCAAGA GGGCATCGTT GACTATGGTG TCCGATTCTT CTTTATACT AGTAAAGAGC
4420          4430          4440          4450          4460          4470          4480

                >< MaeIII
>< SfcI          >< Fnu4HI          >< MnlI
>< AluI          >< AluI          >< AciI          >< MaeIII ><
CTGTAGCTTC TATTATTACG AAGCTGAACT CTCTAAATGA GCCGCTTGTC ACAATGCCAA TTGGTTATGT
4490          4500          4510          4520          4530          4540          4550

                >< ThaI
                >< MvnI
                >< MboII
                >< HinfII
>< HinfII
>< HinfI
>< HinfI
>< HinfI
>< HhaI
                >< Tru9I          >< HhaI
>< NlaIII          >< Fnu4HI
                >< MseI          >< CfoI
                >< MnlI          >< CfoI
                >< Ksp632I          >< BstUI
                >< EarI          >< BssHII>< BspWI          >< Tru9I
                >< Eam1104I          >< Bsp50I          >< MseI
                >< BbvI          >< AccII          >< AluI          HphI ><
GACACATGGT TTTAATCTTG AAGAGGCTGC GCGCTGTATG CGTTCCTTA AAGCTCCTGC CGTAGTGTCA
4560          4570          4580          4590          4600          4610          4620

                >< MaeIII
>< SfaNI          >< AlwNI          >< MnlI >< MnlI>< DdeI
GTATCATCAC CAGATGCTGT TACTACATAT AATGGATACC TCACTTCGTC ATCAAAGACA TCTGAGGAGC
4630          4640          4650          4660          4670          4680          4690

                >< SinI
                >< Sau96I
                >< NspIV
                >< NspHII
>< SduI          >< Eco47I
>< NspII          >< Cfr13I
>< HgiAI          >< BsiZI
>< Bsp1286I          >< Bme18I          >< RsaI
>< BmyI          >< AvaII          >< Csp6I
>< Alw21I          >< AsuI          >< AfaI
ACTTTGTAGA AACAGTTTCT TTGGCTGGCT CTTACAGAGA TTGGTCTTAT TCAGGACAGC GTACAGAGTT
4700          4710          4720          4730          4740          4750          4760

                > < TthHB8I
                > < TaqI
                >< SduI
                >< Van91I          >< NspII
                >< Tru9I          >< RsaI          >< PflMI          >< Eco24I
                >< MseI          >< HphI          >< BslI          >< Bsp1286I
                >< Esp4I          >< Csp6I          >< BsiYI          >< BmyI          >< GsuI ><

```

FIGURE 13.12

```

      >< AflIII >< MaeIII >< AfaI >< AccB7I >< BanIIBpmI ><
AGGTGTTGAA TTTCTTAAGC GTGGTGACAA AATTGTGTAC CACACTCTGG AGAGCCCCGT CGAGTTTCAT
4770 4780 4790 4800 4810 4820 4830

      >< Tru9I
      >< PleI >< EcoNI
      >< MnlI >< BslI
      >< BsmAI >< BsiYI
      >< MnlI >< HphI >< HinfI>< Alw26I>< AciI >< MseI
CTTGACGGTG AGGTTCTTTC ACTTGACAAA CTAAAGAGTC TCTTATCCCT GCGGGAGGTT AAGACTATAA
4840 4850 4860 4870 4880 4890 4900

      >< AluI >< NdeI
AAGTGTTTAC AACTGTGGAC AACACTAATC TCCACACACA GCTTGTGGAT ATGTCTATGA CATATGGACA
4910 4920 4930 4940 4950 4960 4970

      >< SinI
      >< Sau96I
      >< NspIV
      >< NspHII
      >< Eco47I
      >< Cfr13I
      >< BsiZI
      >< Bme18I
      >< AvaII
      >< AsuI
      >< MaeIII >< Tru9I >< MnlI
      >< FokI >< MseI >< BspHI
GCAGTTTGGT CCAACATACT TGGATGGTGC TGATGTTACA AAAATTAAAC CTCATGTAAA TCATGAGGGT
4980 4990 5000 5010 5020 5030 5040

      >< RsaI
      >< RmaI
      >< MaeI
      >< Csp6I
      >< AfaI
      >< RsaI
      >< TaqI
      >< SnaBI
      >< ScaI
      >< MaeII >< HindIII >< RsaI
      >< Eco105I >< Csp6I
      >< BsaAI >< AluI >< AfaI
AAGACTTTCT TTGTACTACC TAGTGATGAC ACACTACGTA GTGAAGCTTT CGAGTACTAC CATACTCTTG
5050 5060 5070 5080 5090 5100 5110

      >< RsaI
      >< NspI
      >< NspHI
      >< NlaIII
      >< Csp6I >< Tru9I
      >< AflIII >< MseI
      >< AfaI >< DraI
      >< MnlI >
      >< BslI ><
      >< BsiYI ><
ATGAGAGTTT TCTTGGTAGG TACATGTCTG CTTTAAACCA CACAAAGAAA TGGAAATTTC CTCAAGTTGG
5120 5130 5140 5150 5160 5170 5180

      >< Tru9I >< Tru9I
      >< MseI >< MseI
      >< MunI
      >< RmaI
      >< MaeI
      >< AluI >
TGGTTTAACT TCAATTAAAT GGGCTGATAA CAATTGTTAT TTGTCTAGTG TTTTATTAGC ACTTCAACAG
5190 5200 5210 5220 5230 5240 5250

      >< SfaNI
      >< SduI
      >< NspII
      >< Eco24I
      >< Bsp1286I
      >< BmyI
      >< BbvI Fnu4HI ><
      >< BanII >< BspWI
      >< HphI >
      >< MnlI

```

FIGURE 13.13

```

CTTGAAGTCA AATTCAATGC ACCAGCACTT CAAGAGGCTT ATTATAGAGC CCGTGCTGGT GATGCTGCTA
5260      5270      5280      5290      5300      5310      5320

>< VneI
>< SnoI
    >< SduI
    >< NspII
    >< HgiAI
    >< Bsp1286I
    >< BmyI
>< ApaLI
>< Alw44I
    >< Alw21I
    >< AluI
    MboII ><
    >< HphI
ACTTTTGTGC ACTCATACTC GCTTACAGTA ATAAAACTGT TGGCGAGCTT GGTGATGTCA GAGAAACTAT
5330      5340      5350      5360      5370      5380      5390

    > < SphI
    > < PaeI
    > < NspI
    > < NspHI >< TfiI
    >< Tru9I
    >< SfcI > < NlaIII>< HinfI
    >< MseI
GACCCATCTT CTACAGCATG CTAATTTGGA ATCTGCAAAG CGAGTTCTTA ATGTGGTGTG TAAACATTGT
5400      5410      5420      5430      5440      5450      5460

    >< RsaI
    >< Tru9I
    > < Csp6I
    Esp4I >
    >< MseI
    >< AluI
    >< AfaI
    AflII >
GGTCAGAAAA CTACTACCTT AACGGGTGTA GAAGCTGTGA TGTATATGGG TACTCTATCT TATGATAATC
5470      5480      5490      5500      5510      5520      5530

    >< RsaI
    >< MboII
    >< RmaI HinfI ><
    >< Csp6I
    >< MaeI >< BbsI
    >< AfaI
>< Tru9I
>< MseI
    >< SfaNI
    >< NlaIII
TTAAGACAGG TGTTCCTT CCATGTGTGT GTGGTCGTGA TGCTACACAA TATCTAGTAC AACAAGAGTC
5540      5550      5560      5570      5580      5590      5600

    >< RsaI
    >< PleI
    > < DdeI
    >< Csp6I
    >< BsgI
    >< BspWI >< BspMI
    >< AfaI
TTCTTTTGTG ATGATGTCTG CACCACCTGC TGAGTATAAA TTACAGCAAG GTACATTCTT ATGTGCGAAT
5610      5620      5630      5640      5650      5660      5670

    >< RsaI
    >< Eco31I
    >< DdeI
    > < MaeIII
    >< BsmAI
    >< Csp6I
    >< BsaI
    MnII ><
    >< AfaI >< BsrI
    >< Alw26I
    >< HphI >
GAGTACACTG GTAACATCA GTGTGGTCAT TACACTCATA TAACTGCTAA GGAGACCCTC TATCGTATTG
5680      5690      5700      5710      5720      5730      5740

    >< SstI
    >< SinI
    >< SduI
    >< Sau96I
    >< SacI
    >< NspIV
    >< NspHII
    >< HgiAI
    > < RsaI
    >< MaeIII
    >< Eco24I
    >< Eco47I
    >< Ecl136II
    >< Cfr13I
    >< Bsp1286I
    >< Bsi2I
    >< BmyI
    >< Bme18I

```

FIGURE 13. 14


```

    >< BanII
    >< Alw21I
    >< AluI
ACGGAGCTCA CCTTACAAAG ATGTCAGAGT ACAAAGGACC AGTGACTGAT GTTTTCTACA AGGAAACATC
    5750      5760      5770      5780      5790      5800      5810

    >< AvaII
    >< Csp6I>< AsuI
    > < AfaI >< BsrI>< AlwNI
    >< TthHB8I
    >< TaqI >< MaeIII
TTACTACTACA ACCATCAAGC CTGTGTCGTA TAAACTCGAT GGAGTTACTT ACACAGAGAT TGAACCAAAA
    5820      5830      5840      5850      5860      5870      5880

    >< RsaI
    >< Csp6I
    >< SfcI >< BbvI
    >< FokI
    >< Fnu4HI
    >< AfaI
TTGGATGGGT ATTATAAAAA GGATAATGCT TACTATACAG AGCAGCCTAT AGACCTTGTA CCAACTCAAC
    5890      5900      5910      5920      5930      5940      5950

    >< Tru9I ><
    >< SwaI ><
    >< MseI ><
    > < NspI
    > < NspHI
    > < NlaIII
    >< AflIII
    >< Tru9I ><
    >< SwaI ><
    >< MseI ><
    >< MamI ><
    >< DraI ><
    >< BsiBI ><
    >< BsaBI ><
CATTACCAAA TGCGAGTTTT GATAATTTCA AACTCACATG TTCTAACACA AAATTTGCTG ATGATTTAAA
    5960      5970      5980      5990      6000      6010      6020

    >< MboII
    >< AluI
    >< AluI>< MaeIII
TCAAATGACA GGCTTCACAA AGCCAGCTTC ACGAGAGCTA TCTGTCACAT TCTTCCCAGA CTTGAATGGC
    6030      6040      6050      6060      6070      6080      6090

    >< SfcI
GATGTAGTGG CTATTGACTA TAGACACTAT TCAGCGAGTT TCAAGAAAGG TGCTAAATTA CTGCATAAGC
    6100      6110      6120      6130      6140      6150      6160

    >< Tru9I
    >< ScrFI
    >< MvaI
    >< MseI
    >< EcoRII
    >< Ecl136I
    >< DsaV
    >< BstOI
    >< BstNI
    >< BsiLI
    >< MuiI
    >< BstXI
    >< ApyI
    >< MaeII
    >< BstXI
    >< MaeII ><
    >< DraIII
    >< BstXI
CAATTGTTTG GCACATTAAC CAGGCTACAA CCAAGACAAC GTTCAAACCA AACACTTGGT GTTTACGTTG
    6170      6180      6190      6200      6210      6220      6230

    > < RsaI
    >< Csp6I
    > < AfaI>< BsrI
    >< MboII ><
    >< BbsI
TCTTTGGAGT ACAAAGCCAG TAGATACTTC AAATTCATTT GAAGTTCTGG CAGTAGAAGA CACACAAGGA
    6240      6250      6260      6270      6280      6290      6300

    >< HindII
    >< HincII
    >< MboII
    >< MnlI
    >< Eco57I
ATGGACAATC TTGCTTGTA AAGTCAACAA CCCACCTCTG AAGAAGTAGT GGAAAATCCT ACCATACAGA
    6310      6320      6330      6340      6350      6360      6370

```

FIGURE 13.15

```

                >< MaeIII
                >< MaeII
AGGAAGTCAT AGAGTGTGAC GTGAAACTA CCGAAGTTGT AGGCAATGTC ATACTTAAAC CATCAGATGA
    6380         6390         6400         6410         6420         6430         6440

                >< XhoII
                >< Sau3AI
                >< NlaIII
                >< NdeII
                >< MflI
                >< MboI
                >< DpnII
                >< DpnI
                >< BstYI
                >< BspAI
    >< Tru9I
    >< MseI
                >< BspHI >< Bsp143I>< Fnu4HI
                > < MaeIII
                >< MnlI >< BbvI
                >< AlwI
AGGTGTTAAA GTAACACAAG AGTTAGGTCA TGAGGATCTT ATGGCTGCTT ATGTGGAAAA CACAAGCATT
    6450         6460         6470         6480         6490         6500         6510

                >< SauI
                >< RmaI
                >< MstII
                >< MaeI
                >< Eco81I
                >< DdeI
                >< CvnI
                >< Bsu36I
                >< Bse21I
                >< BfrI> < Tru9I
    >< Tru9I
    >< MseI
                >< AluI
                >< AxyI> < MseI>< MunI
                >< AocI >< DraI
                >< BbvI Fnu4HI ><
ACCATTAAGA AACCTAATGA GCTTTCAC TA GCTTAGGTT TAAAAACAAT TGCCACTCAT GGTATTGCTG
    6520         6530         6540         6550         6560         6570         6580

    >< VspI
    >< Tru9I
    >< MseI
    >< AsnI
    >< AseI
    >< StyI
    >< EcoT14I
    >< Eco130I
    >< BssT1I
    >< BsaJI
                >< DdeI
                >< BslI
                >< BsiYI
                >< BfrI
                >< Fnu4HI
CAATTAATAG TGTTCCTTGG AGTAAAATTT TGGCTTATGT CAAACCATTC TTAGGACAAG CAGCAATTAC
    6590         6600         6610         6620         6630         6640         6650

                >< HinP1I
                >< Hin6I
                >< HhaI
                >< DdeI
    >< BbvI
                >< CfoI
                >< Tru9I
                >< MaeII>< MseI
                >< DraIII
                >< AflIII
AACATCAAAT TGCCTAAGA GATTAGCACA ACGTGTGTTT AACAAATTATA TGCCTTATGT GTTTACATTA
    6660         6670         6680         6690         6700         6710         6720

                >< RsaI
                >< Csp6I
    >< MunI >< AfaI
                >< RsaI>< XbaI
                >< Csp6I >< RmaI
                >< AfaI >< MaeI
                >< AluI
TTGTTCCAAT TGTGTACTTT TACTAAAAGT ACCAATTCTA GAATTAGAGC TCACTACCT ACAACTATTG
    6730         6740         6750         6760         6770         6780         6790

                >< VspI
                >< Tru9I
                >< NaeI
                >< MspI
                >< MseI

```

FIGURE 13. 16

```

                                >< HpaII
                                >< HapII
                                >< Cfr10I >< FokI
                                >< AsnI
                                >< Tru9I
                                >< MseI
                                >< SfaNI
                                >< AseI>< HphI>< MaeIII
CTAAAAATAG TGTTAAGAGT GTTGCTAAAT TATGTTTGGA TGCCGGCATT AATTATGTGA AGTCACCCAA
6800      6810      6820      6830      6840      6850      6860

                                >< Tru9I      >< DdeI      MaeIII >
                                >< MseI      >< BfrI      >< BbvI
ATTTTCTAAA TTGTTCAAA TCGCTATGTG GCTATTGTTG TTAAGTATTT GCTTAGGTTC TCTAATCTGT
6870      6880      6890      6900      6910      6920      6930

                                >< SduI
                                >< NspII
                                >< HgiAI
                                >< Bsp1286I
                                >< BmyI
                                >< Alw21I
                                > < RsaI
                                >< Csp6I
                                >< Fnu4HI      > < AfaI
GTAAGTGTG CTTTGGTGT ACTCTTATCT AATTTTGGTG CTCCTTCTTA TTGTAATGGC GTTAGAGAAT
6940      6950      6960      6970      6980      6990      7000

                                Tru9I ><
                                MseI ><
                                >< Tru9I      > < MaeIII
                                >< MseI      >< MaeII
                                >< Fnu4HI
                                >< BbvI >
TGTATCTTAA TTCGTCTAAC GTTACTACTA TGGATTCTG TGAAGGTTCT TTCCTTGCA GCATTTGTTT
7010      7020      7030      7040      7050      7060      7070

                                > < TfiI
                                >< MamI
                                >< HinfI
                                >< BsiBI
                                >< XmnI>< MaeIII
                                >< Asp700I
                                >< AluI >
                                >< PleI>< HinfI
                                >< BsaBI >< AluI
                                >< Asp700I
                                >< AfaI ><
AAGTGGATTA GACTCCCTTG ATTCTTATCC AGCTCTTGAA ACCATTGAGG TGACGATTTT ATCGTACAAG
7080      7090      7100      7110      7120      7130      7140

                                >< Pali
                                >< NspBII
                                >< HaeIII
                                >< GdiII
                                >< Fnu4HI
                                >< EaeI
                                >< DdeI
                                >< BsuRI
                                >< BshI >< BslI
                                >< MaeI
                                >< AciI>< BsiYI
CTAGACTTGA CAATTTTAGG TCTGGCCGCT GAGTGGGTTT TGGCATATAT GTTGTTCACA AAATTCITTT
7150      7160      7170      7180      7190      7200      7210

                                >< BspMI
                                >< AluI
                                >< RmaI
                                >< MaeI
ATTATTAGG TCTTTCAGCT ATAATGCAGG TGTTCTTTGG CTATTTTGCT AGTCATTTCA TCAGCAATTC
7220      7230      7240      7250      7260      7270      7280

                                RsaI ><
                                >< MboII
                                >< NlaIV
                                >< Eco64I
                                >< RsaI >< BscBI
                                >< Csp6I >< BanI
                                >< AfaI>< AccBI
                                > < NlaIII
                                >< RmaI
                                >< MamI ><
                                >< Csp6I ><
                                >< BsiBI ><
                                >< BsaBI ><
                                >< AfaI ><

```

FIGURE 13.17

```

TTGGCTCATG TGGTTTATCA TTAGTATTGT ACAAATGGCA CCCGTTTCTG CAATGGTTAG GATGTACATC
7290          7300          7310          7320          7330          7340          7350

TthHB8I ><
>< TaqI
MnlI ><
>< NdeI
>< Ksp632I
>< EarI
>< FokI
>< MboII EarI ><
>< FokI
>< Eam1104I>< AluI>< MboII >< NlaIII Eam1104I ><
TTCTTTGCTT CTTTCTACTA CATATGGAAG AGCTATGTTT ATATCATGGA TGGTTGCACC TCTTCGACTT
7360          7370          7380          7390          7400          7410          7420

XhoII ><
Sau3AI ><
NlaIII ><
NdeII ><
MflI ><
MboI ><
>< ThaI
>< MvnI
>< EarI
>< Eam1104I
DpnII ><
BstYI ><
>< NlaIII >< CfoI >< AflIII >< Csp6I >< Tru9I BspAI ><
>< BspWI >< BspWI >< AccII >< AfaI >< MseI BglII ><
GCATGATGTG CTATAAGCGC AATCGTGCCA CACGCGTTGA GTGTACAAC ATTGTTAATG GCATGAAGAG
7430          7440          7450          7460          7470          7480          7490

>< Pali
>< HaeIII
>< DsaI
>< MnlI
>< BsuRI
>< BshI
>< MunI
>< Bsp143I >< MnlI >< BsaJI >< PleI>< HinfI Alw26I ><
ATCTTTCTAT GTCTATGCAA ATGGAGGCCG TGGCTTCTGC AAGACTCACA ATTGGAATTG TCTCAATTGT
7500          7510          7520          7530          7540          7550          7560

>< RsaI
>< Csp6I
>< EsrI
>< AfaI
>< GsuI
>< BpmI
>< MaeIIIDraI ><
>< BsrI
>< BssHII
Bsp50I ><
>< BsrI
>< AccII
GACCAATCAA CCCTACTGAC CAGTCATCGT ATATTGTTGA TAGTGTTGCT GTGAAAAATG GCGCGCTTCA
7640          7650          7660          7670          7680          7690          7700

```

FIGURE 13. 18

```

                >< FokI
                    >< BsmAI
                >< MnlI                >< Alw26I                >< AclI
CCTCTACTTT GACAAGGCTG GTCAAAAGAC CTATGAGAGA CATCCGCTCT CCCATTTTGT CAATTTAGAC
    7710          7720          7730          7740          7750          7760          7770

                    >< VspI
                    >< Tru9I
                    >< MseI
                    >< AsnI
                > < AluI                >< AseI                >< BcgI/a
AATTTGAGAG CTAACAACAC TAAAGGTTCA CTGCCTATTA ATGTCATAGT TTTTGATGGC AAGTCCAAAT
    7780          7790          7800          7810          7820          7830          7840

                    >< SfcI                >< PvuII
                    >< RsaI                >< Psp5I
                >< PleI                >< Csp6I                >< NspBII
                >< HinfI                >< DdeI                >< BcgI                >< AfaI                >< AluI
GCGACGAGTC TGCTTCTAAG TCTGCTTCTG TGTACTACAG TCAGCTGATG TGCCAACCTA TTCTGTTGCT
    7850          7860          7870          7880          7890          7900          7910

                                                    TthHB8I ><
                                                    TaqI ><
                                                    SalI ><
                                                    RtrI ><
                    >< ScaI                >< Tru9I                HindII >
                    >< RsaI                >< Csp6I                >< SfaNI >< Eco57I
                    >< Csp6I                >< SfaNI >< Eco57I
                >< AluI                >< MaeII                >< AfaI                >< MseI                AccI ><
TGACCAAGCT CTTGTATCAG ACGTTGGAGA TAGTACTGAA GTTTCCGTTA AGATGTTTGA TGCTTATGTC
    7920          7930          7940          7950          7960          7970          7980

                    >< Tru9I
                    >< MseI
                > < Esp4I                >< SfcI
                > < AflII                >< BspWI                >< AluI
GACACCTTTT CAGCAACTTT TAGTGTTTCT ATGGAAAAAC TTAAGGCACT TGTGCTACA GCTCACAGCG
    7990          8000          8010          8020          8030          8040          8050

                                                    >< PvuII
                                                    >< Psp5I
                                                    >< NspBII
                                                    >< Fnu4HI
                >< AluI                >< BbvI                >< AluI
AGTTAGCAAA GGGTGTAGCT TTAGATGGTG TCCTTTCTAC ATTCGTGTCA GCTGCCCCGAC AAGGTGTTGT
    8060          8070          8080          8090          8100          8110          8120

                                                    MaeIII ><
                >< HindII                >< BsmAI                >< DdeI
                >< HincII                >< FokI >< Alw26I                >< BfrI
TGATACCGAT GTTGACACAA AGGATGTTAT TGAATGTCTC AAACTTTCAC ATCACTCTGA CTTAGAAGTG
    8130          8140          8150          8160          8170          8180          8190

                                                    >< XhoII
                                                    Sau3AI ><
                                                    >< NdeII
                                                    >< MflI
                                                    >< MboI
                >< NlaIII >< HgaI
                >< HinfI >< DpnII
                                                    DpnI ><

```

FIGURE 13.19

```

Bsp143I ><
>< BsaHI >< BstYI
>< BbiII >< BspAI
>< AcyI >< BglII
>< MaeIII>< HphI
>< MaeIII >< HphI >< NlaIII
ACAGGTGACA GTTGTAAACA TTTCATGCTC ACCTATAATA AGGTTGAAAA CATGACGCCC AGAGATCTTG
8200 8210 8220 8230 8240 8250 8260

>< NspI
>< NspHI
>< NlaIII
>< HinPII
>< Hin6I
>< HhaI
>< CfoI
>< BspWI >< MaeIII
GCGCATGTAT TGA CTGTAAT GCAAGGCATA TCAATGCCCA AGTAGCAAAA AGTCACAATG TTTCATCAT
8270 8280 8290 8300 8310 8320 8330

>< NspI
>< NspHI >< PvuII
>< NlaIII >< Psp5I
>< Eam1105I >< NspBII
>< BbvI >< Fnu4HI
>< AflIII >< AluI >< BbvI >< Fnu4HI
CTGGAATGTA AAAGACTACA TGTCTTTATC TGAACAGCTG CGTAAACAAA TTCGTAGTGC TGCCAAGAAG
8340 8350 8360 8370 8380 8390 8400

>< RmaI
>< MboII >< MaeI >< Eam1105I
AACACATAC CTTT TAGACT AACTTGTGCT ACAACTAGAC AGGTTGTCAA TGTCATAACT ACTAAAATCT
8410 8420 8430 8440 8450 8460 8470

>< Tru9I
>< Pali
>< MseI
>< HaeIII
>< ScaI >< Esp4I
>< RsaI >< Tru9I >< BsuRI
>< Csp6I >< MseI >< BshI
>< AfaI >< DraI >< AflII >< BbvI
CACTCAAGGG TGGTAAGATT GTTAGTACTT GTTTTAAACT TATGCTTAAG GCCACATTAT TGTGCGTTCT
8480 8490 8500 8510 8520 8530 8540

>< RsaI
>< Csp6I
>< BsrI >< NlaIII
>< Fnu4HI >< AfaI >< MaeIII
TGCTGCATTG GTTTGTTATA TCGTTATGCC AGTACATACA TTGTCAATCC ATGATGGTTA CACAAATGAA
8550 8560 8570 8580 8590 8600 8610

>< MaeIII
>< MaeIII
>< FokI
ATCATTGGTT ACAAGCCAT TCAGGATGGT GTCACTCGTG ACATCATTC TACTGATGAT TGTTTTGCAA
8620 8630 8640 8650 8660 8670 8680

>< NspI
>< NspHI >< NlaIII
>< NlaIII >< HgaI >< BstXI >< BbvI >< AluI
ATAAACATGC TGGTTTTGAC GCATGGTTTA GCCAGCGTGG TGGTTCATAC AAAAATGACA AAAGCTGCCC
8690 8700 8710 8720 8730 8740 8750
SfcI >
Fnu4HI ><
BbvI ><

```

FIGURE 13. 20

```

                                >< ScrFI
                                >< ScrFI    >< RsaI
                                >< MvaI    >< MspI
                                >< EcoRII   >< HpaII
                                >< Ecl136I>< NciI
                                >< DsaV     >< HapII
                                >< BstOI>< DsaV
                                >< BstNI    >< Csp6I
                                >< BsiLI    >< BcnIDdeI ><
                                >< ApyI     >< AfaI
                                >< Fnu4HI
                                >< AluI
TGTAGTAGCT GCTATCATTA CAAGAGAGAT TGGTTTCATA GTGCCGTGGCT TACCGGGTAC TGTGCTGAGA
8760      8770      8780      8790      8800      8810      8820

                                > < MaeIII   >< HphI           >< MnlI           >< BspWI
GCAATCAATG GTGACTTCTT GCATTTTCTA CCTCGTGTTC TTAGTGCTGT TGGCAACATT TGCTACACAC
8830      8840      8850      8860      8870      8880      8890

                                Tru9I >
                                SfaNI ><
                                >< RsaI
                                MseI >
                                >< BspWI           >< Fnu4HI >< Csp6I
                                >< BbvI>< MnlI       >< DdeI >< AfaI
CTTCCAACT CATTGAGTAT AGTGATTTTG CTACCTCTGC TTGCGTTCTT GCTGCTGAGT GTACAATTTT
8900      8910      8920      8930      8940      8950      8960

                                > < RmaI
                                >< MnlI
                                >< FokI
                                > < MaeI
TAAGGATGCT ATGGGCAAAC CTGTGCCATA TTGTTATGAC ACTAATTTGC TAGAGGGTTC TATTTCTTAT
8970      8980      8990      9000      9010      9020      9030

                                ScrFI >
                                MvaI >
                                MnlI ><
                                EcoRII ><
                                Ecl136I >
                                DsaV ><
                                BstOI >
                                >< NlaIV           BstNI >
                                >< FokI           BsiLI >
                                >< BscBI           ApyI >
                                >< AluI
AGTGAGCTTC GTCCAGACAC TCGTTATGTG CTTATGGATG GTTCCATCAT ACAGTTTCCT AACACTTACC
9040      9050      9060      9070      9080      9090      9100

                                >< RsaI
                                >< SfcI           >< NspI
                                >< ScaI           >< NspHI
                                >< RsaI           >< NlaIII
                                >< Csp6I           >< NlaIII
                                >< AfaI           >< Csp6I
                                >< BpmI           >< DdeI   >< AccI   >< AfaI
TGGAGGGTTC TGTTAGAGTA GTAACAACTT TTGATGCTGA GTACTGTAGA CATGGTACAT GCGAAAGGTC
9110      9120      9130      9140      9150      9160      9170

                                >< SstI
                                >< SduI
                                >< SacI
                                NspII ><
                                HgiAI ><
                                Eco24I ><
                                Bsp1286I ><

```

FIGURE 13.21

```

                                Ecl136II ><>< BmyI
                                BanII ><
                                >< Tru9I
                                >< MseI
                                >< AluI
AGAAGTAGGT ATTTGCCTAT CTACCACTGG TAGATGGGTT CTTAATAATG AGCATTACAG AGCTCTATCA
  9180      9190      9200      9210      9220      9230      9240

                                >< TfiI
                                >< HinfI
                                >< AluI
                                >< MnlI
GGAGTTTTCT GTGGTGTGGA TGCGATGAAT CTCATAGCTA ACATCTTTAC TCCTCTTGTTG CAACCTGTGG
  9250      9260      9270      9280      9290      9300      9310

                                >< MaeIII
                                HphI ><
                                > < BbvI Fnu4HI ><
>< Eco57I
GTGCTTTAGA TGTGTCTGCT TCAGTAGTGG CTGGTGGTAT TATTGCCATA TTGGTGACTT GTGCTGCCTA
  9320      9330      9340      9350      9360      9370      9380

                                >< RsaI
                                >< Csp6I
                                >< NlaIII
                                >< MaeII
                                >< BbvI
                                >< Fnu4HI
                                >< AflIII
                                >< AfaI>< HphI
                                >< BspWI
CTACTTTATG AAATTCAGAC GTGTTTTTGG TGAGTACAAC CATGTTGTTG CTGCTAATGC ACTTTTGTTC
  9390      9400      9410      9420      9430      9440      9450

                                >< RsaI
                                >< NlaIV
                                >< KpnI
                                >< Eco64I
                                > < ScrFI
                                >< Csp6I
                                > < NciI
                                >< BscBI
                                >< MspI
                                >< Asp718
                                >< HpaII
                                >< BanI >< AluI
                                >< HinfI
                                >< AfaI
                                >< HapII
                                >< PleI
                                >< AccBII
                                > < BcnI
                                > < DdeI
                                >< Acc65I
                                >< AluI>< DsaV
                                >< AccI
TTGATGTCTT TCACTATACT CTGTCTGGTA CCAGCTTACA GCTTTCTGCC GGGAGTCTAC TCAGTCTTTT
  9460      9470      9480      9490      9500      9510      9520

                                >< RsaI
                                >< Csp6I
                                >< AfaI
                                >< HphI
                                >< HphI
                                NlaIII ><
ACTTGTACTT GACATTCTAT TTCACCAATG ATGTTTCATT CTTGGCTCAC CTTCAATGGT TTGCCATGTT
  9530      9540      9550      9560      9570      9580      9590

TTCTCCTATT GTGCCTTTTT GGATAACAGC AATCTATGTA TTCTGTATTT CTCTGAAGCA CTGCCATTGG
  9600      9610      9620      9630      9640      9650      9660

                                >< TthHB8I
                                >< RsaI
                                >< MnlI
                                >< MnlI
                                >< Csp6I
                                >< Tru9I
                                >< PleI
                                >< BcgI/a
                                >< TaqI
                                >< MseI
                                >< DdeI
                                >< NlaIII
                                >< BbvI
                                >< Eco57I
                                >< BfrI
                                >< HinfI
                                >< MseI
                                >< MaeIII
                                >< AfaI
                                Fnu4HI ><
TTCTTTAACA ACTATCTTAG GAAAAGAGTC ATGTTTAATG GAGTTACATT TAGTACCTTC GAGGAGGCTG
  9670      9680      9690      9700      9710      9720      9730

                                >< RsaI
                                >< Csp6I
                                >< BcgI
                                >< RsaI
                                >< Csp6I
                                >< BsmAI

```

FIGURE 13.22


```

    >< AfaI          >< AfaI          >< Alw26I
CTTTGTGTAC CTTTTTGCTC AACAAGGAAA TGTACCTAAA ATTGCGTAGC GAGACACTGT TGCCACTTAC
  9740      9750      9760      9770      9780      9790      9800

                                >< NlaIV
                                >< RsaI
                                >< DdeI
                                >< Csp6I
                                >< BscBI
                                >< AfaI
                                >< BfrI   AluI ><
ACAGTATAAC AGGTATCTTG CTCTATATAA CAAGTACAAG TATTTTCAGTG GAGCCTTAGA TACTACCAGC
  9810      9820      9830      9840      9850      9860      9870

    >< Fnu4HI
                                >< DdeI
                                >< Fnu4HI   >< BfrI
    >< BbvI   >< AluI   >< BbvI
TATCGTGAAG CAGCTTGCTG CCACTTAGCA AAGGCTCTAA ATGACTTTAG CAACTCAGGT GCTGATGTTT
  9880      9890      9900      9910      9920      9930      9940

                                >< SfcI
                                >< PstI
                                >< BsmI
TCTACCAACC ACCACAGACA TCAATCACTT CTGCTGTTCT GCAGAGTGGT TTTAGGAAAA TGGCATTCCC
  9950      9960      9970      9980      9990      10000     10010

                                >< RsaI
                                >< NlaIII
                                >< MaeIII
                                >< Csp6I
                                >< AfaI
                                >< Tru9I
                                >< MseI
GTCAGGCAAA GTTGAAGGGT GCATGGTACA AGTAACCTGT GGAAGTACAA CTCTTAATGG ATTGTGGTTG
  10020     10030     10040     10050     10060     10070     10080

                                XhoII ><
                                Sau3AI ><
                                >< Tru9I   NdeII ><
                                >< NspI     MflI  ><
                                >< NspHI    MboI  ><
                                >< NlaIII   DpnII ><
                                >< MseI     BstYI ><
                                >< MboII   BspAI ><
                                >< AccI     >< AflIII
                                >< Bst1107I >< NlaIII
                                >< FokI     >< NspHI
                                >< Bsp143I
                                >< BbsI     BglII ><
GATGACACAG TATACTGTCC AAGACATGTC ATTTGCACAG CAGAAGACAT GCTTAATCCT AACTATGAAG
  10090     10100     10110     10120     10130     10140     10150

                                PalI >
                                MscI >
                                HaeIII >
                                EaeI ><
                                BsuRI >
                                BshI >
                                BalI >
    >< DpnI >< MboII
    >< Bsp143I
ATCTGCTCAT TCGCAAATCC AACCATAGCT TTCTTGTTCA GGCTGGCAAT GTTCAACTTC GTGTTATTGG
  10160     10170     10180     10190     10200     10210     10220

                                >< DdeI> < Tru9I
                                >< BfrI> < MseI
                                >< DdeI
CCATTCTATG CAAAATTGTC TGCTTAGGCT TAAAGTTGAT ACTTCTAACC CTAAGACACC CAAGTATAAA
  10230     10240     10250     10260     10270     10280     10290

    >< ScrFI
    >< MvaI
    >< EcoRII
    >< Ecl136I
                                >< SphI

```

FIGURE 13.23

```

>< DsaV
>< BstOI
>< BstNI
>< BsiI
>< ApyI
TTTGTCCGTA TCCAACCTGG TCAACATTT TCAGTTCTAG CATGCTACAA TGGTTCACCA TCTGGTGTTC
10300 10310 10320 10330 10340 10350 10360

>< PaeI
>< NspI
>< NspHI
>< RmaI >< NlaIII
>< MaeI >< HphI
>< Sau3AI
>< NdeII
>< MboI>< NlaIII
>< DpnII
>< Eco31I
>< BsmAI
>< BsaI>< NlaIII
>< Alw26I
ATCAGTGTGC CATGAGACCT AATCATACCA TTAAAGGTTT TTTCTTAAT GGATCATGTG GTAGTGTTCG
10370 10380 10390 10400 10410 10420 10430

>< Zsp2I
>< Ppu10I
>< NsiI>< SfaNI
>< NdeI
>< Mph1103I
>< EcoT22I
>< Tru9I
>< MseI
>< Mph1103I
>< EcoT22I
>< AvaIII
>< AluI
>< RsaI ><
>< Csp6I ><
>< AfaI ><
TTTTAACATT GATTATGATT GCGTGTCTTT CTGCTATATG CATCATATGG AGCTTCCAAC AGGAGTACAC
10440 10450 10460 10470 10480 10490 10500

>< SinI
>< Sau96I
>< NspIV
>< NspHII
>< Eco47I
>< Cfr13I
>< BsiZI
>< Bme18I
>< Bsp6I>< DdeI
>< AfaI>< BfrI
>< HindII
>< HincII
>< BspWI
>< SfcI
>< RsaI ><
>< PstI ><
>< Fnu4HI
>< Csp6I ><
>< BspMI
>< AfaI ><
GCTGGTACTG ACTTAGAAGG TAAATTCTAT GGTCCATTTG TTGACAGACA AACTGCACAG GCTGCAGGTA
10510 10520 10530 10540 10550 10560 10570

>< Tru9I
>< MseI
>< BbvI
>< Fnu4HI
>< HphI ><
CAGACACAAC CATAACATTA AATGTTTTGG CATGGCTGTA TGCTGCTGTT ATCAATGGTG ATAGGTGGTT
10580 10590 10600 10610 10620 10630 10640

>< Tru9I
>< TfiI
>< MseI
>< HphI
>< HinfI
>< Tru9I
>< MseI
>< RsaI
>< Csp6I
>< AfaI
TCTTAATAGA TTCACCACTA CTTTGAATGA CTTTAACCTT GTGGCAATGA AGTACAATA TGAACCTTTG
10650 10660 10670 10680 10690 10700 10710

>< SinI
>< Sau96I
>< PssI
>< Psp5II
>< PpuMI
>< NspIV
>< NspHII
>< NlaIV

```

FIGURE 13. 24

```

                                >< EcoO109I
                                >< Eco47I
                                >< DraII
>< Sau3AI                      >< Cfr13I
>< NdeII                      >< BsiZI
>< MboI                       >< BscBI
>< DpnII>< NlaIII             >< BmeI8I
                                >< DdeI
                                >< BfrI
>< BspAI >< HincII           >< BbvI
>< Bsp143I                    >< MnlI
ACACAAGATC ATGTTGACAT ATTGGGACCT CTTTCTGCTC AAACAGGAAT TGCCGTCTTA GATATGTGTG
10720      10730      10740      10750      10760      10770      10780

                                >< StyI
                                >< RsaI
                                >< EcoT14I
                                >< Eco130I
                                > < Csp6I
                                >< BssT1I
                                >< BsaJI
>< Fnu4HI                      >< Fnu4HI
>< BbvI                        >< Fnu4HI
>< BbvI                        >< AluI >< PstI
CTGCTTTGAA AGAGCTGCTG CAGAATGGTA TGAATGGTCG TACTATCCTT GGTCAGCACTA TTTTAGAAGA
10790      10800      10810      10820      10830      10840      10850

                                >< StyI
                                >< EcoT14I
                                >< Eco130I
                                >< BssT1I
                                >< BsaJI
>< MboII                      > < MaeIII>< BsaJI
TGAGTTTACA CCATTTGATG TTGTTAGACA ATGCTCTGGT GTTACCTTCC AAGGTAAGTT CAAGAAAATT
10860      10870      10880      10890      10900      10910      10920

                                >< SfaNI
                                > < SduI
                                > < NspII
                                >< Tru9I
                                >< MseI
                                >< Bsp1286I
                                >< TfiI
                                >< Csp6I
                                >< RsaI
                                >< HinfI
                                >< AfaI
GTTAAGGGCA CTCATCATTTG GATGCTTTTA ACTTTCTTGA CATCACTATT GATTCTTGTT CAAAGTACAC
10930      10940      10950      10960      10970      10980      10990

                                >< XmnI
                                >< BsmI
                                >< BscCI
                                >< Asp700I
                                >< BbvI
                                >< BbvI
ATGGGTCACT GTTTTCTTTT GTTTACGAGA ATGCTTTCTT GCCATTTACT CTTGGTATTA TGGCAATTGC
11000      11010      11020      11030      11040      11050      11060

                                >< NspI
                                >< NspHI
                                >< NlaIII
                                >< BspWI
                                >< Fnu4HI>< BspWI
                                >< BscCI
                                >< MaeIII
TGCATGTGCT ATGCTGCTTG TTAAGCATAA GCACGCATTC TTGTGCTTGT TTCTGTTACC TTCTCTTGCA
11070      11080      11090      11100      11110      11120      11130

                                >< SfaNI
                                >< RmaI
                                > < NspI
                                > < NlaIII
                                >< NheI
                                >< MaeI
                                >< BsiBI
                                >< NlaIII
                                >< BspWI
                                >< MseI
                                >< AccI> < NspHI>< AluI
                                >< BsaBI >< NlaIII
ACAGTTGCTT ACTTTAATAT GGTCTACATG CCTGCTAGCT GGGTGATGCG TATCATGACA TGGCTTGAAT
11140      11150      11160      11170      11180      11190      11200

```

FIGURE 13.25

```

                                >< Tru9I
                                >< MseI
                                >< Esp4I
                                >< Eco57I
                                >< AluI
                                >< AflIII
                                >< AluI
TGGCTGACAC TAGCTTGTCT GGTATAGGC TTAAGGATTG TGTTATGTAT GCTTCAGCTT TAGTTTGTCT
11210      11220      11230      11240      11250      11260      11270

                                >< RmaI
                                >< MaeII
                                >< MaeI
                                >< Fnu4HI
                                >< AflIII
                                >< BspHI >< AluI >< BbvI >< AflIII
TATTCTCATG ACAGCTCGCA CTGTTTATGA TGATGCTGCT AGACGTGTTT GGACACTGAT GAATGTCATT
11280      11290      11300      11310      11320      11330      11340

                                >< Sau96I
                                >< PstI
                                >< NspIV
                                >< NlaIII
                                >< HaeIII
                                >< DdeI
                                >< Cfr13I
                                >< BsuRI
                                >< BsiZI
                                >< BshI
                                >< BfrI
                                >< Bsp143I
                                >< BspAI>< AluI >< AsuI
                                >< AccI
ACACTTGTTT ACAAAGTCTA CTATGGTAAT GCTTTAGATC AAGCTATTTC CATGTGGGCC TTAGTTATTT
11350      11360      11370      11380      11390      11400      11410

                                >< RmaI
                                >< NlaIII
                                >< MaeI>< SfcI
                                >< AluI>< AluI
                                >< MaeIII
                                >< MnlI >< MaeIII
                                >< BspHI >< BspWI
CTGTAACTCT TAACTATTCT GGTGTCGTTA CGACTATCAT GTTTTATAGT AGAGCTATAG TGTGTGTGTG
11420      11430      11440      11450      11460      11470      11480

                                >< BsrI
                                >< NlaIII BfrI >
                                >< DdeI >
TGTTGAGTAT TACCCATTGT TATTTATTAC TGGCAACACC TTACAGTGTA TCATGCTTGT TTATTGTTTC
11490      11500      11510      11520      11530      11540      11550

                                >< PstI
                                >< HaeIII
                                >< Fnu4HI >< BsuRI
                                >< BbvI >< Fnu4HI >< BspWI
                                >< BbvI >< BspWI >< BshI >< Eco57I >< MaeIII
TTAGGCTATT GTTGCTGCTG CTACTTTGGC CTTTCTGTG TACTCAACCG TTACTTCAGG CTTACTCTTG
11560      11570      11580      11590      11600      11610      11620

                                >< ScrFI
                                >< MvaI
                                >< EcoRII
                                >< Ecl136I
                                >< DsaV
                                >< BstOI
                                >< BstNI
                                >< BsiLI
                                >< BsaJI
                                >< BsaJI
                                >< Eco31I
                                >< BsmAI
                                >< BsaI

```

FIGURE 13.26

```

                >< DrdI   >< Alw26I
GTGTTTATGA CTACTTGGTC TCTACACAAG AATTTAGGTA TATGAACTCC >< ApyI   DdeI ><
11630      11640      11650      11660      11670      11680      11690

                >< Tru9I
                >< MseI
>< SfaNI      > < HindIII> < Tru9I
>< MnlI      >< AluI > < MseI > < MnlI      > < NlaIII
GAGTAGTATT GATGCTTTCA AGCTTAACAT TAAGTTGTTG GGTATTGGAG GTAAACCATG TATCAAGGTT
11700      11710      11720      11730      11740      11750      11760

                >< VneI
                >< SnoI
                >< SduI
                >< NspII
                >< HgiAI
                >< Bsp1286I
                >< BmyI   >< RsaI
                >< RsaI      >< ApaLI      >< MboII
>< Csp6I      >< Alw44I   >< Csp6I      DdeI >
>< AfaI      >< MaeII   >< Alw21I >< AfaI      BfrI >
GCTACTGTAC AGTCTAAAAT GTCTGACGTA AAGTGACAT CTGTGGTACT GCTCTCGGTT CTTCAACAAC
11770      11780      11790      11800      11810      11820      11830

                >< NspII> < RsaI
                >< DraIII
                >< SduI>< Csp6I
                >< Bsp1286I
>< MboII      >< BmyI > < AfaI      >< MboII
>< HinfI >< PleI      CACAATGTGT ACAACTCCAC AATGATATTC TTCTTGCAAA
TTAGAGTAGA GTCATCTTCT AAATTGTGGG
11840      11850      11860      11870      11880      11890      11900

                >< TthHB8I
                >< TaqI
                >< HindIII      >< MboII      SfcI ><
                >< AluI      > < Eco57I      >< NlaIII
AGACACAAC TGAAGCTTTTCG AGAAGATGGT TTCTCTTTTG TCTGTTTTGC TATCCATGCA GGGTGCTGTA
11910      11920      11930      11940      11950      11960      11970

>< VspI
>< Tru9I
>< MseI      >< TthHB8I      > < Ksp632I
>< AsnI      >< TaqI >< MboII      > < EarI
>< AseI>< MnlI >< BcgI/a   >< Eco57I   >< Eco57I >< BcgI
GACATTAATA GGTGTGCGA GGAAATGCTC GATAACCGTG CTACTCTTCA GGCTATTGCT TCAGAATTTA
11980      11990      12000      12010      12020      12030      12040

                >< StuI
                >< ScrFI
                >< Pali
                >< MvaI>< HaeIII
>< EcoRII>< Eco147I
>< Ecl136I
>< DsaV >< BsuRI
>< BstOI
>< BstNI
>< BspWI
>< BsiLI
>< Fnu4HI      >< BsaJI >< BshI      TfiI ><
>< NdeI      >< BspWI>< MnlI >< BglI      >< SfcI HinfI ><
>< AciI      >< ApyI>< AatI      > < AluI

```

FIGURE 13. 27

```

GTTCTTTACC ATCATATGCC GCTTATGCCA CTGCCCAGGA GGCCTATGAG CAGGCTGTAG CTAATGGTGA
12050      12060      12070      12080      12090      12100      12110

      >< XmnI      >< Tru9I      >< SfaNI
      >< HphI      >< MseI      >< DdeI
      >< Asp700I   >< Eco57I   >< BbvI Fnu4HI ><
TTCTGAAGTC GTTCTCAAAA AGTTAAAGAA ATCTTTGAAT GTGGCTAAAT CTGAGTTTGA CCGTGATGCT
12120      12130      12140      12150      12160      12170      12180

                                XhoII ><
                                Sau3AI ><
                                NdeII ><
                                MnlI >
                                >< MnlI
                                >< MflI
                                >< MboI
      > < Sau3AI
      > < NdeII      DpnII ><
      > < MboI      DpnI ><
      > < DpnII      DdeI ><
      >< DpnI      BstYI ><
      >< BspWI      >< RsaIBspAI ><
      > < BspAI      >< Csp6IBsp143I ><
      >< Bsp143I      >< AfaIBglIII ><
      >< NlaIII
GCCATGCAAC GCAAGTTGGA AAAGATGGCA GATCAGGCTA TGACCCAAAT GTACAAACAG GCAAGATCTG
12190      12200      12210      12220      12230      12240      12250

                                >< SpeI
                                >< RmaI
      >< MaeIII      >< MboII      >< Eam1104I >< BspWI
      >< MaeI      >< BspWI      >< EarI>< BfrI >< AluI
AGGACAAGAG GGCAAAAGTA ACTAGTGCTA TGCAAACAAT GCTCTTCACT ATGCTTAGGA AGCTTGATAA
12260      12270      12280      12290      12300      12310      12320

                                >< ThaI
                                >< MvnI
      >< HinPII
      >< Hin6I
      >< HhaI
      >< CfoI
      >< BstUI
      >< Bsp50I
      >< Tru9I
      >< MseI      >< AccII      SfcI ><
TGATGCACTT AACACATTA TCAACAATGC GCGTGATGGT TGTGTTCCAC TCAACATCAT ACCATTGACT
12330      12340      12350      12360      12370      12380      12390

                                >< RsaI
                                >< NlaIV
                                >< Eco64I
                                >< Csp6I
      >< BslI
      >< BsiYI>< KpnI
      >< BscBI
      >< BanI
      >< Asp718
      >< NlaIII
      >< BstXI
      >< AccB1I      >< MaeIII
      >< Fnu4HI >< BbvI      >< Acc65I      BsgI ><
ACAGCAGCCA AACTCATGGT TGTTGTCCCT GATTATGGTA CCTACAAGAA CACTTGTGAT GGTAACACCT
12400      12410      12420      12430      12440      12450      12460

      >< Zsp2I
      >< Ppu10I

```

FIGURE 13. 28

```

    >< NsiI
    >< Mph1103I
    >< NdeI>< EcoT22I
    >< AvaIII >< SfaNI
    >< SfaNI
    >< AciI
    DdeI ><
    BfrI ><
    TTACATATGC ATCTGCACTC TGGGAAATCC AGCAAGTTGT TGATGCGGAT AGCAAGATTG TTCAACTTAG
    12470      12480      12490      12500      12510      12520      12530

    >< PalI
    >< HaeIII >< MnlI >< DdeIDdeI ><
    >< BsuRI >< MaeIII >< BspWI
    >< Tru9I>< NlaIII
    >< MseI>< HphI >< XcmI>< BshI >< AluI BspWI ><
    TGAAATTAAC ATGGACAATT CACCAAATTT GGCTTGGCCT CTTATTGTTA CAGCTCTAAG AGCCAACTCA
    12540      12550      12560      12570      12580      12590      12600

    RsaI ><
    NlaIV ><
    KpnI ><
    >< Fnu4HI
    Eco64I ><
    Csp6I ><
    BscBI ><
    Asp718 ><
    AfaI ><
    >< AciI>< Bani
    AccB1I ><
    >< AluI >< SfcI >< DdeI>< BsrI >< PshAI Acc65I ><
    GCTGTAAAC TACAGAATAA TGAAGTGAAGT CCAGTAGCAC TACGACAGAT GTCCTGTGCG GCTGGTACCA
    12610      12620      12630      12640      12650      12660      12670

    >< TthHB8I
    >< TaqI
    >< SfuI
    >< NspV
    >< MnlI
    >< LspI
    >< Csp45I
    >< BstBI
    >< Bsp119I
    >< BsiCI
    >< Bpu14I
    >< AsuII
    CACAAACAGC TTGTACTGAT GACAATGCAC TTGCCTACTA TAACAATTTCG AAGGGAGGTA GGTTCGTGCT
    12680      12690      12700      12710      12720      12730      12740

    >< XhoII
    >< Sau3AI
    >< NdeII
    >< MflI
    >< MboI
    >< DpnII
    >< DpnI
    >< BstYI >< TfiI >< RsaI
    >< BspAI >< RmaI >< Csp6I
    >< Bsp143I >< HinfI >< Csp6I>< RsaI
    >< BglII >< MaeI >< DdeI >< AfaI>< AfaI
    GGCATTACTA TCAGACCACC AAGATCTCAA ATGGGCTAGA TTCCCTAAGA GTGATGGTAC AGGTACAATT
    12750      12760      12770      12780      12790      12800      12810

    >< Sau96I
    >< PssI
    >< Pali
    >< NspIV

```

FIGURE 13.29

```

                                >< HaeIII
                                >< EcoO109I
                                >< DraII
                                >< Cfr13I
                                >< BsuRI
                                >< BsiZI
                                >< BshI
                                >< AsuI
                                RsaI >
                                Csp6I ><
                                AfaI >
TACACAGAAC TGGAACCACC TTGTAGGTTT GTTACAGACA CACCAAAAGG GCCTAAAGTG AAATACTTGT
12820      12830      12840      12850      12860      12870      12880

                                >< SfcI
                                > < MboII
                                MaeII ><
                                >< Fnu4HI >< RsaI
                                >< Eco57I >< Csp6I
                                > < BbsI
                                >< Tru9I
                                >< MseI >< MnlI
                                >< BbvI
                                >< AluI
                                >< AfaI
ACTTCATCAA AGGCTTAAAC AACCTAAATA GAGGTATGGT GCTGGGCAGT TTAGCTGCTA CAGTACGTCT
12890      12900      12910      12920      12930      12940      12950

                                >< RsaI
                                >< SfcI >< Csp6I
                                >< BspWI >< AfaI
                                >< BspMI
                                AccI ><
TCAGGCTGGA AATGCTACAG AAGTACCTGC CAATTCAACT GTGCTTTCCT TCTGTGCTTT TGCAGTAGAC
12960      12970      12980      12990      13000      13010      13020

                                >< RmaI
                                >< MnlI
                                >< MaeI
                                >< HphI
CCTGCTAAAG CATATAAGGA TTACCTAGCA AGTGGAGGAC AACCAATCAC CAACTGTGTG AAGATGTTGT
13030      13040      13050      13060      13070      13080      13090

                                >< SinI
                                >< Sau96I
                                >< NspIV
                                >< NspHII
                                >< NlaIII
                                >< Eco47I
                                >< Eam1105I
                                >< Cfr13I
                                >< BsiZI
                                >< Bme18I >< XcmI
                                >< AvaII PleI ><
                                >< MaeIII
                                >< AluI >< AsuI >< HinfI
GTACACACAC TGGTACAGGA CAGGCAATTA CTGTAACACC AGAAGCTAAC ATGGACCAAG AGTCCTTTGG
13100      13110      13120      13130      13140      13150      13160

                                >< TfiI
                                >< MaeIII
                                >< SfaNI
                                >< NlaIII
                                >< FokI
                                >< HinfI
TGGTGCTTCA TGTTGTCTGT ATTGTAGATG CCACATTGAC CATCCAAATC CTAAAGGATT CTGTGACTTG
13170      13180      13190      13200      13210      13220      13230

                                > < RsaI
                                >< MaeII
                                >< Csp6I
                                > < AfaI
                                >< BsrI
                                >< DdeI
                                >< BfrI
AAAGGTAAGT ACGTCCAAAT ACCTACCACT TGTGCTAATG ACCCAGTGGG TTTTACACTT AGAAACACAG
13240      13250      13260      13270      13280      13290      13300

                                >< ThaI

```

FIGURE 13.30


```

                                >< SfaNI
                                >< MvnI
                                >< BstUI
                                >< Bsp50I
                                >< AciI
                                >< AccIISfaNI ><
>< RsaI
>< Csp6I
>< AfaI >< AciI                                >< SfcI >< MaeIII
TCTGTACCGT CTGCGGAATG TGGAAAGGTT ATGGCTGTAG TTGTGACCAA CTCCGCGAAC CCTTGATGCA
13310      13320      13330      13340      13350      13360      13370

                                >< Zsp2I
                                > < SfaNI
                                >< Mph1103I>< Tru9I
>< Ppu10I>< MaeII                                Fnu4HI ><
                                BsgI ><
                                >< BbvI
>< AciI>< AvaIII >< DraI >< AciI >< Fnu4HI AciI ><
GTCTGCGGAT GCATCAACGT TTTTAAACGG GTTTGGCGTG TAAGTGCAGC CCGTCTTACA CCGTGCGGCA
13380      13390      13400      13410      13420      13430      13440

>< SpeI
>< ScaI
>< RsaI
>< RmaI
>< MaeI
> < Csp6I >< SfcI                                >< BspWI
>< BspWI >< AfaI >< AccI >< BcgI/a BcgI >
CAGGCACTAG TACTGATGTC GTCTACAGGG CTTTGTATAT TTACAACGAA AAAGTTGCTG GTTTTGCAAA
13450      13460      13470      13480      13490      13500      13510

                                >< ScrFI
                                >< MvaI
                                >< MnlI
>< EcoRII
>< Ecl136I
>< BstOI
>< BstNI
>< BslI
>< DsaV >< BsiYI
>< BsiLI                                >< PleI
>< ApyI > < FokI >< HinfI
GTTCTTAAAA ACTAATTGCT GTCGCTTCCA GGAGAAGGAT GAGGAAGGCA ATTTATTAGA CTCTTACTTT
13520      13530      13540      13550      13560      13570      13580

                                >< NlaIII
                                >< Ksp632I
                                >< EarI
>< Tru9I                                >< Eam1104I
>< MseI                                >< BsmAI >< Tru9I
>< MnlI                                >< Alw26I >< MboII >< MseI
GTAGTTAAGA GGCATACTAT GTCTAACTAC CAACATGAAG AGACTATTTA TAACTTGGTT AAAGATTGTC
13590      13600      13610      13620      13630      13640      13650

                                >< RsaI
                                >< NlaIV
> < NlaIII
                                >< KpnI
                                >< HphI
> < Eco64I
>< Csp6I
>< BscBI
> < BanI
> < Asp718

```

FIGURE 13.31

```

                                >< MaeIII >< AfaI
                                > < AccBII MaeII ><
>< NspBII                                >< NlaIII                                > < Acc65I > < HgaI
>< AciI
CAGCGGTTGC TGTCCATGAC TTTTCAAGT TTAGAGTAGA TGGTGACATG GTACCACATA TATCACGTCA
13660      13670      13680      13690      13700      13710      13720

                                >< MnlI
                                >< MaeII
GCGTCTAACT AAATACACAA TGGCTGATTT AGTCTATGCT CTACGTCATT TTGATGAGGG TAATTGTGAT
13730      13740      13750      13760      13770      13780      13790

>< Tru9I
>< MseI                                >< MaeIII >< MunI
ACATTAAAAG AAATACTCGT CACATACAAT TGCTGTGATG ATGATTATTT CAATAAGAAG GATTGGTATG
13800      13810      13820      13830      13840      13850      13860

                                >< ThaI
                                >< MvnI
                                >< MluI
                                >< BstUI
                                >< Bsp50I
                                >< RsaI
                                >< HphI
>< TfiI                                >< AflIII                                >< DdeI                                >< Csp6I Tru9I ><
>< HinfI                                >< AccII                                >< BfrI                                >< AfaI MseI ><
ACTTCGTAGA GAATCCTGAC ATCTTACGCG TATATGCTAA CTTAGGTGAG CGTGTACGCC AATCATTATT
13870      13880      13890      13900      13910      13920      13930

                                XhoII >
                                Sau3AI >
                                NdeII >
                                MflI >
                                > < SfaNI
                                >< RsaI
                                >< Csp6I
                                >< BspWI
                                >< AfaI
                                >< BspAI >
>< RsaI
>< Csp6I
>< AfaI
AAAGACTGTA CAATTCTGCG ATGCTATGCG TGATGCAGGC ATTGTAGGCG TACTGACATT AGATAATCAG
13940      13950      13960      13970      13980      13990      14000

                                > < ScrFI
                                > < MvaI
                                >< Fnu4HI
                                >< EcoRII
                                > < Ecl136I
                                > < BstOI
                                > < BstNI
>< Tru9I                                >< RsaI                                >< BslI
>< MseI                                >< RsaI                                > < HphI                                >< BsiYI
>< DpnI                                >< Csp6I                                >< Csp6I                                > < BsiLI
>< Bsp143I                                >< BsrI                                > < BbvI                                > < ApyI
                                >< AlwI                                >< AfaI                                >< DsaV >< AciI
GATCTTAATG GGAAGTGGTA CGATTTCGGT GATTTCGTAC AAGTAGCACC AGGCTGCGGA GTTCCTATTG
14010      14020      14030      14040      14050      14060      14070

                                >< SfaNI
                                >< RmaI                                > < HinfI
                                >< MnlI                                >< Fnu4HIpleI ><
>< TfiI                                >< SfaNI                                >< BsiBI                                >< MaeI
>< HinfI                                >< FokI                                >< BsaBI                                >< BbvI                                >< BspWI NdeI ><
TGGATTGATA TTACTCATTG CTGATGCCCA TCCTCACTTT GACTAGGGCA TTGGCTGCTG AGTCCCATAT
14080      14090      14100      14110      14120      14130      14140

>< Sau3AI
>< NdeII

```

FIGURE 13.32

```

    >< MboI
    >< MamI
    >< DpnII
    >< DpnI
    >< BspWI
    >< BspAI
    >< BspI43I
    >< BsiBI
    >< BsaBI >< FokI
    >< XcmI
    >< Tru9I
    >< MseI
    Tth111I ><
    MboII ><
    >< Ksp632I
    >< Eam1104I
    >< BsmAI
    >< EarI AspI ><
    >< Alw26I
    GGATGCTGAT CTCGCAAAAC CACTTATTAA GTGGGATTTG CTGAAATATG ATTTTACGGA AGAGAGACTT
    14150      14160      14170      14180      14190      14200      14210

    > < SinI
    > < Sau96I
    > < NspIV
    >< NspHII
    >< NlaIV
    >< TthHB8I
    >< TaqI
    >< McrI
    > < Ksp632I
    > < EarI
    > < Eam1104I
    >< BsmAI
    > < Tru9I
    >< BsiEI> < MseI
    >< Alw26I
    >< DraI
    >< FokI
    > < Eco47I
    > < Cfr13I
    > < BsiZI
    >< SspI>< BscBI
    > < Bmel8I
    >< MboII
    >< BsiEI> < MseI
    >< Alw26I
    >< DraI
    >< AsuI
    >< Tru9I
    >< MunI
    >< MseI
    TGTCTCTTCG ACCGTTATTT TAAATATTGG GACCAGACAT ACCATCCCAA TTGTATTAAC TGTTCGGATG
    14220      14230      14240      14250      14260      14270      14280

    SinI ><
    Sau96I ><
    NspIV ><
    NspHII >
    Eco47I ><
    Cfr13I ><
    BsiZI ><
    Bmel8I ><
    AvaII ><
    AsuI ><
    >< FokI
    >< MseI
    ATAGGTGTAT CCTTCATTGT GCAAACTTTA ATGTGTTATT TTCTACTGTG TTTCCACCTA CAAGTTTTGG
    14290      14300      14310      14320      14330      14340      14350

    >< SpeI
    >< RmaI
    >< MaeI
    >< SspI
    >< BsrI
    ACCACTAGTA AGAAAAATAT TTGTAGATGG TGTTCCTTTT GTTGTTCCTCAA CTGGATACCA TTTTCGTGAG
    14360      14370      14380      14390      14400      14410      14420

    >< RsaI
    >< HinfI >< PfuI
    > < Csp6I
    >< AfaI
    >< HgaI>< AluI
    >< FokI
    >< AccII
    >< BstUI
    >< Bsp50I
    >< MvnI>< BsmAI
    >< BsmBI
    >< Alw26I
    > < BbvI
    TTAGGAGTCG TACATAATCA GGATGTAAAC TTACATAGCT CGCGTCTCAG TTTCAAGGAA CTTTTAGTGT
    14430      14440      14450      14460      14470      14480      14490

    >< Zsp2I
    >< SphI
    >< Ppu10I
    >< PaeI
    >< NspI

```

FIGURE 13.33

```

    >< Sau3AI          >< NspHI
    >< NdeII           >< NsiI
    >< MboI             >< NlaIII
    >< DpnII           >< Mph1103I
    > < DpnI          >< Fnu4HI
    >< Fnu4HI>< BspWI  >< EcoT22I
    >< BspAI           >< BspWI
    > < Bsp143I> < AvaIII > < AlwNI
    >< AlwI           >< AluI   >< AluI   >< BbvI   >< MaeI
    ATGCTGCTGA TCCAGCTATG CATGCAGCTT CTGGCAATTT ATTGCTAGAT AAACGCACTA CATGCTTTTC
    14500      14510      14520      14530      14540      14550      14560

    >< ScrFI
    >< NciI
    >< MspI
    >< HpaII
    >< HapII
    >< Fnu4HI
    >< AlwNI
    >< AluI
    >< DsaV           >< Tru9I
    >< BcnI           >< MseI
    AGTAGCTGCA CTAACAAACA ATGTTGCTTT TCAAAGTCTC AAACCCGGTA ATTTTAATAA AGACTTTTAT
    14570      14580      14590      14600      14610      14620      14630

    >< Tru9I
    >< MseI
    >< MboII
    GACTTTGCTG TGTCTAAAGG TTTCTTTAAG GAAGGAAGTT CTGTTGAACT AAAACACTTC TTCTTTGCTC
    14640      14650      14660      14670      14680      14690      14700

    >< FokI
    >< Fnu4HI
    AGGATGGCAA CGCTGCTATC AGTGATTATG ACTATTATCG TTATAATCTG CCAACAATGT GTGATATCAG
    14710      14720      14730      14740      14750      14760      14770

    >< VspI
    >< Tru9I
    >< MseI
    >< AsnI
    >< AseI
    >< MaeIII
    ACAACTCCTA TTCGTAGTTG AAGTTGTTGA TAAATACTTT GATTGTTACG ATGGTGGCTG TATTAATGCC
    14780      14790      14800      14810      14820      14830      14840

    >< Tru9I
    >< MseI
    >< PvuII
    >< HpaI
    >< Psp5I
    > < XcmI
    >< HindII
    >< NspBII
    >< Tru9I
    >< RmaI ><
    >< HincII
    >< AluI
    >< MseI
    >< MaeI ><
    AACCAAGTAA TCGTTAACAA TCTGGATAAA TCAGCTGGTT TCCCATTTAA TAAATGGGGT AAGGCTAGAC
    14850      14860      14870      14880      14890      14900      14910

    >< SfaNI
    >< Sau3AI
    >< NdeII
    >< MboI
    >< DpnII
    >< DpnI
    >< Bsp143I
    >< BspAI
    >< AlwI
    >< PleI
    >< HinfI>< MnlI
    >< BspAI
    >< AlwI
    TTTATTATGA CTCAATGAGT TATGAGGATC AAGATGCACT TTTCGCGTAT ACTAAGCGTA ATGTCATCCC
    14920      14930      14940      14950      14960      14970      14980

    >< SstI
    >< SduI
    >< SacI

```

FIGURE 13.34

```

                                >< NspII
                                >< HgiAI
                                >< Eco24I
                                > < Ecl136II
                                >< Bsp1286I
                                >< BmyI
                                >< BanII
                                >< Alw21I
                                >< BspWI
                                > < AluI
                                >< AluI
TACTATAACT CAAATGAATC TTAAGTATGC CATTAGTGCA AAGAATAGAG CTCGCACCGT AGCTGGTGTG
14990      15000      15010      15020      15030      15040      15050

                                >< ScaI
                                >< SfcI>< RsaI
                                >< BsmAI >< Csp6I
                                >< Alw26I >< AfaI
                                >< RmaI ><
                                > < MnlI
                                >< MaeI ><
                                >< Fnu4HI
                                >< AciI
TCTATCTGTA GTACTATGAC AAATAGACAG TTTCATCAGA AATTATTGAA GTCAATAGCC GCCACTAGAG
15060      15070      15080      15090      15100      15110      15120

                                >< Tru9I
                                >< MseI
>< AluI
GAGCTACTGT GGTAATTGGA ACAAGCAAGT TTTACGGTGG CTGGCATAAT ATGTTAAAA CTGTTTACAG
15130      15140      15150      15160      15170      15180      15190

                                >< NspI ><
                                >< NspHI ><
                                >< NlaIII ><
                                >< NlaIII
                                >< DdeI ><
                                >< BspWI ><
                                >< MaeIII
                                >< BfrI ><
TGATGTAGAA ACTCCACACC TTATGGGTTG GGATTATCCA AAATGTGACA GAGCCATGCC TAACATGCTT
15200      15210      15220      15230      15240      15250      15260

                                > < Pali
                                > < HaeIII
                                > < BsuRI
                                > < BshI
                                >< MnlI
                                >< MaeIII
                                >< SfcI ><
AGGATAATGG CCTCTCTTGT TCTTGCTCGC AAACATAACA CTTGCTGTAA CTTATCACAC CGTTTCTACA
15270      15280      15290      15300      15310      15320      15330

                                >< Tru9I ><
                                >< ScrFI >
                                >< MvaI >
                                >< MseI
                                >< FokI ><
                                >< EcoRII ><
                                >< Ecl136I >
                                >< DsaV ><
                                >< BstOI >
                                >< BstNI >
                                >< BsiLI >
                                >< ApyI >
                                >< NlaIII
                                > < Fnu4HI
                                >< AciI
                                >< AluI
                                >< AviII >< MseI
GGTTAGCTAA CGAGTGTGCG CAAGTATTAA GTGAGATGGT CATGTGTGGC GGCTCACTAT ATGTTAAACC
15340      15350      15360      15370      15380      15390      15400

                                > < SfaNI
                                >< MspI
                                >< HpaII
                                >< HapII
                                >< HphI
                                >< BspWI
                                >< Tru9I
                                >< MaeIII ><
                                >< MseI
                                >< AluI ><

```

FIGURE 13.35

```

AGGTGGAACA TCATCCGGTG ATGCTACAAC TGCTTATGCT AATAGTGTCT TTAACATTTG TCAAGCTGTT
15410      15420      15430      15440      15450      15460      15470

                                >< DrdI
>< BspWI                                >< AluI                                > < AciI
ACAGCCAATG TAAATGCACT TCTTTCAACT GATGGTAATA AGATAGCTGA CAAGTATGTC CGCAATCTAC
15480      15490      15500      15510      15520      15530      15540

                                >< Sau3AI
                                >< NdeII
                                >< MboI
                                > < MamI
                                >< FbaI
                                >< DpnII
                                >< DpnI
                                >< BspHI
                                >< BspAI
                                >< Bsp143I
                                >< BsiQI
                                >< BsiBI>< NlaIII
                                >< BsmAI                                > < BsaBI>< FokI
                                >< Alw26I                                >< BclI>< EcoRI                                FokI ><
AACACAGGCT CTATGAGTGT CTCTATAGAA ATAGGGATGT TGATCATGAA TTCGTGGATG AGTTTACGC
15550      15560      15570      15580      15590      15600      15610

                                >< TfiI
                                >< SfaNI
                                >< NlaIII
                                >< BspMI                                >< MaeIII
TTACCTGCGT AAACATTTCT CCATGATGAT TCTTTCTGAT GATGCCGTTG TGTGCTATAA CAGTAACAT
15620      15630      15640      15650      15660      15670      15680

                                > < RmaI
                                >< NheI >< Tru9I
                                > < MaeI                                >< Tru9I
>< Fnu4HI                                >< AluI >< MseI                                >< MseI                                MnlI ><
GCGGCTCAAG GTTTAGTAGC TAGCATTAAG AACTTTAAGG CAGTTCTTTA TTATCAAAAT AATGTGTTCA
15690      15700      15710      15720      15730      15740      15750

                                >< SinI
                                >< Sau96I
                                >< PssI
                                >< Psp5II
                                >< PpuMI
                                >< NspIV
                                >< NspHII
                                >< EcoO109I
                                >< Eco47I
                                >< DraII
                                >< Cfr13I
                                >< Bsi2I
                                >< Bme18I
                                >< AvaII
                                >< AsuI                                >< MnlI
>< NlaIII                                >< BsmAI                                >< DdeI
>< DdeI                                >< Alw26I
TGTCTGAGGC AAAATGTTGG ACTGAGACTG ACCTTACTAA AGGACCTCAC GAATTTTGCT CACAGCATAC
15760      15770      15780      15790      15800      15810      15820

                                >< XhoII
                                >< Sau3AI
                                >< NdeII
                                >< MflI
                                >< MboI

```

FIGURE 13. 36

```

                                >< RsaI          >< DpnII
                                >< MaeII          >< DpnI          > < SspI
                                >< Csp6I          >< BstYI          HinPI ><
                                >< BsaAI          >< BspMI          Hin6I ><
                                >< AflIII         >< BspAI          HhaI ><
>< BspWI>< MseI                >< AfaI          >< AlwI>< Bsp143I    CfoI ><
AATGCTAGTT AAACAAGGAG ATGATTACGT GTACCTGCCT TACCCAGATC CATCAAGAAT ATTAGGCGCA
15830      15840      15850      15860      15870      15880      15890

                                >< RsaI          >< SfaNI
                                >< Csp6I          >< MaeIII
                                >< AfaI          BsrI ><
GGCTGTTTTG TCGATGATAT TGTCAAACA GATGGTACAC TTATGATTGA AAGGTTTCGTG TCACTGGCTA
15900      15910      15920      15930      15940      15950      15960

> < FokI
>< BspWI
TTGATGCTTA CCCACTTACA AAACATCCTA ATCAGGAGTA TGCTGATGTC TTCACTTGT ATTTACAATA
15970      15980      15990      16000      16010      16020      16030

                                >< Van91I
                                >< PflMI
                                >< NspI
                                > < Pali>< NspHI
                                > < MscI>< NlaIII
                                > < HaeIII
                                > < BsuRI
                                >< BsrI
                                >< EaeI >< BslI >< NspI
                                > < BshI>< BsiYI >< NspHI
                                >< NlaIII          >< AflIII >< AflIII
                                >< MaeIII          >< AluI > < BalI>< AccB7I >< NlaIII
CATTAGAAAG TTACATGATG AGCTTACTGG CCACATGTTG GACATGTATT CCGTAATGCT AACTAATGAT
16040      16050      16060      16070      16080      16090      16100

                                >< RsaI> < NlaIV
                                >< MnlI
                                >< Csp6I          >< DdeI          >< RsaI
                                >< BsrI >< MnlI          >< Csp6I
                                >< AfaI> < BscBI          >< AfaI          SfcI ><
AACACCTCAC GGTACTGGGA ACCTGAGTTT TATGAGGCTA TGTACACACC ACATACAGTC TTGCAGGCTG
16110      16120      16130      16140      16150      16160      16170

                                >< NlaIV
                                >< EcoNI
                                >< Eco31I
                                >< Eco64I>< BsmAI
                                >< BscBI >< BslI
                                >< BanI          >< BsiYI
                                >< AciI          >< BsaI
                                >< BspWI          >< AccB1I>< Alw26I BbvI ><
TAGGTGCTTG TGTATTGTGC AATTCACAGA CTTCACTTCG TTGCGGTGCC TGTATTAGGA GACCATTCCCT
16180      16190      16200      16210      16220      16230      16240

                                >< Tth111I
                                >< Fnu4HI          >< NlaIII          > < Tru9I
                                >< BspWI >< AspI          > < MseI
ATGTTGCAAG TGCTGCTATG ACCATGTCAT TTCAACATCA CACAAATTAG TGTTGTCTGT TAATCCCTAT
16250      16260      16270      16280      16290      16300      16310

                                >< ScrFI
                                >< MvaI

```

FIGURE 13.37

```

>< EcoRII
>< Ecl136I
>< DsaV
>< BstOI
>< BstNI
>< BsiLI
>< BsaJI
>< ApyI
>< MaeIII >< MaeIII
>< MaeI
>< AluI
GTTTGCAATG CCCCAGGTTG TGATGTCAC TATGTGACAC AACTGTATCT AGGAGGTATG AGCTATTATT
16320 16330 16340 16350 16360 16370 16380

>< MaeIII >< MnlI
GCAAGTCACA TAAGCCTCCC ATTAGTTTTT CATTATGTGC TAATGGTCAG GTTTTTGGTT TATACAAAAA
16390 16400 16410 16420 16430 16440 16450

>< NspI >< NspI
>< NspHI >< Tth111I >< NspHI
>< NlaIII>< MaeIII>< MaeIII >< NlaIII
>< AflIII >< AspI >< AflIII
CACATGTGTA GGCAGTGACA ATGTCAC TCTCAATGCG ATAGCAACAT GTGATTGGAC TAATGCTGGC
16460 16470 16480 16490 16500 16510 16520

>< RsaI
>< P1eI
>< DdeI
>< Csp6I
>< BsmAI >< HinfI >< MnlI
>< Alw26I >< HindIII DdeI ><
>< AfaI >< AluI >< Fnu4HI >< BbvI
GATTACATAC TTGCCAACAC TTGTACTGAG AGACTCAAGC TTTTCGCAGC AGAAACGCTC AAAGCCACTG
16530 16540 16550 16560 16570 16580 16590

>< ThaI
>< ScaI
>< RsaI >< RsaI
>< MvnI
>< Csp6I >< Csp6I
>< BstUI
>< Bsp50I
>< Tru9I
>< MseI >< NdeI >< AfaI >< AfaI
>< AluI >< AccII MnlI >
AGGAAACATT TAAGCTGTCA TATGGTATTG CCACTGTACG CGAAGTACTC TCTGACAGAG AATTGCATCT
16600 16610 16620 16630 16640 16650 16660

MaeIII ><
>< MaeIII
>< EcoO65I
>< Eco91I
>< BstPI
>< BstEII
>< BsrI
>< SfaNI >< RmaI
>< NlaIII >< MaeI
TTCATGGGAG GTTGGAAAAC CTAGACCACC ATTGAACAGA AACTATGTCT TTACTGGTTA CCGTGTAACT
16670 16680 16690 16700 16710 16720 16730

RsaI ><
>< MnlI
>< HphI
>< RsaI >< RsaI
>< Csp6I >< Csp6I >< SfaNI Csp6I ><
>< AfaI >< AfaI >< MaeIII >< HphI AfaI ><
AAAAATAGTA AAGTACAGAT TGGAGAGTAC ACCTTTGAAA AAGGTGACTA TGGTGATGCT GTTGTGTACA
16740 16750 16760 16770 16780 16790 16800

```

FIGURE 13.38


```

    >< RsaI
    >< Csp6I
    >< AfaI
GAGGTACTAC GACATACAAG TTGAATGTTG GTGATTACTT TGTGTTGACA TCTCACACTG TAATGCCACT
16810      16820      16830      16840      16850      16860      16870

    >< VneI
    >< SnoI
    >< SduI
    >< NspII
    >< HgiAI
    >< DraIII
    >< Bsp1286I
    >< BmyI
    >< ApaLI
    >< Alw44I
    >< Alw21I
    >< RmaI
    >< MaeI
    >< Alw21I
    >< BspWI
    >< DraIII
    >< Bsp1286I
    >< BmyI
    >< Alw21I
    >< RsaI
    >< Csp6I
    >< BsrI
    >< AfaI
    DdeI >
TAGTGCACCT ACTCTAGTGC CACAAGAGCA CTATGTGAGA ATTACTGGCT TGTACCCAAC ACTCAACATC
16880      16890      16900      16910      16920      16930      16940

    StyI ><
    SinI >
    Sau96I >
    NspIV >
    EcoT14I ><
    Eco47I >
    Eco130I ><
    >< ScaI Cfr13I >
    BssT1I ><
    >< SphI >< RsaI BsiZI >
    >< PaeI BsaJI ><
    >< NlaIII Bme18I >
    >< NspI>< Csp6I AvaII >
    >< NspHI>< AfaI AsuI >
    >< RmaI
    >< MaeI
TCAGATGAGT TTTCTAGCAA TGTTGCAAAT TATCAAAAGG TCGGCATGCA AAAGTACTCT ACACTCCAAG
16950      16960      16970      16980      16990      17000      17010

    >< ScrFI
    >< RsaI
    >< MvaI
    >< EcoRII
    >< Ecl136I
    >< Csp6I
    >< BstOI
    >< BstNI
    >< XcmI >< BslI
    >< NspHII >< BsiYI
    >< BsiLI
    >< ApyI >< BsrI
    >< DsaV>< AfaI >< HinfI>< P1eI
GACCACCTGG TACTGGTAAG AGTCATTTTG CCATCGGACT TGCTCTCTAT TACCCATCTG CTCGCATAGT
17020      17030      17040      17050      17060      17070      17080

    >< SfaNI
    >< SphI >< PvuII
    >< PaeI >< Psp5I
    >< NspI >< NspBII
    >< NspHI >< Fnu4HI
    >< Bst1107I >< NlaIII>< BspWI >< SspI
    >< AccI >< NlaIII >< AluI >< BbvI >< MseI
GTATACGGCA TGCTCTCATG CAGCTGTTGA TGCCCTATGT GAAAAGGCAT TAAAATATTT GCCCATAGAT
17090      17100      17110      17120      17130      17140      17150

```

FIGURE 13.39

```

> < ThaI
>< ThaI
> < MvnI
>< MvnI >< ThaI
> < HinP1I
>< HinP1I
>< HinP1I >< MvnI
> < Hin6I
>< Hin6I
> < HhaI
>< HhaI >< HhaI
> < CfoI
>< CfoI >< CfoI
> < BstUI
>< BstUI >< BstUI
>< BssHII
>< BspMI
> < Bsp50I
>< Bsp50I>< Bsp50I
>< TfiI >< Hin6I> < AccII
>< Hinfi >< AccII >< AccII
> < EcoRI
AAATGTAGTA GAATCATACC TGC GCGTAGAGT GTTTTGATAA ATTCAAAGTG AATTC AACAC
17160 17170 17180 17190 17200 17210 17220

>< Zsp2I
>< Ppu10I
>< NsiI
>< Mph1103I
>< EcoT22I
>< BsgI > < AvaIII >< DrdI
TAGAACAGTA TGTTTTCTGC ACTGTAAATG CATTGCCAGA AACAACTGCT GACATTGTAG TCTTTGATGA
17230 17240 17250 17260 17270 17280 17290

>< RmaI
>< MaeI >< MaeII
AATCTCTATG GCTACTAATT ATGACTTGAG TGTTGTCAAT GCTAGACTTC GTGCAAAACA CTACGTCTAT
17300 17310 17320 17330 17340 17350 17360

>< Sau3AI
>< NdeII
>< MboI
>< DpnII
>< DpnI
>< BspAI
>< AlwI>< Bsp143I > < AciI >< RmaI
ATTGGCGATC CTGCTCAATT ACCAGCCCCC CGCACATTGC TGAATAAAGG CACACTAGAA CCAGAATATT
17370 17380 17390 17400 17410 17420 17430

>< SinI
>< Sau96I
>< NspIV >< StyI
>< NspHII >< NspI
>< Eco47I >< NspHI
>< Cfr13I >< NlaIII
>< Bsi2I >< EcoT14I
>< BsgI >< Eco130I
>< Bme18I >< BssTII
>< AvaII >< BsaJI
>< Tru9I
>< MseI
>< AsuI> < AflIII
TTAATTCAGT GTGCAGACTT ATGAAAACAA TAGGTCCAGA CATGTTCCCTT GGAAGTTGTC GCCGTTGTCC
17440 17450 17460 17470 17480 17490 17500

```

FIGURE 13. 40

```

                >< HindII
                >< HincII
                >< AluI
TGCTGAAATT GTTGACACTG TGAGTGCTTT AGTTTATGAC AATAAGCTAA AAGCACACAA GGATAAGTCA
17510      17520      17530      17540      17550      17560      17570

>< AluI                >< NlaIII
GCTCAATGCT TCAAAATGTT CTACAAAGGT GTTATTACAC ATGATGTTTC ATCTGCAATC AACAGACCTC
17580      17590      17600      17610      17620      17630      17640

                >< MnlI
>< EcoNI
                >< BslI
                >< BsiYI
                >< HphI
                >< AluI
AAATAGGCGT TGTAAAGAGAA TTTCTTACAC GCAATCCTGC TTGGAGAAAA GCTGTTTTTA TCTCACCTTA
17650      17660      17670      17680      17690      17700      17710

                >< SfcI                >< DdeI                >< TfiI
                > < AluI                >< BfrI                >< HinfI
TAATTCACAG AACGCTGTAG CTTCAAAAAT CTTAGGATTG CCTACGCAGA CTGTTGATTG ATCACAGGGT
17720      17730      17740      17750      17760      17770      17780

                >< Tth111I                > < HindII
                >< AspI                > < HincII
                >< AcII
TCTGAATATG ACTATGTCAT ATTACACACAA ACTACTGAAA CAGCACACTC TTGTAATGTC AACCGCTTCA
17790      17800      17810      17820      17830      17840      17850

                >< XhoII
                >< Sau3AI
                >< NdeII
                >< MflI
                >< MboI
>< MamI
                >< DpnII
                >< DpnI
                >< BstYI
                >< BspAI
                >< Bsp143I
                >< BsiBI
                >< BsaBI
                >< BglII
                >< BspWI
ATGTGGCTAT CACAAGGGCA AAAATTGGCA TTTTGTGCAT AATGTCTGAT AGAGATCTTT ATGACAACT
17860      17870      17880      17890      17900      17910      17920

                >< XbaI
                >< RmaI
                >< MaeI                >< MaeII                >< MaeIII
                >< BsrI ><
GCAATTTACA AGTCTAGAAA TACCACGTCG CAATGTGGCT ACATTACAAG CAGAAAATGT AACTGGACTT
17930      17940      17950      17960      17970      17980      17990

                >< Sau3AI
                >< NdeII
                >< MboII
                >< MboI
                > < FokI
                >< DpnII
                >< DpnI
                >< BspAI
                >< Bsp143I
                >< NlaIV
                >< Eco64I
                >< BscBI
                >< BanI
                >< MnlI ><
>< Tru9I                >< Bsp143I                >< BanI                >< MnlI ><
>< MseI>< SfcI                >< BbsI > < BsrI                >< AccBI                >< DdeI

```

FIGURE 13. 41

TTTAAAGGACT	GTAAGTAAGAT	CATTACTGGT	CTTCATCCCTA	CACAGGCACC	TACACACCTC	AGCGTTGTATA
18000	18010	18020	18030	18040	18050	18060
			>< ScrFI			
			>< MvaI			
			>< EcoRII			
			>< Eco57I			
			>< Ecl136I			
			>< DsaV			
			>< BstOI		>< PleI	
			>< BstNI		>< NlaIII	
		>< HindII>	BsiLI		HinfI ><	
		>< HincII>	ApyI		AccI ><	
TAAAGTTCAA	GAAGAAGGA	TTATGTGTTG	ACATACCAGG	CATACCAAG	GACATGACCT	ACCGTAGACT
18070	18080	18090	18100	18110	18120	18130
				>< MaeIII	Thai ><	
				>< EcoO651	MvnI ><	
				>< Eco91I	BstUI ><	
			>< BstXI		Bsp50I ><	
			>< BstPI		>< AciI	
			>< BstEII	>< HphI	AccII ><	
CATCTCTATG	ATGGGTTTTCA	AAATGAATTA	CCAAGTCAAT	GGTTACCCTA	ATATGTTTAT	CACCOGCGAA
18140	18150	18160	18170	18180	18190	18200
>< XmnI						
> < MboII					>< SfaNI	
> < MaeIII					>< RmaI	
>< Asp700I					>< NlaIII	
>< AluI	>< MaeII		>< MnlI		>< MaeI	
GAAGCTATTC	GTCACGTTTCG	TGCGTGGAAT	GGCTTTGATG	TAGAGGGCTG	TCATGCAACT	AGAGATGCTG
18210	18220	18230	18240	18250	18260	18270
				>< Tru9I		
				>< MseI		
>< RsaI				>< HpaI		
>< GsuI		>< RmaI		>< HindII	>< RsaI	
>< Csp6I		>< MnlI		>< HincII	>< Csp6I	
>< BpmI		>< MaeI		>< DdeI	>< AluI	BsrI ><
>< AfaI		>< AluI	>< SfcI	>< BfrI	>< AfaI	
TGGGTACTAA	CCTACCTCTC	CAGCTAGGAT	TTTCTACAGG	TGTTAACTTA	G TAGCTGTAC	CGACTGGTTA
18280	18290	18300	18310	18320	18330	18340
					>< ScrFI	
					>< MvaI	
					>< MnlI	
					>< MaeIII	
					>< EcoRII	
					>< EcoO651	
					>< EcoNI	
					>< Eco91I	
					>< Ecl136I	
					>< DsaV	Tru9I ><
					>< DraIII	
					>< BstPI	
					>< BstOI	
					>< BstNI	PmeI ><
					>< BstEII	
					>< Bsil	MseI ><
					>< BsiYI	HphI ><
>< HindII	>< HphI		>< Tru9I		>< BsiLI	DraI ><
>< HincII		>< EcoRI	>< MseI		>< ApyI	>< BsrI

FIGURE 13.42

```

TGTTGACACT GAAAATAACA CAGAATTCAC CAGAGTTAAT GCAAAACCTC CACCAGGTGA CCAGTTTAAA
18350      18360      18370      18380      18390      18400      18410

                >< ScrFI
                >< MvaI
                >< EcoRII
                >< Ecl136I
                >< DsaV
                >< BstOI
                >< BstNI
                >< BsiLI
                >< BsaJI
                >< ApyI
                >< NlaIII
                >< NlaIII
                >< Tth111I
                >< HinPII
                >< Hin6I
                >< HinfI
                >< AspI
                >< PleI
                >< CfoI
                >< AluI
CATCTTATAC CACTCATGTA TAAAGGCTTG CCCTGGAATG TAGTGCGTAT TAAGATAGTA CAAATGCTCA
18420      18430      18440      18450      18460      18470      18480

                >< NlaIII
                >< HinPII
                >< Hin6I
                >< HinfI
                >< AspI
                >< PleI
                >< CfoI
                >< AluI
GTGATACACT GAAAGGATTG TCAGACAGAG TCGTGTTTCGT CCTTTGGGCG CATGGCTTTG AGCTTACATC
18490      18500      18510      18520      18530      18540      18550

                >< SinI
                >< Sau96I
                >< NspIV
                >< NspHII
                >< Eco47I
                >< Cfr13I
                >< ScaI
                >< RsaI
                >< Csp6I
                >< AfaI
                >< BsiZI
                >< Bme18I
                >< AvaII
                >< AsuI
                >< MaeII
                >< AflIII
                >< MaeIII>< MaeII
AATGAAGTAC TTTGTCAAGA TTGGACCTGA AAGAACGTGT TGTCTGTGTG ACAAACGTGC AACTTGCTTT
18560      18570      18580      18590      18600      18610      18620

                >< TfiI
                >< Tth111I
                >< HinfI
                >< AspI
TCTACTTCAT CAGATACTTA TGCCTGCTGG AATCATTCTG TGGGTTTTGA CTATGTCTAT AACCCATTTA
18630      18640      18650      18660      18670      18680      18690

                >< ScrFI
                RsaI ><
                >< MvaI
                >< EcoRII
                Ecl136I ><
                >< DsaV
                Csp6I ><
                BstXI ><
                >< MaeIII
                >< EcoO65I
                >< Eco91I
                >< BstPI
                >< Eco57I> >< BstEII
                >< MaeIII
                >< NlaIII
                AfaI ><
TGATTGATGT TCAGCAGTGG GGCTTTACGG GTAACCTTCA GAGTAACCAT GACCAACATT GCCAGGTACA
18700      18710      18720      18730      18740      18750      18760

                >< SfaNI
                >< RmaI
                >< NspI
                >< NspHI

```

FIGURE 13.43

```

                >< NlaIII                >< RmaI
                >< MaeI                >< NlaIII                Tru9I ><
>< NlaIII    >< BspWI                >< MaeI                >< NlaIII
    > < AflIII                >< BspHI                MseI ><
TGGAAATGCA CATGTGGCTA GTTGTGATGC TATCATGACT AGATGTTTAG CAGTCCATGA GTGCTTTGTT
    18770      18780      18790      18800      18810      18820      18830

    >< ThaI
    >< MvnI
    >< HinPII
    >< Hin6I
    >< HhaI
    >< CfoI
    >< BstUI
    >< Bsp50I
    >< AccII
                >< EcoNI> < MnlI
                >< BslI                >< Tru9I
                >< BsiYI                >< DdeI >< MseI
AAGCGCGTTG ATTGGTCTGT TGAATACCCT ATTATAGGAG ATGAAGTCTG GGTAAATTCT GCTTGCAGAA
    18840      18850      18860      18870      18880      18890      18900

    >< RsaI
    >< Csp6I
    >< AfaI    >< NlaIII                >< BspWI                >< MboII                > < NlaIII
                >< BsrI    >< BspHI
AAGTACAACA CATGGTTGTG AAGTCTGCAT TGCTTGCTGA TAAGTTTCCA GTTCTTCATG ACATTGGAAA
    18910      18920      18930      18940      18950      18960      18970

                >< SauI
                >< MstII
                >< Eco81I
                >< DdeI
                >< CvnI
                >< Bsu36I
                >< Bse21I
                >< AxyI
                >< AocI    >< MnlI    >< SfaNI
                >< Bpu1102I
TCCAAAGGCT ATCAAGTGTG TGCCTCAGGC TGAAGTAGAA TGGAAGTTCT ACGATGCTCA GCCATGTAGT
    18980      18990      19000      19010      19020      19030      19040

                >< MnlI                >< Ksp632I
    >< HindIII                >< EarI
    >< AluI    >< MboII    >< Eam1104I
GACAAAGCTT ACAAATAGA GGAACCTTC TATTCTTATG CTACACATCA CGATAAATTC ACTGATGGTG
    19050      19060      19070      19080      19090      19100      19110

                >< Sau3AI
                >< NdeII
                >< MboI
    >< MaeII> < MaeIII
    >< DpnII
    >< DpnI
    >< BspAI
                >< MaeIII    >< Bsp143I    >< MunI                HinfI >
                >< BspAI                DrdI ><
TTTGTTTGTT TTGGAATTGT AACGTGATC GTTACCCAGC CAATGCAATT GTGTGTAGGT TTGACACAAG
    19120      19130      19140      19150      19160      19170      19180

                Zsp2I ><
                >< SphI
                > < Ppu10I
                >< PaeI
                >< NspI
                >< NspHI
                >< NlaIII
    >< ScrFI
    >< MvaI
    >< EcoRII
                Mph1103I ><

```

FIGURE 1344

```

                >< Ecl136I
                >< DsaV
                >< BstOI
                >< BstNI
                >< BsiLI
                >< ApyI
                >< PleI
AGTCTTGTC AACTTGAACT TACCAGGCTG TGATGGTGGT AGTTTGTATG TGAATAAGCA TGCATTCCAC
19190      19200      19210      19220      19230      19240      19250

                >< Tru9I
                > < MunI
                >< TthHB8I
                >< BcgI/a >< TaqI
                >< AluI
                >< MseI
                >< DraI
                >< BcgI
ACTCCAGCTT TCGATAAAAG TGCATTTACT AATTTAAAGC AATTGCCTTT CTTTTACTAT TCTGATAGTC
19260      19270      19280      19290      19300      19310      19320

                >< PleI
                >< NlaIII
                >< BsmAI
                >< HinFI>< Alw26I
                SfaNI ><
                >< MaeII
                BsaAI ><
                AflIII ><
CTTGTGAGTC TCATGGCAAA CAAGTAGTGT CGGATATTGA TTATGTTCCA CTCAAATCTG CTACGTGTAT
19330      19340      19350      19360      19370      19380      19390

                Zsp2I >
                >< ScaI
                Ppu10I ><
                >< RsaINSiI >
                Mph1103I >
                >< SfaNEcoT22I >
                > < RsaI >< Csp6I
                >< Csp6I
                >< NlaIII> < AfaI >< AfaI
TACACGATGC AATTTAGGTG GTGCTGTTTG CAGACACCAT GCAAATGAGT ACCGACAGTA CTTGGATGCA
19400      19410      19420      19430      19440      19450      19460

                >< FokI
TATAATATGA TGATTTCTGC TGGATTTAGC CTATGGATTT ACAAACAATT TGATACTTAT AACCTGTGGA
19470      19480      19490      19500      19510      19520      19530

                >< ScrFI
                >< MvaI
                >< MaeIII
                >< EcoRII
                >< Ecl136I
                >< DsaV
                >< BstOI
                >< BstNI
                >< BsiLI
                >< ApyI
                >< Tru9I
                >< MseI
ATACATTTAC CAGGTTACAG AGTTTAGAAA ATGTGGCTTA TAATGTTGTT AATAAAGGAC ACTTTGATGG
19540      19550      19560      19570      19580      19590      19600

                >< SgrAI
                >< NaeI
                >< MspI
                >< HpaII
                >< HapII
                >< Cfr10I
                >< BspWI
                > < VspI
                > < Tru9I
                > < MseI
                > < AsnI
                > < AseI
ACACGCCGGC GAAGCACCTG TTTCCATCAT TAATAATGCT GTTTACACAA AGGTAGATGG TATTGATGTG
19610      19620      19630      19640      19650      19660      19670

```

FIGURE 13. 45

```

>< XhoII
>< Sau3AI
>< NdeII
>< MflI
>< MboI
>< DpnII
  >< DpnI
>< BstYI
>< BspAI
  >< Bsp143I
>< BglII
GAGATCTTTG AAAATAAGAC AACACTTCCT GTTAATGTTG CATTGAGCT TTGGGCTAAG CGTAACATTA
19680      19690      19700      19710      19720      19730      19740

                                >< MaeIII
                                >< EspI
                                >< DdeI Tru9I ><
                                >< CeliIMseI ><
                                >< Bpu1102I
                                >< AluI
                                >< Tru9I
                                >< MseI
                                >< Fnu4HI
                                >< EcoRV
                                >< Eco32I
>< BsrI      >< Tru9I      >< MseI      >< BbvI      >< BbvI      >< BbvI
AACCAGTGCC AGAGATTAAG ATACTCAATA ATTTGGGTGT TGATATCGCT GCTAATACTG TAATCTGGGA
19750      19760      19770      19780      19790      19800      19810

                                >< NspI
                                >< NspHI
                                >< NlaIII
                                >< BsgI
                                >< AflIII
CTACAAAAGA GAAGCCCCAG CACATGTATC TACAATAGGT GTCTGCACAA TGA CTGACAT TGCCAAGAAA
19820      19830      19840      19850      19860      19870      19880

>< DdeI>< MboII
CCTACTGAGA GTGCTTGTTT TCACTTACT GTCTTGTTT ATGGTAGAGT GGAAGGACAG GTAGACCTTT
19890      19900      19910      19920      19930      19940      19950

                                SinI ><
                                Sau96I ><
                                NspIV ><
                                NspHII ><
                                NlaIV ><
                                Eco47I ><
                                Cfr13I ><
                                >< BslI
                                BsiZI ><
                                >< BsiYI
                                BscBI ><
                                Bmel8I ><
                                AvaII ><
                                AsuI ><
                                >< Tru9I
                                >< MseI
TTAGAAACGC CCGTAATGGT GTTTTAATAA CAGAAGGTT AGTCAAAGGT CTAACACCTT CAAAGGGACC
19960      19970      19980      19990      20000      20010      20020

                                >< VspI
                                >< Tru9I
                                >< PleI
                                >< MseI
                                >< RmaI
                                >< NheI
                                >< MaeI
                                >< MaeIII
                                >< AsnI
                                >< TfiI
                                >< Tru9I ><
                                >< Tru9I
                                >< MseI ><
                                >< MseI
>< HgaI>< AluI      >< HinfI>< AseI      >< HinfI
AGCACAAGCT AGCGTCAATG GAGTCACATT AATTGGAGAA TCAGTAAAAA CACAGTTTAA CTA CTTTAAG
20030      20040      20050      20060      20070      20080      20090

                                >< DdeI      >< MnlI      Tru9I ><
                                >< BsmAI      >< DdeI

```

FIGURE 1346


```

>< AccI                                     >< Alw26I >< BfrIMseI ><
AAAGTAGACG GCATTATTCA ACAGTTGCCT GAAACCTACT TTACTCAGAG CAGAGACTTA GAGGATTTTA
20100      20110      20120      20130      20140      20150      20160

                                >< TthHB8I
                                >< TaqI
                                    >< SstI
                                    >< SduI
                                    >< SacI
                                > < PaeR7I
                                > < NspIII
                                    >< NspII
                                    >< HgiAI
                                > < Eco88I
                                > < XhoI>< Eco24I
                                >< Ecl136II
>< XcmI                                >< XhoI>< Eco24I
>< Sau3AI                                >< Ecl136II
>< NdeII                                > < SlaI>< Bsp1286I
>< MboI                                > < CcrI>< BmyI
>< DpnII                                > < BcoI>< BanII
>< DpnI                                > < Ama87I
>< BspAI                                > < AvaI>< Alw21I
>< Bsp143I                                >< AluI
AGCCCAGATC ACAAATGGAA ACTGACTTTC TCGAGCTCGC TATGGATGAA TTCATACAGC GATATAAGCT
20170      20180      20190      20200      20210      20220      20230

                                >< TthHB8I
                                >< TaqI
                                >< SfuI
                                >< NspV
                                >< LspI
                                >< Csp45I
                                >< BstBI
                                >< Bsp119I
                                >< BsiCI
                                >< Bpu14I
                                >< AsuII >< BcgI
                                >< MboII
                                >< BbsI Tru9I ><
                                >< NlaIII >< AciIMseI ><
CGAGGGCTAT GCCTTCGAAC ACATCGTTTA TGGAGATTTC AGTCATGGAC AACTGGCGG TCTTCATTTA
20240      20250      20260      20270      20280      20290      20300

                                >< HphI
                                >< HinPII
                                >< Hin6I
>< EspI > < HhaI >< TfiI
>< DdeI >< HaeII
>< CeiII >< Eco47III >< Tru9I
>< Bpu1102I > < CfoI >< HinfI >< MseI
>< BfrI >< Bsp143II >< MnlI
ATGATAGGCT TAGCCAAGCG CTCACAAGAT TCACCACTTA AATTAGAGGA TTTTATCCCT ATGGACAGCA
20310      20320      20330      20340      20350      20360      20370

                                >< MstI
                                >< HinPII
                                >< Hin6I
                                >< HhaI
                                >< FspI
                                >< FdiII
                                >< CfoI
                                >< AviII
>< SfaNI                                >< SfaNI
CAGTGAAAAA TTACTTCATA ACAGATGCGC AAACAGGTTT ATCAAAATGT GTGTGTTCTG TGATTGATCT
20380      20390      20400      20410      20420      20430      20440

                                >< TthHB8I

```

FIGURE 13. 47

```

>< Tth111I
>< TaqI
>< AspI > < MaeIII MaeIII ><
TTTACTTGAT GACTTTGTCG AGATAATAAA GTCACAAGAT TTGTCAGTGA TTTCAAAGT GGTCAAGGTT
20450 20460 20470 20480 20490 20500 20510

>< NspI
>< NspHI
>< NlaIII
>< FokI

>< MunI > < NlaIII >< AflIII
ACAATTGACT ATGCTGAAAT TTCATTCATG CTTTGGTGTA AGGATGGACA TGTGAAACC TTCTACCCAA
20520 20530 20540 20550 20560 20570 20580

>< SfaNI
>< ScrFI
>< MvaI
>< EcoRII
>< Ecl136I
>< DsaV
>< BstOI >< SfaNI
>< BstNI >< RsaI BspWI ><
>< BsiLI > < Csp6I BsmI >
>< BspWI >< ApyI >< AfaI BscCI ><
AACTACAAGC AAGTCAAGCG TGGCAACCAG GTGTTGCGAT GCCTAACTTG TACAAGATGC AAAGAATGCT
20590 20600 20610 20620 20630 20640 20650

>< Eco57I >< MaeIII >< HphI
TCTTGAAAAG TGTGACCTTC AGAATTATGG TGAAAATGCT GTTATACCAA AAGGAATAAT GATGAATGTC
20660 20670 20680 20690 20700 20710 20720

> < RsaI
>< Csp6I

>< Bst1107I >< Tru9I >< AluI
>< AccI >< MseI > < AfaINlaIII ><
GCAAAGTATA CTCAACTGTG TCAATACTTA AATACACTTA CTTTAGCTGT ACCCTACAAC ATGAGAGTTA
20730 20740 20750 20760 20770 20780 20790

>< ScrFI
>< RsaI
>< MvaI
>< EcoRII >< NspBII
>< Ecl136I >< SduI
> < Csp6I >< NspII
>< BstOI >< PvuII>< HgiAI
>< BstNI >< DdeI
>< BsiLI >< Psp5I>< Bsp1286I
>< ApyI >< AluI >< BmyI
>< DsaV>< AfaI >< Alw21I
TTCACTTTGG TGCTGGCTCT GATAAAGGAG TTGCACCAGG TACAGCTGTG CTCAGACAAT GGTGCCAAC
20800 20810 20820 20830 20840 20850 20860

>< XhoII
>< Tru9I
>< Sau3AI
>< NdeII
>< TthHB8I >< MseI
>< MflI
>< MboI
>< MamI
>< DpnII
>< TfiI >< DpnI

```

FIGURE 13. 48

```

                >< BstYI                > < TfiI
                >< BspAI                > < HinfI
                >< HinfI>< Bsp143I        >< Esp3I        >< Tru9I
                >< BsiBI        >< Tth111I    >< BsmBI        >< MseI
                >< BsaBI        >< BsmAI        > < BsmAI
                >< BsrI        >< TaqI >< BglII    >< AspI        >< Alw26I >< HgaI> < Alw26I
TGGCACACTA CTTGTCGATT CAGATCTTAA TGA CTTCGTC TCCGACGCAG ATTCTACTTT AATTGGAGAC
    20870      20880      20890      20900      20910      20920      20930

                >< StyI
                >< SinI
                >< Sau96I
                > < SinI                >< RmaI
                > < Sau96I            >< NspIV
                >< PssI                NspHII ><
                >< Psp5II            >< MaeI
                > < PpuMI            >< EcoT14I
                > < NspIV                >< Eco47I
                >< NspHII            >< Eco130I
                >< NlaIV                >< Cfr13I
                > < EcoO109I        >< BssT1I
                > < Eco47I            >< BsiZI
                > < DraII            >< BsaJI
                > < Cfr13I            >< Bme18I
                > < BsiZI            >< BlnI
                >< BscBI            >< AvrII
                >< RsaI                >< Bme18I        >< AvaII
                > < Csp6I            >< AvaII        >< AsuI
                >< AfaI                > < AsuI        AflIII ><
TGTGCAACAG TACATACGGC TAATAAATGG GACCTTATTA TTAGCGATAT GTATGACCCT AGGACCAAAC
    20940      20950      20960      20970      20980      20990      21000

                >< NspI
                >< NspHI
                >< NlaIII >< PleI                RmaI ><
                >< MaeIII            >< HinfI                MaeI ><
ATGTGACAAA AGAGAATGAC TCTAAAGAAG GGT TTTTCAC TTATCTGTGT GGATTTATAA AGCAAAAAC
    21010      21020      21030      21040      21050      21060      21070

                >< ScrFI
                >< MvaI
                >< EcoRII
                >< Ecl136I
                >< DsaV
                >< BstOI                Sau96I >
                >< BstNI                NspIV >
                >< BsiLI                Cfr13I >
                >< BsaJI                BsiZI >
                >< BsaJI        >< SfcI                >< BsmI        >< BsmI        AsuI >
                >< ApyI            > < AluI            >< BscCI            >< BscCIHindIII ><> < AluI
AGCCCTGGGT GGT TCTATAG CTGTAAAGAT AACAGAGCAT TCTTGGAA TGACCTTTA CAAGCTTATG
    21080      21090      21100      21110      21120      21130      21140

                >< Zsp2I
                >< Ppu10I
                >< Pali
                >< HaeIII            >< NsiI
                >< BsuRI            >< Mph1103I    Tru9I ><
                >< BshI            >< EcoT22I            >< MseI
                >< BshI        >< NlaIII>< AluI        >< BcgI        >< AvaIII >< SfaNIBcgI/a ><
GGCCATT TCT CATGGTGGAC AGCTTTTGT TACAAATGTAA ATGCATCATC ATCGGAAGCA TTTTAAATTG
    21150      21160      21170      21180      21190      21200      21210

```

FIGURE 13.49

```

                                >< Zsp2I
                                >< SphI
                                >< Ppu10I
                                >< PaeI
                                >< NspI
                                >< NspHI
                                >< NsiI
                                >< NlaIII
                                > < NlaIII
                                >< Mph1103I
                                >< EcoT22I
                                > < AvaIII >< MnlI
GGGCTAACTA TCTTGGCAAG CCGAAGGAAC AAATTGATGG CTATACCATG CATGCTAACT ACATTTTCTG
  21220      21230      21240      21250      21260      21270      21280

                                Tru9I ><
                                >< Tru9I
                                >< MboII
                                >< MseI ><
                                >< BsrI
                                >< MseI
                                >< BpmI
                                MnlI ><
                                >< BbsI
                                >< NlaIII >< MnlI
GAGGAACACA AATCCTATCC AGTTGTCTTC CTATTCACCTC TTTGACATGA GCAAATTTCC TCTTAAATTA
  21290      21300      21310      21320      21330      21340      21350

                                >< Tru9I
                                >< MseI
                                >< Esp4I> < TfiI
                                >< BsmAI
                                >< Alw26I
                                >< AflIII> < HinfI
                                Ksp632I ><
                                >< MboII >< EarI
                                Eam1104I ><
AGAGGAACTG CTGTAATGTC TCTTAAGGAG AATCAAATCA ATGATATGAT TTATTCTCTT CTGGAAAAAG
  21360      21370      21380      21390      21400      21410      21420

                                >< Tru9I
                                >< MseI
                                >< HindII
                                >< HincII
                                >< HpaI AflIII >
GTAGGCTTAT CATTAGAGAA AACAACAGAG TTGTGGTTTC AAGTGATATT CTTGTTAACA ACTAAACGAA
  21430      21440      21450      21460      21470      21480      21490

                                >< VneI
                                >< SnoI
                                >< SduI
                                >< NspII
                                >< HpaII
                                >< HgiAI
                                >< HapII
                                >< Cfr10I
                                >< Bsp1286I
                                >< MspI>< BmyI
                                >< ApaLI
                                >< Alw44I
                                >< MaeI >< MaeIII >< AgeI >< Alw21I
                                >< NspI
                                >< NspHI
                                >< NlaIII
CATGTTTATT TTCTTATTAT TTCTTACTCT CACTAGTGGT AGTGACCTTG ACCGGTGCAC CACTTTTGAT
  21500      21510      21520      21530      21540      21550      21560

                                > < AluI
                                >< MnlI
GATGTTCAAG CTCCTAATTA CACTCAACAT ACTTCATCTA TGAGGGGGGT TTACTATCCT GATGAAATTT
  21570      21580      21590      21600      21610      21620      21630

                                >< Sau3AI

```

FIGURE 13. 50

```

>< NdeII
>< MboI
>< DpnII
  >< DpnI          >< Tru9I
>< BspAI          >< MseI > < MboII
  >< Bsp143I       >< DdeI          >< MaeIII
TTAGATCAGA CACTCTTTAT TTAACCTCAGG ATTTATTTCT TCCATTTTAT TCTAATGTTA CAGGGTTTCA
21640      21650      21660      21670      21680      21690      21700

>< VspI
>< Tru9I
>< MseI
>< AsnI          >< Tru9I          >< FokI
>< AseI >< MaeII  >< MseI >< BbvI      > < Fnu4HI
TACTATTAAT CATACGTTTG GCAACCCTGT CATACCTTTT AAGGATGGTA TTTATTTTGC TGCCACAGAG
21710      21720      21730      21740      21750      21760      21770

          >< BslI
          >< DsaI>< BsiYI          >< NlaIII
          >< BsaJI          > < MaeIII
AAATCAAATG TTGTCCTGG TTGGGTTTTT GGTCTACCA TGAACAACAA GTCACAGTCG GTGATTATTA
21780      21790      21800      21810      21820      21830      21840

          >< NspI
>< Tru9I          >< NspHI
>< MseI          >< NlaIII
>< HphI          >< MaeIII          >< MaeIII
TTAACAATTC TACTAATGTT GTTATACGAG CATGTAACCT TGAATTGTGT GACAACCCTT TCTTTGCTGT
21850      21860      21870      21880      21890      21900      21910

  >< StyI          >< Zsp2I
    >< NlaIII          >< Tru9I
  >< NcoI >< RsaI          >< Ppu10I TthHB8I ><
  >< EcoT14I          >< NsiI          >< TaqI
  >< Eco130I          >< MseI          SfaNI ><
  >< DsaI>< Csp6I          >< Mph1103I RsaI ><
  >< BssT1I          >< TthHB8I >< EcoT22I Csp6I ><
  >< BsaJI>< AfaI          >< TaqI >< AvaIII AfaI ><
TTCTAAACCC ATGGGTACAC AGACACATAC TATGATATTC GATAATGCAT TTAATTGCAC TTTCGAGTAC
21920      21930      21940      21950      21960      21970      21980

          >< Tru9I
          >< MseI
          >< DraI
ATATCTGATG CCTTTTCGCT TGATGTTTCA GAAAAGTCAG GTAATTTTAA ACACCTTACGA GAGTTTGTGT
21990      22000      22010      22020      22030      22040      22050

          >< Sau3AI
          >< NdeII
          >< MboI
          >< DpnII
>< Tru9I          >< DpnI
>< MseI          >< BspAI
>< DraI          >< SfcI Bsp143I ><
TTAAAAATAA AGATGGGTTT CTCTATGTTT ATAAGGGCTA TCAACCTATA GATGTAGTTC GTGATCTACC
22060      22070      22080      22090      22100      22110      22120

          >< Tru9I
          > < Tru9I          >< MseI
          >< MseI          > < MseI          >< MnlI
TTCTGGTTTT AACACTTTGA AACCTATTTT TAAGTGCCT CTTGGTATTA ACATTACAAA TTTTAGAGCC
22130      22140      22150      22160      22170      22180      22190

```

FIGURE 13.51

```

> < SduI>< SfcI
>< PvuII
>< Psp5I
> < NspII
>< NspBII
> < MaeII > < Fnu4HI
> < Bsp1286I >< PstI Tru9I >
>< BspMI > < BmyI>< Fnu4HI MseI >
>< HphI >< BbvI >< AluI >< BbvI
ATTCTTACAG CCTTTTCACC TGCTCAAGAC ATTTGGGGCA CGTCAGCTGC AGCCTATTTT GTTGGCTATT
22200 22210 22220 22230 22240 22250 22260

>< SfaNI
>< RsaI
> < Csp6I
>< DraI >< AfaI >< AlwNI
TAAAGCCAAC TACATTTATG CTCAAGTATG ATGAAAATGG TACAATCACA GATGCTGTTG ATTGTTCTCA
22270 22280 22290 22300 22310 22320 22330

> < Tru9I
> < MseI
>< AluI
AAATCCACTT GCTGAACTCA AATGCTCTGT TAAGAGCTTT GAGATTGACA AAGGAATTTA CCAGACCTCT
22340 22350 22360 22370 22380 22390 22400

>< SauI
>< MstII
>< Eco81I
>< DdeI
>< CvnI
>< Bsu36I
>< Bse21I
>< AxyI >< TfiI
>< MnlI >< AocI >< MnlI >< HinfI >< SspI >< MnlI
AATTTTCAGGG TTGTTCCCTC AGGAGATGTT GTGAGATTCC CTAATATTAC AAAGTTGTGT CCTTTTGGAG
22410 22420 22430 22440 22450 22460 22470

>< Zsp2I
>< Ppu10I
>< NsiI
> < NlaIII
>< Mph1103I
>< EcoT22I
>< Tru9I >< AvaIII
>< MseI
AGGTTTTTAA TGCTACTAAA TTCCCTTCTG TCTATGCATG GGAGAGAAAA AAAATTTCTA ATTGTGTTGC
22480 22490 22500 22510 22520 22530 22540

>< SduI
>< NspII
>< HgiAI
>< Bsp1286I
>< BmyI >< Tru9I
>< Alw21I >< MseI DdeI ><
TGATTACTCT GTGCTCTACA ACTCAACATT TTTTCAACC TTTAAGTGCT ATGGCGTTTC TGCCACTAAG
22550 22560 22570 22580 22590 22600 22610

>< Sau3AI
>< NdeII
>< MboI
>< DpnII
>< DpnI

```

FIGURE 13.52

```

    >< BspAI
    >< Bsp143I
    TTGAATGATC TTTGCTTCTC CAATGTCTAT GCAGATTCTT TTGTAGTCAA GGGAGATGAT GTAAGACAAA
    22620      22630      22640      22650      22660      22670      22680

    >< ScrFI
    >< MvaI
    >< HinPII
    >< Hin6I
    >< HhaI
    >< HaeII
    >< EcoRII
    >< Ecl136I
    >< DsaV
    >< CfoI
    >< BstOI
    >< BstNI
    >< Bsp143II
    >< BsiLI
    >< ApyI      > < BsrI
    TAGCGCCAGG ACAACTGGT GTTATTGCTG ATTATAATTA TAAATTGCCA GATGATTTCa TGGGTTGTGT
    22690      22700      22710      22720      22730      22740      22750
    >< SfaNI
    >< RmaI
    >< MaeI
    CCTTGCTTGG AATACTAGGA ACATTGATGC TACTTCAACT GGTAAATTATA ATTATAAATA TAGGTATCTT
    22760      22770      22780      22790      22800      22810      22820
    >< Sau96I
    >< Pali
    >< NspIV
    > < HindIII
    >< HaeIII
    >< Eco0109I
    >< DraII
    >< DdeI
    >< Cfr13I
    >< BsuRI
    >< BsiZI
    >< BshI
    >< BfrI >< PssI
    >< NlaIII >< AsuI>< BsmAI
    >< AluI >< Alw26I
    AGACATGGCA AGCTTAGGCC CTTTGAGAGA GACATATCTA ATGTGCCTTT CTCCCCTGAT GGCAAACCTT
    22830      22840      22850      22860      22870      22880      22890
    >< Tru9I
    >< Pali
    >< MscI
    >< HaeIII
    >< EaeI>< MseI
    >< Tru9I >< BsuRI
    >< MseI >< BshI
    >< BspMI >< BalI
    GCACCCACCC TGCTCTTAAT TGTTATTGGC CATTAAATGA TTATGGTTTT TACACCACTA CTGGCATTGG
    22900      22910      22920      22930      22940      22950      22960
    >< Sau96I ><
    >< PalINspIV ><
    > < MspI NspHII ><
    >< HaeIII

```

FIGURE 13.53

```

                                > < HpaII Eco47I ><
                                >< DsaI
                                > < HapII Cfr13I ><
                                >< BsuRISinI ><
                                >< GdiII BsiZI ><
                                >< BsaJI
                                >< ScaI
                                >< RsaI
                                >< Csp6I
                                >< AfaI
                                >< Tru9I
                                >< MseI >< Cfr10I
                                >< DraI
                                >< BshI AsuI ><
CTACCAACCT TACAGAGTTG TAGTACTTTC TTTTGAACCT TTAAATGCAC CGGCCACGGT TTGTGGACCA
22970      22980      22990      23000      23010      23020      23030

                                >< Tru9I
                                >< Tru9I
                                >< PleI
                                >< MseI
                                >< RsaI
                                >< Csp6I
                                >< BsrI ><
                                >< BsrI
                                >< MseI
                                >< HinfI
                                >< AfaI
AAATTATCCA CTGACCTTAT TAAGAACCAG TGTGTCAATT TTAATTTTAA TGGACTCACT GGTACTGGTG
23040      23050      23060      23070      23080      23090      23100

                                >< Tru9I
                                >< MseI
                                >< MboII
                                >< HpaI
                                >< HindII
                                >< HincII
                                >< Pali
                                >< HaeIII
                                >< GdiII
                                >< EaeI
                                >< BsuRI
                                >< BshI
                                TfiI ><
                                HinfI ><
TGTTAACTCC TTCTTCAAAG AGATTTC AAC CATTTC AACA ATTTGGCCGT GATGTTTCTG ATTTCACTGA
23110      23120      23130      23140      23150      23160      23170

                                > < XhoII
                                >< TthHB8I
                                >< TaqI
                                > < Sau3AI
                                > < NdeII
                                > < MflI
                                > < MboI
                                > < DpnII
                                >< DpnI
                                > < BstYI
                                > < BspAI
                                > < SspI
                                >< AlwI >< Bsp143I
                                >< HphI
TTCCGTTTGA GATCCTAAAA CATCTGAAAT ATTAGACATT TCACCTTGCT CTTTGGGGG TGTAAGTGTA
23180      23190      23200      23210      23220      23230      23240

                                >< ScrFI
                                >< MvaI
                                >< EcoRII
                                >< Ecl136I
                                >< DsaV
                                >< BstOI
                                >< BstNI
                                >< BsiLI
                                >< ApyI
                                >< Tru9I
                                >< MseI
                                >< HpaI
                                >< HindII
                                >< Eco57I
                                >< BsgI
                                >< HincII
ATTACACCTG GAACAAATGC TTCATCTGAA GTTGCTGTTC TATATCAAGA TGTTAACTGC ACTGATGTTT
23250      23260      23270      23280      23290      23300      23310

                                >< Sau3AI
                                >< NlaIII
                                >< NdeII
                                >< MboI
                                >< DpnII
                                >< DpnI
                                >< HinfII

```

FIGURE 13. 54


```

                >< BspWI                >< Hin6I
                >< BspAI                > < HhaI                PleI ><
>< SfcI                >< Bsp143I        >< AluI> < CfoI                >< BsrI
CTACAGCAAT TCATGCAGAT CAACTCACAC CAGCTTGGCG CATATATTCT ACTGGAAACA ATGTATTCCA
    23320          23330          23340          23350          23360          23370          23380

                >< TthHB8I
                >< TaqI
                >< SalI
                >< RtrI
                >< NspI
                >< EspI >< NspHI
                >< DdeI >< NlaIII
                >< CelII >< HindII
                >< Bpu102I>< HincII
>< HinfI                >< AluI                >< AccI
GACTCAAGCA GGCTGTCTTA TAGGAGCTGA GCATGTCGAC ACTTCTTATG AGTGCACAT TCCTATTGGA
    23390          23400          23410          23420          23430          23440          23450

                > < SnaBI
                >< ScaI
                >< RsaI
                >< RmaI
                >< MaeII >< MaeI
                > < Eco105I
                >< Csp6I
                >< RmaI
                >< MaeIII
                > < BsaAI
>< AluI                >< MaeI                >< AfaI
GCTGGCATTT GTGCTAGTTA CCATACAGTT TCTTTATTAC GTAGTACTAG CCAAAAATCT ATTGTGGCTT
    23460          23470          23480          23490          23500          23510          23520

                >< MunI
ATACTATGTC TTTAGGTGCT GATAGTTCAA TTGCTTACTC TAATAACACC ATTGCTATAC CTAATAACTT
    23530          23540          23550          23560          23570          23580          23590

                RsaI ><
                >< MnlI
                Csp6I ><
                AfaI ><
>< SfcI
TTCAATTAGC ATTACTACAG AAGTAATGCC TGTTTCTATG GCTAAAACCT CCGTAGATTG TAATATGTAC
    23600          23610          23620          23630          23640          23650          23660

                > < TfiI
                > < HinfI
                >< AciI                > < AluI
ATCTGCGGAG ATTCTACTGA ATGTGCTAAT TTGCTTCTCC AATATGGTAG CTTTTCGACA CAACTAAATC
    23670          23680          23690          23700          23710          23720          23730

>< VneI
>< SduI
>< NspII
>< HgiAI                >< PmlI
>< SnoI>< DdeI                >< Sau3AI                >< PmaCI
>< Bsp1286I                >< NdeII                >< MaeII
>< BmyI                >< MboI                >< Eco72I
>< BbvI                >< DpnI                >< BsaAI
>< ApaLI                >< Bsp143I                >< BbrPI
>< Alw44I                >< DpnII >< AlwI
>< Alw21I                >< Fnu4HI                >< BspAI                >< AflIII
GTGCACTCTC AGGTATTGCT GCTGAACAGG ATCGCAACAC ACGTGAAGTG TTCGCTCAAG TCAAACAAAT
    23740          23750          23760          23770          23780          23790          23800

```

FIGURE 13.55

```

>< RsaI
>< Csp6I
>< AfaI
GTACAAAACC CCAACTTTGA AATATTTTGG TGGTTTTAAT TTTTCACAAA TATTACCTGA CCCTCTAAAG
23810 23820 23830 23840 23850 23860 23870

>< MnlI
>< MnlI
>< DdeI >< MnlI
CCAAC TAAGA GGTCTTTTAT TGAGGACTTG CTCTTTAATA AGGTGACACT CGCTGATGCT GGCTTCATGA
23880 23890 23900 23910 23920 23930 23940

>< XhoII
>< Sau3AI
>< StyI
>< RmaI
>< MaeI
>< EcoT14I
>< Eco130I
>< BssT1I
>< BsmI
>< BscCI
>< BsaJI
>< BlnI
>< AvrII
>< XhoII
>< Sau3AI
>< RmaI
>< NdeII
>< MflI
>< MboI
>< MaeI
>< MstI
>< HinPII
>< DpnII
>< HphI> < DpnI
>< HhaI
>< BstYI
>< BspAI
>< Bsp143I
>< BglII
>< FspI
>< FdiII
>< CfoI
>< AviII
AGCAATATGG CGAATGCCTA GGTGATATTA ATGCTAGAGA TCTCATTTGT GCGCAGAAGT TCAATGGACT
23950 23960 23970 23980 23990 24000 24010

>< RmaIRsaI ><
>< MnlI
>< Fnu4HI
>< Fnu4HI Csp6I ><
>< BspWI >< BbvI
>< BbvI >< BspWI >< MaeIAfaI ><
TACAGTGTG CCACCTCTGC TCACTGATGA TATGATTGCT GCCTACACTG CTGCTCTAGT TAGTGGTACT
24020 24030 24040 24050 24060 24070 24080

>< MboII
>< HinPII
>< Hin6I
>< HhaI
>< HaeII
>< Fnu4HI >< Ksp632I
>< CfoI >< EarI
>< FokI >< BspWI >< Eam1104I
>< BbvI
>< Bsp143II
GCCACTGCTG GATGGACATT TGGTGCTGGC GCTGCTCTTC AAATACCTTT TGCTATGCAA ATGGCATATA
24090 24100 24110 24120 24130 24140 24150

Tru9I ><
MseI ><
>< MaeIII
GGTTCAATGG CATTGGAGTT ACCCAAAATG TTCTCTATGA GAACCAAAAA CAAATCGCCA ACCAATTTAA
24160 24170 24180 24190 24200 24210 24220

MaeII ><
>< TfiI
>< HinfI
>< BbvI
>< Fnu4HI
>< AluI
CAAGGCGATT AGTCAAATTC AAGAATCACT TACAACAACA TCAACTGCAT TGGGCAAGCT GCAAGACGTT
24230 24240 24250 24260 24270 24280 24290

>< Tru9I
>< MseI
>< HpaI
>< HindII
>< HincII>
>< BsmI
>< BscCI
>< Tru9I
>< MseI
>< DdeI
>< BfrI
>< AluI

```

FIGURE 13. 56

```

GTTAACCAGA ATGCTCAAGC ATTAAACACA CTTGTAAAC AACTTAGCTC TAATTTTGGT GCAATTTCAA
24300      24310      24320      24330      24340      24350      24360

      >< ThaI
      >< SpoI
      >< NruI
      >< MvnI
      >< BstUI      >< TthHB8I
      >< Bsp68I      >< TaqI      >< RsaI
      >< EcoRV      >< Bsp50I      >< MnlI      >< Csp6I      >< Tru9I
      >< Eco32I      >< AccII      >< MnlI      >< AciI      >< AfaI      >< MseI
GTGTGCTAAA TGATATCCTT TCGCGACTTG ATAAAGTCGA GCGCGAGGTA CAAATTGACA GGTTAATTAC
24370      24380      24390      24400      24410      24420      24430

      >< MaeIII      >< BbvI      >< Fnu4HI      BbvI ><
AGGCAGACTT CAAAGCCTTC AACCTATGT AACACAACAA CTAATCAGGG CTGCTGAAAT CAGGGCTTCT
24440      24450      24460      24470      24480      24490      24500

      >< Fnu4HI      >< HindII
      >< BspWI      >< DdeI      >< HincII
GCTAATCTTG CTGCTACTAA AATGTCTGAG TGTGTTCTTG GACAATCAAA AAGAGTTGAC TTTGTGGAA
24510      24520      24530      24540      24550      24560      24570

      > < NspI
      > < NspHI
      > < NlaIII
      >< MaeIII
      >< NlaIII
      >< MboII      >< FokI
      >< Fnu4HI      >< BbsI      BsaAI ><
      >< AciI      >< BbvI      >< AflIII
AGGGCTACCA CCTTATGTCC TTCCCACAAG CAGCCCCGCA TGGTGTGTC TTCCTACATG TCACGTATGT
24580      24590      24600      24610      24620      24630      24640

      >< ScrFI
      >< MvaI
      >< EcoRII
      >< Ecl136I
      >< BstOI
      >< BstNI      >< HinPII
      >< MnlI      >< BslI      >< Hin6I
      >< DsaV      >< BsiYI      >< HhaI
      >< BsiLI      >< HaeII
      >< BsaJI      >< HphI      >< CfoI      >< NlaIII
      >< ApyI      >< Bsp143II      >< BspHI      EcoNI ><
GCCATCCCAG GAGAGGAACT TCACCACAGC GCCAGCAATT TGTCATGAAG GCAAAGCATA CTTCCCTCGT
24650      24660      24670      24680      24690      24700      24710

      >< MnlI
      >< BslI      >< Tru9I
      >< BsiYI      >< MseI      >< MnlI
GAAGGTGTTT TTGTGTTTAA TGGCACTTCT TGGTTTATTA CACAGAGGAA CTTCTTTTCT CCACAAATAA
24720      24730      24740      24750      24760      24770      24780

      >< DdeI      >< Tru9I
      >< BsmAI      >< SfaNI
      >< SfcI      >< Alw26I      >< MseI      >< AlwI ><
TTACTACAGA CAATACATTT GTCTCAGGAA ATTGTGATGT CGTTATTGGC ATCATTAAACA ACACAGTTTA
24790      24800      24810      24820      24830      24840      24850

      >< Sau3AI
      >< NdeII

```

FIGURE 13.57

```

>< MboI          >< P1eI          > < ScaI
>< DpnII         >< MnlI          > < Ksp632I      > < RsaI
  >< DpnI         >< DdeI   >< HinfI      >< MboII
>< BspAI         >< BspWI         > < Eam1104I      >< Csp6I
  >< Bsp143I      >< AluI          > < EarI   > < AluI   > < AfaI   > < HphI
TGATCCTCTG CAACCTGAGC TTGACTCATT CAAAGAAGAG CTGGACAAGT ACTTCAAAAA TCATACATCA
24860      24870      24880      24890      24900      24910      24920

  >< Sau3AI
  >< NdeII
  >< MboI
>< MamI
  >< DpnII
  >< DpnI
  >< BspAI
  >< Bsp143I
  >< BsiBI          >< Tru9I          >< HindII
  >< BsaBI          >< MseI          >< HincII      AciI ><
CCAGATGTTG ATCTTGCGCA CATTTCAGGC ATTAACGCTT CTGTCGTCAA CATTCAAAAA GAAATTGACC
24930      24940      24950      24960      24970      24980      24990

  >< Tru9I
  > < TfiI
  >< MnlI          >< SwaI
  >< EcoNI         >< MseI
  >< BslI          > < HinfI
>< MnlI>< BsiYI      >< DraI
GCCTCAATGA GGTGCTAAA AATTAAATG AATCACTCAT TGACCTTCAA GAATTGGGAA AATATGAGCA
25000      25010      25020      25030      25040      25050      25060

  >< StyI
  >< Pali
  >< HaeIII
  >< EcoT14I
  >< Eco130I
  >< BsuRI
  >< BssTII
  >< Tru9I>< BshI          NlaIII ><
  >< MseI   >< BsaJI          MaeIII ><
  >> BstXI
ATATATTAAA TGGCCTTGGT ATGTTTGGCT CGGCTTCATT GCTGGACTAA TTGCCATCGT CATGGTTACA
25070      25080      25090      25100      25110      25120      25130

  > < SphI
  > < PaeI
  >< SpeI          > < NspI
  > < RmaI          > < NspHI
  >< NlaIII        > < NlaIII
  > < MaeI          >< MnlI>< BbvI   Fnu4HI ><
ATCTTGCTTT GTTGCATGAC TAGTTGTTGC AGTTGCCTCA AGGCTGCATG CTCTTGTTGGT TCTTGCTGCA
25140      25150      25160      25170      25180      25190      25200

  >< FokI
  >< DdeI
>< MnlI >< P1eI>< HinfI >< BsrI
AGTTTGATGA GGATGACTCT GAGCCAGTTC TCAAGGGTGT CAAATTACAT TACACATAAA CGAACTTATG
25210      25220      25230      25240      25250      25260      25270

  >< Sau3AI
  >< NdeII
  >< MboI
  >< DpnII
  > < DpnI

```

FIGURE 13.58

```

                >< BspAI
                > < Bsp143I
                >< BsgI                >< AlwI                >< BsrI                BspWI >
GATTTGTTTA TGAGATTTT TACTCTTGGA TCAATTACTG CACAGCCAGT AAAAATTGAC AATGCTTCTC
25280      25290      25300      25310      25320      25330      25340

                >< ScaI
                >< RsaI
                >< Csp6I                >< SfcI
                >< AfaI                >< NlaIII                >< AciI                >< MnlI                FokI >
CTGCAAGTAC TGTTCATGCT ACAGCAACGA TACCGCTACA AGCCTCACTC CCTTTCGGAT GGCTTGTTAT
25350      25360      25370      25380      25390      25400      25410

                > < HinPII
                > < Hin6I
                >< HhaI                RmaI ><
                >< HaeII                >< HinPII                NheI ><
                >< Eco47III                >< Hin6I                MaeI ><
                >< CfoI                >< HhaI                Fnu4HI ><
                >< BspWI                >< Bsp143II                >< CfoI                AluI ><
TGGCGTTGCA TTTCTTGCTG TTTTTCAGAG CGCTACCAAA ATAATTGCGC TCAATAAAAG ATGGCAGCTA
25420      25430      25440      25450      25460      25470      25480

                >< EcoNI
                >< BslI
                >< BsiYI                >< MaeIII
                >< BbvI                >< BsrI                >< BbvI                > < Fnu4HI                BbvI ><
GCCCTTTATA AGGGCTTCCA GTTCATTTGC AATTTACTGC TGCTATTTGT TACCATCTAT TCACATCTTT
25490      25500      25510      25520      25530      25540      25550

                Zsp2I ><
                Ppu10I ><
                > < SfcI                >< HinPII                NsiI ><
                >< PstI                >< Hin6I                >< RsaI                Mph1103I ><
                > < Fnu4HI                >< HhaI                >< Csp6I                EcoT22I ><
                >< BspMI                >< MnlI                >< CfoI                >< AfaI                >< MnlI                AvaIII ><
TGCTTGTCGC TGCAGGTATG GAGGCGCAAT TTTTGTACCT CTATGCCTTG ATATATTTTC TACAATGCAT
25560      25570      25580      25590      25600      25610      25620

                >< SfaNI
                >< NspI
                >< NspHI
                >< NlaIII                >< SfaNI
CAACGCATGT AGAATTATTA TGAGATGTTG GCTTTGTTGG AAGTGCAAAT CCAAGAACCC ATTACTTTAT
25630      25640      25650      25660      25670      25680      25690

                >< Bst1107I
                >< AccI                MaeIII ><
GATGCCAACT ACTTTGTTTG CTGGCACACA CATAACTATG ACTACTGTAT ACCATATAAC AGTGTCACAG
25700      25710      25720      25730      25740      25750      25760

                >< MboII
                BstXI ><
                >< MunI >< MaeIII                >< MaeIII                >< Eco57I                >< BbsI MnlI >
ATACAATTGT CGTTACTGAA GGTGACGGCA TTTCAACACC AAAACTCAAA GAAGACTACC AAATTGGTGG
25770      25780      25790      25800      25810      25820      25830

                >< RsaI
                > < NlaIII
                >< HphI
                >< Tru9I >< Tth111I >< Csp6I
                >< DdeI                >< DdeI                >< MseI >< AspI                >< AfaI

```

FIGURE 13.59

```

TTATTCTGAG GATAGGCACT CAGGTGTTAA AGACTATGTC GTTGTACATG GCTATTTTCAC CGAAGTTTAC
25840      25850      25860      25870      25880      25890      25900

                > < HinfI>< P1eI                >< BsrI                Tru9I ><
                >< AluI >< AccI                >< SfcI >< AlwNI                MseI ><
                >< MboII                HindIII >
TACCAGCTTG AGTCTACACA AATTACTACA GACACTGGTA TTGAAAATGC TACATTCTTC ATCTTTAACA
25910      25920      25930      25940      25950      25960      25970

                > < TthHB8I
                >< Tru9I                >< TaqI                >< Ksp632I
                >< MseI                >< MboII                >< EarI BspWI ><
                >< AluI                >< Eco57I                >< Eam1104I AlwI ><
AGCTTGTTAA AGACCCACCG AATGTGCAAA TACACACAAT CGACGGCTCT TCAGGAGTTG CTAATCCAGC
25980      25990      26000      26010      26020      26030      26040

                >< XhoII
                >< Sau3AI
                >< NlaIV
                >< NdeII
                >< MflI
                >< MboI
                >< DpnII
                >< DpnI
                >< BstYI
                >< BstI
                >< BspAI
                >< Bsp143I
                >< BscBI
                >< BamHI >< AlwI
                >< RmaI
                >< MaeI
                >< RsaI ><
                >< Csp6I ><
                >< AfaI ><
AATGGATCCA ATTTATGATG AGCCGACGAC GACTACTAGC GTGCCTTTGT AAGCACAAGA AAGTGAGTAC
26050      26060      26070      26080      26090      26100      26110

                > < Tru9I
                >< RsaI
                > < MseI
                >< MboII
                >< MaeII
                >< RsaI
                >< Csp6I
                >< AfaI
                >< Csp6I >< Tru9I >< Csp6I
                >< AfaI >< MseI >< AfaI
GAAC TTATGT ACTCATTCGT TTCGGAAGAA ACAGGTACGT TAATAGTTAA TAGCGTACTT CTTTTCTTG
26120      26130      26140      26150      26160      26170      26180

                >< TthHB8I
                >< TaqI
                >< RmaI
                > < MaeIII
                >< MaeI >< RmaI
                >< FokI >< MaeI
                >< HinP1I
                >< Hin6I
                > < RsaI
                >< HhaI
                >< CfoI >< BbvI > < AfaI
                >< Csp6I
CTTTCGTGGT ATTCTTGCTA GTCACACTAG CCATCCTTAC TGCCTTCGA TTGTGTGCGT ACTGCTGCAA
26190      26200      26210      26220      26230      26240      26250

                >< Tru9I
                >< ThaI
                >< MseI
                >< MvnI
                >< SspI >< MaeII
                >< HpaI
                >< HindII
                >< HincII
                >< BstUI
                >< MaeII >< Bsp50I >< MboII EarI >
                >< AccI >< AccII
                >< Ksp632I >
                >< Eam1104I >
TATTGTTAAC GTGAGTTTAG TAAAACCAAC GGTTTACGTC TACTCGCGTG TTAAAAATCT GAAC TCTTCT
26260      26270      26280      26290      26300      26310      26320

```

FIGURE 13.60

```

        >< Sau3AI
        >< NdeII
        >< MboI
        >< DpnII
    >< MboII>< DpnI
    >< XmnI >< BspAI> < Eco57I
    >< Asp700I>< Bsp143I
GAAGGAGTTC CTGATCTTCT GGTCTAAACG AACTAACTAT TATTATTATT CTGTTTGGAA CTTTAACATT
26330      26340      26350      26360      26370      26380      26390

                                >< ScrFI
                                >< MvaI
                                >< EcoRII
                                >< Ecl136I
                                >< DsaV NlaIV ><
                                >< BstOI
                                >< BstNI RmaI ><
                                >< BsiLI MaeI ><
                                >< ApyIBscBI ><
        >< RsaI
        >< MnlI
        >< Csp6I
        >< AfaI
        >< NlaIII
        >< AluI
GCTTATCATG GCAGACAACG GTACTATTAC CGTTGAGGAG CTTAAACAAC TCCTGGAACA ATGGAACCTA
26400      26410      26420      26430      26440      26450      26460

                                >< ScrFI
                                >< RmaI
                                >< MvaI
                                >< MaeI
                                >< EcoRII
                                >< Ecl136I
                                >< DsaV
                                >< BstOI
                                >< BstNI
                                >< BsiLI
                                >< ApyI >< MaeIII
GTAATAGGTT TCCTATTCCT AGCCTGGATT ATGTTACTAC AATTTGCCTA TTCTAATCGG AACAGGTTTT
26470      26480      26490      26500      26510      26520      26530

                                >< Pali
                                >< MscI
                                >< MnlI >< MaeIII
                                >< HaeIII
                                >< EaeI
                                >< BsuRI
                                >< BsrI
                                >< BspWI
                                >< BshI
                                >< Ball
                                >< BbvI Fnu4HI ><
        >< RsaI
        >< Csp6I >< HindIII
        >< AfaI >< AluI
TGTACATAAT AAAGCTTGTT TTCCTCTGGC TCTTGTGGCC AGTAACACTT GCTTGTTTTG TGCTTGCTGC
26540      26550      26560      26570      26580      26590      26600

                                >< VspI
                                >< Tru9I
                                >< MseI
                                >< HphI
        >< SfcI >< AsnI >< BsrI
        >< AccI >< AseI>< MaeIII>< AciI
TGTCTACAGA ATTAATTGGG TGACTGGCGG GATTGCGATT GCAATGGCTT GTATTGTAGG CTTGATGTGG
26610      26620      26630      26640      26650      26660      26670

    >< EspI
    >< Eco57I
    >< DdeI
    >< CclII
    >< Epu1102I
                                >< RsaI
                                >< Csp6I

```

FIGURE 13.61

```

>< BfrI                                     >< AfaI
  >< AluI                                     >< AciI                                     MboII >
CTTAGCTACT TCGTTGCTTC CTTCAAGGCTG TTTGCTCGTA CCCGCTCAAT GTGGTCATTC AACCCAGAAA
26680      26690      26700      26710      26720      26730      26740

      >< ScrFI
      >< NciI
      >< MspI
      >< HpaII
      >< HapII
      >< DsaV>< MnlI
      >< BslI
      >< BsiYI
      >< BsaJI >< MunI      > < XcmI
      >< BcnI      >< MaeIII >< AciI >< NlaIII
CAAACATTCT TCTCAATGTG CCTCTCCGGG GGACAATTGT GACCAGACCG CTCATGGAAA GTGAACCTGT
26750      26760      26770      26780      26790      26800      26810

      Tru9I ><
      SinI >
      Sau96I >
      PpuMI >
      NspIV >
      MseI ><
      >< MaeIII
      >< Sau3AI      > < RmaI >< HaeII
      >< NdeII      >< Pali      > < MaeI      EcoO109I >
      >< MboI      >< MspI      >< HinP1IEco47I >
      >< FbaI      >< HpaII      >< StyI>< Hin6I DraII >
      >< DpnII      >< HapII      >< EcoT14I      Cfr13I >
      >< DpnI      >< HaeIII      >< Eco130I>< Bsp143II
      >< BspAI      >< GdiII      >< BssT1I      Bsi2I >
      >< Bsp143I      >< EaeI      >< BsaJI      Bmel8I >
      >< BsiQI      >< BsuRI      >< BlnI >< HhaI AvaII >
      >< BclI      >< MaeIII      >< BshI      >< AvrII >< CfoI AsuI >
CATTGGTGCT GTGATCATTC GTGGTCACTT GCGAATGGCC GGACACTCCC TAGGGCGCTG TGACATTAAG
26820      26830      26840      26850      26860      26870      26880

      >< Sau3AI
      >< NdeII
      >< MboI
      >< DpnII
      >< DpnI
      >< PssI >< BspMI
      >< Psp5II      >< BspAI      >< XmnI
      >< NspHII      >< Bsp143I      >< Asp700I > < HgaI      Fnu4HI ><
GACCTGCCAA AAGAGATCAC TGTGGCTACA TCACGAACGC TTTCTTATTA CAAATTAGGA GCGTCGCAGC
26890      26900      26910      26920      26930      26940      26950

      >< TfiI
      >< HinfI
      >< BbvI      > < Tru9I
      >< BbvI      >< Fnu4HI >< AciI      > < MseI
GTGTAGGCAC TGATTCAGGT TTTGCTGCAT ACAACCGCTA CCGTATTGGA AACTATAAAT TAAATACAGA
26960      26970      26980      26990      27000      27010      27020

      >< MspI      >< RsaI
      >< HpaII      >< RmaI
      >< HapII      >< Csp6I
      >< Cfr10I      >< MaeI>< BcgI      HindII ><
      >< BcgI/a      >< SspI      >< AfaI >< MaeIII      HincII ><

```

FIGURE 13.62


```

CCACGCCGGT AGCAACGACA ATATTGCTTT GCTAGTACAG TAAGTGACAA CAGATGTTTC ATCTTGTTGA
  27030      27040      27050      27060      27070      27080      27090

    >< ScrFI
    >< MvaI
      >< MaeIII
    >< EcoRII
      >< Ecl136I
    >< DsaV
      >< BstOI
      >< BstNI
      >< BsiLI
      >< ApyI
      >< MnlI
      >< TfiI
      >< HinfI ><
CTTCCAGGTT ACAATAGCAG AGATATTGAT TATCATTATG AGGACTTTCA GGATTGCTAT TTGGAATCTT
  27100      27110      27120      27130      27140      27150      27160

      >< BsmAI
      >< Tru9I
      > < MnlI
    >< MaeII
      >< Alw26I
      >< MseI
      >< DdeI
      >< MboII
GACGTTATAA TAAGTTCAAT AGTGAGACAA TTATTTAAGC CTCTAACTAA GAAGAATTAT TCGGAGTTAG
  27170      27180      27190      27200      27210      27220      27230

      >< Ksp632I
      >< MboII
      >< EarI
      >< NlaIII Eam1104I ><
ATGATGAAGA ACCTATGGAG TTAGATTATC CATAAACGA ACATGAAAAT TATTCTCTTC CTGACATTGA
  27240      27250      27260      27270      27280      27290      27300

      > < RsaI >< RsaI
      >< Csp6I >< Csp6I
      > < AluI
      >< MnlI
      > < AfaI >< AfaI
TTGTATTTAC ATCTTGCGAG CTATATCACT ATCAGGAGTG TGTTAGAGGT ACGACTGTAC TACTAAAAGA
  27310      27320      27330      27340      27350      27360      27370

      >< MnlI
      >< HphI
      >< HphI
      >< MnlI
ACCTTGCCCA TCAGGAACAT ACGAGGGCAA TTCACCATTT CACCCTCTTG CTGACAATAA ATTTGCACTA
  27380      27390      27400      27410      27420      27430      27440

      >< RmaI
      >< MaeI
      >< TthHB8I
      >< TaqI
      >< RsaI
      >< Csp6I
      >< BbvI
      >< AfaI
      >< AluI
      >< RmaI
      >< MaeI
      >< BspWI
      >< Bsp1286I
      >< BmyI
      >< BanII
      >< Alw21I
      >< SstI
      >< SduI
      >< SacI
      >< NspII
      >< HgiAI
      >< Eco24I
      > < Ecl136II
      >< BspWI
      >< Bsp1286I
      >< BmyI
      >< BanII
      >< Alw21I
      >< HphI
      >< DpnI
      >< MnlI

```

FIGURE 13. 63

```

>< Bsp143I          >< MnlI          > < AluI      BbvI ><
GATCAGTTTC ACCAAACTT TTCATCAGAC AAGAGGAGGT TCAACAAGAG CTCTACTCGC CACTTTTTCT
27520      27530      27540      27550      27560      27570      27580

SstI ><
SduI ><
SacI ><
NspII ><
HgiAI ><
Eco24I ><
Ecl136II ><
Bsp1286I ><
BmyI ><
BanII ><
Alw21I ><
AluI ><
>< RmaI >< Tru9I
>< MaeI >< MseI          >< Tru9I          Alw21I ><
>< Fnu4HI          >< HphI          >< MseI          AluI ><
CATTGTTGCT GCTCTAGTAT TTTTAATACT TTGCTTCACC ATTAAGAGAA AGACAGAATG AATGAGCTCA
27590      27600      27610      27620      27630      27640      27650

>< Tru9I          >< Tru9I
>< MseI          >< MseI
CTTTAATTGA CTTCTATTTG TGCTTTTtag CCTTCTGCT ATTCCTTGTT TTAATAATGC TTATTATATT
27660      27670      27680      27690      27700      27710      27720

>< XhoII
>< XbaI
> < ScrFI
>< Sau3AI
>< RmaI
>< NdeII
> < MvaI
>< MflI
>< MboI
>< EcoRII>< MaeI
> < Ecl136I
>< DpnII
>< DpnI
>< BstYI
> < BstOI
> < BstNI
>< TthHB8I >< BspAI          > < RsaI
>< DsaV>< Bsp143I          >< MboII
> < BsiLI          >< Csp6I
>< TaqI > < ApyI > < AlwI > < AfaI          >< NlaIII
TTGGTTTTCa CTCGAAATCC AGGATCTAGA AGAACCTTGT ACCAAAGTCT AAACGAACAT GAAACTTCTC
27730      27740      27750      27760      27770      27780      27790

>< HinPII
>< Hin6I
>< HhaI
>< RsaI >< HaeII
>< SfcI          >< Eco47III
>< Csp6I>< CfoI SfaNI ><
>< NdeI          >< AfaI >< Bsp143II
ATTGTTTTGA CTTGTATTTT TCTATGCAGT TGCATATGCA CTGTAGTACA GCGCTGTGCA TCTAATAAAC
27800      27810      27820      27830      27840      27850      27860

>< XhoII
>< Sau3AI
>< NdeII
> < MnlI
>< MflI

```

FIGURE 13.64

```

>< MboI
>< DpnII
    >< DpnI    >< RsaI
    >< BstYI    >< MboII
>< NlaIII>< BspAI    >< Csp6I >< RmaI
    >< AlwI >< Bsp143I    >< AfaI >< MaeI
CTCATGTGCT TGAAGATCCT TGTAAGGTAC AACACTAGGG GTAATACTTA TAGCACTGCT TGGCTTTGTG
27870      27880      27890      27900      27910      27920      27930

>< SduI
>< RmaI
>< NspII
>< MaeI
>< HgiAI
>< Bsp1286I
>< BmyI
>< Alw21I
                                >< NspI
                                >< NspHI
                                >< NlaIII >< MaeIII
CTCTAGGAAA GGTTTACCT TTTCATAGAT GGCACACTAT GGTTCAAACA TGCACACCTA ATGTTACTAT
27940      27950      27960      27970      27980      27990      28000

    > < XhoII
    > < Sau3AI > < Van91I
        >< PvuII
        >< Psp5I
    > < NdeII > < PflMI
    > < MflI>< NspBII
    > < DpnII    >< HinP1I
        >< Bsp143I    >< Hin6I
    > < BstYI > < BslI >< HhaI >< RmaI
    > < BspAI > < BsiYI>< CfoI >< MaeI
    > < MboI>< AluI>< BspWI >< BspWI
    >< AlwI >< DpnI > < AccB7I    >< AluI
CAACTGTCAA GATCCAGCTG GTGGTGCCT TATAGCTAGG TGTTGGTACC TTCATGAAGG TCACCAAAC
28010      28020      28030      28040      28050      28060      28070

                                >< SinI
                                >< Sau96I
                                >< NspIV
                                NspHII ><
                                NlaIV ><
                                >< Eco47I
                                >< Cfr13I
                                >< BsiZI
                                BscBI ><
                                >< Bmel8I
                                >< AvaII
                                >< AsuI
>< Fnu4HI    >< RsaI
>< Esp3I    >< Csp6I    >< Tru9I
>< BsmAI    >< BsmBI    >< MseI
>< Alw26I    >< AfaI    >< DraI
GCTGCATTTA GAGACGTACT TGTTGTTTTA AATAAACGAA CAAATTAAAA TGTCTGATAA TGGACCCCAA
28080      28090      28100      28110      28120      28130      28140

                                >< SinI
                                >< Sau96I
                                >< NspIV
                                >< NspHII
                                >< NlaIV
                                >< Eco47I
                                >< Cfr13I
                                >< BsiZI
                                >< BscBI
                                >< Bmel8I
                                >< AvaII
                                >< TfiI
                                >< AsuI
                                >< HinfI
                                >< MnlI
>< SduI
>< NspII
>< Bsp1286I
>< BmyI
>< MaeII
>< AciI

```

FIGURE 13. 65

```

TCAAACCAAC GTAGTGCCCC CCGCATTACA TTTGGTGGAC CCACAGATTC AACTGACAAT AACCAGAATG
28150      28160      28170      28180      28190      28200      28210

      >< HinP1I >< StyI
      >< HaeII
      > < Pali >< Hin6I >< EcoT14I
      > < HaeIII >< HhaI>< Eco130I
      >< BspWI >< BssT1I
      > < BsuRI >< Bsp143II
      >< HgaI> < BshI >< CfoI>< BsaJI >< HgaI
GAGGACGCAA TGGGGCAAGG CCAAAACAGC GCCGACCCCA AGGTTTACCC AATAATACTG CGTCTTGTT
28220      28230      28240      28250      28260      28270      28280

      >< TthHB8I
      > < ScrFI
      >< Pali
      >< PaeR7I
      >< NspIII
      > < MvaI
      >< HaeIII
      >< EcoRII
      >< Eco88I
      >< XhoI > < Ecl136I
      >< DsaV
      >< BsuRI
      >< SlaI > < BstOI
      >< MnlI>< TaqI> < BstNI
      >< CcrI > < BsiLI
      >< HinfI >< BshI
      >< TfiI>< BcoI>< BsaJI
      >< MnlI >< DdeI >< Aval > < ApyI
      >< AluI >< DdeI > < NlaIII >< BfrI >< Ama87I >< MnlI
CACAGCTCTC ACTCAGCATG GCAAGGAGGA ACTTAGATTC CCTCGAGGCC AGGGCGTTCC AATCAACACC
28290      28300      28310      28320      28330      28340      28350

      >< SinI
      >< Sau96I
      >< NspIV
      >< NspHII
      >< Eco47I
      >< Cfr13I
      >< Bsi2I
      >< Bme18I > < Ksp632I
      >< AvalI > < Eam1104I
      >< AsuI > < EarI > < AluI>< MboII >< MaeIII
AATAGTGGTC CAGATGACCA AATTGGCTAC TACCGAAGAG CTACCCGACG AGTTCGTGGT GGTGACGGCA
28360      28370      28380      28390      28400      28410      28420

      >< SstI
      >< SduI
      >< SacI
      >< NspII
      >< HgiAI
      >< EspI
      >< Eco24I
      >< Ecl136II >< StyI >< Sau96I
      >< DdeI >< RmaI >< NspIV
      >< CelII >< MaeI >< HaeIII
      >< Bsp1286I >< EcoT14I >< Cfr13I
      >< Bpu1102I >< Eco130I >< BsuRI
      >< BmyI >< BssT1I > < BsrI
      >< BanII >< RsaI >< BsaJI >< Bsi2I

```

FIGURE 13.66

```

        >< Alw21I      >< Csp6I      >< BlnI      >< BshI>< HindIII
>< HphI  >< AluI      >< AfaI      >< AvrII  >< AsuI  >< AluI
AAATGAAAGA GCTCAGCCCC AGATGGTACT TCTATTACCT AGGAACTGGC CCAGAAGCTT CACTTCCCTA
28430      28440      28450      28460      28470      28480      28490

>< HinP1I
>< Hin6I
>< HhaI
>< HaeII
>< CfoI                      > < MnlI      >< NlaIV
>< Bsp143II                  >< SfaNI >< DdeI >< BscBI
CGGCGCTAAC AAAGAAGGCA TCGTATGGGT TGCAACTGAG GGAGCCTTGA ATACACCCAA AGACCACATT
28500      28510      28520      28530      28540      28550      28560

>< NlaIV
>< Eco64I
>< BscBI
>< BanI
>< AciI
>< AccB1I >< BbvI      >< Fnu4HI                      >< MnlI
GGCACCCGCA ATCCTAATAA CAATGCTGCC ACCGTGCTAC AACTTCCTCA AGGAACAACA TTGCCAAAAG
28570      28580      28590      28600      28610      28620      28630

                                                >< ThaI
                                                >< MnlI
                                                >< MaeII >< MvnI
                                                >< MnlI      BstUI ><
                                                >< Fnu4HI      >< Ksp632I      Bsp50I ><
                                                >< BspWI      >< EarI      >< BsaAI>< AciI
>< MnlI      >< MnlI      >< AciI>< MboII      >< Eam1104I      AccII ><
GCTTCTACGC AGAGGGAAGC AGAGGCGGCA GTCAAGCCTC TTCTCGCTCC TCATCACGTA GTCGCGGTAA
28640      28650      28660      28670      28680      28690      28700

        >< ScrFI
        >< MvaI
        >< EcoRII
        >< Ecl136I
        >< DsaV>< Fnu4HI
        >< BstOI
        >< BstNI
        >< BsiLI
        >< ApyI                      >< BbvI      >< TaqI      >< AciI
TTCAAGAAAT TCAACTCCTG GCAGCAGTAG GGGAAATTCT CCTGCTCGAA TGGCTAGCGG AGGTGGTGAA
28710      28720      28730      28740      28750      28760      28770

        > < ThaI
        > < MvnI
>< HphI  >< MnlI
> < HinP1I
> < Hin6I
>< HhaI
> < BstUI      >< RmaI
> < Bsp50I      >< MaeI
>< BbvI >< CfoI>< Fnu4HI
> < AccII>< BspWI                      >< AluI
ACTGCCCTCG CGCTATTGCT GCTAGACAGA TTGAACCAGC TTGAGAGCAA AGTTTCTGGT AAAGGCCAAC
28780      28790      28800      28810      28820      28830      28840

                                                RsaI ><
                                                >< MnlI
> < PalI>< MaeIII
> < HaeIII
> < BsuRI      >< DdeI      >< Fnu4HI      MaeII ><
                                                >< DdeI      Csp6I ><

```

FIGURE 13.67

```

> < BshI > < BbvI > < MnlI > < BspWI > < SfaNI > < AfaI > <
AACACAAGG CCAAAGTGC ACTAAGAAAT CTGCTGCTGA GGCATCTAAA AAGCCTCGCC AAAAACGTAC
28850 28860 28870 28880 28890 28900 28910

>< Tth111I
>< SinI
>< Sau96I
>< NspIV
>< NspHII
> < MaeII
>< Eco47I
>< Cfr13I
>< BsmBI
>< BsiZI >< StyI
>< Bme18I >< EcoT14I
>< MaeII >< Esp3I >< AvaII >< Eco130I
>< Csp6I >< BsmAI >< AsuI >< BssT1I
>< AfaI >< Alw26I> < AspI >< BsaJI
TGCCACAAAA CAGTACAACG TCACTCAAGC ATTTGGGAGA CGTGGTCCAG AACAAACCCA AGGAAATTTTC
28920 28930 28940 28950 28960 28970 28980

>< SinI
>< Sau96I
>< NspIV
>< NspHII
>< NlaIV
>< Eco47I
>< Cfr13I
>< BsiZI
>< BscBI
>< Bme18I
>< AvaII
>< AsuI
GGGGACCAAG ACCTAATCAG ACAAGGAAGT GATTACAAAC ATTGGCCGCA AATTGCACAA TTTGCTCCAA
28990 29000 29010 29020 29030 29040 29050

>< BsmI
>< BscCI >< MnlI >< MaeIII
>< NlaIII
>< MaeIII
>< NlaIII
GTGCCTCTGC ATTCTTTGGA ATGTCACGCA TTGGCATGGA AGTCACACCT TCGGGAACAT GGCTGACTTA
29060 29070 29080 29090 29100 29110 29120

>< XhoII
>< Sau3AI
>< NdeII
>< MflI
>< MboI
>< FokI
>< Tru9I
>< NlaIV
>< NlaIII
>< MseI
>< BscBI >< BstXI>< AlwI> < Bsp143I
>< AspI
>< BspWI ><
TCATGGAGCC ATTAAATTGG ATGACAAAGA TCCACAATTC AAAGACAACG TCATACTGCT GAACAAGCAC
29130 29140 29150 29160 29170 29180 29190

EspI ><
DdeI ><
CclII ><
Bpu1102I ><
AluI ><
>< HgaI
ATTGACGCAT ACAAACATT CCCACCAACA GAGCCTAAAA AGGACAAAAA GAAAAAGACT GATGAAGCTC
29200 29210 29220 29230 29240 29250 29260

```

FIGURE 13.68

```

                                >< PleI
                                >< MboII
                                >< Ksp632I >< GsuI
                                >< EarI>< Fnu4HI
                                >< HinfI >< Eam1104I>< BpmI
                                >< Fnu4HI >< BbvI >< AciI >< NlaIII
AGCCTTTGCC GCAGAGACAA AAGAAGCAGC CCACTGTGAC TCTTCTTCCT GCGGCTGACA TGGATGATTT
29270      29280      29290      29300      29310      29320      29330

                                >< NlaIII >< HinfI >< NlaIII ><
                                >< FokI >< AluI >< TfiI>< DdeI >< BspHI
CTCCAGACAA CTTCAAATTT CCATGAGTGG AGCTTCTGCT GATTCAACTC AGGCATAAAC ACTCATGATG
29340      29350      29360      29370      29380      29390      29400

                                >< MaeII >< AccI
ACCACACAAG GCAGATGGGC TATGTAAACG TTTTCGCAAT TCCGTTTACG ATACATAGTC TACTCTTGTG
29410      29420      29430      29440      29450      29460      29470

                                >< Tru9I
                                >< Tru9I
                                >< MseI
                                >< MseI
                                >< HpaI
                                >< HindII
                                >< HincII
                                >< Tru9I ><
                                >< MseI ><
CAGAATGAAT TCTCGTAACT AAACAGCACA AGTAGGTTTA GTTAACTTTA ATCTCACATA GCAATCTTTA
29480      29490      29500      29510      29520      29530      29540

                                XorII >
                                TthHB8I >
                                TaqI >
                                Sau3AI ><
                                RsaI ><
                                >< ThaIPvuI >
                                NdeII ><
                                >< MnlI
                                >< MvnIMcrI >
                                MboI ><
                                DpnII ><
                                DpnI ><
                                Csp6I ><
                                >< BstUI
                                >< HaeIII BspCI >
                                BspAI ><
                                >< TthHB8I >< Bsp50I
                                >< Pali Bsp143I ><
                                >< BsuRI BsiEI >
                                >< BshIAfaI ><
                                >< MnlI
                                >< MaeIII
                                >< MnlI >< AccII
ATCAATGTGT AACATTAGGG AGGACTTGAA AGAGCCACCA CATTTTTCATC GAGGCCACGC GGAGTACGAT
29550      29560      29570      29580      29590      29600      29610

                                >< SduI
                                >< NspII
                                >< MboII >< VspI
                                >< Ksp632I >< Eco24I >< Tru9I
                                >< Bsp1286I >< MseI
                                >< BmyI >< AsnI
                                >< AfaI >< BbvI >< AluI>< Eam1104I >< BanII >< AseI

```

FIGURE 13.69

```
CGAGGGTACA GTGAATAATG CTAGGGAGAG CTGCCTATAT GGAAGAGCCC TAATGTGTAA AATTAATTTT
29620      29630      29640      29650      29660      29670      29680

                >< Tru9I    >< DdeI
                >< MseI    >< BfrI
                >< NlaIII   > < AluI
AGTAGTGCTA TCCCCATGTG ATTTTAATAG CTTCTTAGGA GAATGACAAA AAAAAAAAAA AAAAAA
29690      29700      29710      29720      29730      29740
```


SRAS serology : Indirect N technique(first set)

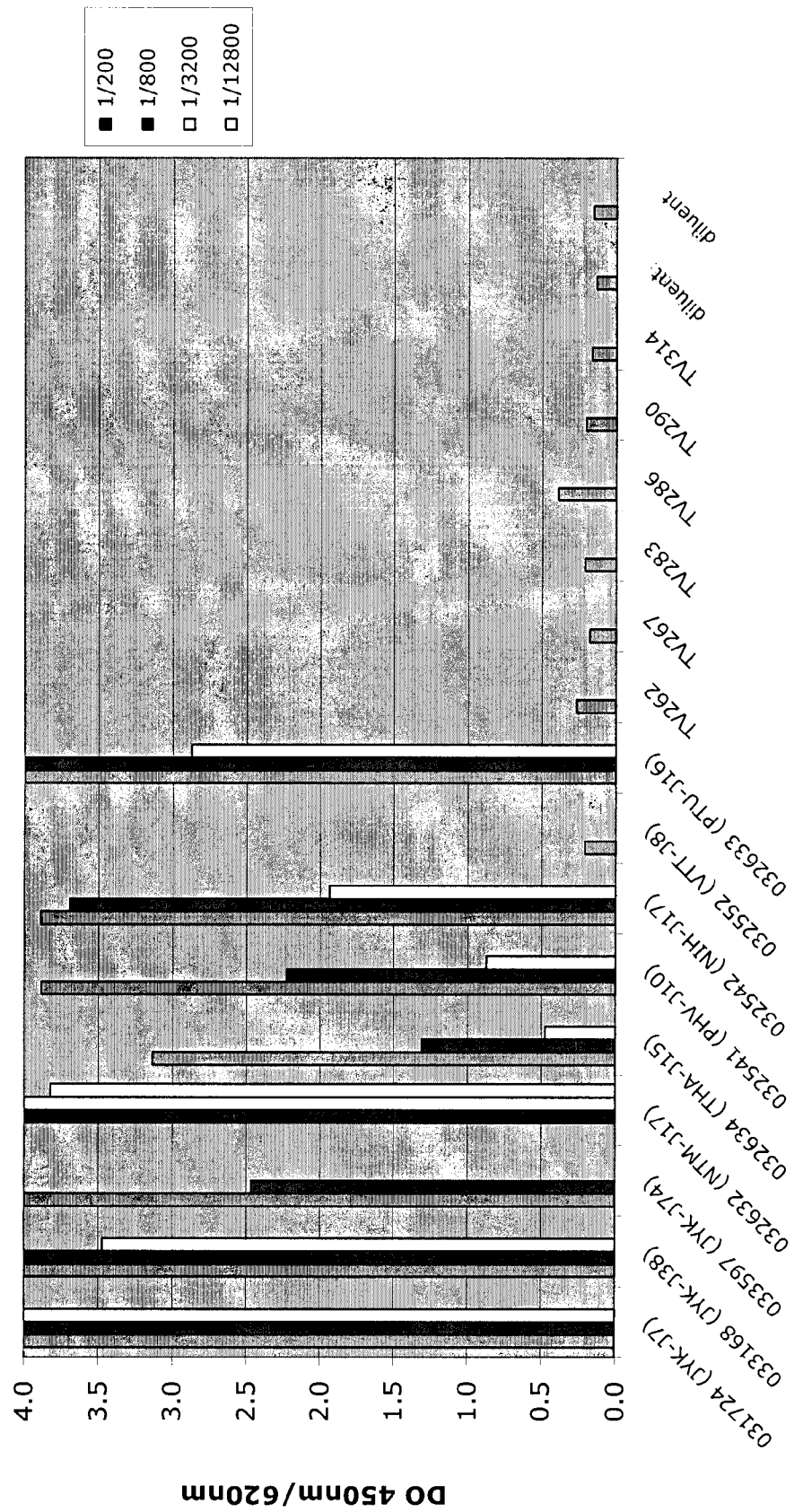
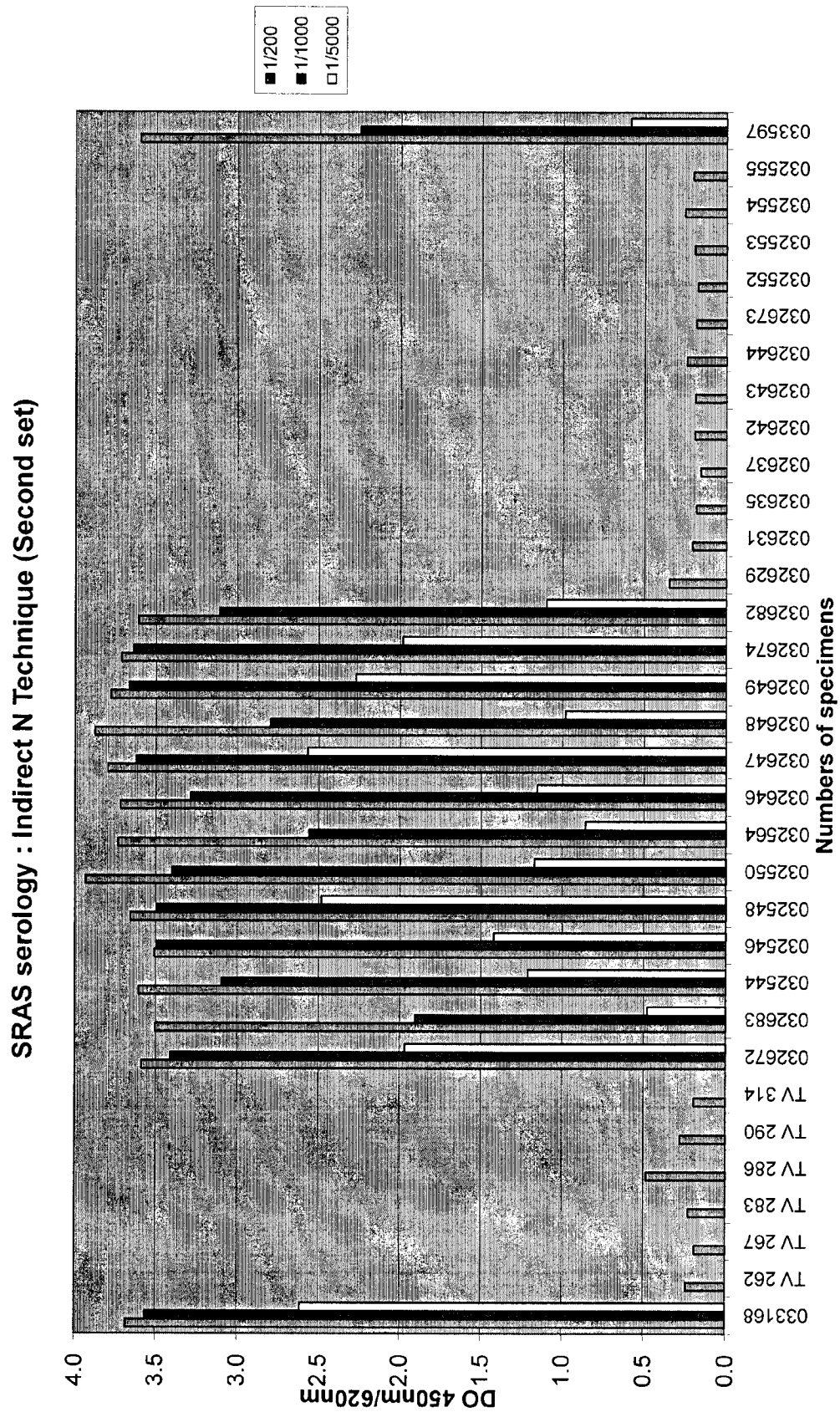


FIGURE 14



SRAS serology: Double Epitope Technique (First set)

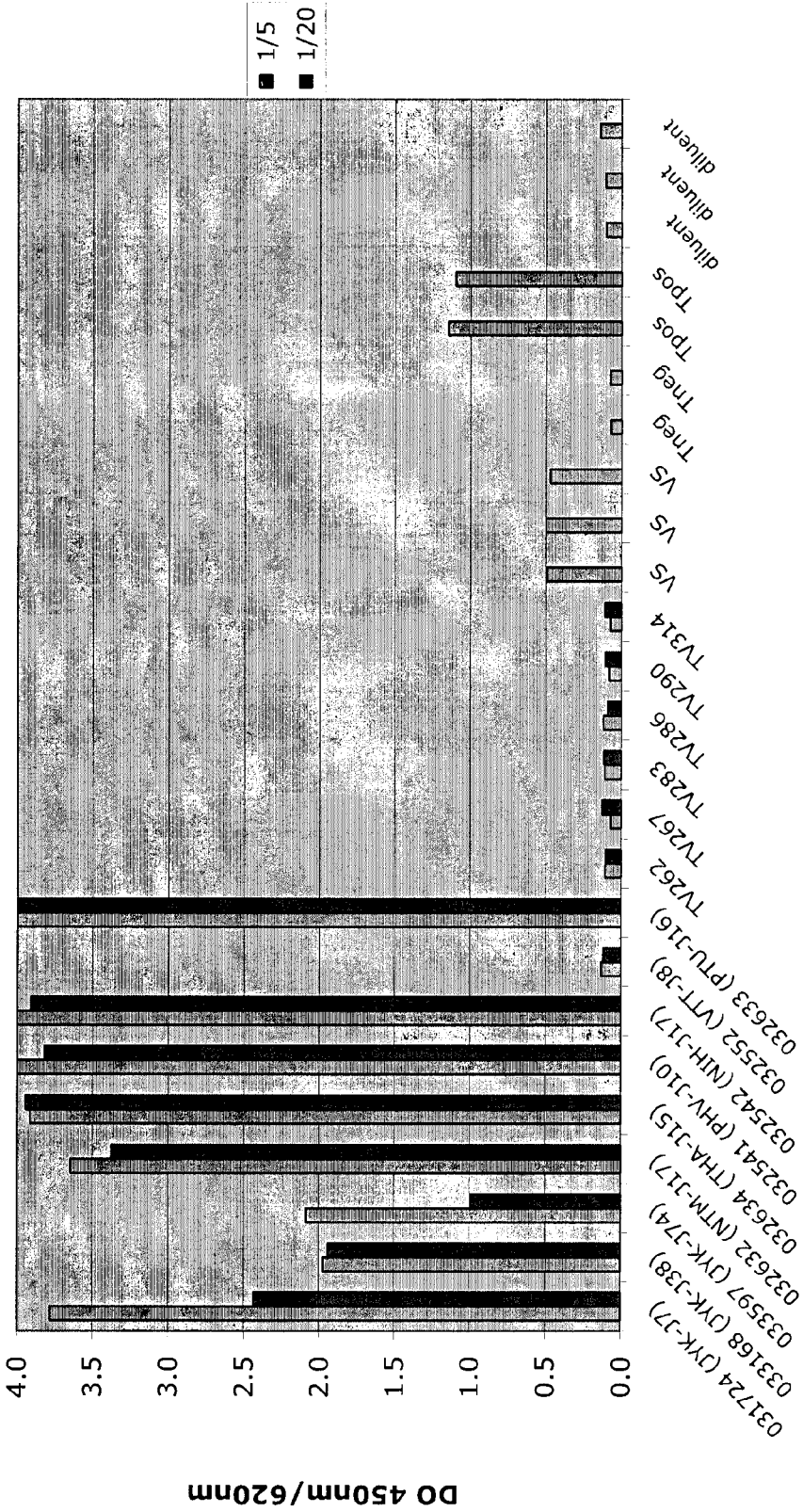


FIGURE 16

SRAS serology : Double Epitope Technique (Second set)

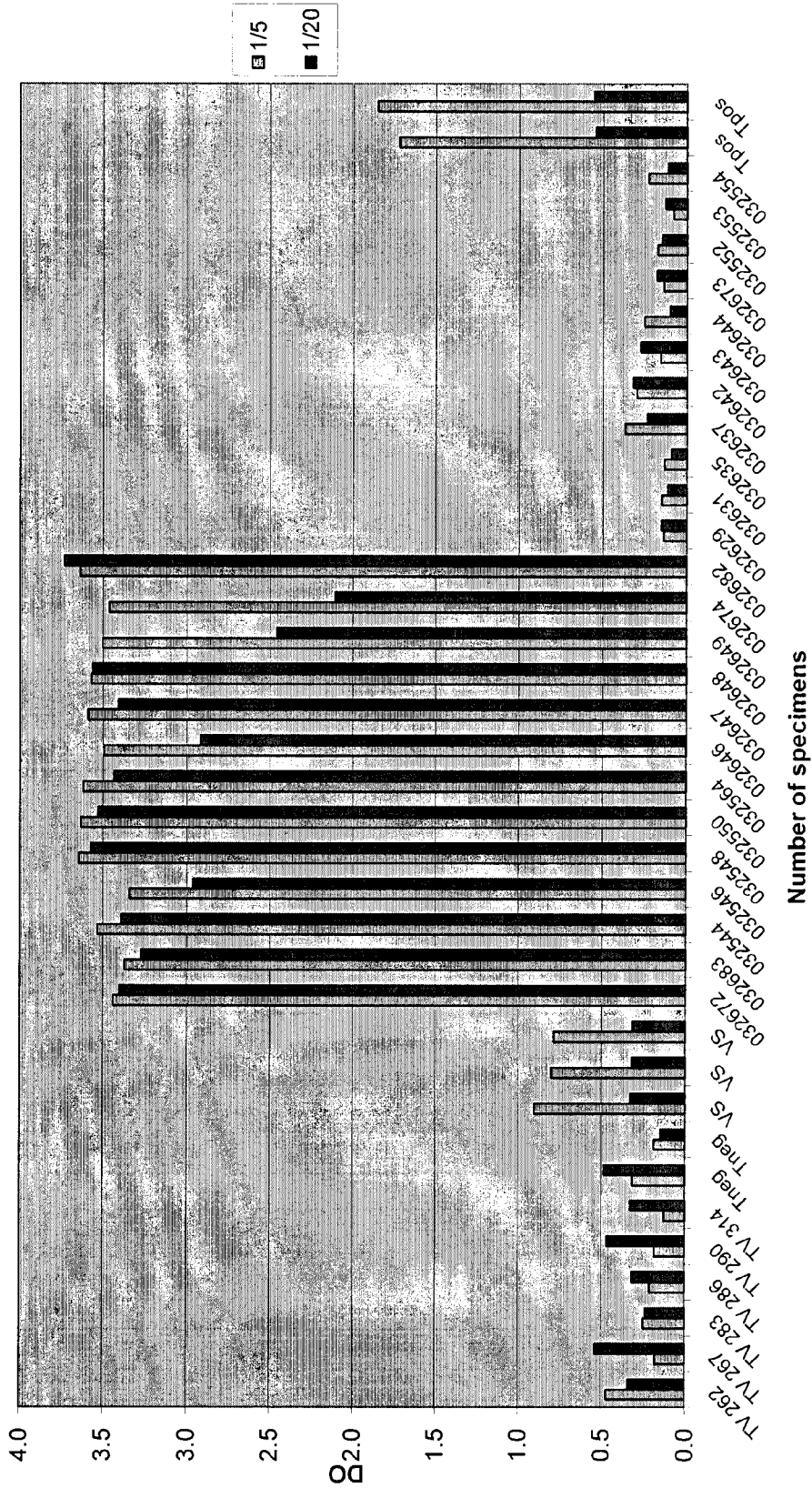


FIGURE 17

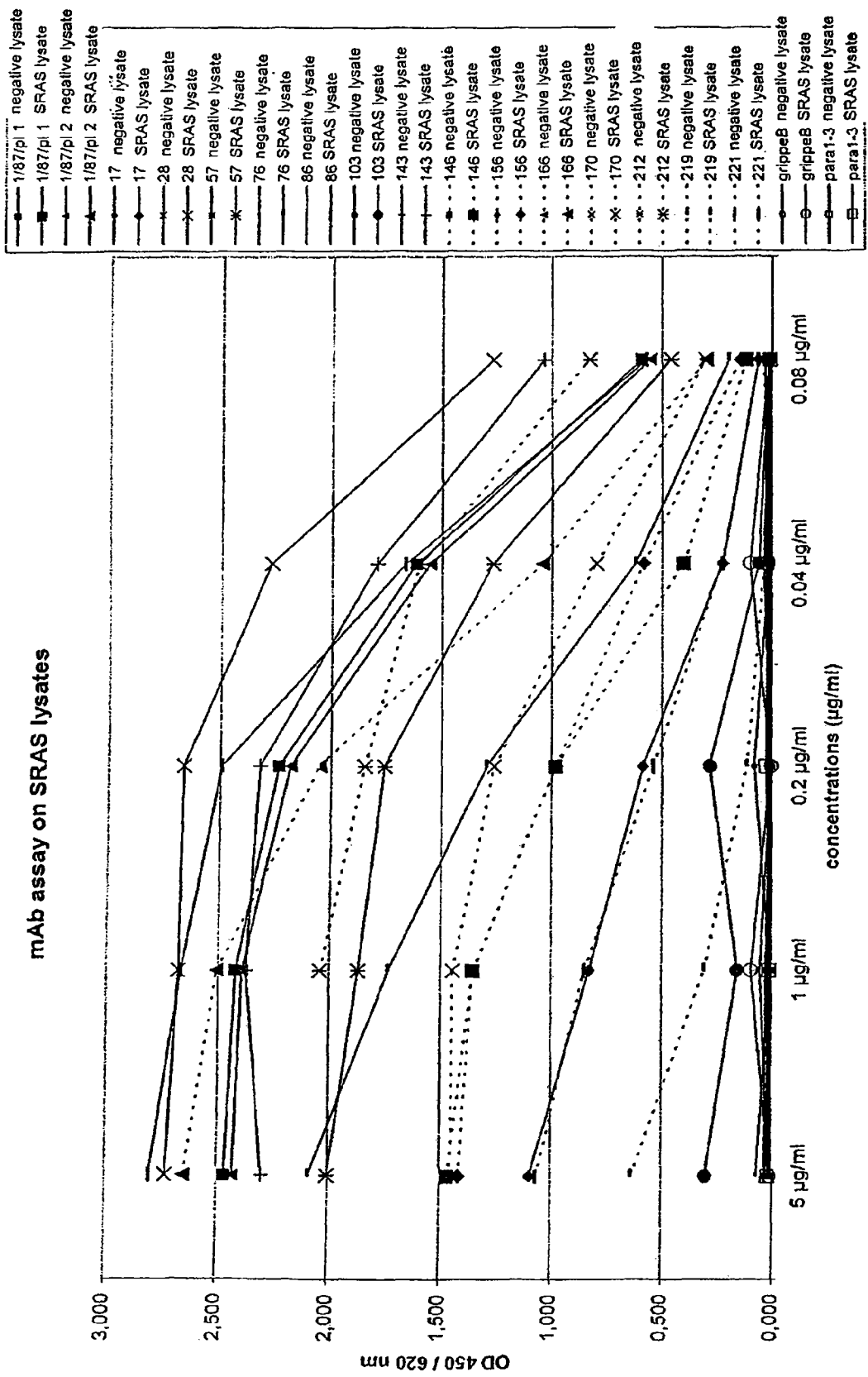
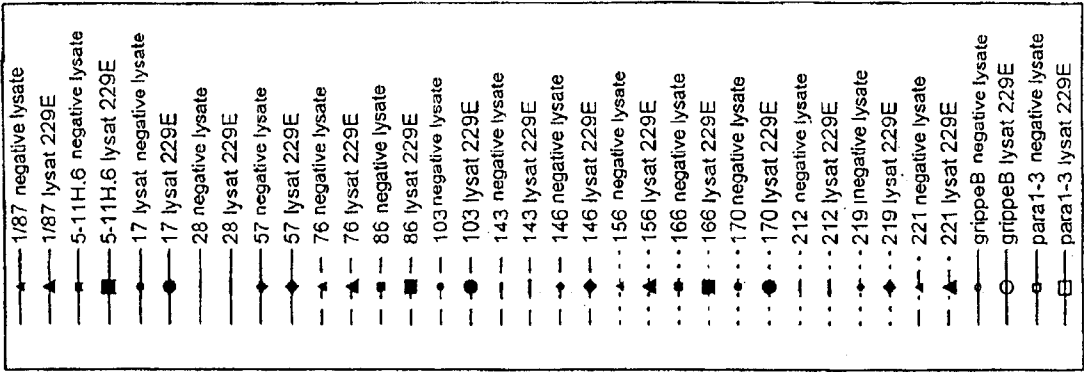


FIGURE 18



mAb assay on HCoV-229E lysates

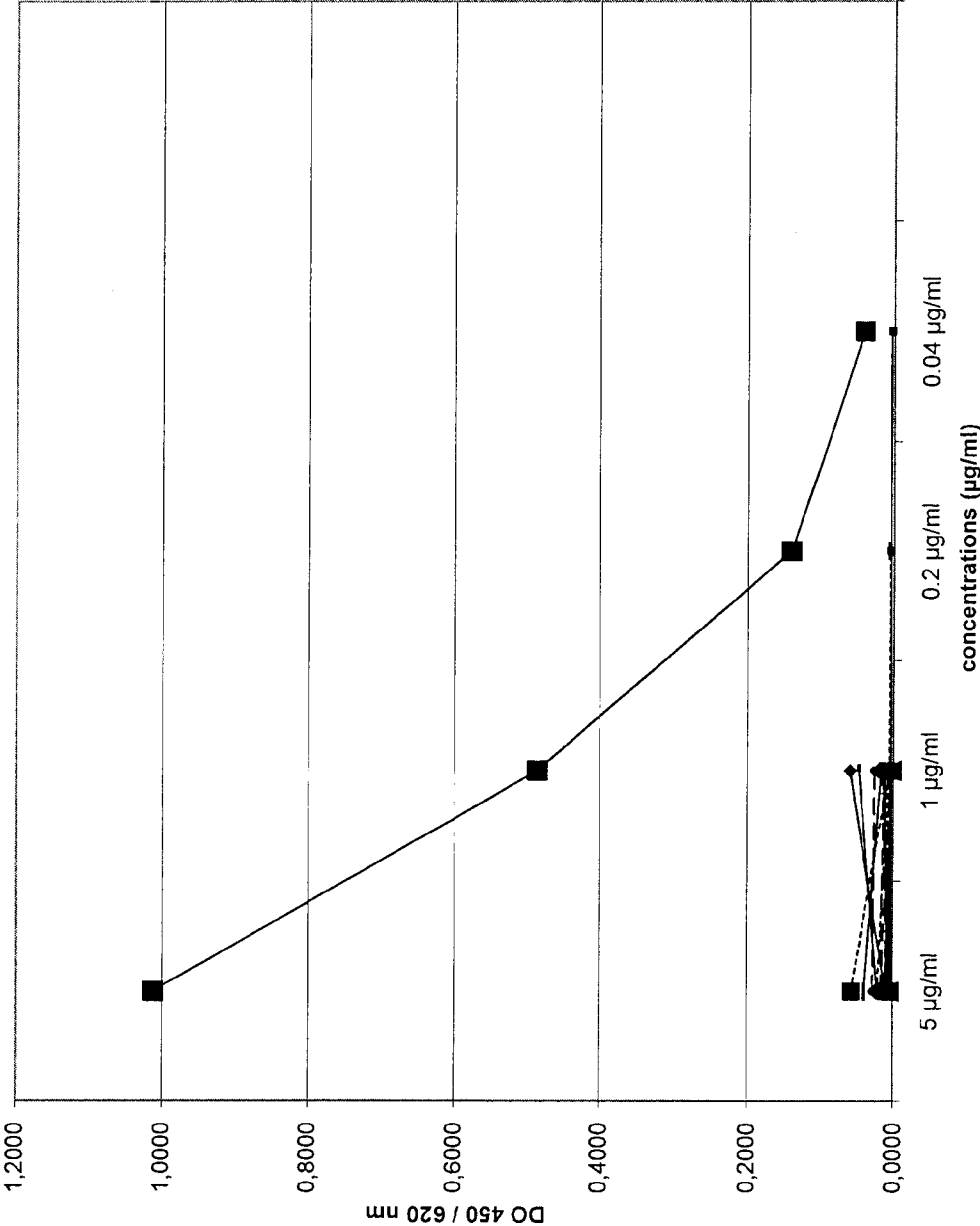


FIGURE 19

#para1-3

#grippeB

#221

#219

#212

#170

#166

#156

#146

#143

#103

#86

#76

#57

#28

#17

1/87

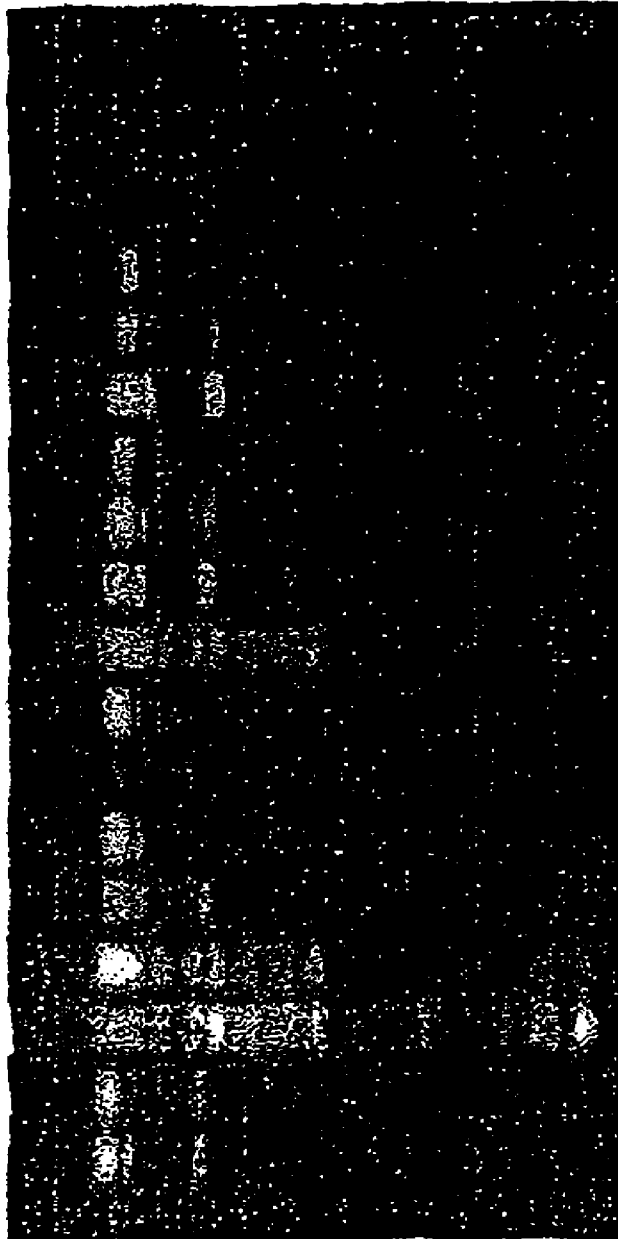


FIGURE 20

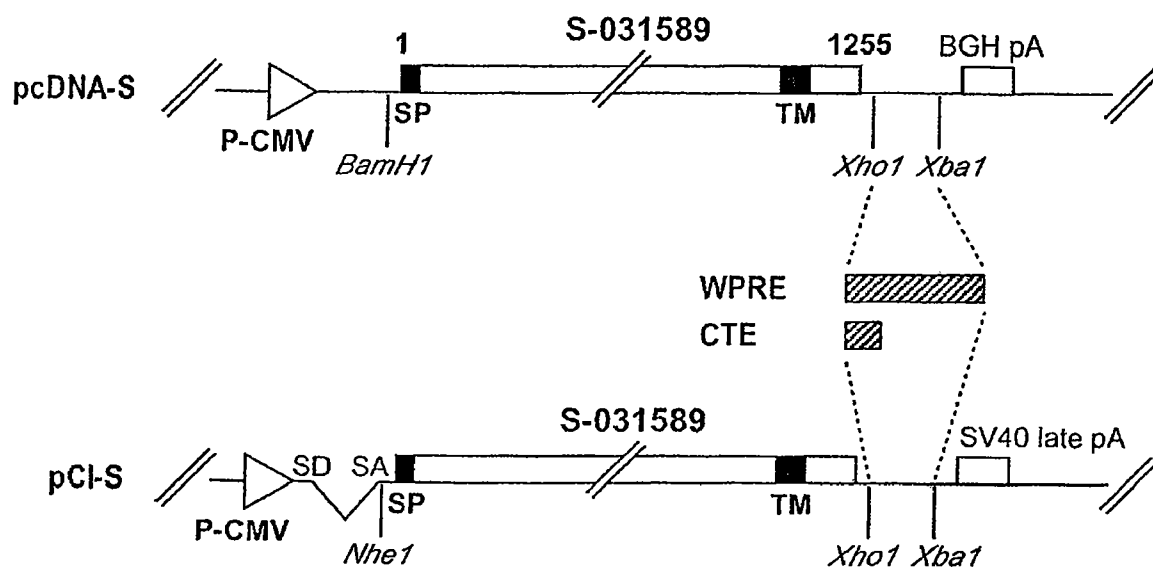
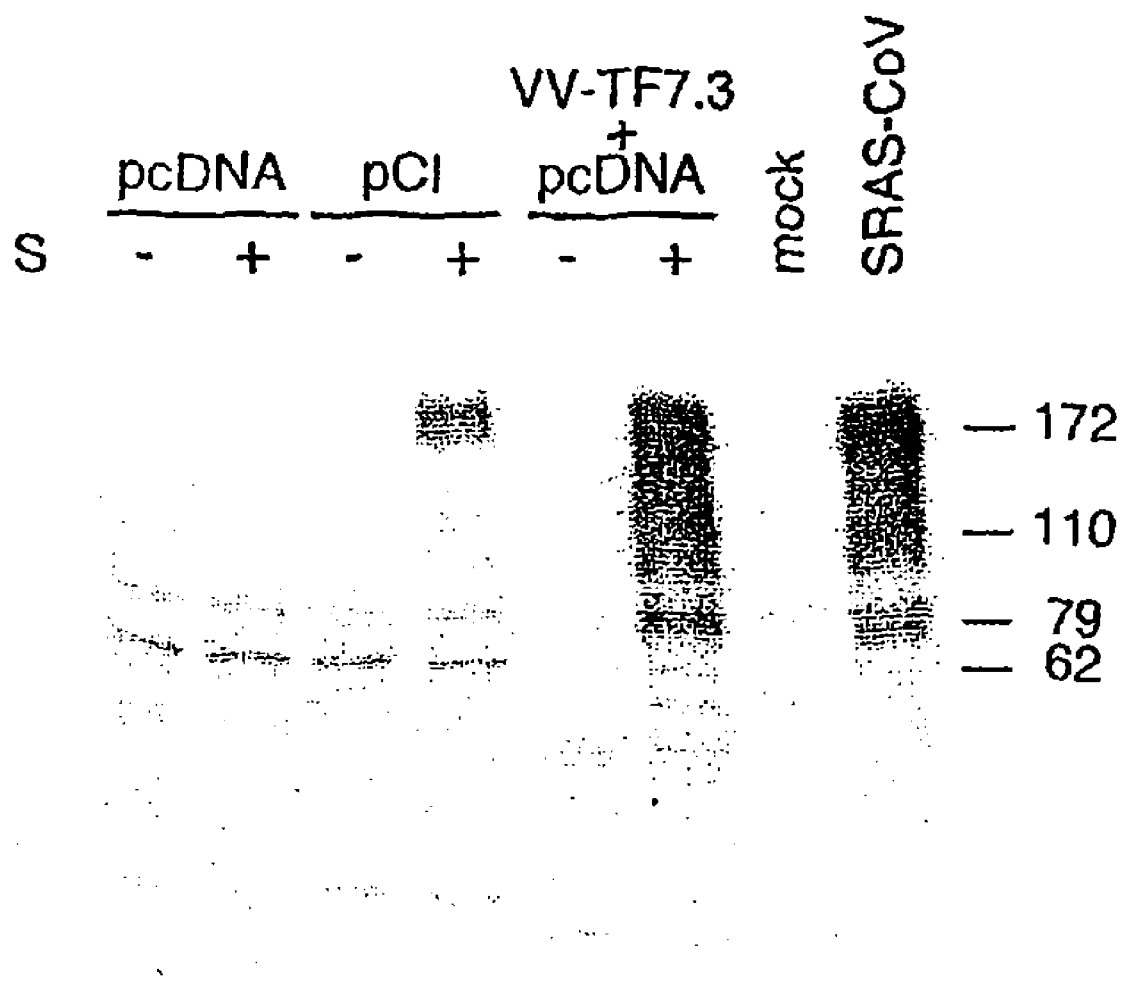
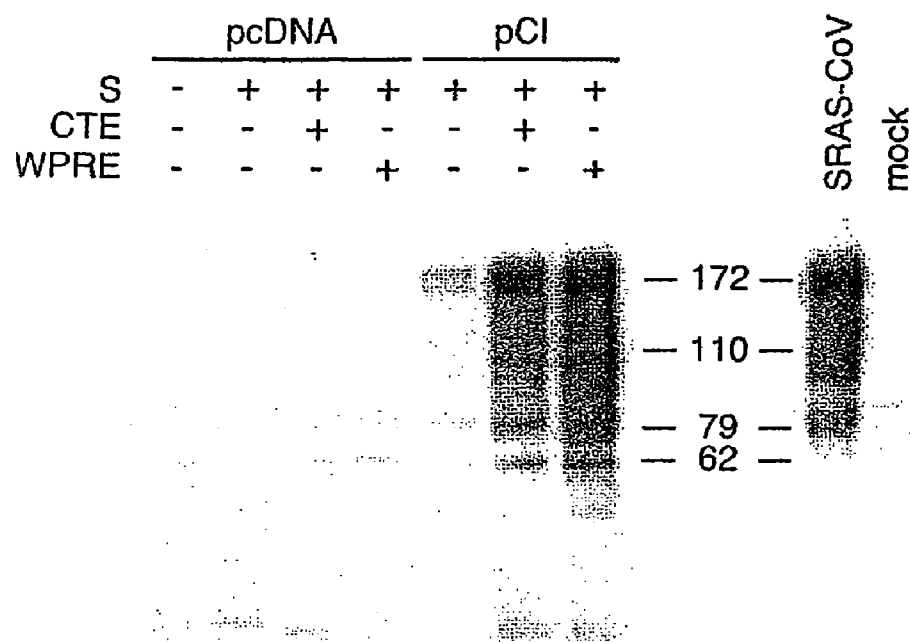
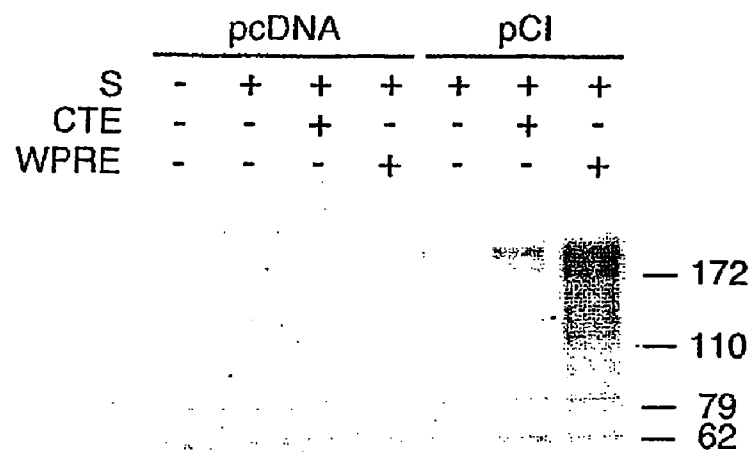
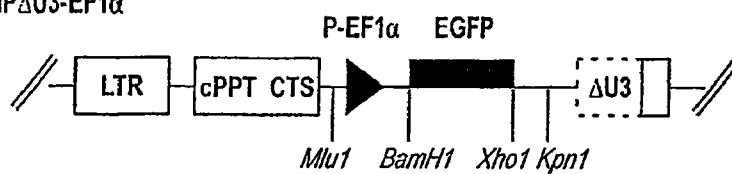


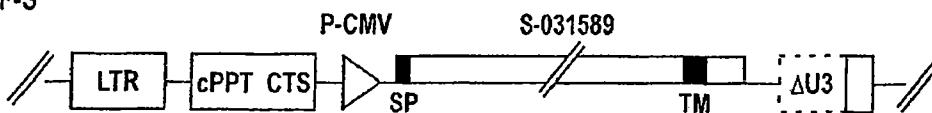
FIGURE 21

**FIGURE 22**

A.**B.****FIGURE 23**

pTRIP Δ U3-EF1 α 

pTRIP-S



pTRIP-SD/SA-S-CTE



pTRIP-SD/SA-S-WPRE

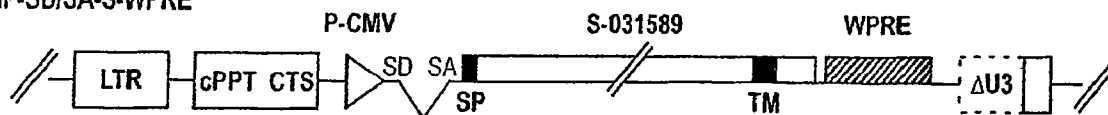


FIGURE 24

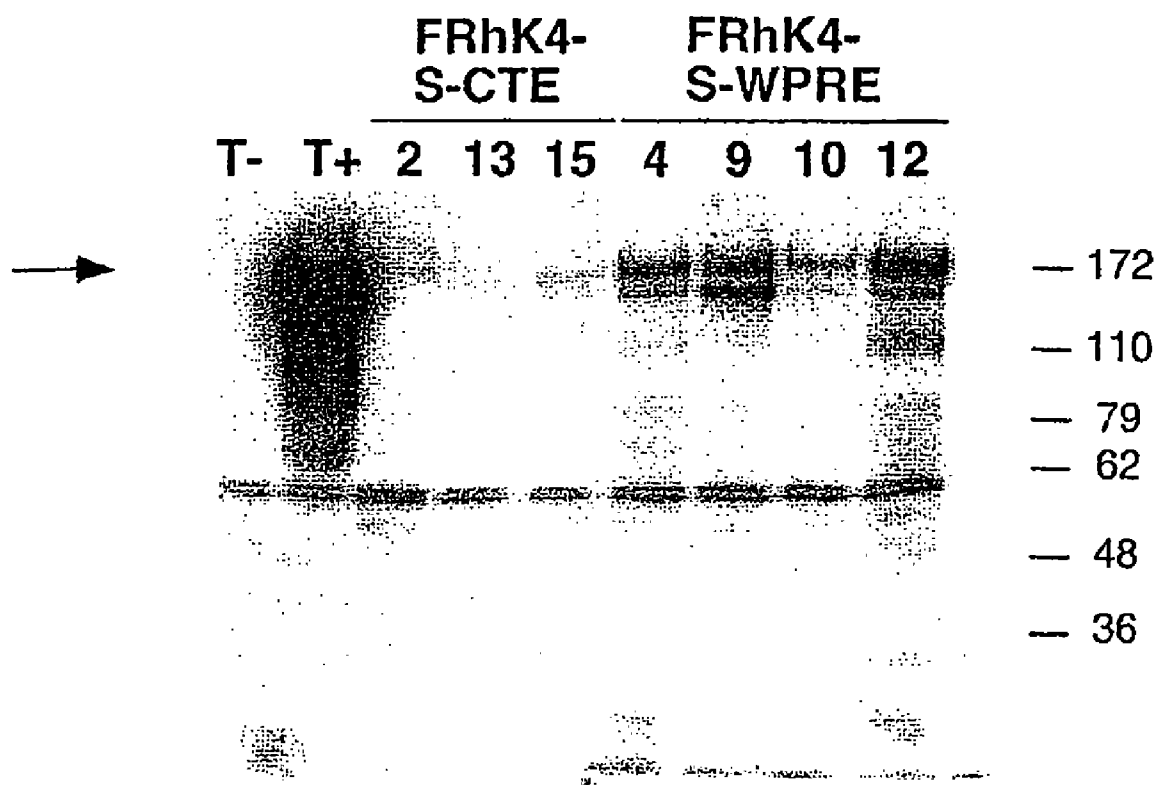


FIGURE 25

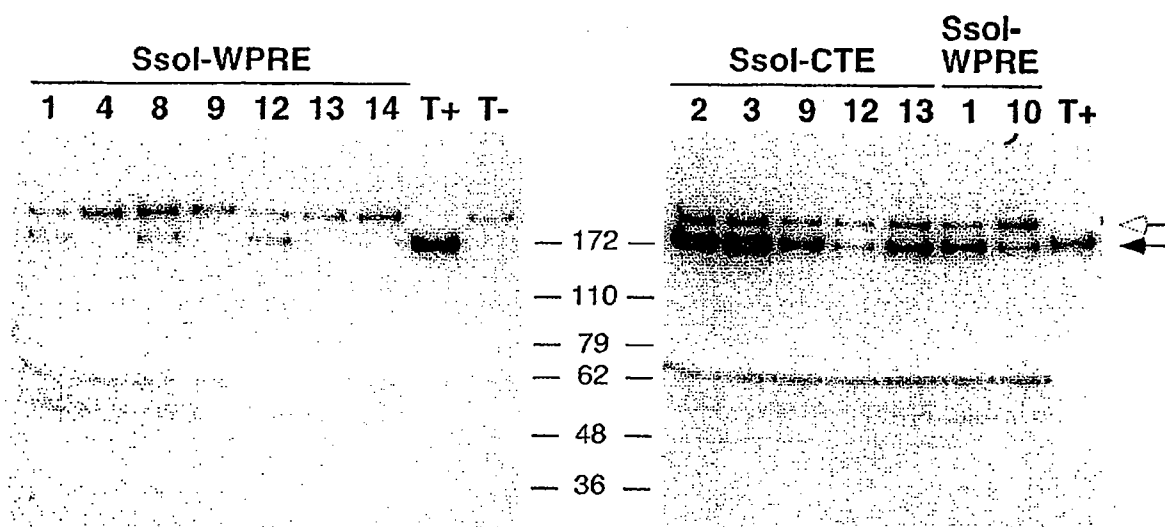
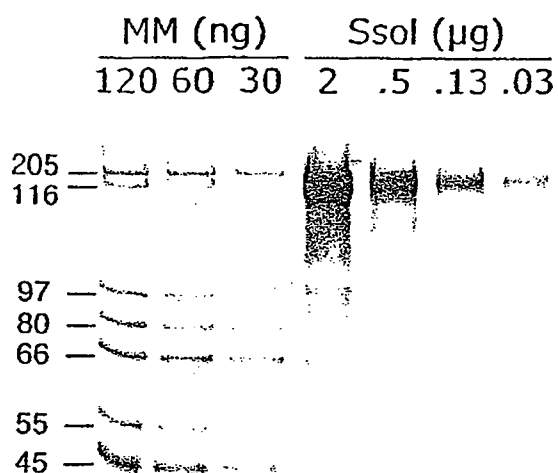
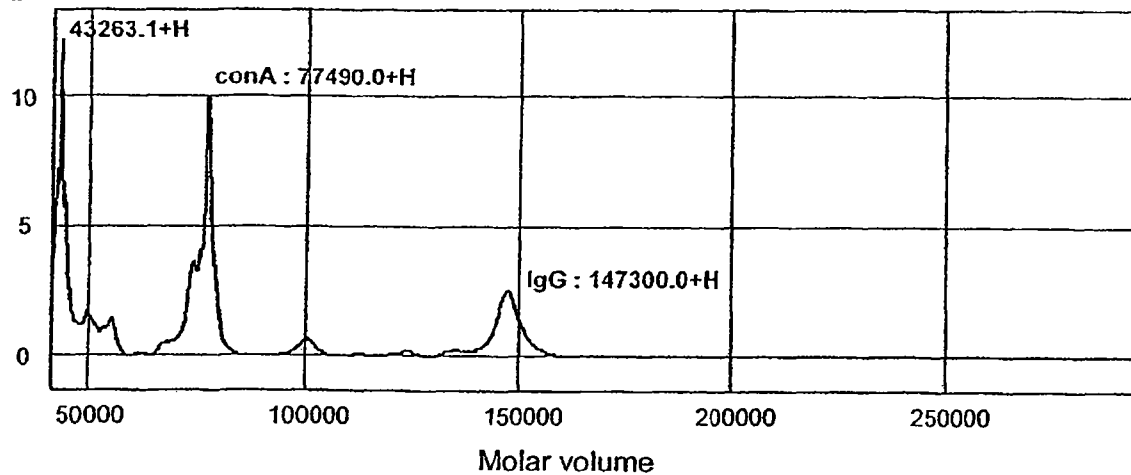


FIGURE 26

A.



B.



C.

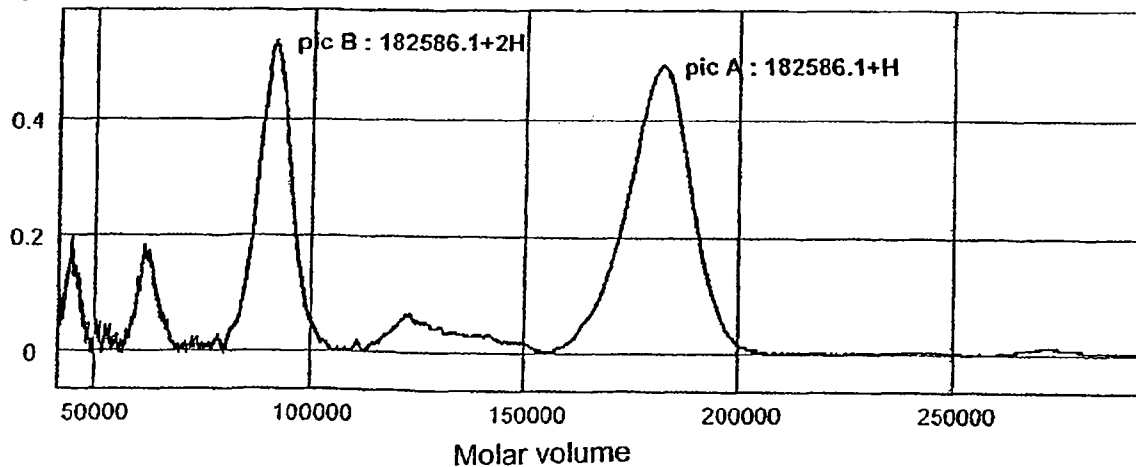


FIGURE 27 A-C

D.

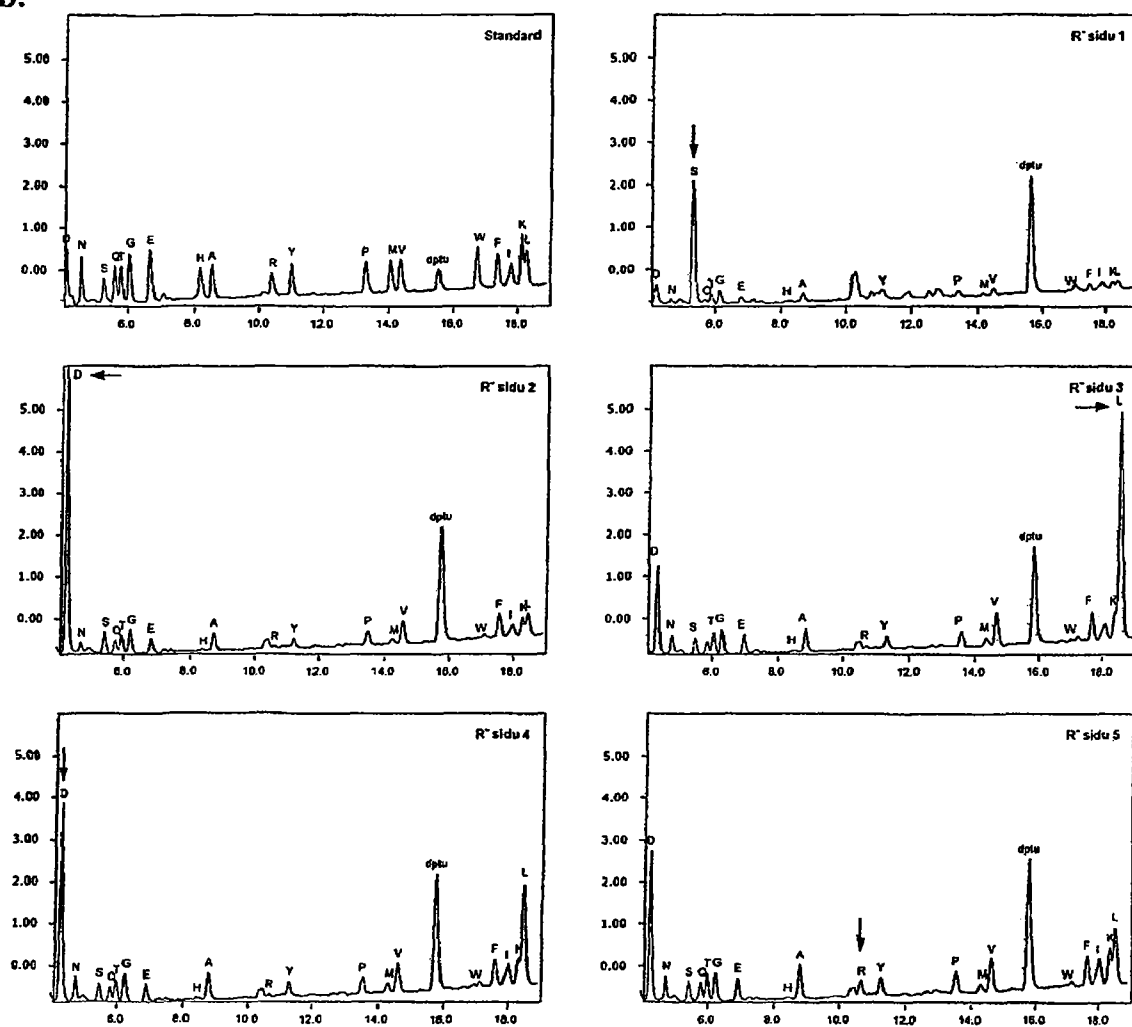


FIGURE 27 D

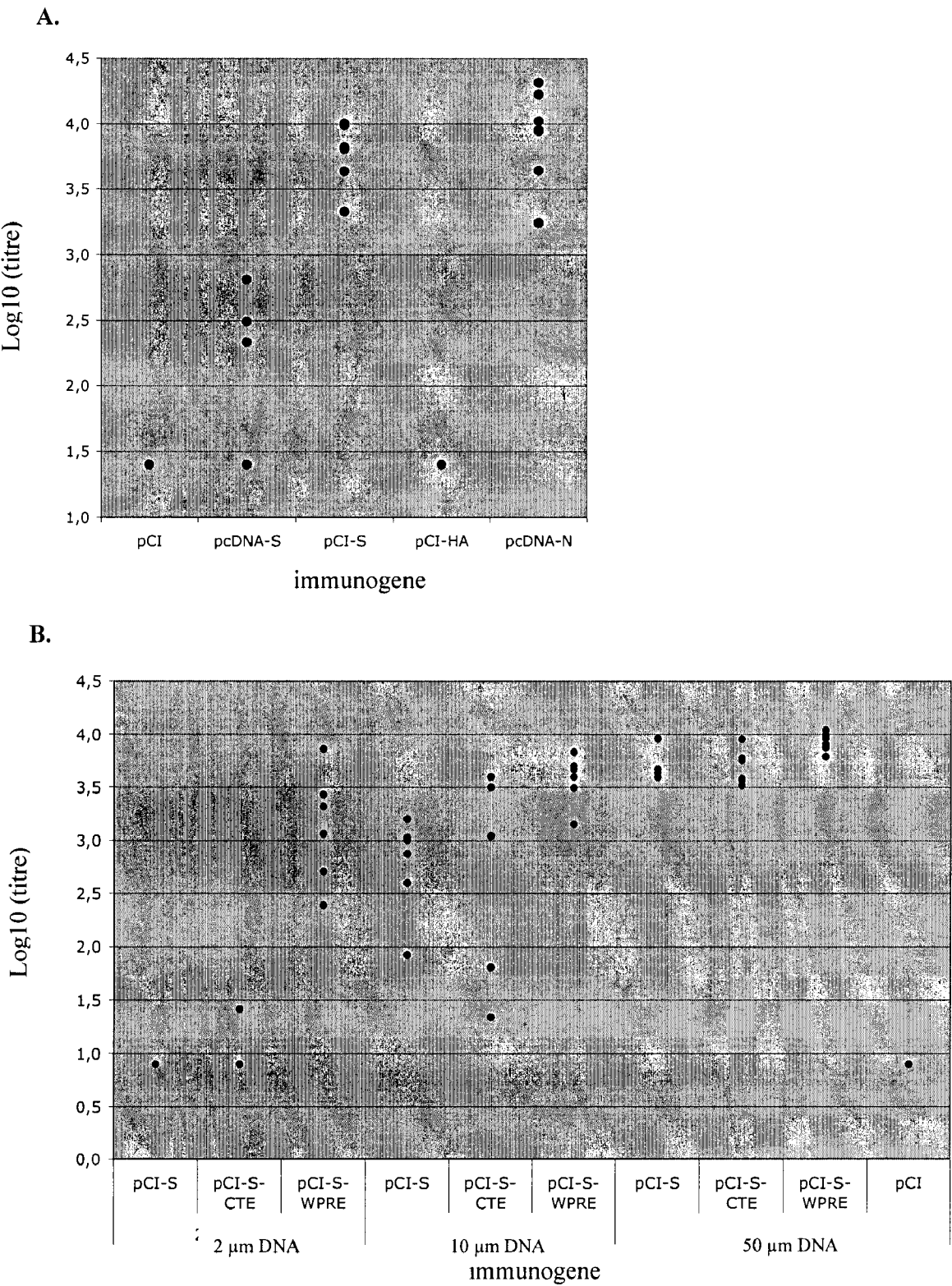
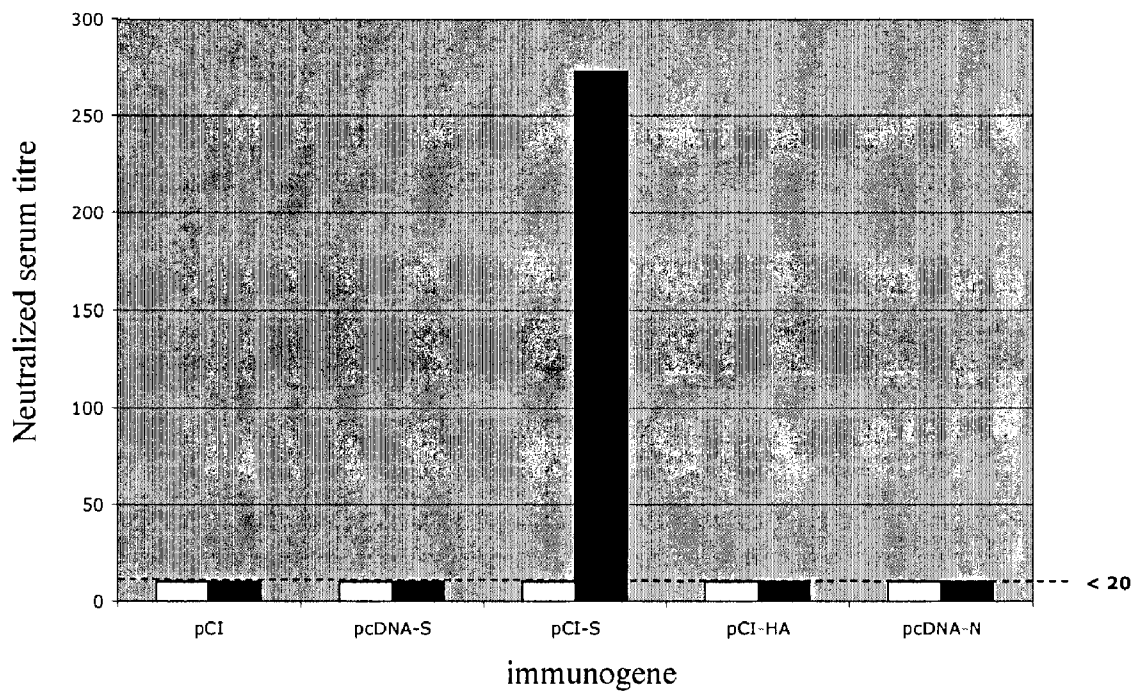


FIGURE 28

A.



B.

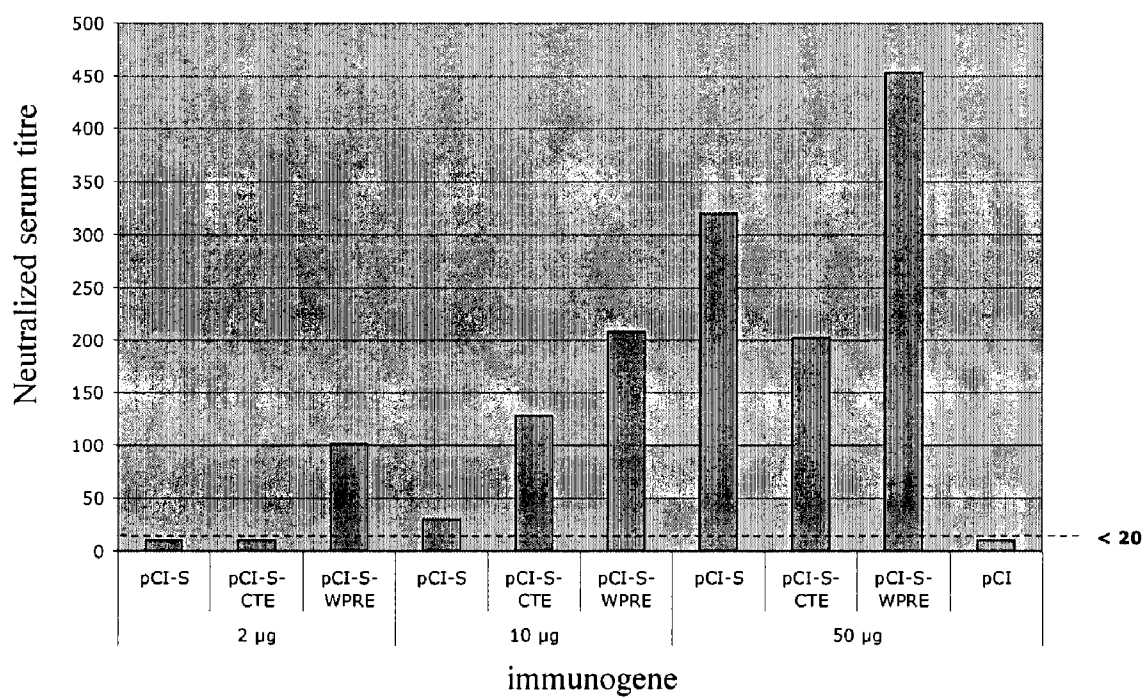


FIGURE 29

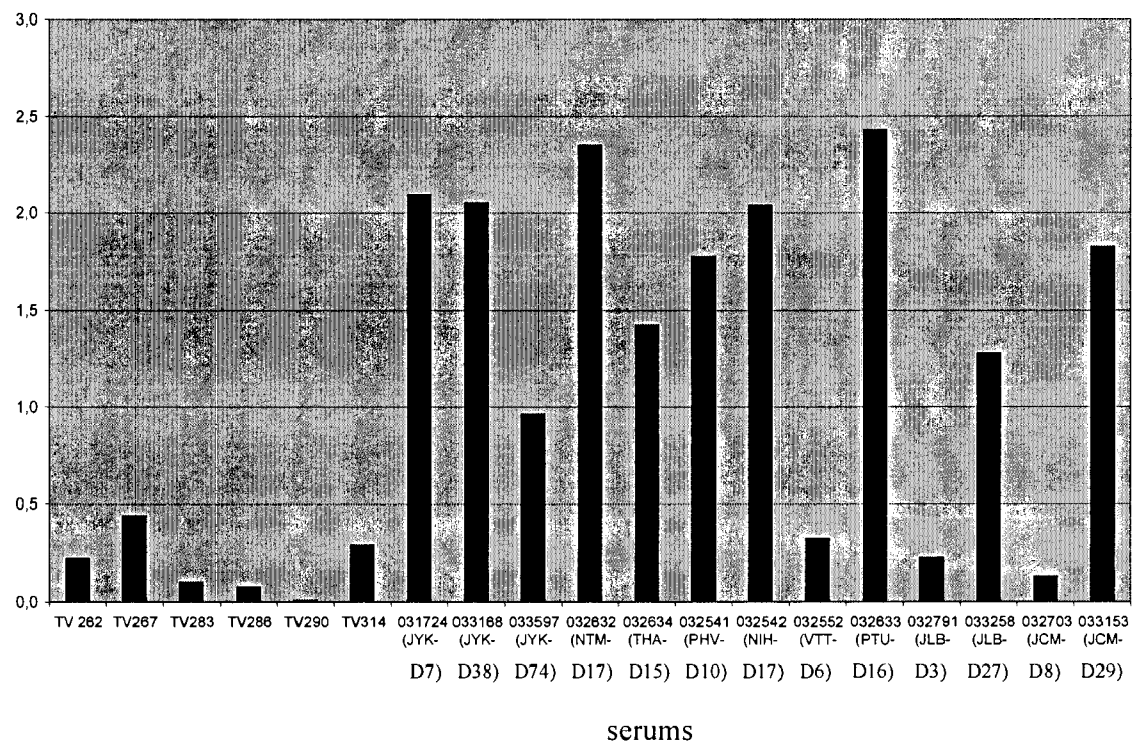


FIGURE 30

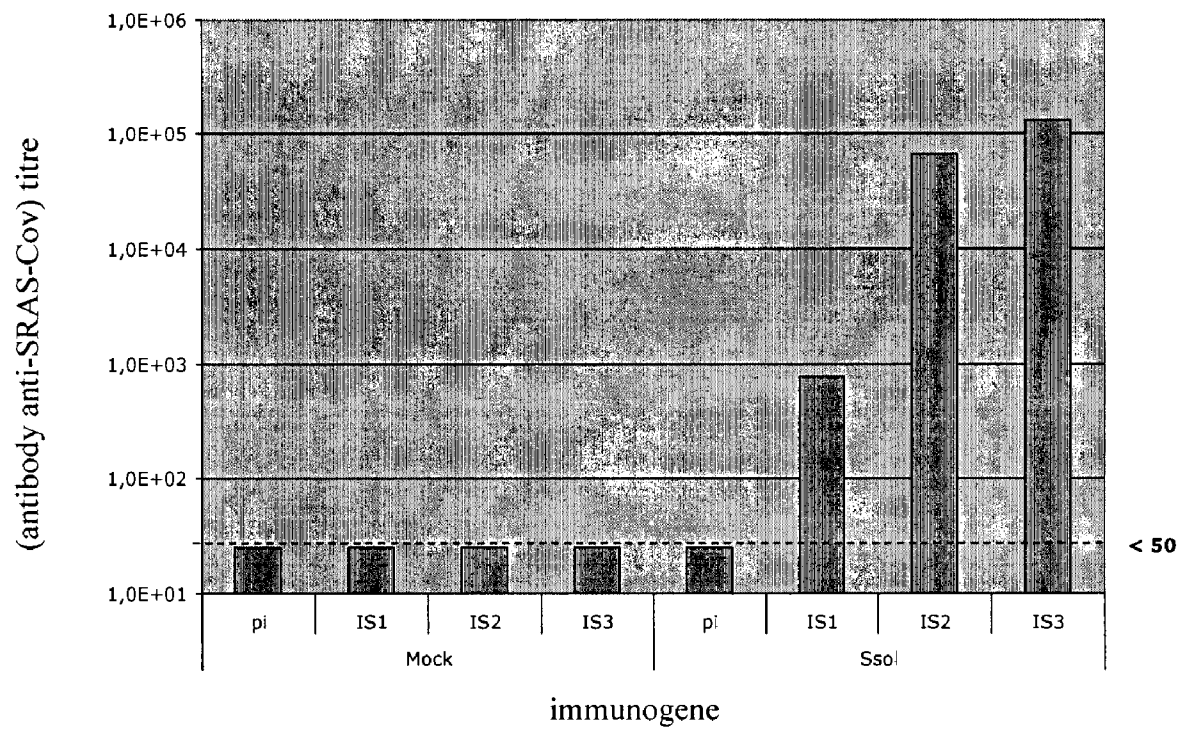


FIGURE 31

FIGURE 32.1

I-3059 S-040530	1320 1243	TTGCCAGATGATTTCATGGGTTGTCTCCTTGCTTGGAACTAGGAACATTGATGCTACT C""""C""C""C""""""C""C""G""G""C""""C""CC""""""C""C""C""A
I-3059 S-040530	1380 1303	TCAACTGGTAATTATAATTATAAATATAGGTATCTTAGACATGGCAAGCTTAGGCCCTTT AGC""C""C""C""C""""C""G""CC""C""C""G""C""""""GC""""""C
I-3059 S-040530	1440 1363	GAGAGAGACATATCTAATGTGCCTTTCTCCCTGATGGCAAACCTTGACCCACCTGCT ""C""G""""C""C""C""C""""C""AG""C""C""""G""C""""""C""""C
I-3059 S-040530	1500 1423	CTTAATTGTTATTGGCCATTAAATGATTATGGTTTTTACACCACTACTGGCATTGGCTAC ""G""C""C""C""C""""CC""G""C""C""C""C""C""""C""C""C""C""""T
I-3059 S-040530	1560 1483	CAACCTTACAGAGTTGTAGTACTTTCTTTTGAACCTTTAAATGCACCGGCCACGGTTTGT ""G""C""""""G""G""G""GAGC""C""G""GC""G""C""C""T""""C""G""C
I-3059 S-040530	1620 1543	GGACCAAATATCCACTGACCTTATTAAGAACCAGTGTGTCAATTTTAATTTTAATGGA ""C""C""C""GC""GAG""C""""G""C""""""C""G""C""C""C""C""C""C
I-3059 S-040530	1680 1603	CTCACTGGTACTGGTGTGTTAACTCCTTCTTCA _AAGAGATTTCAACCATTTCACAAT ""G""C""C""C""C""C""C""G""C"" _AG""GC""C""C""C""G""C""C""G""G
I-3059 S-040530	1738 1661	TTGGCCGTGATGTCTCTGATTTCAGTATCCGTTTCGAGATCCTAAACATCTGAAATAT C""""G""""GAGC""C""""C""CAG""G""G""C""C""G""CAGC""G""CC
I-3059 S-040530	1798 1721	TAGACATTTACCTTGCTCTTTTGGGGGTGTAAGTGTAAATTACACCTGGAACAAATGCTT G""""CAGC""C""AGC""C""C""C""GTCC""G""C""C""C""C""C""C""CA
I-3059 S-040530	1858 1781	CACTGGAAGTTGCTGTTCTATATCAAGATGTTAACTGCACTGATGTTTCTACAGCAATC G""G"" _ ""G""C""G""G""C""G""C""G""""C""C""GAGC""C""C""
I-3059 S-040530	1917 1840	CATGCAGATCAACTCACACCAGCTTGGCGCATATATTCTACTGGAACAATGTATTCAG ""C""C""C""G""G""C""C""C""""G""C""CAGC""C""G""""C""G""""
I-3059 S-040530	1977 1900	ACTCAAGCAGGCTGTCTTATAGGAGCTGAGCATGTGACACTTCTTATGAGTGGGACATT ""C""G""C""""C""G""C""C""C""""C""G""""CAGC""C""""C
I-3059 S-040530	2037 1960	CCTATTGGAGCTGGCATTGTGCTAGTTACCATACAGTTTCTTTATTACGTAGTACTAGC ""C""""C""C""A""C""C""C""C""C""C""C""GAGCC""GC""G""G""C""C""
I-3059 S-040530	2097 2020	CAAAAATCTATTGTGGCTTATACTATGTCTTTAGGTGCTGATAGTTCAATTGCTTACTCT ""G""G""C""C""""C""C""C""AGCC""G""C""C""C""CAGC""C""C""AGC
I-3059 S-040530	2157 2080	AATAACACCATTGCTATACCTACTAACTTTTCAATTAGCATTACTACAGAAGTAATGCCT ""C""""C""C""C""C""C""C""C""C""CAGC""CTC""C""C""C""G""C
I-3059 S-040530	2217 2140	GTTTCTATGGCTAAAACCTCCGTAGATTGTAATATGTACATCTGCGGAGATTCTACTGAA ""GAGC""""C""G""AAG""G""""C""C""""C""C""CAGC""C""G
I-3059 S-040530	2277 2200	TGTGCTAATTTGCTTCTCCAATATGGTAGCTTTTGCACACAATAAATCGTCACTCTCA ""C""C""CC""G""G""G""C""C""C""C""C""G""G""C""G""C""GAGC
I-3059 S-040530	2337 2260	GGTATTGCTGCTGAACAGGATCGCAACACACGTGAAGTGTTCGCTCAAGTCAAACAAATG ""C""C""C""C""G""""C""G""""CA""A""""C""C""G""G""G""
I-3059 S-040530	2397 2320	TACAAAACCCCAACTTTGAAATATTTTGGTGGTTTTAATTTTTCACAAATATTACCTGAC ""T""G""""C""CC""G""C""C""G""C""C""C""C""T""G""CC""G""C""
I-3059 S-040530	2457 2380	CCTCTAAGCCAACTAAGAGGTCTTTTATTGAGGACTTGCTCTTTAATAAGGTGACACTC ""G""C""C""C""C""C""C""C""C""C""G""C""C""C""A""C""G
I-3059 S-040530	2517 2440	GCTGATGCTGGCTTCATGAACCAATATGGCGAATGCCTAGGTGATATTAACTGCTAGAGAT ""C""C""C""C""T""G""C""C""G""C""C""G""C""C""C""C""C""CC""G""C
I-3059 S-040530	2577 2500	CTCATTGTGTCGCGAGAAGTTCAATGGGCTTACAGTGTGCCACCTCTGCTCACTGATGAT ""G""C""C""C""C""""T""C""""G""C""C""C""C""C""C""G""C""C""C
I-3059 S-040530	2637 2560	ATGATTGCTGCCTACACTGCTGCTCTAGTTAGTGGTACTGCCACTGCTGGATGGACATTT ""C""C""C""C""T""A""C""C""G""G""C""C""C""C""C""C""C""C""C""C

FIGURE 32.2

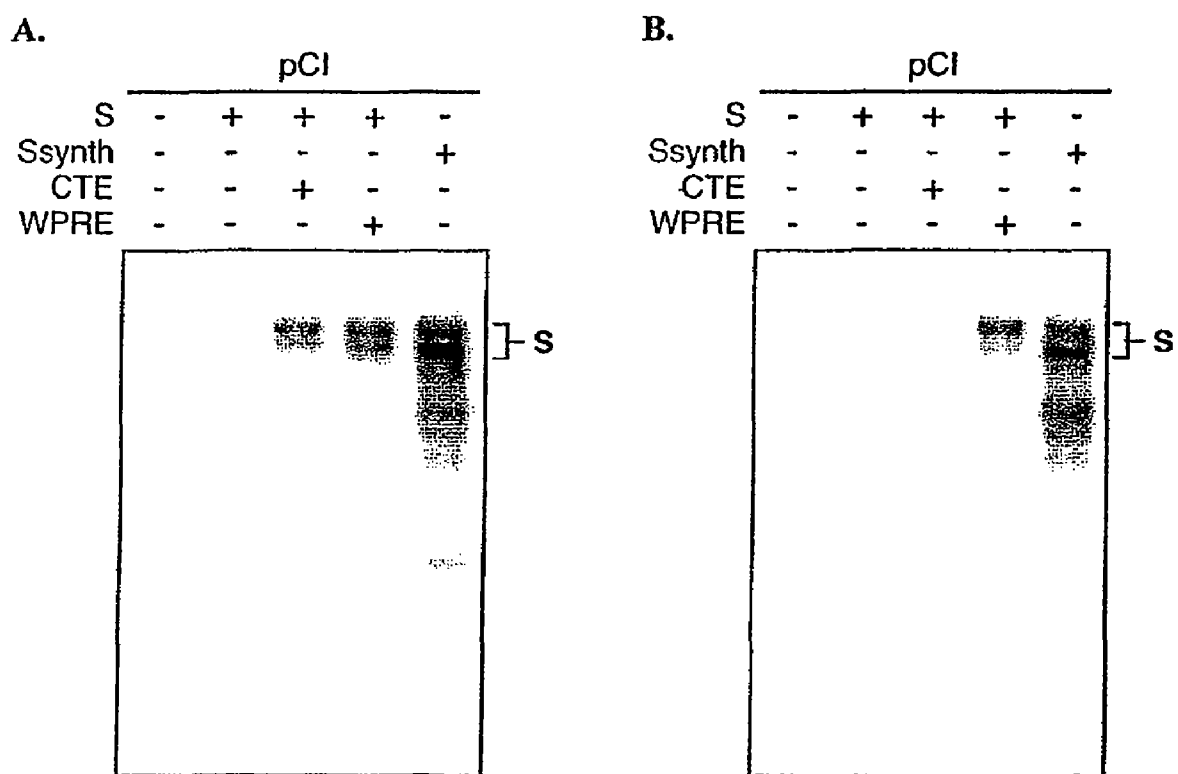
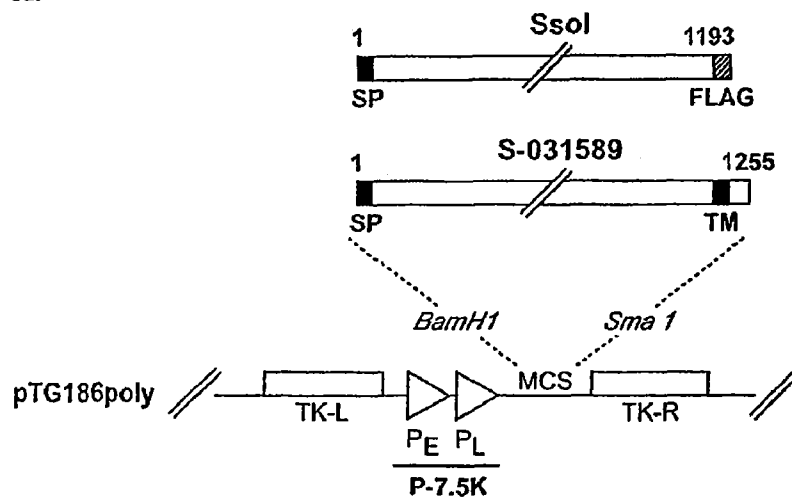
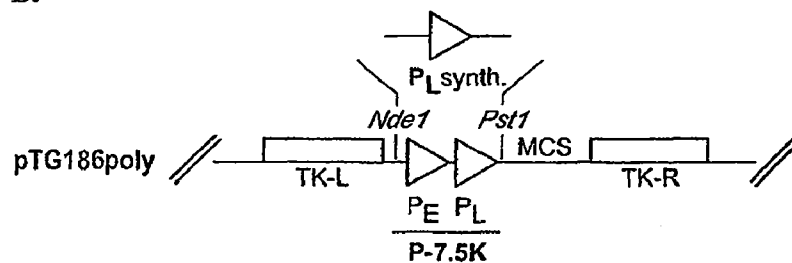


FIGURE 33

A.



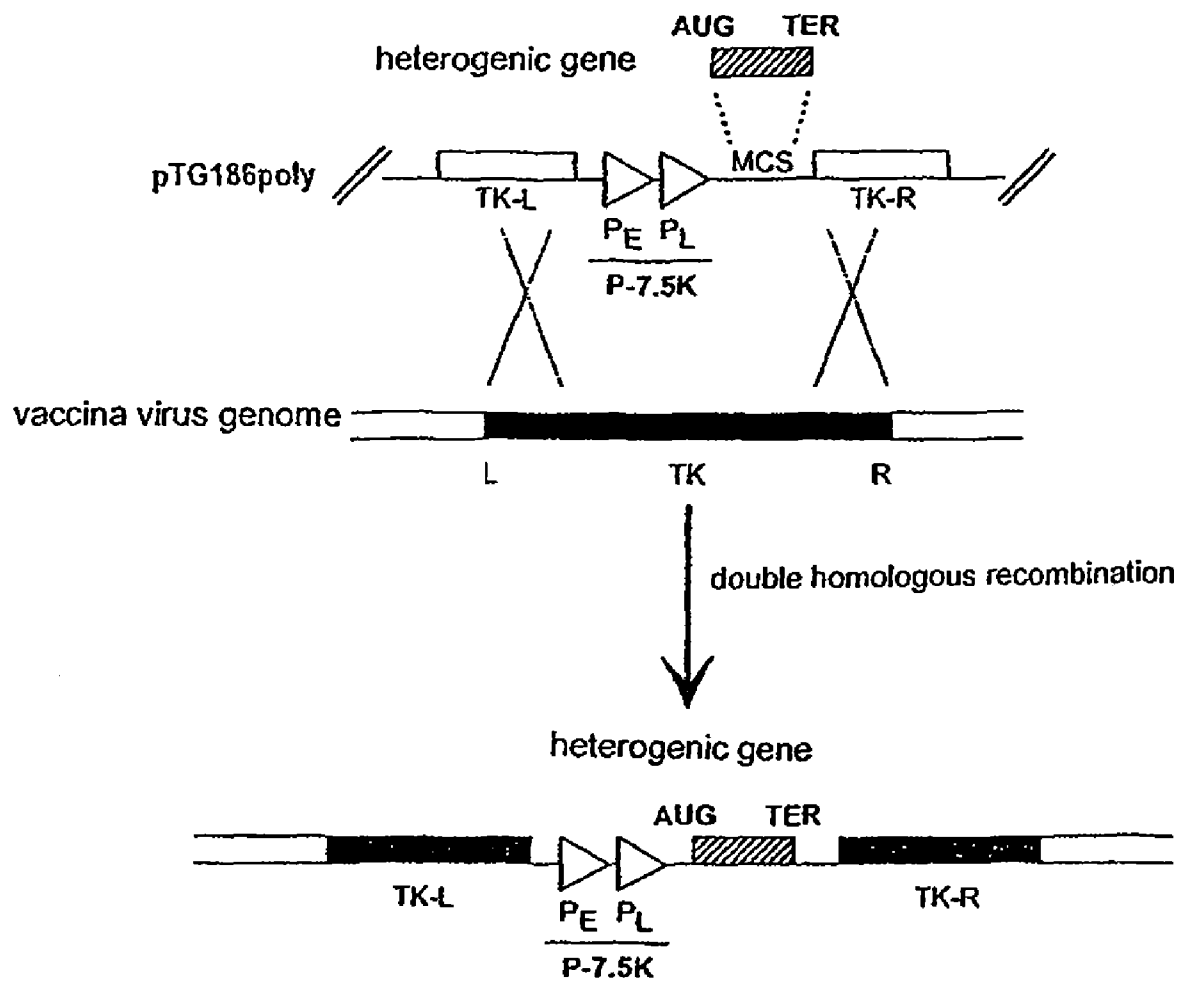
B.



C.

CATATG AGC [T]₂₀GGCATATAAATA GACTC GGCGCGCC AT CTGCAG
NdeI *promoteur 480* *AscI* *PstI*

FIGURE 34 A-C

D.**FIGURE 34 D**

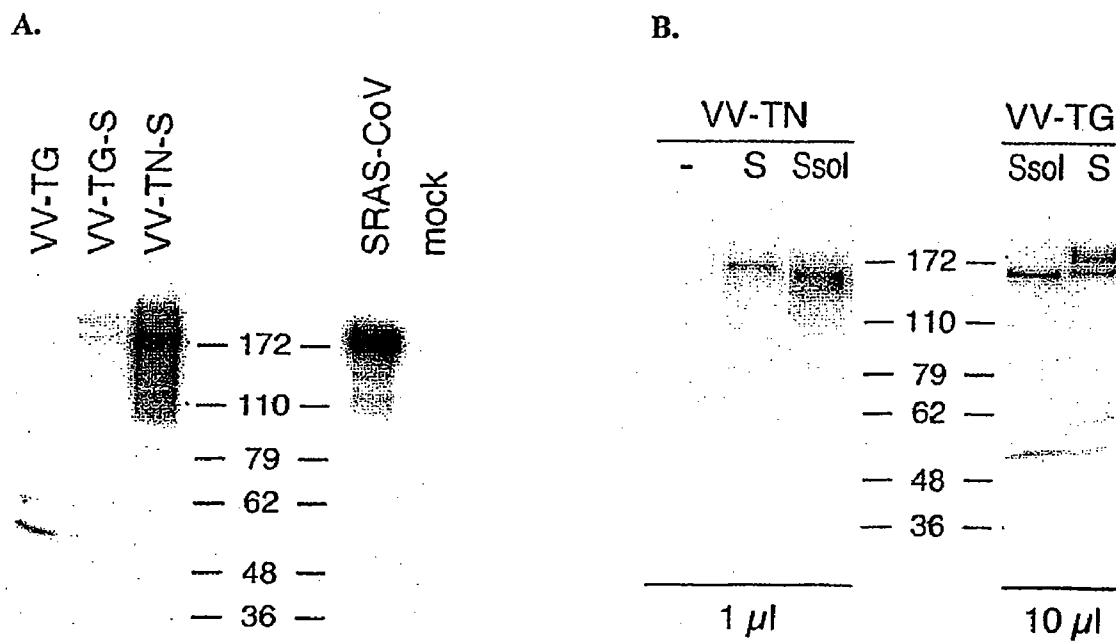
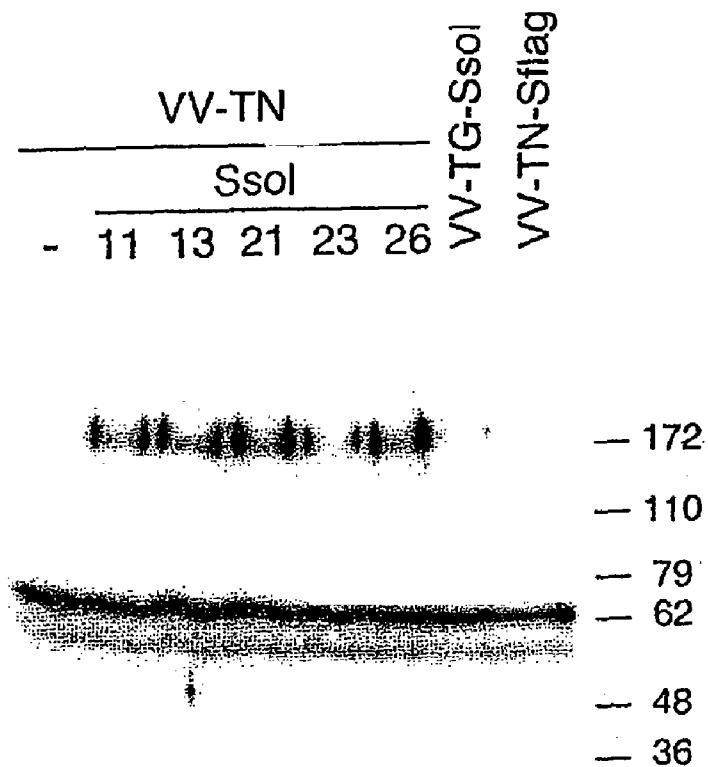


FIGURE 35

A.



B.

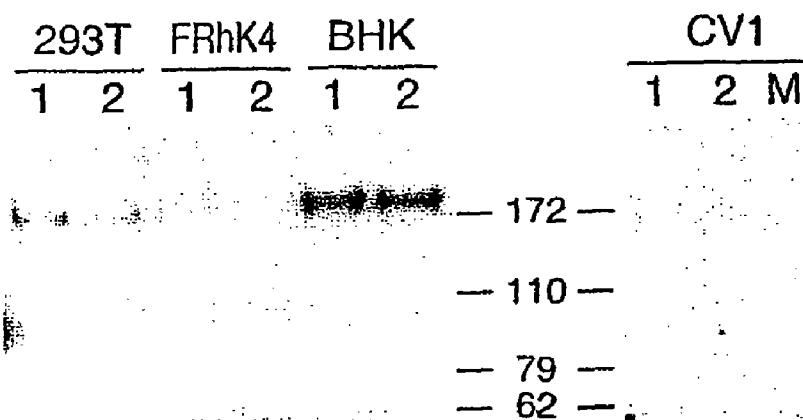


FIGURE 36

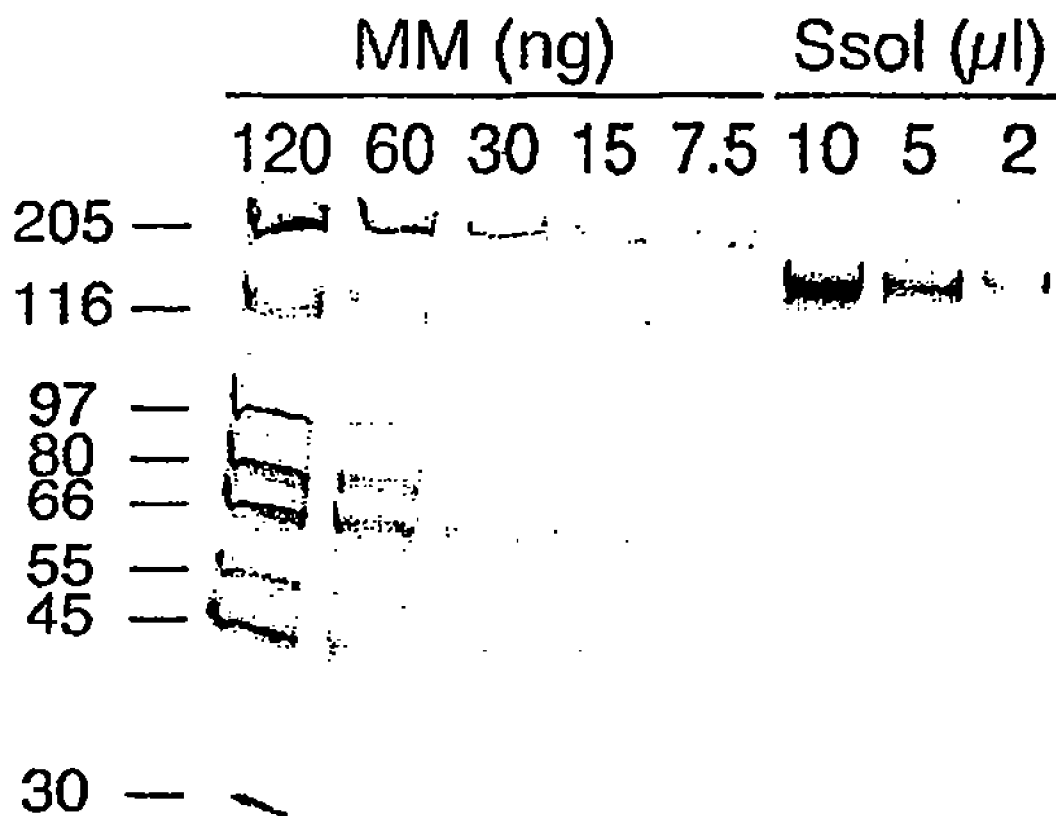


FIGURE 37

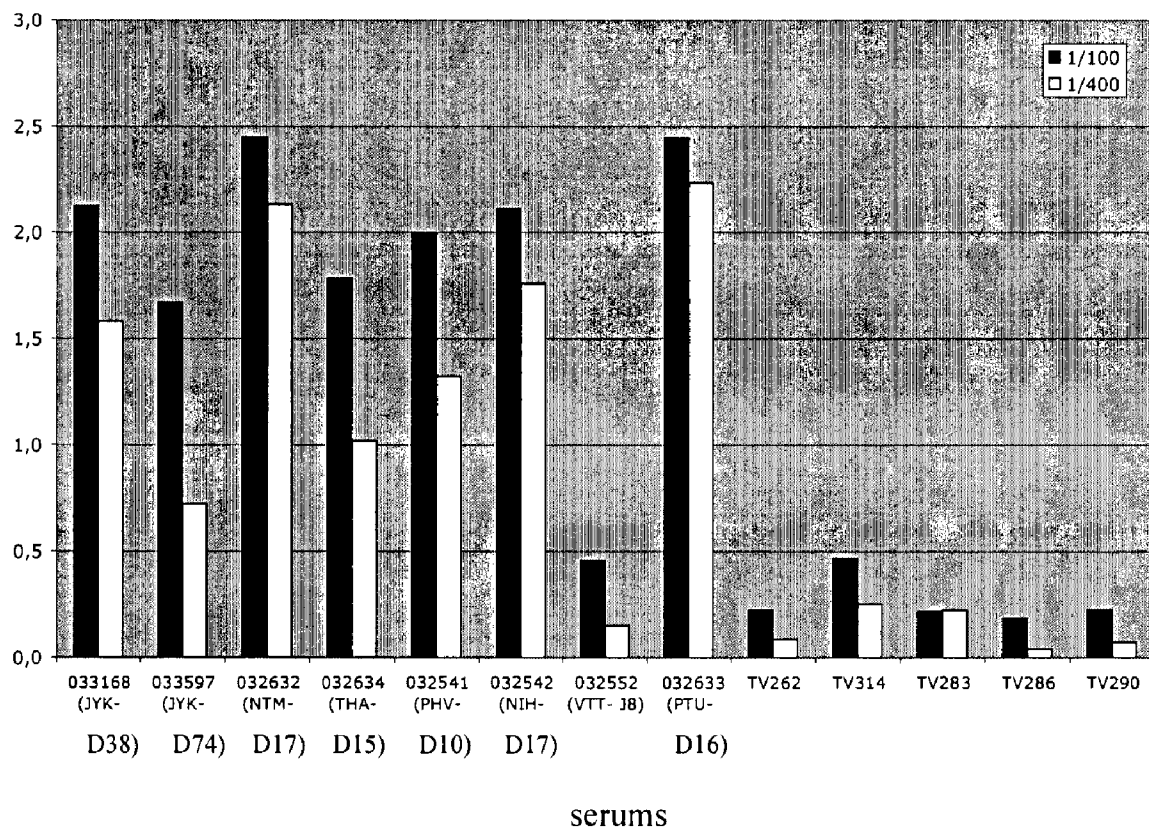
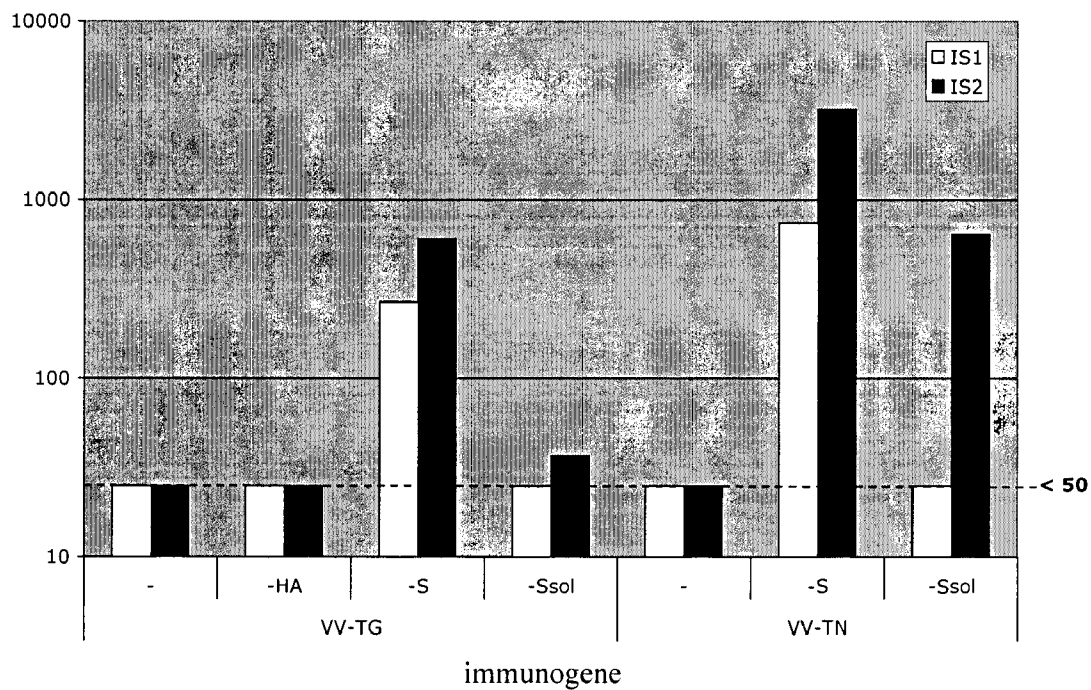


FIGURE 38

A.



B.

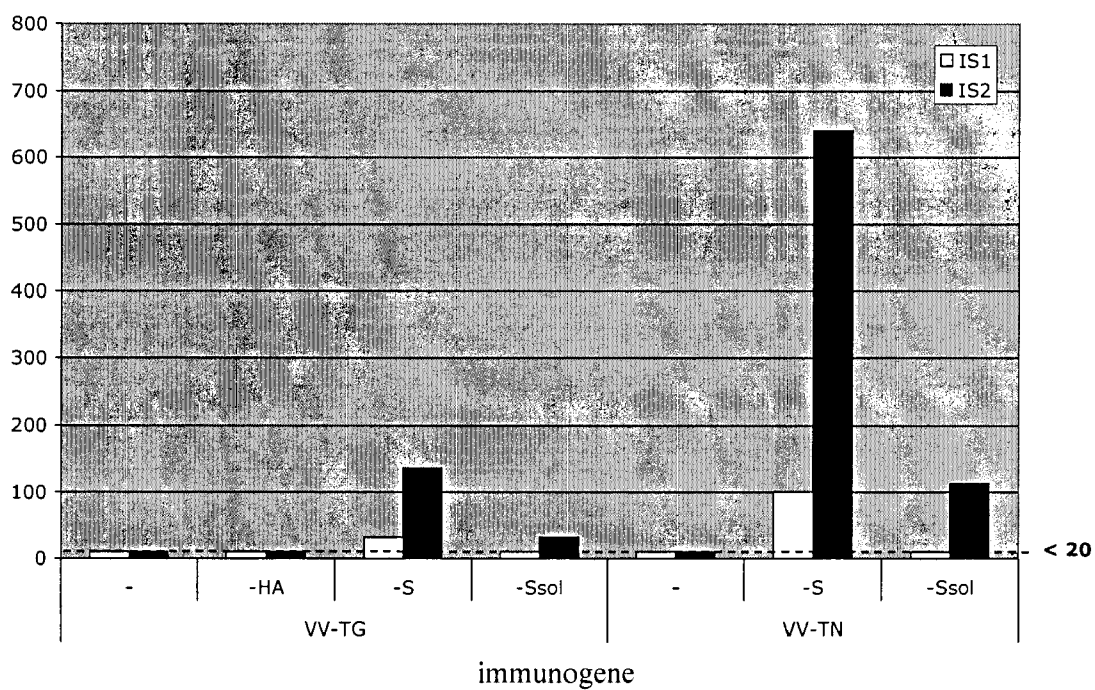


FIGURE 39

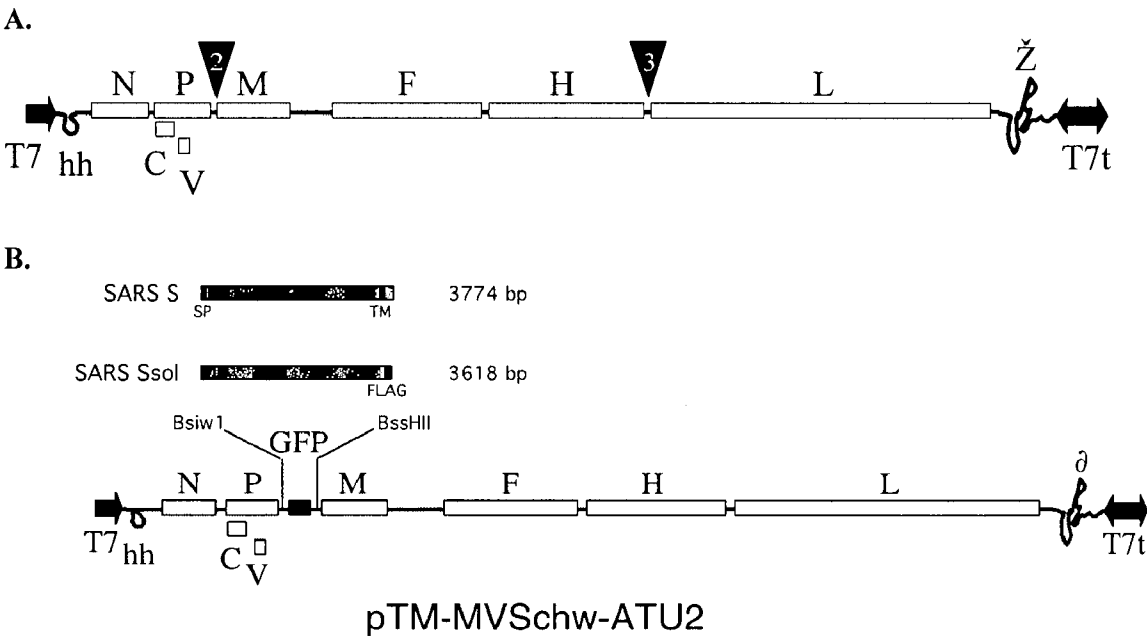


FIGURE 40

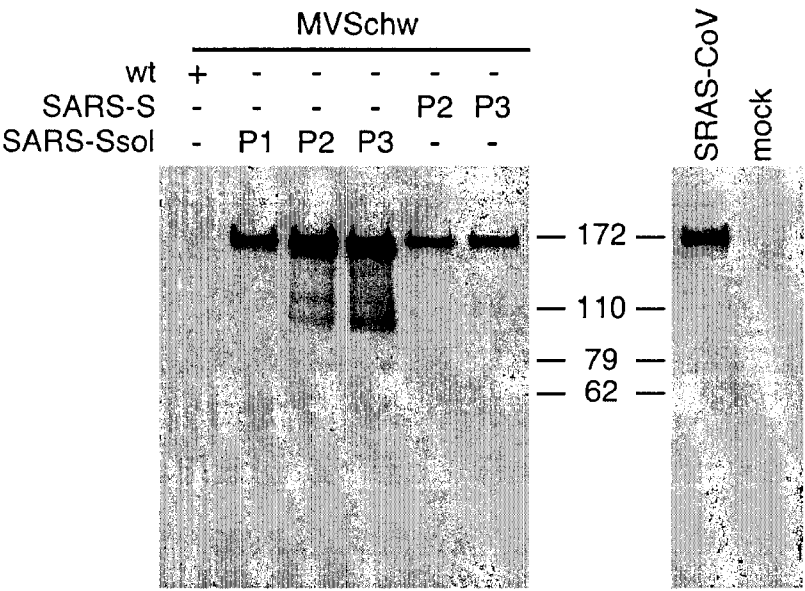


FIGURE 41

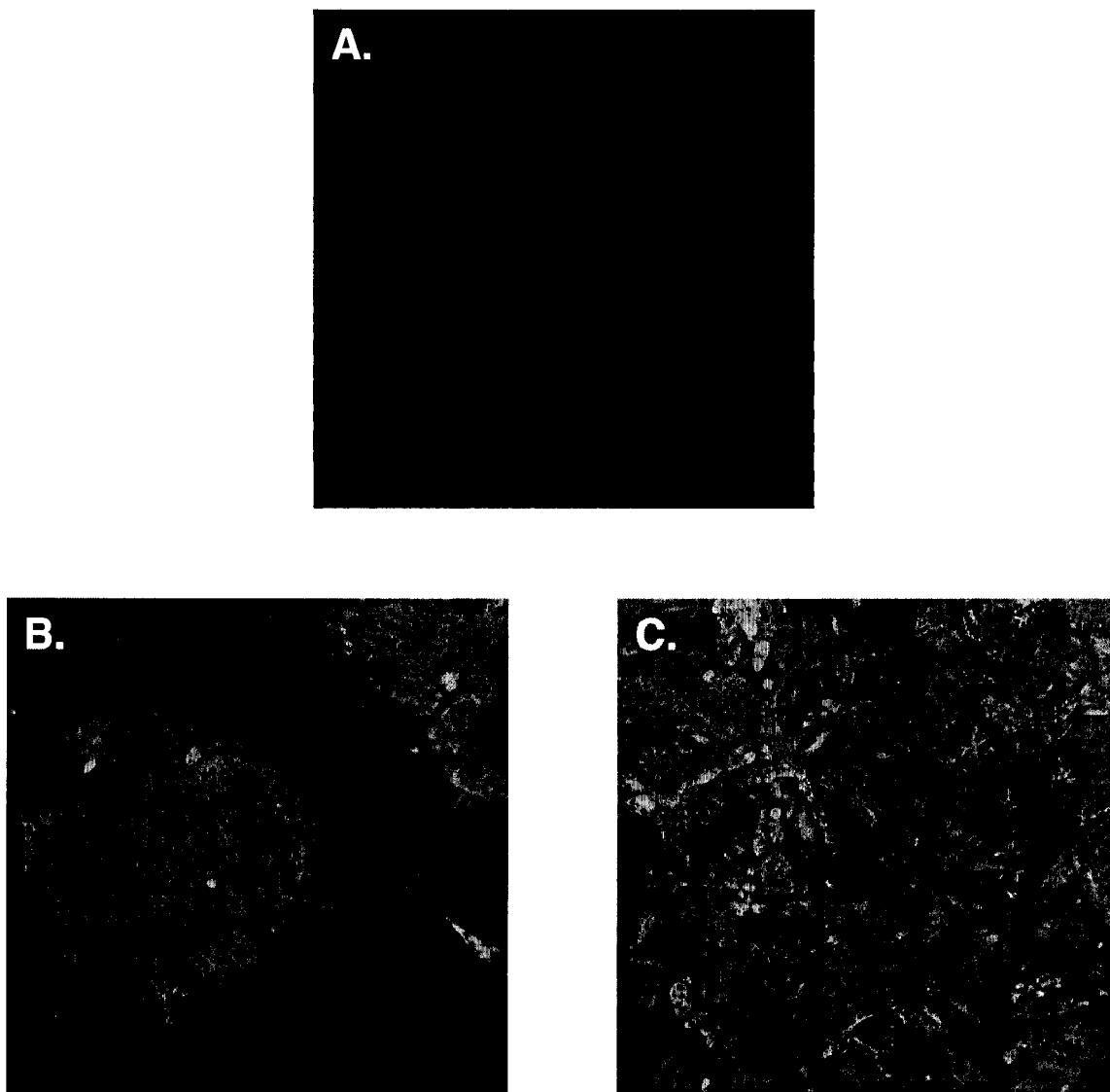


FIGURE 42

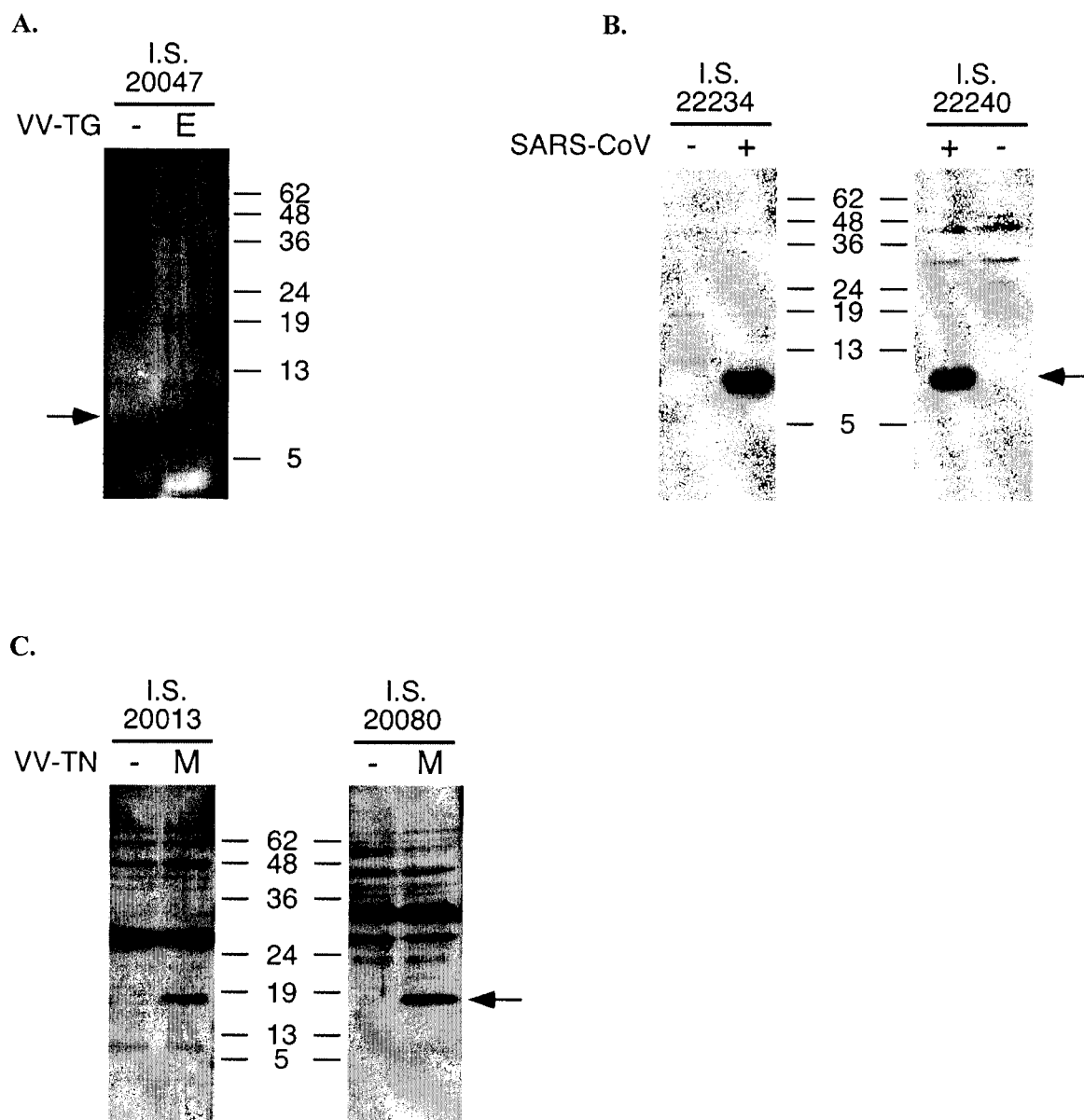


FIGURE 43

1

STRAIN OF SARS-ASSOCIATED CORONAVIRUS AND APPLICATIONS THEREOF

This is a division of application Ser. No. 10/581,356, filed Feb. 8, 2007, now U.S. Pat. No. 7,736,850, which is a continuation of International Application No. PCT/FR2004/003106, filed Dec. 2, 2004, both of which are incorporated herein by reference.

The present invention relates to a novel strain of severe acute respiratory syndrome (SARS)-associated coronavirus derived from a sample recorded under No. 031589 and collected in Hanoi (Vietnam), to nucleic acid molecules derived from its genome, to the proteins and peptides encoded by said nucleic acid molecules and to their applications, in particular as diagnostic reagents and/or as vaccine.

Coronavirus is a virus containing single-stranded RNA, of positive polarity, of approximately 30 kilobases which replicates in the cytoplasm of the host cells; the 5' end of the genome has a capped structure and the 3' end contains a polyA tail. This virus is enveloped and comprises, at its surface, peplomer structures called spicules.

The genome comprises the following open reading frames or ORFs, from its 5' end to its 3' end: ORF1a and ORF1b corresponding to the proteins of the transcription-replication complex, and ORF-S, ORF-E, ORF-M and ORF-N corresponding to the structural proteins S, E, M and N. It also comprises ORFs corresponding to proteins of unknown function encoded by: the region situated between ORF-S and ORF-E and overlapping the latter, the region situated between ORF-M and ORF-N, and the region included in ORF-N.

The S protein is a membrane glycoprotein (200-220 kDa) which exists in the form of spicules or spikes emerging from the surface of the viral envelope. It is responsible for the attachment of the virus to the receptors of the host cell and for inducing the fusion of the viral envelope with the cell membrane.

The small envelope protein (E), also called sM (small membrane), which is a nonglycosylated transmembrane protein of about 10 kDa, is the protein present in the smallest quantity in the virion. It plays a powerful role in the coronavirus budding process which occurs at the level of the intermediate compartment in the endoplasmic reticulum and the Golgi apparatus.

The M protein or matrix protein (25-30 kDa) is a more abundant membrane glycoprotein which is integrated into the viral particle by an M/E interaction, whereas the incorporation of S into the particles is directed by an S/M interaction. It appears to be important for the viral maturation of coronaviruses and for the determination of the site where the viral particles are assembled.

The N protein or nucleocapsid protein (45-50 kDa) which is the most conserved among the coronavirus structural proteins is necessary for encapsidating the genomic RNA and then for directing its incorporation into the virion. This protein is probably also involved in the replication of the RNA.

When the host cell is infected, the reading frame (ORF) situated in 5' of the viral genome is translated into a polypeptide which is cleaved by the viral proteases and then releases several nonstructural proteins such as the RNA-dependent RNA polymerase (Rep) and the ATPase helicase (Hel). These two proteins are involved in the replication of the viral genome and in the generation of transcripts which are used in the synthesis of the viral proteins. The mechanisms by which these subgenomic mRNAs are produced are not completely understood; however, recent facts indicate that the sequences for regulation of transcription at the 5' end of each gene

2

represent signals which regulate the discontinuous transcription of the subgenomic mRNAs.

The proteins of the viral membrane (S, E and M proteins) are inserted into the intermediate compartment, whereas the replicated RNA (+ strand) is assembled with the N (nucleocapsid) protein. This protein-RNA complex then combines with the M protein contained in the membranes of the endoplasmic reticulum and the viral particles form when the nucleocapsid complex buds into the endoplasmic reticulum. The virus then migrates across the Golgi complex and eventually leaves the cell, for example by exocytosis. The site of attachment of the virus to the host cell is at the level of the S protein.

Coronaviruses are responsible for 15 to 30% of colds in humans and for respiratory and digestive infections in animals, especially cats (FIPV: Feline infectious peritonitis virus), poultry (IBV: Avian infectious bronchitis virus), mice (MHV: Mouse hepatitis virus), pigs (TGEV: Transmissible gastroenteritis virus, PEDV: Porcine Epidemic diarrhea virus, PRCoV: Porcine Respiratory Coronavirus, HEV: Hemagglutinating encephalomyelitis Virus) and bovines (BCoV: Bovine coronavirus).

In general, each coronavirus affects only one species; in immunocompetent individuals, the infection induces optionally neutralizing antibodies and cell immunity, capable of destroying the infected cells.

An epidemic of atypical pneumonia, called severe acute respiratory syndrome (SARS) has spread in various countries (Vietnam, Hong Kong, Singapore, Thailand and Canada) during the first quarter of 2003, from an initial focus which appeared in China in the last quarter of 2002. The severity of this disease is such that its mortality rate is about 3 to 6%. The determination of the causative agent of this disease is underway by numerous laboratories worldwide.

In March 2003, a new coronavirus (SARS-CoV or SARS virus) was isolated, in association with cases of severe acute respiratory syndrome (T. G. KSIAZEK et al., *The New England Journal of Medicine*, 2003, 348, 1319-1330; C. DROSTEN et al., *The New England Journal of Medicine*, 2003, 348, 1967-1976; Peiris et al., *Lancet*, 2003, 361, 1319).

Genomic sequences of this new coronavirus have thus been obtained, in particular those of the Urbani isolate (Genbank accession No. AY274119.3 and A. MARRA et al., *Science*, May 1, 2003, 300, 1399-1404) and the Toronto isolate (Tor2, Genbank accession No. AY278741 and A. ROTA et al., *Science*, 2003, 300, 1394-1399).

The organization of the genome is comparable with that of other known coronaviruses, thus making it possible to confirm that SARS-CoV belongs to the Coronaviridae family; open reading frames ORF1a and 1b and open reading frames corresponding to the S, E, M and N proteins, and to proteins encoded by: the region situated between ORF-S and ORF-E (ORF3), the region situated between ORF-S and ORF-E and overlapping ORF-E (ORF4), the region situated between ORF-M and ORF-N (ORF7 to ORF11) and the region corresponding to ORF-N (ORF13 and ORF14), have in particular been identified.

Seven differences have been identified between the sequences of the Tor2 and Urbani isolates; 3 correspond to silent mutations (c/t at position 16622 and a/g at position 19064 of ORF1b, t/c at position 24872 of ORF-S) and 4 modify the amino acid sequence of respectively: the proteins encoded by ORF1a (c/t at position 7919 corresponding to the A/V mutation), the S protein (g/t at position 23220 corresponding to the A/S mutation), the protein encoded by ORF3

(a/g at position 25298 corresponding to the R/G mutation) and the M protein (t/c at position 26857 corresponding to the S/P mutation).

In addition, phylogenetic analysis shows that SARS-CoV is distant from other coronaviruses and that it did not appear by mutation of human respiratory coronaviruses nor by recombination between known coronaviruses (for a review, see Holmes, J. C. I., 2003, 111, 1605-1609).

The determination and the taking into account of new variants are important for the development of reagents for the detection and diagnosis of SARS which are sufficiently sensitive and specific, and immunogenic compositions capable of protecting populations against epidemics of SARS.

The inventors have now identified another strain of SARS-associated coronavirus which is distinguishable from the Tor2 and Urbani isolates.

The subject of the present invention is therefore an isolated or purified strain of severe acute respiratory syndrome-associated human coronavirus, characterized in that its genome has, in the form of complementary DNA, a serine codon at position 23220-23222 of the gene for the S protein or a glycine codon at position 25298-25300 of the gene for ORF3, and an alanine codon at position 7918-7920 of ORF1a or a serine codon at position 26857-26859 of the gene for the M protein, said positions being indicated in terms of reference to the Genbank sequence AY274119.3.

According to an advantageous embodiment of said strain, the DNA equivalent of its genome has a sequence corresponding to the sequence SEQ ID No: 1; this coronavirus strain is derived from the sample collected from the bronchoalveolar washings from a patient suffering from SARS, recorded under the No. 031589 and collected at the Hanoi (Vietnam) French hospital.

In accordance with the invention, said sequence SEQ ID No: 1 is that of the deoxyribonucleic acid corresponding to the ribonucleic acid molecule of the genome of the isolated coronavirus strain as defined above.

The sequence SEQ ID No: 1 is distinguishable from the Genbank sequence AY274119.3 (Tor2 isolate) in that it possesses the following mutations:

g/t at position 23220; the alanine codon (gct) at position 577 of the amino acid sequence of the Tor2 S protein is replaced by a serine codon (tct),

a/g at position 25298; the arginine codon (aga) at position 11 of the amino acid sequence of the protein encoded by the Tor2 ORF3 is replaced by a glycine codon (gga).

In addition, the sequence SEQ ID No: 1 is distinguishable from the Genbank sequence AY278741 (Urbani isolate) in that it possesses the following mutations:

t/c at position 7919; the valine codon (gtt) in position 2552 of the amino acid sequence of the protein encoded by ORF1a is replaced by an alanine codon (gct),

t/c at position 16622: this mutation does not modify the amino acid sequence of the proteins encoded by ORF1b (silent mutation),

g/a at position 19064: this mutation does not modify the amino acid sequence of the proteins encoded by ORF1b (silent mutation),

c/t at position 24872: this mutation does not modify the amino acid sequence of the S protein, and

c/t at position 26857: the proline codon (ccc) at position 154 of the amino acid sequence of the M protein is replaced by a serine codon (tcc).

Unless otherwise stated, the positions of the nucleotide and peptide sequences are indicated with reference to the Genbank sequence AY274119.3.

The subject of the present invention is also an isolated or purified polynucleotide, characterized in that its sequence is that of the genome of the isolated coronavirus strain as defined above.

According to an advantageous embodiment of said polynucleotide, it has the sequence SEQ ID No: 1.

The subject of the present invention is also an isolated or purified polynucleotide, characterized in that its sequence hybridizes under high stringency conditions with the sequence of the polynucleotide as defined above.

The terms "isolated or purified" mean modified "by the hand of humans" from the natural state; in other words if an object exists in nature, it is said to be isolated or purified if it is modified or extracted from its natural environment or both. For example, a polynucleotide or a protein/peptide naturally present in a living organism is neither isolated nor purified; on the other hand, the same polynucleotide or protein/peptide separated from coexisting molecules in its natural environment, obtained by cloning, amplification and/or chemical synthesis is isolated for the purposes of the present invention. Furthermore, a polynucleotide or a protein/peptide which is introduced into an organism by transformation, genetic manipulation or by any other method, is "isolated" even if it is present in said organism. The term purified as used in the present invention means that the proteins/peptides according to the invention are essentially free of association with the other proteins or polypeptides, as is for example the product purified from the culture of recombinant host cells or the product purified from a nonrecombinant source.

For the purposes of the present invention, high stringency hybridization conditions are understood to mean temperature and ionic strength conditions chosen such that they make it possible to maintain the specific and selective hybridization between complementary polynucleotides.

By way of illustration, high stringency conditions for the purposes of defining the above polynucleotides are advantageously the following: the DNA-DNA or DNA-RNA hybridization is performed in two steps: (1) prehybridization at 42° C. for 3 hours in phosphate buffer (20 mM, pH 7.5) containing 5×SSC (1×SSC corresponds to a 0.15 M NaCl+0.015 M sodium citrate solution), 50% formamide, 7% sodium dodecyl sulfate (SDS), 10×Denhardt's, 5% dextran sulfate and 1% salmon sperm DNA; (2) hybridization for 20 hours at 42° C. followed by 2 washings of 20 minutes at 20° C. in 2×SSC+2% SDS, 1 washing of 20 minutes at 20° C. in 0.1×SSC+0.1% SDS. The final washing is performed in 0.1×SSC+0.1% SDS for 30 minutes at 60° C.

The subject of the present invention is also a representative fragment of the polynucleotide as defined above, characterized in that it is capable of being obtained either by the use of restriction enzymes whose recognition and cleavage sites are present in said polynucleotide as defined above, or by amplification with the aid of oligonucleotide primers specific for said polynucleotide as defined above, or by transcription in vitro, or by chemical synthesis.

According to an advantageous embodiment of said fragment, it is selected from the group consisting of: the cDNA corresponding to at least one open reading frame (ORF) chosen from: ORF1a, ORF1b, ORF-S, ORF-E, ORF-M, ORF-N, ORF3, ORF4, ORF7 to ORF11, ORF13 and ORF14 and the cDNA corresponding to the noncoding 5' or 3' ends of said polynucleotide.

According to an advantageous feature of this embodiment, said fragment has a sequence selected from the group consisting of:

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the sequences SEQ ID NO: 2 and 4 representing the cDNA corresponding to the ORF-S which encodes the S protein,

the sequences SEQ ID NO: 13 and 15 representing the cDNA corresponding to the ORF-E which encodes the E protein,

the sequences SEQ ID NO: 16 and 18 representing the cDNA corresponding to the ORF-M which encodes the M protein,

the sequences SEQ ID NO: 36 and 38 representing the cDNA corresponding to the ORF-N which encodes the N protein,

the sequences representing the cDNA corresponding respectively: to ORF1a and ORF1b (ORF1ab, SEQ ID NO: 31), to ORF3 and ORF4 (SEQ ID NO: 7, 8), to ORF7 to 11 (SEQ ID NO: 19, 20) to ORF13 (SEQ ID NO: 32) and to ORF14 (SEQ ID NO: 34), and

the sequences representing the cDNAs corresponding respectively to the noncoding 5' (SEQ ID NO: 39 and 72) and 3' (SEQ ID NO: 40, 73) ends of said polynucleotide.

The subject of the present invention is also a cDNA fragment encoding the S protein, as defined above, characterized in that it has a sequence selected from the group consisting of the sequences SEQ ID NO: 5 and 6 (Sa and Sb fragments).

The subject of the present invention is also a cDNA fragment corresponding to ORF1a and ORF1b as defined above, characterized in that it has a sequence selected from the group consisting of the sequences SEQ ID NO: 41 to 54 (L0 to L12 fragments).

The subject of the present invention is also a polynucleotide fragment as defined above, characterized in that it has at least 15 consecutive bases or base pairs of the sequence of the genome of said strain including at least one of those situated in position 7979, 16622, 19064, 23220, 24872, 25298 and 26857. Preferably this is a fragment of 20 to 2500 bases or base pairs, preferably from 20 to 400.

According to an advantageous embodiment of said fragment, it includes at least one pair of bases or base pairs corresponding to the following positions: 7919 and 23220, 7919 and 25298, 16622 and 23220, 19064 and 23220, 16622 and 25298, 19064 and 25298, 23220 and 24872, 23220 and 26857, 24872 and 25298, 25298 and 26857.

The subject of the present invention is also primers of at least 18 bases capable of amplifying a fragment of the genome of a SARS-associated coronavirus or of the DNA equivalent thereof.

According to an embodiment of said primers, they are selected from the group consisting of:

the pair of primers No. 1 corresponding respectively to positions 28507 to 28522 (sense primer, SEQ ID NO: 60) and 28774 to 28759 (antisense primer, SEQ ID NO: 61) of the sequence of the polynucleotide as defined above,

the pair of primers No. 2 corresponding respectively to positions 28375 to 28390 (sense primer, SEQ ID NO: 62) and 28702 to 28687 (antisense primer, SEQ ID NO: 63) of the sequence of the polynucleotide as defined above, and

the pair of primers consisting of the primers SEQ ID Nos: 55 and 56.

The subject of the present invention is also a probe capable of detecting the presence of the genome of a SARS-associated coronavirus or of a fragment thereof, characterized in that it is selected from the group consisting of: the fragments as defined above and the fragments corresponding to the following positions of the polynucleotide sequence as defined

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above: 28561 to 28586, 28588 to 28608, 28541 to 28563 and 28565 to 28589 (SEQ ID NO: 64 to 67).

The probes and primers according to the invention may be labeled directly or indirectly with a radioactive or nonradioactive compound by methods well known to persons skilled in the art so as to obtain a detectable and/or quantifiable signal. Among the radioactive isotopes used, there may be mentioned ^{32}P , ^{33}P , ^{35}S , ^3H or ^{125}I . The nonradioactive entities are selected from ligands such as biotin, avidin, streptavidin, digoxigenin, haptens, dyes, luminescent agents such as radioluminescent, chemoluminescent, bioluminescent, fluorescent and phosphorescent agents.

The invention encompasses the labeled probes and primers derived from the preceding sequences.

Such probes and primers are useful for the diagnosis of infection by a SARS-associated coronavirus.

The subject of the present invention is also a method for the detection of a SARS-associated coronavirus, from a biological sample, which method is characterized in that it comprises at least:

(a) the extraction of nucleic acids present in said biological sample,

(b) the amplification of a fragment of ORF-N by RT-PCR with the aid of a pair of primers as defined above, and

(c) the detection, by any appropriate means, of the amplification products obtained in (b).

The amplification products (amplicons) in (b) are 268 bp for the pair of primers No. 1 and 328 bp for the pair of primers No. 2.

According to an advantageous embodiment of said method, the step (b) of detection is carried out with the aid of at least one probe corresponding to positions 28561 to 28586, 28588 to 28608, 28541 to 28563 and 28565 to 28589 of the sequence of the polynucleotide as defined above.

Preferably, the SARS-associated coronavirus genome is detected and optionally quantified by PCR in real time with the aid of the pair of primers No. 2 and probes corresponding to positions 28541 to 28563 and 28565 to 28589 labeled with different compounds, in particular different fluorescent agents.

The real time RT-PCR which uses this pair of primers and this probe is very sensitive since it makes it possible to detect 10^2 copies of RNA and up to 10 copies, of RNA; it is in addition reliable and reproducible.

The invention encompasses the single-stranded, double-stranded and triple-stranded polydeoxyribonucleotides and polyribonucleotides corresponding to the sequence of the genome of the isolated strain of coronavirus and its fragments as defined above, and to their sense or antisense complementary sequences, in particular the RNAs and cDNAs corresponding to the sequence of the genome and of its fragments as defined above.

The present invention also encompasses the amplification fragments obtained with the aid of primers specific for the genome of the purified or isolated strain as defined above, in particular with the aid of primers or pairs of primers as defined above, the restriction fragments formed by or comprising the sequence of fragments as defined above, the fragments obtained by transcription in vitro from a vector containing the sequence SEQ ID NO: 1 or a fragment as defined above, and fragments obtained by chemical synthesis. Examples of restriction fragments are deduced from the restriction map of the sequence SEQ ID NO: 1 illustrated by FIG. 13. In accordance with the invention, said fragments are either in the form of isolated fragments, or in the form of mixtures of fragments. The invention also encompasses fragments modified, in relation to the preceding ones, by removal

or addition of nucleotides in a proportion of about 15%, relative to the length of the above fragments and/or modified in terms of the nature of the nucleotides, as long as the modified nucleotide fragments retain a capacity for hybridization with the genomic or antigenomic RNA sequences of the isolate as defined above.

The nucleic acid molecules according to the invention are obtained by conventional methods, known per se, following standard protocols such as those described in *Current Protocols in Molecular Biology* (Frederick M. AUSUBEL, 2000, Wiley and son Inc., Library of Congress, USA). For example, they may be obtained by amplification of a nucleic sequence by PCR or RT-PCR or alternatively by total or partial chemical synthesis.

The subject of the present invention is also a DNA or RNA chip or filter, characterized in that it comprises at least one polynucleotide or one of its fragments as defined above.

The DNA or RNA chips or filters according to the invention are prepared by conventional methods, known per se, such as for example chemical or electrochemical grafting of oligonucleotides on a glass or nylon support.

The subject of the present invention is also a recombinant cloning and/or expression vector, in particular a plasmid, a virus, a viral vector or a phage comprising a nucleic acid fragment as defined above. Preferably, said recombinant vector is an expression vector in which said nucleic acid fragment is placed under the control of appropriate elements for regulating transcription and translation. In addition, said vector may comprise sequences (tags) fused in phase with the 5' and/or 3' end of said insert, which are useful for the immobilization and/or detection and/or purification of the protein expressed from said vector.

These vectors are constructed and introduced into host cells by conventional recombinant DNA and genetic engineering methods which are known per se. Numerous vectors into which a nucleic acid molecule of interest may be inserted in order to introduce it and to maintain it in a host cell are known per se; the choice of an appropriate vector depends on the use envisaged for this vector (for example replication of the sequence of interest, expression of this sequence, maintenance of the sequence in extrachromosomal form or alternatively integration into the chromosomal material of the host), and on the nature of the host cell.

In accordance with the invention, said plasmid is selected in particular from the following plasmids:

the plasmid, called SARS-S, contained in the bacterial strain deposited under the No. I-3059, on Jun. 20, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA sequence encoding the S protein of the SARS-CoV strain derived from the sample recorded under the No. 031589, said sequence corresponding to the nucleotides at positions 21406 to 25348 (SEQ ID NO: 4), with reference to the Genbank sequence AY274119.3,

the plasmid, called SARS-S1, contained in the bacterial strain deposited under the No. I-3020, on May 12, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains a 5' fragment of the cDNA sequence encoding the S protein of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above, said fragment corresponding to the nucleotides at positions 21406 to 23454 (SEQ ID NO: 5), with reference to the Genbank sequence AY274119.3 Tor2,

the plasmid, called SARS-S2, contained in the bacterial strain deposited under the No. I-3019, on May 12, 2003,

at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains a 3' fragment of the cDNA sequence encoding the S protein of the SARS-CoV strain derived from the sample recorded under the number No. 031589, as defined above, said fragment corresponding to the nucleotides at positions 23322 to 25348 (SEQ ID NO: 6), with reference to the Genbank sequence accession No. AY274119.3,

the plasmid, called SARS-SE, contained in the bacterial strain deposited under the No. I-3126, on Nov. 13, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA corresponding to the region situated between ORF-S and ORF-E and overlapping ORF-E of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above, said region corresponding to the nucleotides at positions 25110 to 26244 (SEQ ID NO: 8), with reference to the Genbank sequence accession No. AY274119.3,

the plasmid, called SARS-E, contained in the bacterial strain deposited under the No. I-3046, on May 28, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA sequence encoding the E protein of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above, said sequence corresponding to the nucleotides at positions 26082 to 26413 (SEQ ID NO: 15), with reference to the Genbank sequence accession No. AY274119.3,

the plasmid, called SARS-M, contained in the bacterial strain deposited under the No. I-3047, on May 28, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA sequence encoding the M protein of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above; said sequence corresponding to the nucleotides at positions 26330 to 27098 (SEQ ID NO: 18), with reference to the Genbank sequence accession No. AY274119.3,

the plasmid, called SARS-MN, contained in the bacterial strain deposited under the No. I-3125, on Nov. 13, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA sequence corresponding to the region situated between ORF-M and ORF-N of the SARS-CoV strain derived from the sample recorded under the No. 031589 and collected in Hanoi, as defined above, said sequence corresponding to the nucleotides at positions 26977 to 28218 (SEQ ID NO: 20), with reference to the Genbank accession No. AY274119.3,

the plasmid, called SARS-N, contained in the bacterial strain deposited under the No. I-3048, on Jun. 5, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA encoding the N protein of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above, said sequence corresponding to the nucleotides at positions 28054 to 29430 (SEQ ID NO: 38), with reference to the Genbank sequence accession No. AY274119.3; thus, this plasmid comprises an insert of sequence SEQ ID NO: 38 and is contained in a bacterial strain which was deposited under the No. I-3048, on Jun. 5, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15,

the plasmid, called SARS-5'NC, contained in the bacterial strain deposited under the No. I-3124, on Nov. 7, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA corresponding to the noncoding 5' end of the genome of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above, said sequence corresponding to the nucleotides at positions 1 to 204 (SEQ ID NO: 39), with reference to the Genbank sequence accession No. AY274119.3,

the plasmid called SARS-3'NC, contained in the bacterial strain deposited under the No. I-3123 on Nov. 7, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA sequence corresponding to the non-coding 3' end of the genome of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above, said sequence corresponding to that situated between the nucleotide and position 28933 to 29727 (SEQ ID NO: 40), with reference to the Genbank sequence accession No. AY274119.3, ends with a series of nucleotides a,

the expression plasmid, called pIV2.3N, containing a cDNA fragment encoding a C-terminal fusion of the N protein (SEQ ID NO: 37) with a polyhistidine tag,

the expression plasmid, called pIV2.3S_C, containing a cDNA fragment encoding a C-terminal fusion of the fragment corresponding to positions 475 to 1193 of the amino acid sequence of the S protein (SEQ ID NO: 3) with a polyhistidine tag,

the expression plasmid, pIV2.3S_L, containing a cDNA fragment encoding a C-terminal fusion of the fragment corresponding to positions 14 to 1193 of the amino acid sequence of the S protein (SEQ ID NO: 3) with a polyhistidine tag,

the expression plasmid, called pIV2.4N, containing a cDNA fragment encoding a N-terminal fusion of the N protein (SEQ ID NO: 3) with a polyhistidine tag,

the expression plasmid, called pIV2.4S_C or pIV2.4S_L, containing an insert encoding a N-terminal fusion of the fragment corresponding to positions 475 to 1193 of the amino acid sequence of the S protein (SEQ ID NO: 3) with a polyhistidine tag, and

the expression plasmid, called pIV2.4S_L, containing a cDNA fragment encoding an N-terminal fusion of the fragment corresponding to positions 14 to 1193 of the amino acid sequence of the S protein (SEQ ID NO: 3) with a polyhistidine tag.

According to an advantageous feature of the expression plasmid as defined above, it is contained in a bacterial strain which was deposited under the No. I-3117, on Oct. 23, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15.

According to another advantageous feature of the expression plasmid as defined above, it is contained in a bacterial strain which was deposited under the No. I-3118, on Oct. 23, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15.

According to another feature of the expression plasmid as defined above, it is contained in a bacterial strain which was deposited at the CNCM, 25 rue du Docteur Roux, 75724 Paris Cedex 15 under the following numbers:

- a) strain No. I-3118, deposited on Oct. 23, 2003,
- b) strain No. I-3019, deposited on May 12, 2003,
- c) strain No. I-3020, deposited on May 12, 2003,
- d) strain No. I-3059, deposited on Jun. 20, 2003,
- e) strain No. I-3323, deposited on Nov. 22, 2004,

- f) strain No. I-3324, deposited on Nov. 22, 2004,
- g) strain No. I-332, deposited on Dec. 1, 2004,
- h) strain No. I-3327, deposited on Dec. 1, 2004,
- i) strain No. I-3332, deposited on Dec. 1, 2004,
- j) strain No. I-3333, deposited on Dec. 1, 2004,
- k) strain No. I-3334, deposited on Dec. 1, 2004,
- l) strain No. I-3335, deposited on Dec. 1, 2004,
- m) strain No. I-3336, deposited on Dec. 1, 2004,
- n) strain No. I-3337, deposited on Dec. 1, 2004,
- o) strain No. I-3338, deposited on Dec. 2, 2004,
- p) strain No. I-3339, deposited on Dec. 2, 2004,
- q) strain No. I-3340, deposited on Dec. 2, 2004,
- r) strain No. I-3341, deposited on Dec. 2, 2004.

The subject of the present invention is also a nucleic acid insert of viral origin, characterized in that it is contained in any of the strains as defined above in a)-r).

The subject of the present invention is also a nucleic acid containing a synthetic gene allowing optimized expression of the S protein in eukaryotic cells, characterized in that it possesses the sequence SEQ ID NO: 140.

The subject of the present invention is also an expression vector containing a nucleic acid containing a synthetic gene allowing optimized expression of the S protein, which vector is contained in the bacterial strain deposited at the CNCM, on Dec. 1, 2004, under the No. I-3333.

According to one embodiment of said expression vector, it is a viral vector, in the form of a viral particle or in the form of a recombinant genome.

According to an advantageous feature of this embodiment, this is a recombinant viral particle or a recombinant viral genome capable of being obtained by transfection of a plasmid according to paragraphs g), h) and k) to r) as defined above, in an appropriate cellular system, that is to say, for example, cells transfected with one or more other plasmids intended to transcomplement certain functions of the virus that are deleted in the vector and that are necessary for the formation of the viral particles.

The expression "S protein family" is understood here to mean the complete S protein, its ectodomain and fragments of this ectodomain which are preferably produced in a eukaryotic system.

The subject of the present invention is also a lentiviral vector encoding a polypeptide of the S protein family, as defined above.

The subject of the present invention is also a recombinant measles virus encoding a polypeptide of the S protein family, as defined above.

The subject of the present invention is also a recombinant vaccinia virus encoding a polypeptide of the S protein family, as defined above.

The subject of the present invention is also the use of a vector according to paragraphs e) to r) as defined above, or of a vector containing a synthetic gene for the S protein, as defined above, for the production, in a eukaryotic system, of the SARS-associated coronavirus S protein or of a fragment of this protein.

The subject of the present invention is also a method for producing the S protein in a eukaryotic system, comprising a step of transfecting eukaryotic cells in culture with a vector chosen from the vectors contained in the bacterial strains mentioned in paragraphs e) to r) above or a vector containing a synthetic gene allowing optimized expression of the S protein.

The subject of the present invention is also a cDNA library characterized in that it comprises fragments as defined above,

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in particular amplification fragments or restriction fragments, cloned into a recombinant vector, in particular an expression vector (expression library).

The subject of the present invention is also cells, in particular prokaryotic cells, modified by a recombinant vector as defined above.

The subject of the present invention is also a genetically modified eukaryotic cell expressing a protein or a polypeptide as defined above. Quite obviously, the terms "genetically modified eukaryotic cell" do not denote a cell modified with a wild-type virus.

According to an advantageous embodiment of said cell, it is capable of being obtained by transfection with any of the vectors mentioned in paragraphs i) to l) above.

According to an advantageous feature of this embodiment, this is the cell FRhK4-Ssol-30, deposited at the CNCM on Nov. 22, 2004, under the No. I-3325.

The recombinant vectors as defined above and the cells transformed with said expression vectors are advantageously used for the production of the corresponding proteins and peptides. The expression libraries derived from said vectors, and the cells transformed with said expression libraries are advantageously used to identify the immunogenic epitopes (B and T epitopes) of the SARS-associated coronavirus proteins.

The subject of the present invention is also the purified or isolated proteins and peptides, characterized in that they are encoded by the polynucleotide or one of its fragments as defined above.

According to an advantageous embodiment of the invention, said protein is selected from the group consisting of:

the S protein having the sequence SEQ ID NO: 3 or its ectodomaine

the E protein having the sequence SEQ ID NO: 14

the M protein having the sequence SEQ ID NO: 17

the N protein having the sequence SEQ ID NO: 37

the proteins encoded by the ORFs: ORF1a, ORF1b, ORF3, ORF4 and ORF7 to ORF11, ORF13 and ORF14 and having the respective sequence, SEQ ID NO: 74, 75, 10, 12, 22, 24, 26, 28, 30, 33 and 35.

The terms "ectodomaine of the S protein" and "soluble form of the S protein" will be used interchangeably below.

According to an advantageous embodiment of the invention, said polypeptide consists of the amino acids corresponding to positions 1 to 1193 of the amino acid sequence of the S protein.

According to another advantageous embodiment of the invention, said peptide is selected from the group consisting of:

a) the peptides corresponding to positions 14 to 1193 and 475 to 1193 of the amino acid sequence of the S protein,

b) the peptides corresponding to positions 2 to 14 (SEQ ID NO: 69) and 100 to 221 of the amino acid sequence of the M protein; these peptides correspond respectively to the ectodomaine and to the endodomaine of the M protein, and

c) the peptides corresponding to positions 1 to 12 (SEQ ID NO: 70) and 53 to 76 (SEQ ID NO: 71) of the amino acid sequence of the E protein; these peptides correspond respectively to the ectodomaine and to the C-terminal end of the E protein, and

d) the peptides of 5 to 50 consecutive amino acids, preferably of 10 to 30 amino acids, inclusive or partially or completely overlapping the sequence of the peptides as defined in a), b) or c).

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The subject of the present invention is also a peptide, characterized in that it has a sequence of 7 to 50 amino acids including an amino acid residue selected from the group consisting of:

the alanine situated at position 2552 of the amino acid sequence of the protein encoded by ORF1a,

the serine situated at position 577 of the amino acid sequence of the S protein of the SARS-CoV strain as defined above,

the glycine at position 11 of the amino acid sequence, of the protein encoded by ORF3 of the SARS-CoV strain as defined above,

the serine at position 154 of the amino acid sequence of the M protein of the SARS-CoV strain as defined above.

The subject of the present invention is also an antibody or a polyclonal or monoclonal antibody fragment which can be obtained by immunization of an animal with a recombinant vector as defined above, a cDNA library as defined above or alternatively a protein or a peptide as defined above, characterized in that it binds to at least one of the proteins encoded by SARS-CoV as defined above.

The invention encompasses the polyclonal antibodies, the monoclonal antibodies, the chimeric antibodies such as the humanized antibodies, and fragments thereof (Fab, Fv, scFv).

A subject of the present invention is also a hybridoma producing a monoclonal antibody against the N protein, characterized in that it is chosen from the following hybridomas: the hybridoma producing the monoclonal antibody 87, deposited at the CNCM on Dec. 1, 2004 under the number I-3328,

the hybridoma producing the monoclonal antibody 86, deposited at the CNCM on Dec. 1, 2004 under the number I-3329,

the hybridoma producing the monoclonal antibody 57, deposited at the CNCM on Dec. 1, 2004 under the number I-3330, and

the hybridoma producing the monoclonal antibody 156, deposited at the CNCM on Dec. 1, 2004 under the number I-3331.

The subject of the present invention is also a polyclonal or monoclonal antibody or antibody fragment directed against the N protein, characterized in that it is produced by a hybridoma as defined above.

For the purposes of the present invention, the expression chimeric antibody is understood to mean, in relation to an antibody of a particular animal species or of a particular class of antibody, an antibody comprising all or part of a heavy chain and/or of a light chain of an antibody of another animal species or of another class of antibody.

For the purposes of the present invention, the expression humanized antibody is understood to mean a human immunoglobulin in which the residues of the CDRs (Complementary Determining Regions) which form the antigen-binding site are replaced by those of a nonhuman monoclonal antibody possessing the desired specificity, affinity or activity. Compared with the nonhuman antibodies, the humanized antibodies are less immunogenic and possess a prolonged half-life in humans because they possess only a small proportion of nonhuman sequences given that practically all the residues of the FR (Framework) regions and of the constant (Fc) region of these antibodies are those of a consensus sequence of human immunoglobulins.

A subject of the present invention is also a protein chip or filter, characterized in that it comprises a protein, a peptide or alternatively an antibody as defined above.

The protein chips according to the invention are prepared by conventional methods known per se. Among the appropri-

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ate supports on which proteins may be immobilized, there may be mentioned those made of plastic or glass, in particular in the form of microplates.

The subject of the present invention is also reagents derived from the isolated strain of SARS-associated coronavirus, derived from the sample recorded under the No. 031589, which are useful for the study and diagnosis of the infection caused by a SARS-associated coronavirus, said reagents are selected from the group consisting of:

- (a) a pair of primers, a probe or a DNA chip as defined above,
- (b) a recombinant vector or a modified cell as defined above,
- (c) an isolated coronavirus strain or a polynucleotide as defined above,
- (d) a protein or a peptide as defined above,
- (e) an antibody or an antibody fragment as defined above, and
- (f) a protein chip as defined above.

These various reagents are prepared and used according to conventional molecular biology and immunology techniques following standard protocols such as those described in *Current Protocols in Molecular Biology* (Frederick M. AUSUBEL, 2000, Wiley and Son Inc., Library of Congress, USA), in *Current Protocols in Immunology* (John E. Coligan, 2000, Wiley and Son Inc., Library of Congress, USA) and in *Antibodies: A Laboratory Manual* (E. Howell and D. Lane, Cold Spring Harbor Laboratory, 1988).

The nucleic acid fragments according to the invention are prepared and used according to conventional techniques as defined above. The peptides and proteins according to the invention are prepared by recombinant DNA techniques, known to persons skilled in the art, in particular with the aid of the recombinant vectors as defined above. Alternatively, the peptides according to the invention may be prepared by conventional techniques of solid or liquid phase synthesis, known to persons skilled in the art.

The polyclonal antibodies are prepared by immunizing an appropriate animal with a protein or a peptide as defined above, optionally coupled to KLH or to albumin and/or combined with an appropriate adjuvant such as (complete or incomplete) Freund's adjuvant or aluminum hydroxide; after obtaining a satisfactory antibody titer, the antibodies are harvested by collecting serum from the immunized animals and enriched with IgG by precipitation, according to conventional techniques, and then the IgGs specific for the SARS-CoV proteins are optionally purified by affinity chromatography on an appropriate column to which said peptide or said protein is attached, as defined above, so as to obtain a monospecific IgG preparation.

The monoclonal antibodies are produced from hybridomas obtained by fusion of B lymphocytes from an animal immunized with a protein or a peptide as defined above with myelomas, according to the Köhler and Milstein technique (Nature, 1975, 256, 495-497); the hybridomas are cultured in vitro, in particular in fermenters or produced in vivo, in the form of ascites; alternatively, said monoclonal antibodies are produced by genetic engineering as described in American patent U.S. Pat. No. 4,816,567.

The humanized antibodies are produced by general methods such as those described in International application WO 98/45332.

The antibody fragments are produced from the cloned V_H and V_L regions, from the mRNAs of hybridomas or splenic lymphocytes of an immunized mouse; for example, the Fv, scFv or Fab fragments are expressed at the surface of filamentous phages according to the Winter and Milstein tech-

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nique (Nature, 1991, 349, 293-299); after several selection steps, the antibody fragments specific for the antigen are isolated and expressed in an appropriate expression system, by conventional techniques for cloning and expression of recombinant DNA.

The antibodies or fragments thereof as defined above are purified by conventional techniques known to persons skilled in the art, such as affinity chromatography.

The subject of the present invention is additionally the use of a product selected from the group consisting of: a pair of primers, a probe, a DNA chip, a recombinant vector, a modified cell, an isolated coronavirus strain, a polynucleotide, a protein or a peptide, an antibody or an antibody fragment and a protein chip as defined above, for the preparation of a reagent for the detection and optionally genotyping/serotyping of a SARS-associated coronavirus.

The proteins and peptides according to the invention, which are capable of being recognized and/or of inducing the production of antibodies specific for the SARS-associated coronavirus, are useful for the diagnosis of infection with such a coronavirus; the infection is detected, by an appropriate technique—in particular EIA, ELISA, RIA, immunofluorescence—in a biological sample collected from an individual capable of being infected.

According to an advantageous feature of said use, said proteins are selected from the group consisting of the S, E, M and/or N proteins and the peptides as defined above.

The S, E, M and/or N proteins and the peptides derived from these proteins as defined above, for example the N protein, are used for the indirect diagnosis of a SARS-associated coronavirus infection (serological diagnosis; detection of an antibody specific for SARS-CoV), in particular by an immunoenzymatic method (ELISA).

The antibodies and antibody fragments according to the invention, in particular those directed against the S, E, M and/or N proteins and the derived peptides as defined above, are useful for the direct diagnosis of a SARS-associated coronavirus infection; the detection of the protein(s) of SARS-CoV is carried out by an appropriate technique, in particular EIA, ELISA, RIA, immunofluorescence, in a biological sample collected from an individual capable of being infected.

The subject of the present invention is also a method for the detection of a SARS-associated coronavirus, from a biological sample, which method is characterized in that it comprises at least:

- (a) bringing said biological sample into contact with at least one antibody or one antibody fragment, one protein, one peptide or alternatively one protein or peptide chip or filter as defined above, and
- (b) visualizing by any appropriate means antigen-antibody complexes formed in (a), for example by EIA, ELISA, RIA, or by immunofluorescence.

According to one advantageous embodiment of said process, step (a) comprises:

- (a₁) bringing said biological sample into contact with at least a first antibody or an antibody fragment which is attached to an appropriate support, in particular a microplate,
- (a₂) washing the solid phase, and
- (a₃) adding at least a second antibody or an antibody fragment, different from the first, said antibody or antibody fragment being optionally appropriately labeled.

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This method, which makes it possible to capture the viral particles present in the biological sample, is also called immunocapture method.

For example:

step (a₁) is carried out with at least a first monoclonal or polyclonal antibody or a fragment thereof, directed against the S, M and/or E protein, and/or a peptide corresponding to the ectodomain of one of these proteins (M2-14 or E1-12 peptides)

step (a₃) is carried out with at least one antibody or an antibody fragment directed against another epitope of the same protein or preferably against another protein, preferably against an inner protein such as the N nucleoprotein or the endodomain of the E or M protein, more preferably still these are antibodies or antibody fragments directed against the N protein which is very abundant in the viral particle; when an antibody or an antibody fragment directed against an inner protein (N) or against the endodomain of the E or M proteins is used, said antibody is incubated in the presence of detergent, such as Tween 20 for example, at concentrations of the order of 0.1%.

step (b) for visualizing the antigen-antibody complexes formed is carried out, either directly with the aid of a second antibody labeled for example with biotin or an appropriate enzyme such as peroxidase or alkaline phosphatase, or indirectly with the aid of an anti-immunoglobulin serum labeled as above. The complexes thus formed are visualized with the aid of an appropriate substrate.

According to a preferred embodiment of this aspect of the invention, the biological sample is mixed with the visualizing monoclonal antibody prior to its being brought into contact with the capture monoclonal antibodies. Where appropriate, the serum-visualizing antibody mixture is incubated for at least 10 minutes at room temperature before being applied to the plate.

The subject of the present invention is also an immunocapture test intended to detect an infection by the SARS-associated coronavirus by detecting the native nucleoprotein (N protein), in particular characterized in that the antibody used for the capture of the native viral nucleoprotein is a monoclonal antibody specific for the central region and/or for a conformational epitope.

According to one embodiment of said test, the antibody used for the capture of the N protein is the monoclonal antibody mAb87, produced by the hybridoma deposited at the CNCM on Dec. 1, 2004 under the number I-3328.

According to another embodiment of said immunocapture test, the antibody used for the capture of the N protein is the monoclonal antibody mAb86, produced by the hybridoma deposited at the CNCM on Dec. 1, 2004 under the number I-3329.

According to another embodiment of said immunocapture test, the monoclonal antibodies mAb86 and mAb87 are used for the capture of the N protein.

In the immunocapture tests according to the invention, it is possible to use, for visualizing the N protein, the monoclonal antibody mAb57, produced by the hybridoma deposited at the CNCM on Dec. 1, 2004 under the number I-3330, said antibody being conjugated with a visualizing molecule or particle.

In accordance with said immunocapture test, a combination of the antibodies mAb57 and mAb87, conjugated with a visualizing molecule or particle, is used for the visualization of the N protein.

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A visualizing molecule may be a radioactive atom, a dye, a fluorescent molecule, a fluorophore, an enzyme; a visualizing particle may be for example: colloidal gold, a magnetic particle or a latex bead.

The subject of the present invention is also a reagent for detecting a SARS-associated coronavirus, characterized in that it is selected from the group consisting of:

- (a) a pair of primers or a probe as defined above,
- (b) a recombinant vector as defined above or a modified cell as defined above,
- (c) an isolated coronavirus strain as defined above or a polynucleotide as defined above,
- (d) an antibody or an antibody fragment as defined above,
- (e) a combination of antibodies comprising the monoclonal antibodies mAb86 and/or mAb87, and the monoclonal antibody mAb57, as defined above,
- (f) a chip or a filter as defined above.

The subject of the present invention is also a method for the detection of a SARS-associated coronavirus infection, from a biological sample, by indirect IgG ELISA using the N protein, which method is characterized in that the plates are sensitized with an N protein solution at a concentration of between 0.5 and 4 µg/ml, preferably to 2 µg/ml, in a 10 mM PBS buffer pH 7.2, phenol red at 0.25 ml/l.

The subject of the present invention is additionally a method for the detection of a SARS-associated coronavirus infection, from a biological sample, by double epitope ELISA, characterized in that the serum to be tested is mixed with the visualizing antigen, said mixture then being brought into contact with the antigen attached to a solid support.

According to one variant of the tests for detecting SARS-associated coronaviruses, these tests combine an ELISA using the N protein, and another ELISA using the S protein, as described below.

The subject of the present invention is also an immune complex formed of a polyclonal or monoclonal antibody or antibody fragment as defined above, and of a SARS-associated coronavirus protein or peptide.

The subject of the present invention is additionally a SARS-associated coronavirus detection kit, characterized in that it comprises at least one reagent selected from the group consisting of: a pair of primers, a probe, a DNA or RNA chip, a recombinant vector, a modified cell, an isolated coronavirus strain, a polynucleotide, a protein or a peptide, an antibody, and a protein chip as defined above.

The subject of the present invention is additionally an immunogenic composition, characterized in that it comprises at least one product selected from the group consisting of:

- a) a protein or a peptide as defined above,
 - b) a polynucleotide of the DNA or RNA type or one of its representative fragments as defined above, having a sequence chosen from:
 - (i) the sequence SEQ ID NO: 1 or its RNA equivalent
 - (ii) the sequence hybridizing under high stringency conditions with the sequence SEQ ID NO: 1,
 - (iii) the sequence complementary to the sequence SEQ ID NO: 1 or to the sequence hybridizing under high stringency conditions with the sequence SEQ ID NO: 1,
 - (iv) the nucleotide sequence of a representative fragment of the polynucleotide as defined in (i), (ii) or (iii),
 - (v) the sequence as defined in (i), (ii), (iii) or (iv), modified, and
 - c) a recombinant expression vector comprising a polynucleotide as defined in b), and
 - d) a cDNA library as defined above,
- said immunogenic composition being capable of inducing protective humoral or cellular immunity specific for the

SARS-associated coronavirus, in particular the production of an antibody directed against a specific epitope of the SARS-associated coronavirus.

The proteins and peptides as defined above, in particular the S, M, E and/or N proteins and the derived peptides, and the nucleic acid (DNA or RNA) molecules encoding said proteins or said peptides are good candidate vaccines and may be used in immunogenic compositions for the production of a vaccine against the SARS-associated coronavirus.

According to an advantageous embodiment of the compositions according to the invention, they additionally contain at least one pharmaceutically acceptable vehicle and optionally carrier substances and/or adjuvants.

The pharmaceutically acceptable vehicles, the carrier substances and the adjuvants are those conventionally used.

The adjuvants are advantageously chosen from the group consisting of oily emulsions, saponin, mineral substances, bacterial extracts, aluminum hydroxide and squalene.

The carrier substances are advantageously selected from the group consisting of unilamellar liposomes, multilamellar liposomes, micelles of saponin or solid microspheres of a saccharide or auriferous nature.

The compositions according to the invention are administered by the general route, in particular by the intramuscular or subcutaneous route or alternatively by the local, in particular nasal (aerosol) route.

The subject of the present invention is also the use of an isolated or purified protein or peptide having a sequence selected from the group consisting of the sequences SEQ ID NO: 3, 10, 12, 14, 17, 22, 24, 26, 28, 30, 33, 35, 37, 69, 70, 71, 74 and 75 to form an immune complex with an antibody specifically directed against an epitope of the SARS-associated coronavirus.

The subject of the present invention is also an immune complex consisting of an isolated or purified protein or peptide having a sequence selected from the group consisting of the sequences SEQ ID NO: 3, 10, 12, 14, 17, 22, 24, 26, 28, 30, 33, 35, 37, 69, 70, 71, 74 and 75, and of an antibody specifically directed against an epitope of the SARS-associated coronavirus.

The subject of the present invention is also the use of an isolated or purified protein or peptide having a sequence selected from the group consisting of the sequences SEQ ID NO: 3, 10, 12, 14, 17, 22, 24, 26, 28, 30, 33, 35, 37, 69, 70, 71, 74 and 75 to induce the production of an antibody capable of specifically recognizing an epitope of the SARS-associated coronavirus.

The subject of the present invention is also the use of an isolated or purified polynucleotide having a sequence selected from the group consisting of the sequences SEQ ID NO: 1, 2, 4, 7, 8, 13, 15, 16, 18, 19, 20, 31, 36 and 38 to induce the production of an antibody directed against the protein encoded by said polynucleotide and capable of specifically recognizing an epitope of the SARS-associated coronavirus.

The subject of the present invention is also monoclonal antibodies recognizing the native S protein of a SARS-associated coronavirus.

The subject of the present invention is also the use of a protein or a polypeptide of the S protein family, as defined above, or of an antibody recognizing the native S protein, as defined above, to detect an infection by a SARS-associated coronavirus, in a biological sample.

The subject of the present invention is also a method for detecting an infection by a SARS-associated coronavirus, in a biological sample, characterized in that the detection is carried out by ELISA using the recombinant S protein, expressed in a eukaryotic system.

According to an advantageous embodiment of said method, it is a double epitope ELISA method, and the serum to be tested is mixed with the visualizing antigen, said mixture then being brought into contact with the antigen attached to a solid support.

The subject of the present invention is also an immune complex consisting of a monoclonal antibody or antibody fragment recognizing the native S protein, and of a protein or a peptide of the SARS-associated coronavirus.

The subject of the present invention is also an immune complex consisting of a protein or a polypeptide of the S protein family, as defined above, and of an antibody specifically directed against an epitope of the SARS-associated coronavirus.

The subject of the present invention is additionally a SARS-associated coronavirus detection kit or box, characterized in that it comprises at least one reagent selected from the group consisting of: a protein or polypeptide of the S protein family, as defined above, a nucleic acid encoding a protein or peptide of the S protein family, as defined above, a cell expressing a protein or polypeptide of the S protein family, as defined above, or an antibody recognizing the native S protein of a SARS-associated coronavirus.

The subject of the present invention is an immunogenic and/or vaccine composition, characterized in that it comprises a polypeptide or a recombinant protein of the S protein family, as defined above, obtained in a eukaryotic expression system.

The subject of the present invention is also an immunogenic and/or vaccine composition, characterized in that it comprises a vector or recombinant virus, expressing a protein or a polypeptide of the S protein family, as defined above.

In addition to the preceding features, the invention further comprises other features, which will emerge from the description which follows, which refers to examples of use of the polynucleotide representing the genome of the SARS-CoV strain derived from the sample recorded under the number 031589, and derived cDNA fragments which are the subject of the present invention, and to Table I presenting the sequence listing:

TABLE I

Sequence listing			
Identification number	Sequence	Position of the cDNA with reference to Genbank AY274119.3	Deposit number at the CNCM of the corresponding plasmid
SEQ ID NO: 1	genome of the strain derived from the sample 031589	—	—
SEQ ID NO: 2	ORF-S*	21406-25348	—
SEQ ID NO: 3	S protein	—	—
SEQ ID NO: 4	ORF-S**	21406-25348	I-3059
SEQ ID NO: 5	Sa fragment	21406-23454	I-3020
SEQ ID NO: 6	Sb fragment	23322-25348	I-3019
SEQ ID NO: 7	ORF-3 + ORF-4*	25110-26244	—
SEQ ID NO: 8	ORF-3 + ORF-4**	25110-26244	I-3126
SEQ ID NO: 9	ORF3	—	—
SEQ ID NO: 10	ORF-3 protein	—	—
SEQ ID NO: 11	ORF4	—	—
SEQ ID NO: 12	ORF-4 protein	—	—
SEQ ID NO: 13	ORF-E*	26082-26413	—
SEQ ID NO: 14	E protein	—	—
SEQ ID NO: 15	ORF-E**	26082-26413	I-3046
SEQ ID NO: 16	ORF-M*	26330-27098	—
SEQ ID NO: 17	M protein	—	—

TABLE I-continued

Sequence listing		Position of the cDNA with reference to Genbank AY274119.3	Deposit number at the CNCM of the cor- responding plasmid
Identification number	Sequence		
SEQ ID NO: 18	ORF-M**	26330-27098	I-3047
SEQ ID NO: 19	ORF7 to 11*	26977-28218	—
SEQ ID NO: 20	ORF7 to 11**	26977-28218	I-3125
SEQ ID NO: 21	ORF7	—	—
SEQ ID NO: 22	ORF7 protein	—	—
SEQ ID NO: 23	ORF8	—	—
SEQ ID NO: 24	ORF8 protein	—	—
SEQ ID NO: 25	ORF9	—	—
SEQ ID NO: 26	ORF9 protein	—	—
SEQ ID NO: 27	ORF10	—	—
SEQ ID NO: 28	ORF10 protein	—	—
SEQ ID NO: 29	ORF11	—	—
SEQ ID NO: 30	ORF11 protein	—	—
SEQ ID NO: 31	Orf1ab	265-21485	—
SEQ ID NO: 32	ORF13	28130-28426	—
SEQ ID NO: 33	ORF13 protein	—	—
SEQ ID NO: 34	ORF14	—	—
SEQ ID NO: 35	ORF14 protein	28583-28795	—
SEQ ID NO: 36	ORF-N*	28054-29430	—
SEQ ID NO: 37	N protein	—	—
SEQ ID NO: 38	ORF-N**	28054-29430	I-3048
SEQ ID NO: 39	noncoding 5**	1-204	I-3124
SEQ ID NO: 40	noncoding 3**	28933-29727	I-3123
SEQ ID NO: 41	ORF1ab	30-500	—
SEQ ID NO: 42	Fragment L0	—	—
SEQ ID NO: 43	Fragment L1	211-2260	—
SEQ ID NO: 44	Fragment L2	2136-4187	—
SEQ ID NO: 45	Fragment L3	3892-5344	—
SEQ ID NO: 46	Fragment L4b	4932-6043	—
SEQ ID NO: 47	Fragment L4	5305-7318	—
SEQ ID NO: 48	Fragment L5	7275-9176	—
SEQ ID NO: 49	Fragment L6	9032-11086	—
SEQ ID NO: 50	Fragment L7	10298-12982	—
SEQ ID NO: 51	Fragment L8	12815-14854	—
SEQ ID NO: 52	Fragment L9	14745-16646	—
SEQ ID NO: 53	Fragment L10	16514-18590	—
SEQ ID NO: 54	Fragment L11	18500-20602	—
SEQ ID NO: 55	Fragment L12	20319-22224	—
SEQ ID NO: 56	Sense N primer	—	—
SEQ ID NO: 57	Antisense	—	—
SEQ ID NO: 58	N primer	—	—
SEQ ID NO: 59	Sense S _C primer	—	—
SEQ ID NO: 60	Sense S ₂ primer	—	—
SEQ ID NO: 61	Antisense S _C	—	—
SEQ ID NO: 62	and S ₂ primer	—	—
SEQ ID NO: 63	Sense primer series 1	28507-28522	—
SEQ ID NO: 64	Antisense primer series 1	28774-28759	—
SEQ ID NO: 65	Sense primer series 2	28375-28390	—
SEQ ID NO: 66	Antisense primer series 2	28702-28687	—
SEQ ID NO: 67	Probe 1/series 1	28561-28586	—
SEQ ID NO: 68	Probe 2/series 1	28588-28608	—
SEQ ID NO: 69	Probe 1/series 2	28541-28563	—
SEQ ID NO: 70	Probe 2/series 2	28565-28589	—
SEQ ID NO: 71	Anchor primer	—	—
SEQ ID NO: 72	Peptide M2-14	—	—
SEQ ID NO: 73	Peptide E1-12	—	—
SEQ ID NO: 74	Peptide E53-76	—	—
SEQ ID NO: 75	Noncoding 5**	1-204	—
SEQ ID NO: 76	Noncoding 3**	28933-29727	—
SEQ ID NO: 77	ORF1a protein	—	—
SEQ ID NO: 78	ORF1b protein	—	—
SEQ ID NO: 79	Primers	—	—
SEQ ID NO: 80	Pseudogene of S	—	—
SEQ ID NO: 81	Primers	—	—
SEQ ID NO: 82	Aa1-13 of S	—	—

TABLE I-continued

Sequence listing		Position of the cDNA with reference to Genbank AY274119.3	Deposit number at the CNCM of the cor- responding plasmid
Identification number	Sequence		
SEQ ID NO: 150	Polypeptide		
SEQ ID NO: 151-158	Primers		
*PCR amplification product (amplicon)			
**Insert cloned into the plasmid deposited at the CNCM and to the appended drawings in which:			
FIG. 1 illustrates Western-blot analysis of the expression in vitro of the recombinant proteins N, S _C and S ₂ from the expression vectors pVEX. Lane 1: pV2.3N. Lane 2: pV2.3S _C . Lane 3: pV2.3S ₂ . Lane 4: pV2.4N. Lane 5: pV2.4S ₁ or pV2.4S _C . Lane 6: pV2.4S ₂ . The expression of the GFP protein expressed from the same vector is used as a control.			
FIG. 2 illustrates the analysis, by polyacrylamide gel electrophoresis under denaturing conditions (SDS-PAGE) and staining with Coomassie blue, of the expression in vivo of the N protein from the expression vectors pVEX. The <i>E. coli</i> BL21(DE3)pDIA17 strain transformed with the recombinant vectors pVEX is cultured at 30° C. in LB medium, in the presence or in the absence of inducer (IPTG 1 mM). Lane 1: pV2.3N. Lane 2: pV2.4N.			
FIG. 3 illustrates the analysis, by polyacrylamide gel electrophoresis under denaturing conditions (SDS-PAGE) and staining with Coomassie blue, of the expression in vivo of the S ₂ and S _C polypeptides from the expression vectors pVEX. The <i>E. coli</i> BL21(DE3)pDIA17 strain transformed with the recombinant vectors pVEX is cultured at 30° C. in LB medium, in the presence or in the absence of inducer (IPTG 1 mM). Lane 1: pV2.3S _C . Lane 2: pV2.3S ₂ . Lane 3: pV2.4S ₁ . Lane 4: pV2.4S ₂ .			
FIG. 4 illustrates the antigenic activity of the recombinant N, S ₂ and S _C proteins produced in the <i>E. coli</i> BL21(DE3)pDIA17 strain transformed with the recombinant vectors pVEX. A: electrophoresis (SDS-PAGE) of the bacterial lysates. B and C: Western-blot with the sera, obtained from the same patient infected with SARS-CoV, collected 8 days (B: serum M12) and 29 days (C: serum M13) respectively after the onset of the SARS symptoms. Lane 1: pV2.3N. Lane 2: pV2.4N. Lane 3: pV2.3S _C . Lane 4: pV2.4S ₁ . Lane 5: pV2.4S ₂ . Lane 6: pV2.4S ₂ .			
FIG. 5 illustrates the purification on an Ni-NTA agarose column of the recombinant N protein produced in the <i>E. coli</i> BL21(DE3)pDIA17 strain from the vector pV2.3N. Lane 1: total bacterial extract. Lane 2: soluble extract. Lane 3: insoluble extract. Lane 4: extract deposited on the Ni-NTA column. Lane 5: unbound proteins. Lane 6: fractions of peak 1. Lane 7: fractions of peak 2.			
FIG. 6 illustrates the purification of the recombinant S _C protein from the inclusion bodies produced in the <i>E. coli</i> BL21(DE3)pDIA17 strain transformed with pV2.4S ₁ . A: Treatment with Triton X-100 (2%): Lane 1: total bacterial extract. Lane 2: soluble extract. Lane 3: insoluble extract. Lane 4: supernatant after treatment with Triton X-100 (2%). Lanes 5 and 6: pellet after treatment with Triton X-100 (2%). B: Treatment with 4 M, 5 M, 6 M and 7 M urea of the soluble and insoluble extracts.			
FIG. 7 represents the immunoblot produced with the aid of a lysate of cells infected with SARS-CoV and a serum from a patient suffering from a typical pneumonia.			
FIG. 8 represents immunoblots produced with the aid of a lysate of cells infected with SARS-CoV and rabbit immunosera specific for the nucleoprotein N (A) and for the spike protein S (B). L.S.: immune serum. p.i.: preimmune serum. The anti-N immune serum was used at 1/50 000 and the anti-S immune serum at 1/10 000.			
FIG. 9 illustrates the ELISA reactivity of the rabbit monospecific polyclonal sera directed against the N protein or the short fragment of the S protein (S ₂), toward the corresponding recombinant proteins used for immunization. A: rabbits P13097, P13081 and P13031 immunized with the purified recombinant N protein. B: rabbits P11135, P13042 and P14001 immunized with a preparation of inclusion bodies corresponding to the short fragment of the S protein (S ₂). L.S.: immune serum. p.i.: preimmune serum.			
FIG. 10 illustrates the ELISA reactivity of the purified recombinant N protein, toward sera from patients suffering from a typical pneumonia caused by SARS-CoV. FIG. 10a: ELISA plates prepared with the N protein at the concentration of 4 µg/ml and 2 µg/ml. FIG. 10b: ELISA plate prepared with the N protein at the concentration of 1 µg/ml. The sera designated A, B, D, E, F, G, H correspond to those of Table IV.			
FIG. 11 illustrates the amplification by RT-PCR of decreasing quantities of synthetic RNA of the SARS-CoV N gene (10 ¹ to 1 copy), with the aid of pairs of primers No. 1 (N+/28507, N/-/28774) (A) and No. 2 (N+/28375, N/-/28702) (B). T: amplification performed in the absence of RNA. MW: DNA marker.			
FIG. 12 illustrates the amplification by RT-PCR in real time of synthetic RNA for the SARS-CoV N gene: decreasing quantities of synthetic RNA as replica (repli.; lanes 16 to 29) and of viral RNA diluted 1/20 × 10 ⁻⁴ (lane 32) were amplified by RT-PCR in real time with the aid of the kit "Light Cycler RNA Amplification Kit Hybridization Probes" and pairs of primers and probes of the No. 2 series, under the conditions described in Example 8.			
FIG. 13 (FIG. 13.1 to 13.7) represents the restriction map of the sequence SEQ ID NO: 1 corresponding to the DNA equivalent of the genome of the SARS-CoV strain derived from the sample recorded under the number 031589.			
FIG. 14 shows the result of the SARS serology test by indirect N ELISA (1 st series of sera tested).			
FIG. 15 shows the result of the SARS serology test by indirect N ELISA (2 nd series of sera tested).			
FIG. 16 presents the result of the SARS serology test by double epitope N ELISA (1 st series of sera tested).			
FIG. 17 shows the result of the SARS serology test by double epitope N ELISA (2 nd series of sera tested).			
FIG. 18 illustrates the test of reactivity of the anti-N monoclonal antibodies by ELISA on the native nucleoprotein N of SARS-CoV. The antibodies were tested in the form of hybridoma culture supernatants by indirect ELISA using an irradiated lysate of VeroE6 cells infected with SARS-CoV as antigen (SARS lysate curves). A negative control for reactivity is performed for each antibody on a lysate of uninfected VeroE6 cells (negative lysate curves).			
Several monoclonal antibodies of known specificity were used as negative control antibodies: para1-3 directed against the antigens of the parainfluenza viruses type 1-3 (Bio-Rad) and influenza B directed against the antigens of the influenza virus type B (Bio-Rad).			

TABLE I-continued

Identification number	Sequence	Sequence listing	
		Position of the cDNA with reference to Genbank	Deposit number at the CNCM of the corresponding plasmid
		AY274119.3	

FIG. 19 illustrates the test of reactivity of the anti-N of SARS-CoV monoclonal antibodies by ELISA on the native antigens of the human coronavirus 229E (HCoV-229E). The antibodies were tested in the form of hybridoma culture supernatants by an indirect ELISA test using a lysate of MRC-5 cells infected with the human coronavirus 229E as antigen (229E lysate curves). A negative control for immunoreactivity was performed for each antibody on a lysate of noninfected MRC-5 cells (negative lysate curves). The monoclonal antibody 5-11H.6 directed against the S protein of the human coronavirus 229E (Sizun et al. 1998, J. Virol. Met. 72: 145-152) is used as positive control antibody. The antibodies para-1-3 directed against the antigens of the parainfluenza virus type 1-3 (Bio-Rad) and influenza B directed against the antigens of the influenza virus type B (Bio-Rad) were added to the panel of monoclonal antibodies tested.

FIG. 20 shows a test of reactivity of the anti-N of SARS-CoV monoclonal antibodies by Western blotting on the denatured native nucleoprotein N of SARS-CoV. A lysate of VeroE6 cells infected with SARS-CoV was prepared in the loading buffer according to Laemmli and caused to migrate in a 12% SDS polyacrylamide gel and then the proteins were transferred onto PVDF membrane. The anti-N monoclonal antibodies tested were used for the immunoblotting at the concentration of 0.05 µg/ml. The visualization is carried out with anti-mouse IgG(H + L) antibodies coupled to peroxidase (NA931V, Amersham) and the ECL+ system. Two monoclonal antibodies were used as negative controls for reactivity: influenza B directed against the antigens of the influenza virus type B (Bio-Rad) and para-1-3 directed against the antigens of the parainfluenza virus type 1-3 (Bio-Rad).

FIG. 21 presents the plasmids for expression in mammalian cells of the SARS-CoV S protein. The cDNA for the SARS-CoV S was inserted between the BamHI and XhoI sites of the expression plasmid pcDNA3.1(+) (Clontech) in order to obtain the plasmid pcDNA-S and between the NheI and XhoI sites of the expression plasmid pCI (Promega) in order to obtain the plasmid pCI-S. The WPRE and CTE sequences were inserted between each of the two plasmids pcDNA-S and pCI-S between the XhoI and XbaI sites in order to obtain the plasmids pcDNA-S-CTE, pcDNA-S-WPRE, pCI-S-CTE and pCI-S-WPRE, respectively. SP: signal peptide predicted (aa 1-13) with the software signalP v2.0 (Nielsen et al., 1997, Protein Engineering, 10: 1-6)

TM: transmembrane region predicted (aa 1196-1218) with the software TMHMM v2.0 (Sonnhammer et al., 1998, Proc. of Sixth Int. Conf. on Intelligent Systems for Molecular Biology, pp. 175-182, AAAI Press). It should be noted that the amino acids W1194 and P1195 are possibly part of the transmembrane region with the respective probabilities of 0.13 and 0.42

P-CMV: cytomegalovirus immediate/early promoter.

BGH pA: polyadenylation signal of the bovine growth hormone gene

SV40 late pA: SV40 virus late polyadenylation signal

SD/SA: splice donor and acceptor sites

WPRES: sequences of the "Woodchuck Hepatitis Virus posttranscriptional regulatory element" of the woodchuck hepatitis virus

CTE: sequences of the "constitutive transport element" of the Mason-Pfizer simian retrovirus

FIG. 22 illustrates the expression of the S protein after transfection of VeroE6 cells. Cellular extracts were prepared 48 hours after transfection of VeroE6 cells with the plasmids pcDNA, pcDNA-S, pCI and pCI-S. Cellular extracts were also prepared 18 hours after infection with the recombinant vaccinia virus VV-TF7.3 and transfection with the plasmids pcDNA or pcDNA-S. As a control, extracts of VeroE6 cells were prepared 8 hours after infection with SARS-CoV at a multiplicity of infection of 3. They were separated on an 8% SDS acrylamide gel and analyzed by Western blotting with the aid of an anti-S rabbit polyclonal antibody and an anti-rabbit IgG(H + L) polyclonal antibody coupled to peroxidase (NA934V, Amersham). A molecular mass ladder (kDa) is presented in the figure.

SARS-CoV: extract of VeroE6 cells infected with SARS-CoV

Mock: control extract of noninfected cells

FIG. 23 illustrates the effect of the CTE and WPRES sequences on the expression of the S protein after transfection of VeroE6 and 293T cells. Cellular extracts were prepared 48 hours after transfection of VeroE6 cells (A) or 293T cells (B) with the plasmids pcDNA, pcDNA-S, pcDNA-S-CTE, pcDNA-S-WPRE, pCI-S, pCI-S-CTE and pCI-S-WPRE separated on 8% SDS polyacrylamide gel and analyzed by Western blotting with the aid of an anti-S rabbit polyclonal antibody and an anti-rabbit IgG(H + L) polyclonal antibody coupled to peroxidase (NA934V, Amersham). A molecular mass ladder (kDa) is presented in the figure.

SARS-CoV: extract of VeroE6 cells prepared 8 hours after infection with SARS-CoV at a multiplicity of infection of 3.

Mock: control extract of noninfected VeroE6 cells

FIG. 24 presents defective lentiviral vectors with central DNA flap for the expression of SARS-CoV S. The cDNA for the SARS-CoV S protein was cloned in the form of a BamHI-XhoI fragment into the plasmid pTRIPAU3-CMV containing a defective lentiviral vector TRIP with central DNA flap (Sirven et al., 2001, Mol. Ther., 3: 438-448) in order to obtain the plasmid pTRIP-S. The optimum expression cassettes consisting of the CMV virus immediate/early promoter, a splice signal, cDNA for S and either of the posttranscriptional signals CTE or WPRES were substituted for the cassette EF1α-EGFP of the defective lentiviral expression vector with central DNA flap TRIPAU3-EF1α (Sirven et al., 2001, Mol. Ther., 3: 438-448) in order to obtain the plasmids pTRIP-SD/SA-S-CTE and pTRIP-SD/SA-S-WPRE.

SP: signal peptide

TM: transmembrane region

P-CMV: cytomegalovirus immediate/early promoter

P-EF1α: EF1α gene promoter

SD/SA: splice donor and acceptor sites

WPRES: sequences of the "Woodchuck Hepatitis Virus posttranscriptional regulatory element" of the woodchuck hepatitis virus

CTE: sequences of the "constitutive transport element" of the Mason-Pfizer simian retrovirus

LTR: long terminal repeat AU3: LTR deleted for the "promoter/enhancer" sequences

cPPT: "polypurine tract cis-active sequence"

CTS: "central termination sequence"

TABLE I-continued

Identification number	Sequence	Sequence listing	
		Position of the cDNA with reference to Genbank	Deposit number at the CNCM of the corresponding plasmid
		AY274119.3	

5

Identification number

FIG. 25 shows the Western-blot analysis of the expression of the SARS-CoV S by cell lines transduced with the lentiviral vectors TRIP-SD/SA-S-WPRE and TRIP-SD/SA-S-CTE. Cellular extracts were prepared from established lines FrhK4-S-CTE and FrhK4-S-WPRE after transduction with the lentiviral vectors TRIP-SD/SA-S-CTE and TRIP-SD/SA-S-WPRE respectively. They were separated on an 8% SDS acrylamide gel and analyzed by Western blotting with the aid of an anti-S rabbit polyclonal antibody and an anti-rabbit IgG(H + L) conjugate coupled to peroxidase. A molecular mass ladder (kDa) is presented in the figure.

T-: control extract of FrhK-4 cells

T+: extract of FrhK-4 cells prepared 24 hours after infection with SARS-CoV at a multiplicity of infection of 3.

FIG. 26 relates to the analysis of the expression of Ssol polypeptide by cell lines transduced with the lentiviral vectors TRIP-SD/SA-Ssol-WPRE and TRIP-SD/SA-Ssol-CTE. The secretion of the Ssol polypeptide was determined in the supernatant of a series of cell clones isolated after transduction of FrhK-4 cells with the lentiviral vectors TRIP-SD/SA-Ssol-WPRE and TRIP-SD/SA-Ssol-CTE. 5 µl of supernatant, diluted 1/2 in loading buffer according to Laemmli, were analyzed by Western blotting, visualized with an anti-FLAG monoclonal antibody (M2, Sigma) and an anti-mouse IgG(H + L) conjugate coupled to peroxidase.

T-: supernatant of the parental FrhK-4 line.

T+: supernatant of FBK cells infected with a recombinant vaccinia virus expressing the Ssol polypeptide. The solid arrow indicates the Ssol polypeptide, while the empty arrow indicates a cross reaction with a protein of cellular origin.

FIG. 27 shows the results relating to the analysis of the purified Ssol polypeptide

A. 8, 2, 0.5 and 0.125 µg of recombinant Ssol polypeptide purified by anti-FLAG affinity chromatography and gel filtration (G75) were separated on 8% SDS polyacrylamide gel. The Ssol polypeptide and variable quantities of molecular mass markers (MM) were visualized by staining with silver nitrate (Gelcode SilverSNAP stain kit II, Pierce).

B. Standard markers for analysis by SELDI-TOF mass spectrometry

IgG: bovine IgG of MM 147300

ConA: conalbumin of MM 77490

HRP: horseradish peroxidase analyzed as a control and of MM 43240

C. Analysis by mass spectrometry (SELDI-TOF) of the recombinant Ssol polypeptide.

The peaks A and B correspond to the single and double charged Ssol polypeptide.

D. Sequencing of the N-terminal end of the recombinant Ssol polypeptide. 5 Edman degradation cycles in liquid phase were carried out on an ABI494 sequencer (Applied Biosystems).

FIG. 28 illustrates the influence of a splicing signal and of the CTE and WPRES sequences on the efficacy of the gene immunization with the aid of plasmid DNA encoding the SARS-CoV S

A. Groups of 7 BALB/c mice were immunized twice at 4 weeks' interval with the aid of 50 µg of plasmid DNA of pCI, pcDNA-S, pCI-S, pcDNA-N and pCI-HA.

B. Groups of 6 BALB/c mice were immunized twice at 4 weeks' interval with the aid of 2 µg, 10 µg or 50 µg of plasmid DNA of pCI, pCI-S, pCI-S-CTE and pCI-S-WPRE.

The immune sera collected 3 weeks after the second immunization were analyzed by indirect ELISA using a lysate of VeroE6 cells infected with SARS-CoV as antigen. The anti-SARS-CoV antibody titers are calculated as the reciprocal of the dilution producing a specific OD of 0.5 after visualization with an anti-mouse IgG polyclonal antibody coupled to peroxidase (NA931V, Amersham) and TMB (KPL).

FIG. 29 shows the seroneutralization of the

infectivity of SARS-CoV with the antibodies induced in

mice after gene immunization with the aid of plasmid

DNA encoding SARS-CoV S. Pools of immune sera collected

3 weeks after the second immunization were prepared for

each of the groups of experiments described in

FIG. 28 and evaluated for their capacity to seroneutralize the infectivity of 100 TCID₅₀ of SARS-CoV on FRhK-4 cells. 4 points are produced for each of the 2-fold dilutions tested from 1/20. The seroneutralizing titer is calculated according to the Reed and Munch method as the reciprocal of the dilution neutralizing the infectivity of 2 wells out of 4.

A. Groups by BALB/c mice immunized twice at 4 weeks' interval with the aid of 50 µg of plasmid DNA of pCI, pcDNA-S, pCI-S, pcDNA-N and pCI-HA. □: preimmune serum. ■: immune serum.

B. Groups of BALB/c mice immunized twice at 4 weeks' interval with the aid of 2 µg, 10 µg or 50 µg of plasmid DNA of pCI, pCI-S, pCI-S-CTE and pCI-S-WPRE.

FIG. 30 illustrates the immunoreactivity of the recombinant Ssol polypeptide toward sera from patients suffering from SARS. The reactivity of sera from patients was analyzed by indirect ELISA test against solid phases prepared with the aid of the purified recombinant Ssol polypeptide. The antibodies from patients reacting with the solid phase at a dilution of 1/400 are visualized with a human anti-IgG(H + L) polyclonal antibody coupled to peroxidase (Amersham NA933V) and TMB plus H202 (KPL). The sera of probable SARS cases are identified by a National Reference Center for Influenza Viruses serial number and by the initials of the patient and the number of days elapsed since the onset of symptoms, where appropriate. The TV sera are control sera from subjects which were collected in France before the SARS epidemic which occurred in 2003.

FIG. 31 shows the induction of antibodies directed against SARS-CoV after immunization with the recombinant Ssol polypeptide. Two groups of 6 mice were immunized at 3 weeks' interval with 10 µg of recombinant Ssol polypeptide (Ssol group) adjuvanted with aluminum hydroxide or, as a control, of adjuvant alone (mock group). Three successive immunizations were performed and the immune sera were collected 3 weeks after each of the three immunizations (IS1, IS2, IS3). The immune sera were analyzed per pool for each of the 2 groups by indirect ELISA using a lysate of VeroE6 cells infected with SARS-CoV as antigen. The anti-SARS-CoV antibody titers are calculated as the reciprocal of the dilution producing a specific OD of 0.5 after visualization with an anti-mouse IgG polyclonal antibody coupled to peroxidase (Amersham) and TMB (KPL).

FIG. 32 presents the nucleotide alignment of the sequences of the synthetic gene 040530 with the sequence of the wild-type gene of the SARS-CoV isolate 031589. I-3059 corresponds to nucleotides 21406-25348 of the SARS-CoV isolate 031589 deposited at the C.N.C.M. under the number I-3059 (SEQ ID NO: 4, plasmid pSARS-S) S-040530 is the sequence of the synthetic gene 040530.

65

TABLE I-continued

Sequence listing		Position of the cDNA with reference to Genbank	Deposit number at the CNCM of the cor- responding plasmid
Identification number	Sequence	AY274119.3	

FIG. 33 illustrates the use of a synthetic gene for the expression of the SARS-CoV S. Cellular extracts prepared 48 hours after transfection of VeroE6 cells (A) or 293T cells (B) with the plasmids pCI, pCI-S, pCI-S-CTE, pCI-S-WPRE and pCI-S-synth were separated on 8% SDS acrylamide gel and analyzed by Western blotting with the aid of an anti-S rabbit polyclonal antibody and an anti-rabbit IgG(H + L) polyclonal antibody coupled to peroxidase (NA934V, Amersham). The Western blot is visualized by luminescence (ECL+, Amersham) and acquisition on a digital imaging device (FluorS, BioRad). The levels of expression of the S protein were measured by quantifying the 2 predominant bands identified on the image.

FIG. 34 presents a diagram for the construction of recombinant vaccinia viruses VV-TG-S, VV-TG-Ssol, VV-TN-S and VV-TN-Ssol.

A. The cDNAs for the S protein and the Ssol polypeptide of SARS-CoV were inserted between the BamHI and SmaI sites of the transfer plasmid pTG186 in order to obtain the plasmids pTG-S and pTG-Ssol.

B. The sequences of the synthetic promoter 480 were then substituted for those of the 7.5 promoter by exchange of the NdeI-PstI fragments of the plasmids pTG186poly, pTG-S and pTG-Ssol in order to obtain the transfer plasmids pTN480, pTN-S and pTN-Ssol.

C. Sequence of the synthetic promoter 480 as contained between the NdeI and PstI sites of the transfer plasmids of the pTN series. An AscI site was inserted in order to facilitate subsequent handling. The restriction sites and the promoter sequence are underlined.

D. The recombinant vaccinia viruses are obtained by double homologous recombination in vivo between the TK cassette of the transfer plasmids of the pTG and pTN series and the TK gene of the Copenhagen strain of the vaccinia virus.

SP: signal peptide predicted (aa 1-13) with the software signalP v2.0 (Nielsen et al., 1997, Protein Engineering, 10: 1-6)

TM: transmembrane region predicted (aa 1196-1218) with the software TMHMM v2.0 (Sonnhammer et al., 1998, Proc. of Sixth Int. Conf. on Intelligent Systems for Molecular Biology, pp. 175-182, AAAI Press). It should be noted that the amino acids W1194 and P1195 possibly form part of the transmembrane region with respective probabilities of 0.13 and 0.42.

TK-L, TK-R: left- and right-hand parts of the vaccinia virus thymidine kinase gene

MCS: multiple cloning site

PE: early promoter

PL: late promoter

PL synth: synthetic late promoter 480

FIG. 35 illustrates the expression of the S protein by recombinant vaccinia viruses, analyzed by Western blotting. Cellular extracts were prepared 18 hours after infection of CV1 cells with the recombinant vaccinia viruses VV-TG, VV-TG-S and VV-TN-S at an M.O.I. of 2 (A). As a control, extracts of VeroE6 cells were prepared 8 hours after infection with SARS-CoV at a multiplicity of infection of 2. Cellular extracts were also prepared 18 hours after infection of CV1 cells with the recombinant vaccinia viruses VV-TG-S, VV-TG-Ssol, VV-TN, VV-TN-S and VV-TN-Ssol (B). They were separated on 8% SDS acrylamide gels and analyzed by Western blotting with the aid of an anti-S rabbit polyclonal antibody and an anti-rabbit IgG(H + L) polyclonal antibody coupled to peroxidase (NA934V, Amersham). "1 μ l" and "10 μ l" indicates the quantities of cellular extracts deposited on the gel. A molecular mass ladder (kDa) is presented in the figure.

SARS-CoV: extract of VeroE6 cells infected with SARS-CoV

Mock: control extract of noninfected cells

FIG. 36 shows the result of a Western-blot analysis of the secretion of the Ssol polypeptide by the recombinant vaccinia viruses.

A. Supernatants of CV1 cells infected with the recombinant vaccinia virus VV-TN, various clones of the VV-TN-Ssol virus and with the viruses VV-TG-Ssol or VV-TN-Sflag were harvested 18 hours after infection of CV1 cells at an M.O.I. of 2.

B. Supernatants of 293T, FRhK-4, BHK-21 and CV1 cells infected in duplicate (1:2) with the recombinant vaccinia virus VV-TN-Ssol at an M.O.I. of 2 were harvested 18 hours after infection. The supernatant of CV1 cells infected with the virus VV-TN was also harvested as a control (M).

All the supernatants were separated on 8% SDS acrylamide gel according to Laemmli and analyzed by Western blotting with the aid of an anti-FLAG mouse monoclonal antibody and an anti-mouse IgG(H + L) polyclonal antibody coupled to peroxidase (NA931V, Amersham) (A) or with the aid of an anti-S rabbit polyclonal antibody and an anti-rabbit IgG(H + L) polyclonal antibody coupled to peroxidase (NA934V, Amersham) (B).

A molecular mass ladder (kDa) is presented in the figure.

FIG. 37 shows the analysis of the Ssol polypeptide, purified on SDS polyacrylamide gel 10, 5 and 2 μ l of recombinant Ssol polypeptide purified by anti-FLAG affinity chromatography were separated on 4 to 15% gradient SDS polyacrylamide gel. The Ssol polypeptide and variable quantities of molecular mass markers (MM) were visualized by staining with silver nitrate (Gelcode SilverSNAP stain kit II, Pierce).

FIG. 38 illustrates the immunoreactivity of the recombinant Ssol polypeptide produced by the recombinant vaccinia virus VV-TN-Ssol toward sera of patients suffering from SARS. The reactivity of sera from patients was analyzed by indirect ELISA test against solid phases prepared with the aid of the purified recombinant Ssol polypeptide. The antibodies from patients reacting with the solid phase at a dilution of 1/100 and 1/400 are visualized with a human anti-IgG(H + L) polyclonal antibody coupled to peroxidase (Amersham NA933V) and TMB plus H202 (KPL). The sera of probable SARS cases are identified by a National Reference Center for Influenza Virus serial number and by the initials of the patient and the number of days elapsed since the onset of symptoms, where appropriate. The TV sera are control sera from subjects which were collected in France before the SARS epidemic which occurred in 2003.

FIG. 39 shows the anti-SARS-CoV antibody response in mice after immunization with the recombinant vaccinia viruses. Groups of 7 BALB/c mice were immunized by the i.v. route twice at 4 weeks' interval with 106 pfu of recombinant vaccinia viruses VV-TG, VV-TG-HA, VV-TG-S, VV-TG-Ssol, VV-TN, VV-TN-S, VV-TN-Ssol.

A. Pools of immune sera collected 3 weeks after each of the two immunizations were prepared for each of the groups and were analyzed by indirect ELISA using a lysate of VeroE6 cells infected with SARS-CoV as antigen. The anti-SARS-CoV antibody titers are calculated as the reciprocal of the dilution producing a specific OD of 0.5 after visualization with an anti-mouse IgG polyclonal antibody coupled to peroxidase (NA931V, Amersham) and TMB (KPL).

TABLE I-continued

Sequence listing		Position of the cDNA with reference to Genbank	Deposit number at the CNCM of the cor- responding plasmid
Identification number	Sequence	AY274119.3	

5 B. The pools of immune sera were evaluated for their capacity to seroneutralize the infectivity of 100 TCID₅₀ of SARS-CoV on FRhK-4 cells. 4 points are produced for each of the 2-fold dilutions tested from 1/20. The seroneutralizing titer is calculated according to the Reed and Munch method as the reciprocal of the dilution neutralizing the infectivity of 2 wells out of 4.

FIG. 40 describes the construction of the recombinant viruses MVSchw2-SARS-S and MVSchw2-SARS-Ssol.

A. The measles vector is a complete genome of the Schwarz vaccine strain of the measles virus (MV) into which an additional transcription unit has been introduced (Combedet, 2003, Journal of Virology, 77: 11546-11554). The expression of the additional open reading frames (ORF) is controlled by cis-acting elements necessary for the transcription, for the formation of the cap and for the polyadenylation of the transgene which were copied from the elements present at the N/P junction. 2 different vectors allow the insertion between the P (phosphoprotein) and M (matrix) genes on the one hand and the H (hemagglutinin) and L (polymerase) genes on the other hand.

B. The recombinant genomes MVSchw2-SARS-S and MVSchw2-SARS-Ssol of the measles virus were constructed by inserting the ORFs of the S protein and of the Ssol polypeptide into an additional transcription unit located between the P and M genes of the vector.

The various genes of the measles virus (MV) are indicated: N (nucleoprotein), PVC (V/C phosphoprotein and protein), M (matrix), F (fusion), H (hemagglutinin), L (polymerase), T7 = T7 RNA polymerase promoter, hh = hammerhead ribozyme, T7t = T7 phage RNA polymerase terminator sequence, δ = ribozyme of the hepatitis δ virus, (2), (3) = additional transcription units (ATU).

Size of the MV genome: 15 894 nt.

25 SP: signal peptide

TM: transmembrane region

FLAG: FLAG tag

FIG. 41 illustrates the expression of the S protein by the recombinant measles viruses, analyzed by Western blotting.

Cytoplasmic extracts were prepared after infection of Vero cells by different passages of the viruses MVSchw2-SARS-S and MVSchw2-SARS-Ssol and the wild-type virus MWSchw as control. Cellular extracts in loading buffer according to Laemmli were also prepared 8 hours after infection of VeroE6 cells with SARS-CoV at a multiplicity of infection of 3. They were separated on 8% SDS acrylamide gel and analyzed by Western blotting with the aid of an anti-S rabbit polyclonal antibody and an anti-rabbit IgG(H + L) polyclonal antibody coupled to peroxidase (NA934V, Amersham).

A molecular mass ladder (kDa) is presented in the figure.

Pn: nth passage of the virus after coculture of 293-3-46 and Vero cells

SARS-CoV: extract of VeroE6 cells infected with SARS-CoV

Mock: control extract of noninfected VeroE6 cells

FIG. 42 shows the expression of the S protein by the recombinant measles viruses, analyzed by immunofluorescence

Vero cells in monolayers on glass slides were infected with the wild-type virus MWSchw (A) or the viruses MVSchw2-SARS-S (B) and MVSchw2-SARS-Ssol (C). When the syncytia have reached 30 to 40% confluence (A, B.) or 90-100% (C), the cells were fixed, permeabilized and labeled with anti-SARS-CoV rabbit polyclonal antibodies and an anti-rabbit IgG(H + L) conjugate coupled to FITC (Jackson).

FIG. 43 illustrates the Western-blot analysis of the immunoreactivity of rabbit sera directed against the peptides E1-12, E53-76 and M2-14. The rabbit 20047 was immunized with the peptide E1-12 coupled to KLH. The rabbits 22234 and 22240 were immunized with the peptide E53-76 coupled to KLH. The rabbits 20013 and 20080 were immunized with the peptide M2-14 coupled to KLH. The immune sera were analyzed by Western blotting with the aid of extracts of cells infected with SARS-CoV (B) or with the aid of extracts of cells infected with a recombinant vaccinia virus expressing the protein E (A) or M (C) of the SARS-CoV 031589 isolate. The immunoblots were visualized with the aid of an anti-rabbit IgG(H + L) conjugate coupled to peroxidase (NA934V, Amersham).

The position of the E and M proteins is indicated by an arrow.

A molecular mass ladder (kDa) is presented in the figure.

It should be understood, however, that these examples are given solely by way of illustration of the subject of the invention, and do not constitute in any manner a limitation thereto.

EXAMPLE 1

Cloning and Sequencing of the Genome of the SARS-CoV Strain Derived from the Sample Recorded Under the Number 031589

The RNA of the SARS-CoV strain was extracted from the sample of bronchoalveolar washing recorded under the number 031589, performed on a patient at the Hanoi (Vietnam) French hospital suffering from SARS.

The isolated RNA was used as template to amplify the cDNAs corresponding to the various open reading frames of the genome (ORF1a, ORF1b, ORF-S, ORF-E, ORF-M, ORF-N (including ORF-13 and ORF-14), ORF3, ORF4, ORF7 to ORF11), and at the noncoding 5' and 3' ends. The sequences of the primers and of the probes used for the amplification/detection were defined based on the available SARS-CoV nucleotide sequence.

In the text which follows, the primers and the probes are identified by: the letter S, followed by a letter which indicates the corresponding region of the genome (L for the 5' end including ORF1a and ORF1b; S, N and N for ORF-S, ORF-M, ORF-N, SE and MN for the corresponding intergene regions), and then optionally by Fn, Rn, with n between 1 and 6 corresponding to the primers used for the nested PCR (F1+R1 pair for the first amplification, F2+R2 pair for the second amplification, and the like), and then by +/- or -/- corresponding to a sense or antisense primer and finally by the positions of the primers with reference to the Genbank sequence AY27411.3; for the sense and antisense S and N primers and the other sense primers only, when a single position is indicated, it corresponds to that of the 5' end of a probe or of a primer of about 20 bases; for the antisense primers other than the S and N primers, when a single position is indicated, it corresponds to that of the 3' end of a probe or of a primer of about 20 bases.

The amplification products thus generated were sequenced with the aid of specific primers in order to determine the complete sequence of the genome of the SARS-CoV strain derived from the sample recorded under the number 031589. These amplification products, with the exception of those corresponding to ORF1a and ORF1b, were then cloned into expression vectors in order to produce the corresponding viral proteins and the antibodies directed against these proteins, in particular by DNA-based immunization.

1. Extraction of the RNAs

The RNAs were extracted with the aid of the QIamp viral RNA extraction mini kit (QIAGEN) according to the manufacturer's recommendations. More specifically: 140 µl of the sample and 560 µl of AVL buffer were vigorously mixed for 15 seconds, incubated for 10 minutes at room temperature and then briefly centrifuged at maximum speed. 560 µl of 100% ethanol were added to the supernatant and the mixture thus obtained was very vigorously stirred for 15 sec. 630 µl of the mixture were then deposited on the column.

The column was placed on a 2 ml tube, centrifuged for 1 min at 8000 rpm, and then the remainder of the preceding mixture was deposited on the same column, centrifuged again, for 1 min at 8000 rpm, and the column was transferred over a clean 2 ml tube. Next, 500 µl of AW1 buffer were added to the column, and then the column was centrifuged for 1 min at 8000 rpm and the eluate was discarded. 500 µl of AW2 buffer were added to the column which was then centrifuged for 3 min at 14 000 rpm and transferred onto a 1.5 ml tube. Finally, 60 µl of AVE buffer were added to the column which was incubated for 1 to 2 min at room temperature and then centrifuged for 1 min at 8000 rpm. The eluate corresponding to the purified RNA was recovered and frozen at -20° C.

2. Amplification, Sequencing and Cloning of the cDNAs

2.1) cDNA Encoding the S Protein

The RNAs extracted from the sample were subjected to reverse transcription with the aid of random sequence hexameric oligonucleotides (pdN6), so as to produce cDNA fragments.

The sequence encoding the SARS-CoV S glycoprotein was amplified in the form of two overlapping DNA fragments: 5' fragment (SARS-Sa, SEQ ID NO: 5) and 3' fragment (SARS-Sb, SEQ ID NO: 6), by carrying out two successive amplifications with the aid of nested primers. The amplicons thus obtained were sequenced, cloned into the PCR plasmid vector 2.1-TOPO™ (INVITROGEN), and then the sequence of the cloned cDNAs was determined.

a) Cloning and Sequencing of the Sa and Sb Fragments

a.1) Synthesis of the cDNA

The reaction mixture containing: RNA (5 µl), H₂O for injection (3.5 µl), 5× reverse transcriptase buffer (4 µl), 5 mM dNTP (2 µl), pdN6 100 µg/ml (4 µl), RNasin 40 IU/µl (0.5 µl) and reverse transcriptase AMV-RT, 10 IU/µl, PROMEGA (1 µl) was incubated in a thermocycler under the following conditions: 45 min at 42° C., 15 min at 55° C., 5 min at 95° C., and then the cDNA obtained was kept at +4° C.

a.2) First PCR Amplification

The 5' and 3' ends of the S gene were respectively amplified with the pairs of primers S/F1+/21350-21372 and S/R1/-/23518-23498, S/F3+/23258-23277 and S/R3/-/25382-25363. The 50 µl reaction mixture containing: cDNA (2 µl), 50 µM primers (0.5 µl), 10× buffer (5 µl), 5 mM dNTP (2 µl), Taq Expand High Fidelity, Roche (0.75 µl) and H₂O (39, 75 µl) was amplified in a thermocycler, under the following conditions: an initial step of denaturation at 94° C. for 2 min was followed by 40 cycles comprising: a step of denaturation at 94° C. for 30 sec, a step of annealing at 55° C. for 30 sec and then a step of extension at 72° C. for 2 min 30 sec, with 10 sec of additional extension at each cycle, and then a final step of extension at 72° C. for 5 min.

a.3) Second PCR Amplification

The products of the first PCR amplification (5' and 3' amplicons) were subjected to a second PCR amplification step (nested PCR) under conditions identical to those of the first amplification, with the pairs of primers S/F2+/21406-21426 and S/R2/-/23454-23435 and S/F4+/23322-23341 and S/R4/-/25348-25329, respectively for the 5' amplicon and the 3' amplicon.

a.4) Cloning and Sequencing of the Sa and Sb Fragments

The Sa (5' end) and Sb (3' end) amplicons thus obtained were purified with the aid of the QIAquick PCR purification kit (QIAGEN), following the manufacturer's instructions, and then they were cloned into the vector PCR2.1-TOPO (Invitrogen kit), to give the plasmids called SARS-S1 and SARS-S2.

The DNA of the Sa and Sb clones was isolated and then the corresponding insert was sequenced with the aid of the Big Dye kit, Applied Biosystem® and universal primers M13 forward and M13 reverse, and primers: S/S+/21867, S/S+/22353, S/S+/22811, S/S+/23754, S/S+/24207, S/S+/24699, S/S+/24348, S/S-/24209, S/S-/23630, S/S-/23038, S/S-/22454, S/S-/21815, S/S-/24784, S/S+/21556, S/S+/23130 and S/S+/24465 following the manufacturer's instructions; the sequences of the Sa and Sb fragments thus obtained correspond to the sequences SEQ ID NO: 5 and SEQ ID NO: 6 in the sequence listing appended as an annex.

The plasmid, called SARS-S1, was deposited under the No. I-3020, on May 12, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains a 5' fragment of the sequence of the S gene of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above, said fragment called Sa corresponding to the nucleotides at positions 21406 to 23454 (SEQ ID NO: 5), with reference to the Genbank sequence AY274119.3 Tor2.

The plasmid, called TOP10F⁺-SARS-S2, was deposited under the No. I-3019, on May 12, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains a 3' fragment of the sequence of the S gene of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above, said fragment called Sb corresponding to the nucleotides at positions 23322 to 25348 (SEQ ID NO: 6), with reference to the Genbank sequence accession No. AY274119.3.

b) Cloning and Sequencing of the Complete cDNA (SARS-S Clone of 4 kb)

The complete S cDNA was obtained from the abovementioned clones SARS-S1 and SARS-S2, in the following manner:

1) A PCR amplification reaction was carried out on a SARS-S2 clone in the presence of the abovementioned primer S/R4/-/25348-25329 and of the primer S/S/+24696-24715: an amplicon of 633 bp was obtained,

2) Another PCR amplification reaction was carried out on another SARS-S2 clone, in the presence of the primers S/F4/+23322-23341 mentioned above and S/S/-/24803-24784: an amplicon of 1481 bp was obtained.

The amplification reaction was carried out under the conditions as defined above for the amplification of the Sa and Sb fragments, with the exception that 30 amplification cycles comprising a step of denaturation at 94° C. for 20 sec and a step of extension at 72° C. for 2 min 30 sec were carried out.

3) The 2 amplicons (633 bp and 1481 bp) were purified under the conditions as defined above for the Sa and Sb fragments.

4) Another PCR amplification reaction with the aid of the abovementioned primers S/F4/+23322-23341 and S/R4/-/25348-25329 was carried out on the purified amplicons obtained in 3). The amplification reaction was carried out under the conditions as defined above for the amplification of the Sa and Sb fragments, except that 30 amplification cycles were performed.

The 2026 bp amplicon thus obtained was purified, cloned into the vector PCR2.1-TOPO and then sequenced as above, with the aid of the primers as defined above for the Sa and Sb fragments. The clone thus obtained was called clone 3'.

5) The clone SARS-S1 obtained above and the clone 3' were digested with EcoR I, the bands of about 2 kb thus obtained were gel purified and then amplified by PCR with the abovementioned primers S/F2/+21406-21426 and S/R4/-/25348-25329. The amplification reaction was carried out under the conditions as defined above for the amplification of the Sa and Sb fragments, except that 30 amplification cycles were performed. The amplicon of about 4 kb was purified and sequenced. It was then cloned into the vector PCR2.1-TOPO in order to give the plasmid, called SARS-S, and the insert obtained in this plasmid was sequenced as above, with the aid of the primers as defined above for the Sa and Sb fragments. The cDNA sequences of the insert and of the amplicon encoding the S protein correspond respectively to the sequences SEQ ID NO: 4 and SEQ ID NO: 2 in the sequence listing appended as an annex, they encode the S protein (SEQ ID NO: 3).

The sequence of the amplicon corresponding to the cDNA encoding the S protein of the SARS-CoV strain derived from the sample No. 031589 has the following two mutations compared with the corresponding sequences of respectively the Tor2 and Urbani isolates, the positions of the mutations being indicated with reference to the complete sequence of the genome of the Tor2 isolate (Genbank AY274119.3):

g/t in position 23220; the alanine codon (gct) in position 577 of the amino acid sequence of the S protein of Tor2 is replaced with a serine codon (tct),

c/t in position 24872: this mutation does not modify the amino acid sequence of the S protein, and

the plasmid, called SARS-S, was deposited under the No. I-3059, on Jun. 20, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA sequence encoding the S protein of the SARS-CoV strain derived from the sample recorded under the No. 031589, said sequence corre-

sponding to the nucleotides at positions 21406 to 25348 (SEQ ID NO: 4), with reference to the Genbank sequence AY274119.3.

2.2) cDNA Encoding the M and E Proteins

The RNAs derived from the sample 031589, extracted as above, were subjected to a reverse transcription, combined, during the same step (Titan One Step RT-PCR® kit, Roche), with a PCR amplification reaction, with the aid of the pairs of primers:

S/E/F1/+26051-26070 and S/E/R1/-/26455-26436 in order to amplify ORF-E, and

S/M/F1/+26225-26244 and S/M/R1/-/27148-27129 in order to amplify ORF-M.

A first reaction mixture containing: 8.6 µl of H₂O for injection, 1 µl of dNTP (5 mM), 0.2 µl of each of the primers (50 µM), 1.25 µl of DTT (100 mM) and 0.25 µl of RNasin (40 IU/µl) was combined with a second reaction mixture containing: 1 µl of RNA, 7 µl of H₂O for injection, 5 µl of 5×RT-PCR buffer and 0.5 µl of enzyme mixture and the combined mixtures were incubated in a thermocycler under the following conditions: 30 min at 42° C., 10 min at 55° C., 2 min at 94° C. followed by 40 cycles comprising a step of denaturation at 94° C. for 10 sec, a step of annealing at 55° C. for 30 sec and a step of extension at 68° C. for 45 sec, with 3 sec increment per cycle and finally a step of terminal extension at 68° C. for 7 min.

The amplification products thus obtained (M and E amplicons) were subjected to a second PCR amplification (nested PCR) using the Expand High-Fi® kit, Roche), with the aid of the pairs of primers:

S/E/F2/+26082-26101 and S/E/R2/-/26413-26394 for the amplicon E, and

S/M/F2/+26330-26350 and S/M/R2/-/27098-27078 for the amplicon M.

The reaction mixture containing: 2 µl of the product of the first PCR, 39.25 µl of H₂O for injection, 5 µl of 10× buffer containing MgCl₂, 2 µl of dNTP (5 mM), 0.5 µl of each of the primers (50 µM) and 0.75 µl of enzyme mixture was incubated in a thermocycler under the following conditions: a step of denaturation at 94° C. for 2 min was followed by 30 cycles comprising a step of denaturation at 94° C. for 15 sec, a step of annealing at 60° C. for 30 sec and a step of extension at 72° C. for 45 sec, with 3 sec increment per cycle, and finally a step of terminal extension at 72° C. for 7 min. The amplification products obtained corresponding to the cDNAs encoding the E and M proteins were sequenced as above, with the aid of the primers: S/E/F2/+26082 and S/E/R2/-/126394, S/M/F2/+26330, S/M/R2/-/27078 cited above and the primers S/M/+26636-26655 and S/M/-/26567-26548. They were then cloned, as above, in order to give the plasmids called SARS-E and SARS-M. The DNA of these clones was then isolated and sequenced with the aid of the universal primers M13 forward and M13 reverse and the primers S/M/+26636 and S/M/-/26548 mentioned above.

The sequence of the amplicon representing the cDNA encoding the E protein (SEQ ID NO: 13) of the SARS-CoV strain derived from the sample No. 031589 does not contain differences in relation to the corresponding sequences of the isolates AY274119.3-Tor2 and AY278741-Urbani. The sequence of the E protein of the SARS-CoV 031589 strain corresponds to the sequence SEQ ID NO: 14 in the sequence listing appended as an annex.

The plasmid, called SARS-E, was deposited under the No. I-3046, on May 28, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA sequence encoding the E protein of the SARS-CoV strain derived from the

sample recorded under the No. 031589, as defined above, said sequence corresponding to the nucleotides at positions 26082 to 26413 (SEQ ID NO: 15), with reference to the Genbank sequence accession No. AY274119.3.

The sequence of the amplicon representing the cDNA encoding M (SEQ ID NO: 16) from the SARS-CoV strain derived from the sample No. 031589 does not contain differences in relation to the corresponding sequence of the isolate AY274119.3-Tor2. By contrast, at position 26857, the isolate AY278741-Urbani contains a c and the sequence of the SARS-CoV strain derived from the sample recorded under the No. 031589 contains a t. This mutation results in a modification of the amino acid sequence of the corresponding protein: at position 154, a proline (AY278741-Urbani) is changed to serine in the SARS-CoV strain derived from the sample recorded under the No. 031589. The sequence of the M protein of the SARS-CoV strain derived from the sample recorded under the No. 031589 corresponds to the sequence SEQ ID NO: 17 in the sequence listing appended as an annex.

The plasmid, called SARS-M, was deposited under the No. I-3047, on May 28, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA sequence encoding the M protein of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above; said sequence corresponding to the nucleotides at positions 26330 to 27098 (SEQ ID NO: 18), with reference to the Genbank sequence accession No. AY274119.3.

2.3) cDNA Corresponding to ORF3, ORF4, ORF7 to ORF11

The same amplification, cloning and sequencing strategy was used to obtain the cDNA fragments corresponding respectively to the following ORFs: ORF3, ORF4, ORF7, ORF8, ORF9, ORF10 and ORF11. The pairs of primers used for the first amplification are:

ORF3 and ORF4: S/SE/F1/+25069-25088 and S/SE/R1/-26300-26281

ORF7 to ORF11: S/MN/F1/+26898-26917 and S/MN/R1/-28287-28266

The pairs of primers used for the second amplification are: ORF3 and ORF4: S/SE/F2/+25110-25129 and S/SE/R2/-26244-26225

ORF7 to ORF11: S/NN/F2/+26977-26996 and S/MN/R2/-28218-28199

The conditions for the first amplification (RT-PCR) are the following: 45 min at 42° C., 10 min at 55° C., 2 min at 94° C. followed by 40 cycles comprising a step of denaturation at 94° C. for 15 sec, a step of annealing at 58° C. for 30 sec and a step of extension at 68° C. for 1 min, with 5 sec increment per cycle and finally a step of terminal extension at 68° C. for 7 min.

The conditions for the nested PCR are the following: a step of denaturation at 94° C. for 2 min was followed by 40 cycles comprising a step of denaturation at 94° C. for 20 sec, a step of annealing at 58° C. for 30 sec and a step of extension at 72° C. for 50 sec, with 4 sec increment per cycle and finally a step of terminal extension at 72° C. for 7 min.

The amplification products obtained corresponding to the cDNAs containing respectively ORF3 and 4 and ORF7 to 11 were sequenced with the aid of the primers: S/SE/+25363, S/SE/+25835, S/SE/-25494, S/SE/-25875, S/MN/+27839, S/MN/+27409, S/MN/-27836, S/MN/-27799 and cloned as above for the other ORFs, to give the plasmids called SARS-SE and SARS-MN. The DNA of these clones was isolated and sequenced with the aid of these same primers and of the universal primers M13 sense and M13 antisense.

The sequence of the amplicon representing the cDNA of the region containing ORF3 and ORF4 (SEQ ID NO: 7) of the SARS-CoV strain derived from the sample No. 031589 contains a nucleotide difference in relation to the corresponding sequence of the isolate AY274119-Tor2. This mutation at position 25298 results in a modification of the amino acid sequence of the corresponding protein (ORF3): at position 11, an arginine (AY274119-Tor2) is changed to glycine in the SARS-CoV strain derived from the sample No. 031589. By contrast, no mutation was identified in relation to the corresponding sequence of the isolate AY278741-Urbani. The sequences of ORF3 and 4 of the SARS-CoV strain derived from the sample No. 031589 correspond respectively to the sequences SEQ ID NO: 10 and 12 in the sequence listing appended as an annex.

The plasmid, called SARS-SE, was deposited under the No. I-3126, on Nov. 13, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA corresponding to the region situated between ORF-S and ORF-E and overlapping ORF-E of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above, said region corresponding to the nucleotides at positions 25110 to 26244 (SEQ ID NO: 8), with reference to the Genbank sequence accession No. AY274119.3.

The sequence of the amplicon representing the cDNA corresponding to the region containing ORF7 to ORF11 (SEQ ID NO: 19) of the SARS-CoV strain derived from the sample No. 031589 does not contain differences in relation to the corresponding sequences of the isolates AY274119-Tor2 and AY278741-Urbani. The sequences of ORF7 to 11 of the SARS-CoV strain derived from the sample No. 031589 correspond respectively to the sequences SEQ ID NO: 22, 24, 26, 28 and 30 in the sequence listing appended as an annex.

The plasmid, called SARS-MN, was deposited under the No. I-3125, on Nov. 13, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA sequence corresponding to the region situated between ORF-M and ORF-N of the SARS-CoV strain derived from the sample recorded under the No. 031589 and collected in Hanoi, as defined above, said sequence corresponding to the nucleotides at positions 26977 to 28218 (SEQ ID NO: 20), with reference to the Genbank sequence accession No. AY274119.3.

The sequence of the amplicon representing the cDNA corresponding to the region containing ORF7 to ORF11 (SEQ ID NO: 19) of the SARS-CoV strain derived from the sample No. 031589 does not contain differences in relation to the corresponding sequences of the isolates AY274119-Tor2 and AY278741-Urbani. The sequences of ORF7 to 11 of the SARS-CoV strain derived from the sample No. 031589 correspond respectively to the sequences SEQ ID NO: 22, 24, 26, 28 and 30 in the sequence listing appended as an annex.

2.4) cDNA Encoding the N Protein and Including ORF13 and ORF14

The cDNA was synthesized and amplified as described above for the fragments Sa and Sb. More specifically, the reaction mixture containing: 5 µl of RNA, 5 µl of H₂O for injection, 4 µl of 5× reverse transcriptase buffer, 2 µl of dNTP (5 mM), 2 µl of oligo 20 T (5 µM), 0.5 µl of RNasin (40 IU/µl) and 1.5 µl of AMV-RT (10 IU/µl Promega) was incubated in a thermocycler under the following conditions: 45 min at 42° C., 15 min at 55° C., 5 min at 95° C., and it was then kept at 44° C.

A first PCR amplification was performed with the pair of primers S/N/F3/+28023 and S/N/R3/-29460.

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The reaction mixture as above for the amplification of the S1 and S2 fragments was incubated in a thermo-cycler, under the following conditions: an initial step of denaturation at 94° C. for 2 min was followed by 40 cycles comprising a step of denaturation at 94° C. for 20 sec, a step of annealing at 55° C. for 30 sec and then a step of extension at 72° C. for 1 min 30 sec with 10 sec of additional extension at each cycle, and then a final step of extension at 72° C. for 5 min.

The amplicon obtained at the first PCR amplification was subjected to a second PCR amplification step (nested PCR) with the pairs of primer S/N/F4/+28054 and S/N/R4/-29430 under conditions identical to those of the first amplification.

The amplification product obtained, corresponding to the cDNA encoding the N protein of the SARS-CoV strain derived from the sample No. 031589, was sequenced with the aid of the primers: S/N/F4/+28054, S/N/R4/-29430, S/N/+28468, S/N/+28918 and S/N/-28607 and cloned as above for the other ORFs, to give the plasmid called SARS-N. The DNA of these clones was isolated and sequenced with the aid of the universal primers M13 sense and M13 antisense, and the primers S/N/+28468, S/N/+28918 and S/N/-28607.

The sequence of the amplicon representing the cDNA corresponding to ORF-N and including ORF13 and ORF14 (SEQ ID NO: 36) of the SARS-CoV strain derived from the sample No. 031589 does not contain differences in relation to the corresponding sequences of the isolates AY274119.3-Tor2 and AY278741-Urbani. The sequence of the N protein of the SARS-CoV strain derived from the sample No. 031589 corresponds to the sequence SEQ ID NO: 37 in the sequence listing appended as an annex.

The sequences of ORF13 and 14 of the SARS-CoV strain derived from the sample No. 031589 correspond respectively to the sequences SEQ ID NO: 32 and 34 in the sequence listing appended as an annex.

The plasmid, called SARS-N, was deposited under the No. I-3048, on Jun. 5, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA encoding the N protein of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above, said sequence corresponding to the nucleotides at positions 28054 to 29430 (SEQ ID NO: 38), with reference to the Genbank sequence accession No. AY274119.3.

2.5) Noncoding 5' and 3' Ends

a) Noncoding end (5'NC)

a₁) Synthesis of the cDNA

The RNAs derived from the sample 031589, extracted as above, were subjected to reverse transcription under the following conditions:

The RNA (15 µl) and the primer S/L/-443 (3 µl at the concentration of 5 µM) were incubated for 10 min at 75° C.

Next, the 5× reverse transcriptase buffer (6 µl, INVITROGEN), 10 mM dNTP (1 µl), 0.1 M DTT (3 µl) were added and the mixture was incubated at, 50° C. for 3 min.

Finally, the reverse transcriptase (3 µl of Superscript®, INVITROGEN) was added to the preceding mixture which was incubated at 50° C. for 1 h 30 min and then at 90° C. for 2 min.

The cDNA thus obtained was purified with the aid of the QIAquick PCR purification kit (QIAGEN), according to the manufacturer's recommendations.

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b₁) Terminal Transferase Reaction (TdT)

The cDNA (10 µl) is incubated for 2 min at 100° C., stored in ice, and the following are then added: H₂O (2.5 µl), 5× TdT buffer (4 µl, AMERSHAM), 5 mM dATP (2 µl) and TdT (1.5 µl, AMERSHAM). The mixture thus obtained is incubated for 45 min at 37° C. and then for 2 min at 65° C.

The product obtained is amplified by a first PCR reaction with the aid of the primers: S/L/-225-206 and anchor 14T: 5'-AGATGAATTCGGTACCTTTTTTTTTTTTTTTT-3' (SEQ ID NO: 68). The amplification conditions are the following: an initial step of denaturation at 94° C. for 2 min is followed by 10 cycles comprising a step of denaturation at 94° C. for 10 sec, a step of annealing at 45° C. for 30 sec and then a step of extension at 72° C. for 30 sec and then by 30 cycles comprising a step of denaturation at 94° C. for 10 sec, a step of annealing at 50° C. for 30 sec and then a step of extension at 72° C. for 30 sec, and then a final step of extension at 72° C. for 5 min.

The product of the first PCR amplification was subjected to a second amplification step with the aid of the primers: S/L/-204-185 and anchor 14 T mentioned above under conditions identical to those of the first amplification. The amplicon thus obtained was purified, sequenced with the aid of the primer S/L/-182-163 and it was then cloned as above for the different ORFs, to give the plasmid called SARS-5'NC. The DNA of this clone was isolated and sequenced with the aid of the universal primers M13 sense and M13 antisense and the primer S/L/-182-163 mentioned above.

The amplicon representing the cDNA corresponding to the 5'NC end of the SARS-CoV strain derived from the sample recorded under the No. 031589 corresponds to the sequence SEQ ID NO: 72 in the sequence listing appended as an annex; this sequence does not contain differences in relation to the corresponding sequences of the isolates AY274119.3-Tor2 and AY278741-Urbani.

The plasmid, called SARS-5'NC, was deposited under the No. I-3124, on Nov. 7, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA corresponding to the noncoding 5' end of the genome of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above, said sequence corresponding to the nucleotides at positions 1 to 204 (SEQ ID NO: 39), with reference to the Genbank sequence accession No. AY274119.3.

b) Noncoding 3' End (3'NC)

a₁) Synthesis of the cDNA

The RNAs derived from the sample 031589, extracted as above, were subjected to reverse transcription, according to the following protocol: the reaction mixture containing: RNA (5 µl), H₂O (5 µl), 5× reverse transcriptase buffer (4 µl), 5 mM dNTP (2 µl), 5 µM Oligo 20 T (2 µl), 40 U/µl RNasin (0.5 µl) and 10 IU/µl RT-AMV (1.5 µl, PROMEGA) was incubated in a thermo-cycler, under the following conditions: 45 min at 42° C., 15 min at 55° C., 5 min at 95° C., and it was then kept at +4° C.

The cDNA obtained was amplified by a first PCR reaction with the aid of the primers S/N/+28468-28487 and anchor 14 T mentioned above. The amplification conditions are the following: an initial step of denaturation at 94° C. for 2 min is followed by 10 cycles comprising a step of denaturation at 94° C. for 20 sec, a step of annealing at 45° C. for 30 sec and then a step of extension at 72° C. for 50 sec and then 30 cycles comprising a step of denaturation at 94° C. for 20 sec, a step of annealing at 50° C. for 30 sec and then a step of extension at 72° C. for 50 sec, and then a final step of extension at 72° C. for 5 min.

The product of the first PCR amplification was subjected to a second amplification step with the aid of the primers S/N/+28933-28952 and anchor 14 T mentioned above, under conditions identical to those of the first amplification. The amplicon thus obtained was purified, sequenced with the aid of the primer S/N/+29257-29278 and cloned as above for the different ORFs, to give the plasmid called SARS-3'NC. The DNA of this clone was isolated and sequenced with the aid of the universal primers M13 sense and M13 antisense and the primer S/N/+29257-29278 mentioned above.

031589 was performed by carrying out RT-PCR reactions followed by nested PCRs according to the same principles as those described above for the other ORFs. The amplified fragments overlap over several tenths of bases, thus allowing computer reconstruction of the complete sequence of this part of the genome. On average, the amplified fragments are of two kilobases.

14 overlapping fragments, called L0 to L12, were thus amplified with the aid of the following primers:

TABLE II

Primers used for the amplification of the 5' region (ORF1a and ORF1b)				
REGION AMPLIFIED AND SEQUENCED (does not include the primers)	RT-PCR sense primer	RT-PCR antisense primer	Nested PCR sense primer	Nested PCR antisense primer
L0	S/L0/F1/+30	S/L0/R1/-481		
50-480				
L1	S/L1/F1/+147	S/L1/R1/-2336	S/L1/F2/+211	S/L1/R2/-2241
231-2240				
L2	S/L2/F1/+2033	S/L2/R1/-4192	S/L2/F2/+2136	S/L2/R2/-4168
2156-4167				
L3	S/L3bis/F1/+3850	S/L3bis/R1/-5365	S/L3bis/F2/+3892	S/L3bis/R2/-5325
3913-5324				
L4b	S/L4b/F1/+4878	S/L4b/R1/-6061	S/L4b/F2/+4932	S/L4b/R2/-6024
4952-6023				
L4	S/L4/F1/+5272	S/L4/R1/-7392	S/L4/F2/+5305	S/L4/R2/-7323
5325-7318				
L5	S/L5/F1/+7111	S/L5/R1/-9253	S/L5/F2/+7275	S/L5/R2/-9157
7296-9156				
L6	S/L6/F1/+8975	S/L6/R1/-11151	S/L6/F2/+9032	S/L6/R2/-11067
9053-11066				
L7	S/L7/F1/+10883	S/L7/R1/-13050	S/L7/F2/+10928	S/L7/R2/-12963
10928-12962				
L8	S/L8/F1/+12690	S/L8/R1/-14857	S/L8/F2/+12815	S/L8/R2/-14835
12835-14834				
L9	S/L9/F1/+14688	S/L9/R1/-16678	S/L9/F2/+14745	S/L9/R2/-16625
14765-16624				
L10	S/L10/F1/+16451	S/L10/R1/-18594	S/L10/F2/+16514	S/L10/R2/-18571
16534-18570				
L11	S/L11/F1/+18441	S/L11/R1/-20612	S/L11/F2/+18500	S/L11/R2/-20583
18521-20582				
L12	S/L12/F1/+20279	S/L12/R1/-22229	S/L12/F2/+20319	S/L12/R2/-22206
20338-22205				

The amplicon representing the cDNA corresponding to the 3'NC end of the SARS-CoV strain derived from the sample recorded under the No. 031589 corresponds to the sequence SEQ ID NO: 73 in the sequence listing appended as an annex; this sequence does not contain differences in relation to the corresponding sequences of the isolates AY274119.3-Tor2 and AY278741-Urbani.

The plasmid called SARS-3'NC was deposited under the No. I-3123 on Nov. 7, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA sequence corresponding to the noncoding 3' end of the genome of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above, said sequence corresponding to that situated between the nucleotide at positions 28933 to 29272 (SEQ ID NO: 40), with reference to the Genbank sequence accession No. AY274119.3, ends with a series of nucleotides a.

2.6) ORF1a and ORF1b

The amplification of the 5' region containing ORF1a and ORF1b of the SARS-CoV genome derived from the sample

All the fragments were amplified under the following conditions, except fragment L0 which was amplified as described above for ORF-M:

RT-PCR: 30 min at 42° C., 15 min at 55° C., 2 min at 94° C., and then the cDNA obtained is amplified under the following conditions: 40 cycles comprising: a step of denaturation at 94° C. for 15 sec, a step of annealing at 58° C. for 30 sec and then a step of extension at 68° C. for 1 min 30 sec, with 5 sec additional extension at each cycle, and then a final step of extension at 68° C. for 7 min.

Nested PCR: An initial step of denaturation at 94° C. for 2 min is followed by 35 cycles comprising: a step of denaturation at 94° C. for 15 sec, a step of annealing at 60° C. for 30 sec and then a step of extension at 72° C. for 1 min 30 sec, with 5 sec of additional extension at each cycle, and then a final step of extension at 72° C. for 7 min.

The amplification products were sequenced with the aid of the primers defined in table III below:

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TABLE III

Primers used for the sequencing of the 5' region (ORF1a and ORF1b)	
Names	Sequences (SEQ ID NO: 76 to 139)
S/L3/+4932	5'-CCACACACAGCTTGTGGATA-3'
S/L4/+6401	5'-CCGAAGTTGTAGGCAATGTC-3'
S/L4/+6964	5'-TTTGGTGCTCCTTCTTATTG -3'
S/L4/-6817	5'-CCGGCATCCAAACATAATTT-3'
S/L5/-7633	5'-TGCTCAGTAGGGTTGATTGG-3'
S/L5/-8127	5'-CATCCTTTGTGTCAACATCG-3'
S/L5/-8633	5'-GTCACGAGTGACACCATCCT-3'
S/L5/+7839	5'-ATGCGACGAGTCTGCTTCTA-3'
S/L5/+8785	5'-TTCATAGTGCTGGCTTACC-3'
S/L5/+8255	5'-ATCTTGGCGCATGTATTGAC-3'
S/L6/-9422	5'-TGCATTAGCAGCAACAACAT-3'
S/L6/-9966	5'-TCTGCAGAACAGCAGAAGTG-3'
S/L6/-10542	5'-CCTGTGCAGTTTGTCTGTCA-3'
S/L6/+10677	5'-CCTTGTGGCAATGAAGTACA-3'
S/L6/+10106	5'-ATGTCATTTCACAGCAGAA-3'
S/L6/+9571	5'-CTTCAATGGTTTGCCATGTT-3'
S/L7/-11271	5'-TGCGAGCTGTCATGAGAATA-3'
S/L7/-11801	5'-AACCGAGAGCAGTACCACAG-3'
S/L7/-12383	5'-TTTGGCTGCTGTAGTCAATG-3'
S/L7/+12640	5'-CTACGACAGATGTCTGTGC-3'
S/L7/+12088	5'-GAGCAGGCTGTAGCTAATGG-3'
S/L7/+11551	5'-TTAGGCTATTGTTGCTGCTG-3'
S/L8/-13160	5'-CAGACAACATGAAGCACCAC-3'
S/L8/-13704	5'-CGCTGACGTGATATATGTGG-3'
S/L8/-14284	5'-TGCACAATGAAGGATACACC-3'
S/L8/+14453	5'-ACATAGCTCGCGTCTCAGTT-3'
S/L8/+13968	5'-GGCATTGTAGGCGTACTGAC-3'
S/L8/+13401	5'-GTTTGCAGGTGAAGTGCAG-3'
S/L9/-15099	5'-TAGTGGCGGCTATTGACTTC-3'
S/L9/-15677	5'-CTAAACCTTGAGCCGATAG-3'
S/L9/-16247	5'-CATGGTCATAGCAGCACTTG-3'
S/L9/+16323	5'-CCAGGTTGTGATGTCACTGAT-3'
S/L9/+15858	5'-CCTTACCAGATCCATCAAG-3'
S/L9/+15288	5'-CGCAAACATAACACTTGCTG-3'
S/L10/-16914	5'-AGTGTGGGTACAAGCCAGT-3'
S/L10/-17466	5'-GTTCCAAGGAACATGTCTGG-3'
S/L10/-18022	5'-AGGTGCCTGTGTAGGATGAA-3'
S/L10/+18245	5'-GGGCTGTCTGCAACTAGAG-3'

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TABLE III-continued

Primers used for the sequencing of the 5' region (ORF1a and ORF1b)	
Names	Sequences (SEQ ID NO: 76 to 139)
S/L10/+17663	5'-TCTTACACGCAATCTGCTT-3'
S/L10/+17061	5'-TACCCATCTGCTCGCATAGT-3'
S/L11/-18877	5'-GCAAGCAGAATTAACCTCA-3'
S/L11/-19396	5'-AGCACCACCTAAATTGCATC-3'
S/L11/-20002	5'-TGGTCCCTTTGAAGGTGTTA-3'
S/L11/+20245	5'-TCGAACACATCGTTTATGGA-3'
S/L11/+19611	5'-GAAGCACCTGTTTCCATCAT-3'
S/L11/+19021	5'-ACGATGCTCAGCCATGTAGT-3'
SARS/L1/F3/+800	5'-GAGGTGCAGTCACTCGCTAT-3'
SARS/L1/F4/+1391	5'-CAGAGATTGGACCTGAGCAT-3'
SARS/L1/F5/+1925	5'-CAGCAAACCACTCAATTCCT-3'
SARS/L1/R3/-1674	5'-AAATGATGGCAACCTCTTCA-3'
SARS/L1/R4/-1107	5'-CACGTGGTTGAATGACTTTG-3'
SARS/L1/R5/-520	5'-ATTTCTGCAACCAGCTCAAC-3'
SARS/L2/F3/+2664	5'-CGCATGTCTCTCTGTTTAC-3'
SARS/L2/F4/+3232	5'-GAGATTGAGCCAGAACCAGA-3'
SARS/L2/F5/+3746	5'-ATGAGCAGGTTGTCATGGAT-3'
SARS/L2/R3/-3579	5'-CTGCCTTAAGAAGCTGGATG-3'
SARS/L2/R4/-2991	5'-TTTCTTCACCAGCATCATCA-3'
SARS/L2/R5/-2529	5'-CACCGTCTTGAGAACAACC-3'
SARS/L3/F3/+4708	5'-TCTTTGGCTGGCTCTTACAG-3'
SARS/L3/F4/+5305	5'-GCTGGTGATGCTGCTAACTT-3'
SARS/L3/F5/+5822	5'-CCATCAAGCCTGTGTCGTAT-3'
SARS/L3/R3/-5610	5'-CAGGTGGTGACAGATCATA-3'
SARS/L3/R4/-4988	5'-AACATCAGCACCATCCAAGT-3'
SARS/L3/R5/-4437	5'-ATCGGACACCATAGTCAACG-3'

The sequences of the fragments L0 to L12 of the SARS-CoV strain derived from the sample recorded under the No. 031589 correspond respectively to the sequences SEQ ID NO: 41 to SEQ ID NO: 54 in the sequence listing appended as an annex. Among these sequences, only that corresponding to the fragments L5 contains a nucleotide difference in relation to the corresponding sequence of the isolate AY278741-Urbani. This t/c mutation at position 7919 results in a modification of the amino acid sequence of the corresponding protein, encoded by ORF1a: at position 2552, a valine (gtt codon; AY278741) is changed to alanine (gct codon) in the SARS-CoV strain 031589. By contrast, no mutation was identified in relation to the corresponding sequence of the isolate AY274119.3-Urbani. The other fragments do not exhibit differences in relation to the corresponding sequences of the isolates Tor2 and Urbani.

Production and Purification of the Recombinant N
and S Proteins of the SARS-CoV Strain Derived
from the Sample Recorded Under the Number
031589

The entire N protein and two polypeptide fragments of the S protein of the SARS-CoV strain derived from the sample recorded under the number 031589 were produced in *E. coli*, in the form of fusion proteins comprising an N- or C-terminal polyhistidine tag. In the two S polypeptides, the N- and C-terminal, hydrophobic sequences of the S protein (signal peptide: positions 1 to 13 and transmembrane helix: positions 1196 to 1218) were deleted whereas the β helix (positions 565 to 687) and the two motifs of the coiled-coil type (positions 895 to 980 and 1155 to 1186) of the S protein were preserved. These two polypeptides consist of: a long fragment (S_L) corresponding to positions 14 to 1193 of the amino acid sequence of the S protein and a short fragment (S_C) corresponding to positions 475 to 1193 of the amino acid sequence of the S protein.

1) Cloning of the cDNAs N, S_L and S_C into the Expression Vectors pIVEX2.3 and pIVEX2.4

The cDNAs corresponding to the N protein and to the S_L and S_C fragments were amplified by PCR under standard conditions, with the aid of the DNA polymerase Platinum Pfx® (INVITROGEN). The plasmids SRAS-N and SRAS-S were used as template and the following oligo-nucleotides as primers:

5'-CCCATATGCTCTGATAATGGACCCCAATCAAAC-3' (Nsense,

SEQ ID NO: 55)

5'-CCCCCGGGTGCCTGAGTTGAATCAGCAGAAGC-3' (N

antisense, SEQ ID NO: 56)

5'-CCCATATGAGTGACCTTGACCGGTGCACCAC-3' (S_C sense,

SEQ ID NO: 57)

5'-CCCATATGAAACCTTGACCCCCACCTGCTC-3' (S_L sense,

SEQ ID NO: 58)

5'-CCCCCGGGTTTAATATATATGCTCATATTTTCCC-3' (S_C and

S_L antisense, SEQ ID NO: 29).

The sense primers introduce an NdeI site (underlined) while the antisense primers introduce an XmaI or SmaI site (underlined). The 3 amplification products were column purified (QIAquick PCR Purification kit, QIAGEN) and cloned into an appropriate vector. The plasmid DNA purified from the 3 constructs (QIAfilter Midi Plasmid kit, QIAGEN) was verified by sequencing and digested with the enzymes NdeI and XmaI. The 3 fragments corresponding to the cDNAs N, S_L and S_C were purified on agarose gel and then inserted into the plasmids pIVEX2.3MCS (C-terminal polyhistidine tag) and pIVEX2.4d (N-terminal polyhistidine tag) digested beforehand with the same enzymes. After verification of the constructs, the 6 expression vectors thus obtained (pIV2.3N, pIV2.3 S_C , pIV2.3 S_L , pIV2.4N, pIV2.4 S_C , also called pIV2.4 S_1 , pIV2.4 S_2) were then used, on the one hand to test the expression of the proteins in vitro, and on the other hand to transform the bacterial strain BL21(DE3)pDIA17 (NOVAGEN). These constructs encode proteins whose expected molecular mass is the following: pIV2.3N (47174

Da), pIV2.3 S_C (82897 Da), pIV2.3 S_L (132056 Da), pIV2.4N (48996 Da), pIV2.4 S_1 (81076 Da) and pIV2.4 S_2 (133877 Da). Bacteria transformed with pIV2.3N were deposited at the CNCM on Oct. 23, 2003, under the number I-3117, and bacteria transformed with pIV2.4 S_1 were deposited at the CNCM on Oct. 23, 2003, under the number I-3118.

2) Analysis of the Expression of the Recombinant Proteins In Vitro and In Vivo

The expression of recombinant proteins from the 6 recombinant vectors was tested, in a first instance, in a system in vitro (RTS100, Roche). The proteins produced in vitro, after incubation of the recombinant vectors pIVEX for 4 h at 30° C., in the RTS100 system, were analyzed by Western blotting with the aid of an anti-(his)₆ antibody coupled to peroxidase. The result of expression in vitro (FIG. 1) shows that only the N protein is expressed in large quantities, regardless of the position, N- or C-terminal, of the polyhistidine tag. In a second step, the expression of the N and S proteins was tested in vivo at 30° C. in LB medium in the presence or in the absence of inducer (1 mM IPTG). The N protein is very well produced in this bacterial system (FIG. 2) and is found mainly in a soluble fraction after lysis of the bacteria. By contrast, the long version of S (S_L) is very weakly produced and is completely insoluble (FIG. 3). The short version (S_C) also exhibits a very weak solubility, but an expression level that is much higher than that of the long version. Moreover, the construct S_C fused with a polyhistidine tag at the C-terminal position has a smaller size than that expected. An immunodetection experiment with an anti-polyhistidine antibody has shown that this construct was incomplete. In conclusion, the two constructs, pIV2.3N and pIV2.4 S_1 , which express respectively the entire N protein fused with the C-terminal polyhistidine tag and the short S protein fused with the N-terminal polyhistidine tag, were selected in order to produce the two proteins in a large quantity so as to purify them. The plasmids pIV2.3N and pIV2.4 S_1 were deposited respectively under the No. I-3117 and I-3118 at the CNCM, 25 rue du Docteur Roux, 75724 PARIS 15, on Oct. 23, 2003.

3) Analysis of the Antigenic Activity of the Recombinant Proteins

The antigenic activity of the N, S_L and S_C proteins was tested by Western blotting with the aid of two serum samples, obtained from the same patient infected with SARS-CoV, collected 8 days (M12) and 29 days (M13) after the onset of the SARS symptoms. The experimental protocol is as described in example 3. The results illustrated by FIG. 4 show (i) the seroconversion of the patient, and (ii) that the N protein possesses a higher antigenic reactivity than the short S protein.

4) Purification of the N protein from pIV2.3N

Several experiments for purifying the N protein, produced from the vector pIV2.3N, were carried out according to the following protocol. The bacteria BL21(DE3)pDIA17, transformed with the expression vector pIV2.3N, were cultured at 30° C. in 1 liter of culture medium containing 0.1 mg/ml of ampicillin, and induced with 1 mM IPTG when the cell density equivalent to $A_{600}=0.8$ is reached (about 3 hours). After 2 hours of culture in the presence of inducer, the cells were recovered by centrifugation (10 min at 5000 rpm), resuspended in the lysis buffer (50 mM NaH₂PO₄, 0.3 M NaCl, 20 mM imidazole, pH 8, containing the mixture of protease inhibitors Complete®, Roche), and lysed with the French press (12 000 psi). After centrifugation of the bacterial lysate (15 min at 12 000 rpm), the supernatant (50 ml) was deposited at a flow rate of 1 ml/min on a metal chelation column (15 ml) (Ni-NTA superflow, Qiagen), equilibrated with the lysis buffer. After washing the column with 200 ml of lysis buffer,

the N protein was eluted with an imidazole gradient (20→250 mM) in 10 column volumes. The fractions containing the N protein were assembled and analyzed by polyacrylamide gel electrophoresis under denaturing conditions followed by staining with Coomassie blue. The results illustrated by FIG. 5 show that the protocol used makes it possible to purify the N protein with a very satisfactory homogeneity (95%) and a mean yield of 15 mg of protein per liter of culture.

5) Purification of the S_c Protein from pIV2.4S_c (pIV2.4S₁)

The protocol followed for purifying the short S protein is very different from that described above because the protein is highly aggregated in the bacterial system (inclusion bodies). The bacteria BL21(DE3)pDIA17, transformed with the expression vector pIV2.4S₁, were cultured at 30° C. in 1 liter of culture medium containing 0.1 mg/ml of ampicillin, and induced with 1 mM IPTG when the cell density equivalent to A₆₀₀=0.8 is reached (about 3 hours). After 2 hours of culture in the presence of inducer, the cells were recovered by centrifugation (10 min at 5000 rpm), resuspended in the lysis buffer (0.1 M Tris-HCl, 1 mM EDTA, pH 7.5), and lysed with the French press (1200 psi). After centrifugation of the bacterial lysate (15 min at 12 000 rpm), the pellet was resuspended in 25 ml of lysis buffer containing 2% Triton X100 and 10 mM β-mercaptoethanol, and then centrifuged for 20 min at 12 000 rpm. The pellet was resuspended in 10 mM Tris-HCl buffer containing 7 M urea, and gently stirred for 30 min at room temperature. This final washing of the inclusion bodies with 7 M urea is necessary in order to remove most of the *E. coli* membrane proteins which co-sediment with the aggregated S_c protein. After a final centrifugation for 20 min at 12 000 rpm, the final pellet is resuspended in the 10 mM Tris-HCl buffer. The electrophoretic analysis of this preparation (FIG. 6) shows that the short S protein may be purified with a satisfactory homogeneity (about 90%) from the inclusion bodies (insoluble extract).

EXAMPLE 3

Immunodominance of the N Protein

The reactivity of the antibodies present in the serum of patients suffering from atypical pneumopathy caused by the SARS-associated coronavirus (SARS-CoV), toward the various proteins of this virus, was analyzed by Western blotting under the conditions described below.

1) Materials

a) Lysate of Cells Infected with SARS-CoV

Vero E6 cells (2×10⁶) were infected with SARS-CoV (isolate recorded under the number FFM/MA104) at a multiplicity of infection (M.O.I.) of 10⁻¹ or 10⁻² and then incubated in DMEM medium containing 2% FCS, at 35° C. in an atmosphere containing 5% CO₂. 48 hours later, the cellular lawn was washed with PBS and then lysed with 500 μl of loading buffer prepared according to Laemmli and containing β-mercaptoethanol. The samples were then boiled for 10 minutes and then sonicated for 3 times 20 seconds.

b) Antibodies

b₁) Serum from a Patient Suffering from Atypical Pneumopathy

The serum designated by a reference at the National Reference Center for Influenza Viruses (Northern region) under the No. 20033168 is that from a French patient suffering from atypical pneumopathy caused by SARS-CoV collected on day 38 after the onset of the symptoms; the diagnosis of SARS-CoV infection was performed by nested RT-PCR and quantitative PCR.

b₂) Monospecific Rabbit Polyclonal sera Directed Against the N Protein or the S Protein

The sera are those produced from the recombinant N and S_c proteins (example 2), according to the immunization protocol described in example 4; they are the rabbit P13097 serum (anti-N serum) and the rabbit P11135 serum (anti-S serum).

2) Method

20 μl of lysate of cells infected with SARS-CoV at M.O.I. values of 10⁻¹ and 10⁻² and, as a control, 20 μl of a lysate of noninfected cells (mock) were separated on 10% SDS polyacrylamide gel and then transferred onto a nitrocellulose membrane. After blocking in a solution of PBS/5% milk/0.1% Tween and washing in PBS/0.1% Tween, this membrane was hybridized overnight at 4° C. with: (1) the immune serum No. 20033168 diluted 1/300, 1/1000 and 1/3000 in the buffer PBS/1% BSA/0.1% Tween, (ii) the rabbit P13097 serum (anti-N serum) diluted 1/50 000 in the same buffer and (iii) the rabbit P11135 serum (anti-S serum) diluted 1/10 000 in the same buffer. After washing in PBS/Tween, a secondary hybridization was performed with the aid of either sheep polyclonal antibodies directed against the heavy and light chains of human G immunoglobulins and coupled with peroxidase (NA933V, Amersham), or of donkey polyclonal antibodies directed against the heavy and light chains of the rabbit G immunoglobulins and coupled with peroxidase (NA934V, Amersham). The bound antibodies were visualized with the aid of the ECL+ kit (Amersham) and of Hyperfilm MP autoradiography films (Amersham). A molecular mass ladder (kDa) is presented in the figure.

3) Results

FIG. 7 shows that three polypeptides of apparent molecular mass 35, 55 and 200 kDa are specifically detected in the extracts of cells infected with SARS-CoV.

In order to identify these polypeptides, two other immunoblots (FIG. 8) were prepared on the same samples and under the same conditions with rabbit polyclonal antibodies specific for the nucleoprotein N (rabbit P13097, FIG. 8A) and for the spike protein S (rabbit P11135, FIG. 8B). This experiment shows that the 200 kDa polypeptide corresponds to the SARS-CoV spike glycoprotein S, that the 55 kDa polypeptide corresponds to the nucleoprotein N while the 35 kDa polypeptide probably represents a truncated or degraded form of N.

The data presented in FIG. 7 therefore show that the serum 20033168 strongly reacts with N and a lot more weakly with the SARS-CoV S since the 35 and 55 kDa polypeptides are visualized in the form of intense bands for 1/300, 1/1000 and 1/3000 dilutions of the immunoserum whereas the 200 kDa polypeptide is only weakly visualized for a dilution of 1/300. It is also possible to note that no other SARS-CoV polypeptide is detected for dilutions greater than 1/300 of the serum 20033168.

This experiment indicates that the antibody response specific for the SARS-CoV N dominates the antibody responses specific for the other SARS-CoV polypeptides and in particular the antibody response directed against the S glycoprotein. It indicates an immunodominance of the nucleoprotein N during human infections with SARS-CoV.

Preparation of Monospecific Polyclonal Antibodies
Directed Against the SRAS-Associated Coronavirus
(SARS-CoV) N and S Proteins

1) Materials and Method

Three rabbits (P13097, P13081, P13031) were immunized with the purified recombinant polypeptide corresponding to the entire nucleoprotein (N), prepared according to the protocol described in example 2. After a first injection of 0.35 mg per rabbit of protein emulsified in complete Freund's adjuvant (intradermal route), the animals received 3 booster injections at 3 and then 4 weeks' interval, of 0.35 mg of recombinant protein emulsified in incomplete Freund's adjuvant.

Three rabbits (P11135, P13042, P14001) were immunized with the recombinant polypeptide corresponding to the short fragment of the S protein (S_c) produced as described in example 2. As this polypeptide is found mainly in the form of inclusion bodies in the bacterial cytoplasm, the animals received 4 intradermal injections at 3-4 weeks' interval of a preparation of inclusion bodies corresponding to 0.5 mg of recombinant protein emulsified in incomplete Freund's adjuvant. The first 3 injections were made with a preparation of inclusion bodies prepared according to the protocol described in example 2, while the fourth injection was made with a preparation of inclusion bodies which were prepared according to the protocol described in example 2 and then purified on sucrose gradient and washed in 2% Triton X100.

For each rabbit, a preimmune (p.i.) serum was prepared before the first immunization and an immune serum (I.S.) 5 weeks after the fourth immunization.

In a first instance, the reactivity of the sera was analyzed by ELISA test on preparations of recombinant proteins similar to those used for the immunizations; the ELISA tests were carried out according to the protocol and with the reagents as described in example 6.

In a second instance, the reactivity of the sera was analyzed by preparing an immunoblot (Western blot) of a lysate of cells infected with SARS-CoV, according to the protocol as described in example 3.

2) Results

The ELISA tests (FIG. 9) demonstrate that the preparations of recombinant N protein and of inclusion bodies of the short fragment of the S protein (S_c) are immunogenic in animals and that the titer of the immune sera is high (more than 1/25 000).

The immunoblot (FIG. 8) shows that the rabbit P13097 immune serum recognizes two polypeptides present in the lysates of cells infected with SARS-CoV: a polypeptide whose apparent molecular mass (50-55 kDa based on experiments) is compatible with that of the nucleoprotein N (422 residues, predicted molecular mass of 46 kDa) and a polypeptide of 35 kDa, which probably represents a truncated or degraded form of N.

This experiment also shows that the rabbit P11135 serum mainly recognizes a polypeptide whose apparent molecular mass (180-220 kDa based on experiments) is compatible with a glycosylated form of S (1255 residues, nonglycosylated polypeptide chain of 139 kDa), as well as lighter polypeptides, which probably represent truncated and/or nonglycosylated forms of S.

In conclusion, all these experiments demonstrate that the recombinant polypeptides expressed in *E. coli* and corresponding to the SARS-CoV N and S proteins make it possible to induce, in animals, polyclonal antibodies capable of recognizing the native forms of these proteins.

Preparation of Monospecific Polyclonal Antibodies
Directed Against the SARS-Associated Coronavirus
(SARS-CoV) M and E Proteins

1) Analysis of the Structure of the M and E Proteins

a) E Protein

The structure of the SARS-CoV E protein (76 amino acids) was analyzed in silico, with the aid of various software packages such as signalP v1.1, NetNGlyc 1.0, THMM 1.0 and 2.0 (Krogh et al., 2001, J. Mol. Biol., 305(3):567-580) or alternatively TOPPRED (von Heijne, 1992, J. Mol. Biol. 225, 487-494). The analysis shows that this nonglycosylated polypeptide is a type 1 membrane protein, containing a single transmembrane helix (aa 12-34 according to THMM), and in which the majority of the hydrophilic domain (42 residues) is located at the C-terminal end and probably inside the viral particle (endodomain). It is possible to note an inversion in the topology predicted by versions 1.0 (N-ter is external) and 2.0 (N-ter is internal) of the THMM software, but that other algorithms, in particular TOPPRED and THUMBUP (Zhou et Zhou, 2003, Protein Science 12:1547-1555) confirm an external location of the N-terminal end of E.

b) M Protein

A similar analysis carried out on the SARS-CoV M protein (221 amino acids) shows that this polypeptide does not possess a signal peptide (according to the software signalP v1.1) but three transmembrane domains (residues 15-37, 50-72, 77-99 according to THMM2.0) and a large hydrophilic domain (aa 100-221) located inside the viral particle (endodomain). It is probably glycosylated on the asparagine at position 4 (according to NetNGlyc 1.0).

Thus, in agreement with the experimental data known for the other coronaviruses, it is remarkable that the two M and E proteins exhibit endodomains corresponding to the majority of the polypeptides and of the ectodomains that are very small in size.

The ectodomain of E probably corresponds to residues 1 to 11 or 1 to 12 of the protein: MYSFVSEETGT(L), SEQ ID NO: 70. Indeed, the probability associated with the transmembrane location of residue 12 is intermediate (0.56 according to THMM 2.0).

The ectodomain of M probably corresponds to residues 2 to 14 of the protein: ADNGTITVEELKQ, SEQ ID NO: 69. Indeed, the N-terminal methionine of M is very probably cleaved from the mature polypeptide because the residue at position 2 is an alanine (Varshaysky, 1996, 93:12142-12149).

Moreover, the analysis of the hydrophobicity (Kyte & Doolittle Hopp & Woods) of the E protein demonstrates that the C-terminal end of the endodomain of E is hydrophilic and therefore probably exposed at the surface of this domain. Thus, a synthetic peptide corresponding to this end is a good immunogenic candidate for inducing, in animals, antibodies directed against the endodomain of E. Consequently, a peptide corresponding to 24 C-terminal residues of E was synthesized.

2) Preparation of Antibodies Directed Against the Ectodomain of the M and E Proteins and the Endodomain of the E Protein

The peptides M2-14 (ADNGTITVEELKQ, SEQ. ID NO: 69), E1-12 (MYSFVSEETGTL, SEQ ID NO: 70) and E53-76 (KPTVYVYSRV KNLNSSEGVP DLLV, SEQ ID NO: 71) were synthesized by Neosystem. They were coupled with KLH (Keyhole Limpet Hemocyanin) with the aid of MBS (m-maleimido-benzoyl-N-hydroxysuccinimide ester) via a

cysteine added during the synthesis either at the N-terminus of the peptide (case for E53-76) or at the C-terminus (case of M2-14 and E1-12).

Two rabbits were immunized with each of the conjugates, according to the following immunization protocol: after a first injection of 0.5 mg of peptide coupled with KLH and emulsified in complete Freund's adjuvant (intradermal route), the animals receive 2 to 4 booster injections at 3 or 4 weeks' interval of 0.25 mg of peptide coupled to KLH and emulsified in incomplete Freund's adjuvant.

For each rabbit, a preimmune (p.i.) serum was prepared before the first immunization and an immune serum (I.S.) is prepared 3 to 5 weeks after the booster injections.

The reactivity of the sera was analyzed by Western blotting with the aid of extracts of cells infected with SARS-CoV (FIG. 43B) or with the aid of extracts of cells infected with a recombinant vaccinia virus expressing the protein E (VV-TG-E, FIG. 43A) or M (VV-TN-M, FIG. 43C) of the SARS-CoV 031589 isolate.

The immune sera of the rabbits 22234 and 22240, immunized with the conjugate KLH-E53-76, recognize a polypeptide of about 9 to 10 kD, which is present in the extracts of cells infected with SARS-CoV but absent from the extracts, of noninfected cells (FIG. 43B). The apparent mass of this polypeptide is compatible with the predicted mass of the E protein, which is 8.4 kD. Similarly, the immune serum of the rabbit 20047, immunized with the conjugate KLH-E1-12, recognizes a polypeptide present in the extracts of cells infected with the VV-TG-E virus, whose apparent molar mass is compatible with that of the E protein (FIG. 43A).

The immune serum of the rabbits 20013 and 20080, immunized with the conjugate KLH-M2-14, recognizes a polypeptide present in the extracts of cells infected with the VV-TN-M virus (FIG. 43C), whose apparent molar mass (about 18 kD) is compatible with that of the glycoprotein M, which is 25.1 kD and has a high iso-electric point (9.1 for the naked polypeptide).

These results demonstrate that the peptides E1-12 and E53-76, on the one hand, and the peptide M2-14, on the other hand, make it possible to induce, in animals, polyclonal antibodies capable of recognizing the native forms of the SARS-CoV E and M proteins, respectively.

EXAMPLE 6

Analysis of the ELISA Reactivity of the Recombinant N Protein Toward Sera from Patients Suffering from SARS

1) Materials

The antigen used to prepare the solid phases is the purified recombinant nucleoprotein N prepared according to the protocol described in example 2.

The sera to be tested (table IV) were chosen on the basis of the results of analysis of their reactivity by immunofluorescence (IF-SARS titer), toward cells infected with SARS-CoV.

TABLE IV

Sera tested by ELISA				
Reference	Serum No.	Type of serum	Date of the serum***	IF-SARS titer
3050	A	Control	na*	nt**
3048	B	Control	na	nt
033168	D	Patient 1-SARS	Apr. 27, 2003 (D38)	320
033397	E	Patient-1 SARS	May 11, 2005 (D52)	320

TABLE IV-continued

Sera tested by ELISA				
Reference	Serum No.	Type of serum	Date of the serum***	IF-SARS titer
032632	F	Patient-2 SARS	Mar. 21, 2003 (D17)	2500
032791	G	Patient-3 SARS	Apr. 04, 2003 (D3)	<40
033258	H	Patient-3 SARS	Apr. 28, 2003 (D27)	160

*na: not applicable.

**nt: not tested.

***the dates indicated correspond to the number of days after the onset of the SARS symptoms.

2) Method

The N protein (100 µl) diluted at various concentrations in 0.1 M carbonate buffer, pH 9.6 (1, 2 or 4 µg/ml) is distributed into the wells of ELISA plates, and then the plates are incubated overnight at laboratory temperature. The plates are washed with PBS-Tween buffer saturated with PBS-skimmed milk-sucrose (5%) buffer. The test sera (100 µl), diluted beforehand (1/50, 1/100, 1/200, 1/400, 1/800, 1/1600 and 1/3200) are added and then the plates are incubated for 1 h at 37° C. After 3 washings, the peroxidase-labeled anti-human IgG conjugate (reference 209-035-098, JACKSON) diluted 1/18 000 is added and then the plates are incubated for 1 h at 37° C. After 4 washings, the chromogen (TMB) and the substrate (H₂O₂) are added and the plates are incubated for 30 min at room temperature, protected from light. The reaction is then stopped and then the absorbance at 450 nm is measured with the aid of an automated reader.

3) Results

The ELISA tests (FIG. 10) demonstrate that the recombinant N protein preparation is specifically recognized by the antibodies of sera from patients suffering from SARS collected in the late phase of the infection (≥17 days after the onset of the symptoms) whereas it is not significantly recognized by the antibodies of a patient's serum collected in the early phase of the infection (3 days after the onset of the symptoms) or by control sera from subjects not suffering from SARS.

EXAMPLE 7

ELISA Tests Prepared for a Very Specific and Sensitive Detection of a SARS-Associated Coronavirus Infection, from Sera of Patients

1) Indirect ELISA IgG Test

a) Reagents

Preparation of the Plates

The plates are sensitized with a solution of N protein at 2 µg/ml in a 10 mM PBS buffer, pH 7.2, phenol red at 0.25 ml/l. 100 µl of solution are deposited in the wells and left to incubate at room temperature overnight. Saturation is obtained by prewashing in 10 mM PBS/0.1% Tween buffer, followed by washing with a saturation solution PBS, 25% milk/sucrose.

Diluent Sera

Buffer 0.48 g/l TRIS, 10 mM PBS, 3.7 g/l EDTA, 15% v/v milk, pH 6.7

Diluent Conjugate

Citrate buffer (15 g/l), 0.5% Tween, 25% bovine serum, 12% NaCl, 6% v/v skimmed milk pH 6.5

Conjugate

50× anti-human IgG conjugate, marketed by Bio-Rad: Platelia H. pylori kit ref 72778

Other Solutions:

Washing solution R2, solutions for visualizing with TMB R8 diluent, R9 chromogen, R10 stopping solution: reagents marketed by Bio-Rad (e.g.: Platelia *pylori* kit, ref 72778)

b) Procedure

Dilute the sera 1/200 in the sample diluent

Distribute 100 μ l/well

Incubation 1 h at 37° C.

3 washings in 10 \times WASHING solution R2 diluted beforehand 10-fold in demineralized water (i.e., 1 \times washing solution)

Distribute 100 μ l of conjugate (50 \times conjugate to be diluted immediately before use in the diluent conjugate provided)

Incubation 1 h at 37° C.

4 washings in 1 \times washing solution

Distribute 200 μ l/well of visualization solution (to be diluted immediately before use e.g.: 1 ml of R9 in 10 ml of R8)

Incubation for 30 min at room temperature in the dark

Stop the reaction with 100 μ l/well of R10

READING at 450/620 nm

The results can be interpreted by taking a THRESHOLD serum giving a response above which the sera tested would be considered as positive. This serum is chosen and diluted so as to give a significantly higher signal than the background noise.

2) Double Epitope Elisa Test

Reagents

Preparation of the Plates

The plates are sensitized with a solution of N protein at 1 μ g/ml in a 10 mM PBS buffer, pH 7.2, phenol red at 0.25 ml/l. 100 μ l of solution are deposited in the wells and left to incubate at room temperature overnight. Saturation is obtained by prewashing in 10 mM PBS/0.1% Tween buffer, followed by washing with a saturation solution 10 mM PBS, 25% (V/V) milk.

Diluent sera and Conjugate

Buffer 50 mM TRIS saline, pH 8, 2% milk

Conjugate

This is the purified recombinant N protein coupled with peroxidase according to the Nakane protocol (Nakane P. K. and Kawaoi A.; (1974): *Peroxydase-labeled antibody, a new method of conjugation*. The Journal of Histochemistry and Cytochemistry Vol. 22, N) 23, pp. 1084-1091), in respective molar ratios 1/2. This ProtN POD conjugate is used at a concentration of 2 μ g/ml in serum/conjugate diluent.

Other Solutions:

Washing solution R2, solutions for visualization with TMB R8, diluent, R9 chromogen, R10 stopping solution: reagents marketed by Bio-Rad (e.g. Platelia *pylori* kit, ref 72778).

b) Procedure

1st step in "predilution" plate

Dilute each serum 1/5 in the predilution plate (48 μ l of diluent+12 μ l of serum).

After having diluted all the sera, distribute 60 μ l of conjugate.

Where appropriate, the serum+conjugate mix is left to incubate.

2nd step in "reaction" plate

Transfer 100 μ l of mixture/well into the reaction plate

Incubation 1 h 37° C.

5 washings in 10 \times WASHING solution R2 diluted 10-fold beforehand in demineralized water (\rightarrow 1 \times washing solution)

Distribute 200 μ l/well of visualization solution (to be diluted immediately before use e.g.: 1 ml of R9 in 10 ml of R8)

Incubation 30 min at room temperature and protected from light

Stop the reaction with 100 μ l/well of R10

READING at 450/620 nm

Likewise as for the indirect ELISA test, the results can be interpreted using a "threshold value" serum. Any serum having a response greater than the threshold value serum will be considered as positive.

2) Results

The sera of patients classified as probable cases of SARS from the French hospital of Hanoi, Vietnam or in relation with the French hospital of Hanoi (JYK) were analyzed using the indirect IgG-N test and the double epitope N test.

The results of the indirect IgG-N test (FIGS. 14 and 15) and double epitope N test (FIGS. 16 and 17) show an excellent correlation between them and with an indirect ELISA test comparing the reactivity of the sera toward a lysate of VeroE6 cells infected or not infected with SARS-CoV (ELISA-SARS-CoV lysate; see table V below). All the sera collected 12 days or more after the onset of the symptoms were found to be positive, including in patients for whom it had not been possible to document the SARS-CoV virus infection by analyzing respiratory samples by RT-PCR, probably because of a sample being collected too late during the infection (\geq D12). In the case of the patient TTH for whom a nasal sample collected on D7 was found to be negative by RT-PCR, the quality of the sample may be in question.

Some sera were found to be negative whereas the presence of SARS-CoV was detected by RT-PCR. They are in all cases early sera collected less than 10 days after the onset of the symptoms (e.g.: serum #032637). In the case of a patient PTTH (serum #032673), only a suspicion of SARS was raised at the time the samples were collected.

In conclusion, the indirect IgG-N and N-double epitope serological tests make it possible to document the SARS-CoV infection in all the patients for the sera collected 12 days or more after the infection.

TABLE V

Results of the ELISA tests						
Sample Num	Patient	Day	PCR-SARS (1)	ELISA SARS-CoV lysate (2)	IgG-N (2nd series)	2Xepitope (2nd series)
033168	JYK	38	POS	+++	>5000	NT
033597	J K	74	POS	NT	\approx 5000	NT
032552	VTT	8	NEG-D3&D8&D12	NEG	<200	<5
032544	CTP	16	NEG-D16&D20	++	>5000	>>20
032546	CJF	15	NEG-D15&D19	++	>5000	>>20

TABLE V-continued

Results of the ELISA tests						
Sample Num	Patient	Day	PCR-SARS (1)	ELISA SARS-CoV lysate (2)	IgG-N (2nd series)	2Xepitope (2nd series)
032548	PTL	17	NEG	++	>5000	>>20
			D17&D21			
032550	NTH	17	NEG-	++	>5000	>>20
			D17&D21			
032553	VTT	8	NEG-	NEG	<200	<5
			D3&D8&D12			
032554	NTBV	4	POS	NEG	<200	<5
032555	NTBV	4	POS	NEG	<200	<5
032564	NTP	15	POS	++	>5000	>>20
032629	NVH	4	POS	NEG	<200	<5
032631	BTTX	9	POS	NEG	<200	<5
032635	NHH	4	POS	NEG	<200	<5
032637	NHB	10	POS	NEG	<200	<5
032642	BTTX	9	POS	NEG	<200	<5
032643	LTDH	1	POS	NEG	<200	<5
032644	NTBV	4	POS	NEG	<200	<5
032646	TTH	12	NEG	++	>5000	>>20
			D7&D12&D16			
032647	DTH	17	NEG	++	>5000	>>20
			D17&D21			
032648	NNT	15	NEG	++	>5000	>>20
			D15&D19			
032649	PTH	17	NEG	++	>5000	>>20
			D17&D21			
032672	LVV	16	NEG	+	>5000	>>20
			D16&D20			
032673	PTTH	NA	NEG	NEG	<200	<5
032674	PNB	17	NEG	++	>5000	>>20
			D17&D21			
032682	VTH	12	NEG	++	>5000	>>20
			D12&D16			
032683	DTV	17	NEG	+	>1000	>>20
			D17&D21			

Remarks:

(1): The RT-PCR analyses were carried out by nested RT-PCR BNI, LC Artus and LC-N on nasal or pharyngeal swabs; POS means that at least one sample was found to be positive in this patient.

(2): The reactivity of the sera in the ELISA test using a lysate of cells infected with SARS-CoV was classified as very highly reactive (+++), highly reactive (++), reactive (+) and negative according to the OD value obtained at the dilutions tested.

EXAMPLE 8

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-continued

Detection of SARS-Associated Coronavirus (SARS-CoV) by RT-PCR

1) Real Time Development of RT-PCR Conditions with the Aid of Primers Specific for the Gene for the Nucleocapsid Protein—"Light Cycler N" Test

a) Design of the Primers and Probes

The primers and probes were designed from the sequence of the genome of the SARS-CoV strain derived from the sample recorded under the number 031589, with the aid of the programme "Light Cycler Probe Design (Roche)". Thus, the following two series of primers and probes were selected:

series 1 (SEQ ID NO: 60, 61, 64, 65):

sense primer: N/+ /28507:
5'-GGC ATC GTA TGG GTT G-3' [28507-28522]

antisense primer: N/- /28774:
5'-CAG TTT CAC CAC CTC C-3' [28774-28759]

probe 1:
5'-GGC ACC CGC AAT CCT AAT AAC AAT GC-fluorescein
3' [28561-28586]

probe 2:

5' Red705-GCC ACC GTG CTA CAA CTT CCT-phosphate

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[28588-28608]

series 2 (SEQ ID NO: 62, 63, 66, 67)

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sense primer: N/+ /28375:
5'-GGC TAC TAC CGA AGA G-3' [28375-28390]

antisense primer: N/- /28702:
5'-AAT TAC CGC GAC TAC G-3' [28702-28687]

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probe 1: SARS/N/FL:
5'-ATA CAC CCA AAG ACC ACA TTG GC-fluorescein 3'
[28541-28563]

60

probe 2: SARS/N/LC705:
5' Red705-CCC GCA ATC CTA ATA ACA ATG CTG C-phosphate 3' [28565-28589]

b) Analysis of the Efficacy of the Two Primer Pairs

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In order to test the respective efficacy of the two pairs of primers, an RT-PCR amplification was carried out on a synthetic RNA corresponding to nucleotides 28054-29430 of the

genome of the SARS-CoV strain derived from the sample recorded under the number 031589 and containing the sequence of the N gene.

More specifically:

This synthetic RNA was prepared by in vitro transcription with the aid of the T7 phage RNA polymerase, of a DNA template obtained by linearization of the plasmid SRAS-N with the enzyme Bam HI. After eliminating the DNA template by digestion with the aid of DNase I, the synthetic RNAs are purified by a phenol-chloroform extraction, followed by two successive precipitations in ammonium acetate and isopropanol. They are then quantified by measuring the absorbance at 260 nm and their quality is checked by the ratio of the absorbances at 260 and 280 nm and by agarose gel electrophoresis. Thus, the concentration of the synthetic RNA preparation used for these studies is 1.6 mg/ml, which corresponds to 2.1×10^{15} copies/ml of RNA.

Decreasing quantities of synthetic RNA were amplified by RT-PCR with the aid of the "Superscript™ One-Step RT-PCR with Platinum® Taq" kit and the pairs of primers No. 1 (N+/28507, N-/28774) (FIG. 1A) and No. 2 (N+/28375, N-/28702) (FIG. 1B), according to the supplier's instructions. The amplification conditions used are the following: the cDNA was synthesized by incubation for 30 min at 45° C., 15 min at 55° C. and then 2 min at 94° C. and it was then amplified by 5 cycles comprising: a step of denaturation at 94° C. for 15 sec, a step of annealing at 45° C. for 30 sec and, then a step of extension at 72° C. for 30 sec, followed by 35 cycles comprising: a step of denaturation at 94° C. for 15 sec, a step of annealing at 55° C. for 30 sec and then a step of extension at 72° C. for 30 sec, with 2 sec of additional extension at each cycle, and a final step of extension at 72° C. for 5 min. The amplification products obtained were then kept at 10° C.

The results presented in FIG. 11 show that the pair of primers No. 2 (N+/28375, N-/28702) makes it possible to detect up to 10 copies of RNA (band of weak intensity) or 10^2 copies (band of good intensity) against 10^4 copies for the pair of primers No. 1 (N+/28507, N-/28774). The amplicons are respectively 268 bp (pair 1) and 328 bp (pair 2).

c) Development of Real Time RT-PCR

A real time RT-PCR was developed with the aid of the pair of primers No. 2 and of the pair of probes consisting of SRAS/N/FL and SRAS/N/LC705 (FIG. 2).

The amplification was carried out on a LightCycler™ (Roche) with the aid of the "Light Cycler RNA Amplification Kit Hybridization Probes" kit (reference 2 015 145, Roche) under the following optimized conditions. A reaction mixture containing: H₂O (6.8 µl), 25 mM MgCl₂ (0.8 µl, 4 µM Mg2+ final), 5× reaction mixture (4 µl), 3 µM probe SRAS/N/FL (0.5 µl, 0.075 µM final), 3 µM probe SRAS/N/LC705 (0.5 µl, 0.075 µM final), 10 µM primer N+/28375 (1 µl, 0.5 µM final), 10 µM primer N-/28702 (1 µl, 0.5 µM final), enzyme mixture (0.4 µl) and sample (viral RNA, 5 µl) was amplified according to the following program:

Reverse transcription: 50° C. 10:00 min analysis mode: none

Denaturation: 95° C. 30 sec×1 analysis mode: none

Amplification: 95° C. 2 sec

50° C. 15 sec analysis mode: quantification*{×45

72° C. 13 sec thermal ramp 2.0° C./sec}

* The fluorescence is measured at the end of the annealing and at each cycle (in SINGLE mode).

Annealing: 40° C. 30 sec×1 analysis mode: none

The results presented in FIG. 12 show that this real time RT-PCR is very sensitive since it makes it possible to detect 10^2 copies of synthetic RNA in 100% of the 5 samples ana-

lyzed (29/29 samples in 8 experiments) and up to 10 copies of RNA in 100% of the 5 samples analyzed (40/45 samples in 8 experiments). It also shows that this RT-PCR makes it possible to detect the presence of the SARS-CoV genome in a sample and to quantify the number of genomes present. By way of example, the viral RNA of a SARS-CoV stock cultured on Vero E6 cells was extracted with the aid of the "QIamp viral RNA extraction" kit (Qiagen), diluted to 0.05×10^{-14} and analyzed by real time RT-PCR according to the protocol described above; the analysis presented in FIG. 12 shows that this virus stock contains 6.5×10^9 genome-equivalents/ml (geq/ml), which is entirely similar to the 1.0×10^{10} geq/ml value measured with the aid of the "RealArt™ HPA-Coronavirus LC RT PCR Reagents" kit marketed by Artus.

2) Development of Nested RT-PCR Conditions Targeting the Gene for RNA Polymerase—"CDC (Centers for Disease Control and Prevention)/IP Nested RT-PCR" Test

a) Extraction of the Viral RNA

Clinical sample: QIamp viral RNA Mini Kit (QIAGEN) according to the manufacturer's instructions, or an equivalent technique. The RNA is eluted in a volume of 60 µl.

b) "SNE/SAR" Nested RT-PCR

First step: "SNE" coupled RT-PCR

The Invitrogen "Superscript™ One-Step RT-PCR with Platinum® Taq" kit was used, but the "Titan" kit from Roche Boehringer can be used in its place with similar results.

Oligonucleotides:

SNE-S1 5' GGT TGG GAT TAT CCA AAA TGT GA 3'
SNE-AS1 5' GCA TCA TCA GAA AGA ATC ATC ATG 3'

→Expected size: 440 bp

1. Prepare a mix:

H2O	6.5 µl
Reaction mix 2X	12.5 µl
Oligo SNE-S1 50 µM	0.2 µl
Oligo SNE-AS1 50 µM	0.2 µl
RNAasin 40 U/µl	0.12 µl
RT/Platinum Taq mix	0.5 µl

2. To 20 µl of the mix, add 5 µl of RNA and carry out the amplification on a thermocycler (ABI 9600 conditions):

2.1	45° C.	30 min.		
	55° C.	15 min.		
	94° C.	2 min.		
2.2.	94° C.	15 sec.	}	× 5 cycles
	45° C.	30 sec.		
	72° C.	30 sec.		
2.3.	94° C.	15 sec.	}	× 35 cycles
	55° C.	30 sec.		
	72° C.	30 sec. + 2 sec./cycle		
2.4.	72° C.	5 min.		
2.5	10° C.	∞		

Storage at +4° C.

The RNAasin (N2511/N2515) from Promega was used as RNase inhibitors.

Synthetic RNAs served as positive control. As the control, 10^3 , 10^2 and 10 copies of synthetic RNA R_{SNE} were amplified in each experiment.

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Second step: "SAR" nested PCR
Oligonucleotides:

SAR1-S 5' CCT CTC TTG TTC TTG CTC GCA 3' 5
SAR1-AS 5' TAT AGT GAG CCG CCA CAC ATG 3'

→Expected size: 121 bp

1. Prepare a mix:

H2O	35.8 µl
Taq buffer 10X	5 µl
MgCl ₂ 25 mM	4 µl
Mix dNTPs 5 mM	2 µl
Oligo SAR1-S 50 µM	0.5 µl
Oligo SAR1-AS 50 µM	0.5 µl
Taq DNA pol 5 U/µl	0.25 µl

AmpliTaq DNA Pot from Applied Biosystems was used (10x buffer without MgCl₂, ref 27216601).

2. To 48 µl of the mix, add 2 µl of the product from the first PCR and carry out the amplification (ABI 9600 conditions):

2.1.	94° C.	2 min.	} × 5 cycles
2.2.	94° C.	30 sec.	
	45° C.	45 sec.	
	72° C.	30 sec.	
2.3.	94° C.	30 sec.	} × 35 cycles
	55° C.	30 sec.	
	72° C.	30 sec. + 1 sec./cycle	
	72° C.	5 min.	
2.4.	72° C.	5 min.	
2.5	10° C.	∞	

3. Analyze 10 µl of the reaction product on "low-melting" gel (Seakem GTG type) containing 3% agarose.

The sensitivity of the nested test is routinely, under the conditions described, 10 copies of RNA.

4. The fragments can then be purified on QIAquick PCR kit (QIAGEN) and sequenced with the oligos SAR1-S and SAR1-AS.

3) Detection of the SARS-CoV RNA by PCR from Respiratory Samples

a) First Comparative Study

A comparative study was carried out on a series of respiratory samples received by the National Reference Center for the Influenza Virus (Northern region) and likely to contain SARS-CoV. To do this, the RNA was extracted from the samples with the aid of the "Qiamp viral RNA extraction" kit (Qiagen) and analyzed by real time RT-PCR, on the one hand with the aid of the pairs of primers and probes of the No. 2 series under the conditions described above on the one hand, and on the other hand with the aid of the kit "LightCycler SARS-CoV quantification kit" marketed by Roche (reference 03 604 438). The results are summarized in table VI below. They show that 18 of the 26 samples are negative and 5 of the 26 samples are positive for the two kits, while one sample is positive for the Roche kit alone and two for the "series 2" N reagents alone. Additionally, for 3 samples (20032701, 20032712, 20032714) the quantities of RNA detected are markedly higher with the reagents (probes and primers) of the No. 2 series. These results indicate that the "series 2" N primers and probes are more sensitive for the detection of the SARS-CoV genome in biological samples than those of the kit currently available.

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TABLE VI

Real time RT-PCR analysis of the RNAs extracted from a series of samples from 5 patients with the aid of the pairs of primers and probes of the No. 2 series ("series 2" N) or of the kit "Lightcycler SARS-CoV quantification kit" (Roche). The type of sample is indicated as well as the number of copies of viral genome measured in each of the two tests. NEG: negative RT-PCR.

10	Sample No.	Patient	Type of sample	ROCHE KIT	"Series 2" N
	20033082	K	nasal	NEG	NEG
	20033083	K	pharyngeal	NEG	NEG
	20033086	K	nasal	NEG	NEG
	20033087	K	pharyngeal	NEG	NEG
15	20032802	M	nasal	NEG	NEG
	20032803	M	expectoration	NEG	NEG
	20032806	M	nasal or pharyngeal	NEG	NEG
	20031746ARN2	C	pharyngeal	NEG	NEG
	20032711	C	nasal or pharyngeal	39	NEG
20	20032910	B	nasal	NEG	NEG
	20032911	B	pharyngeal	NEG	NEG
	20033356	V	expectoration	NEG	NEG
	20033357	V	expectoration	NEG	NEG
	20031725	K	endotracheal asp.	NEG	150
25	20032657	K	endotracheal asp.	NEG	NEG
	20032698	K	endotracheal asp.	NEG	NEG
	20032720	K	endotracheal asp.	3	5
30	20033074	K	stools	115	257
	20032701	M	pharyngeal	443	1676
	20032702	M	expectoration	NEG	249
	20031747ARN2	C	pharyngeal	NEG	NEG
	20032712	C	unknown	634	6914
	20032714	C	pharyngeal	17	223
35	20032800	B	nasal	NEG	NEG
	20033353	V	nasal	NEG	NEG
	20033384	V	nasal	NEG	NEG

b) Second Comparative Study

The performance of various nested RT-PCR and real time RT-PCR methods were then compared for 121 respiratory samples from possible cases of SARS at the French hospital in Hanoi, Vietnam, taken between the 4th and the 17th day after the onset of the symptoms. Among these samples, 14 were found to be positive during a first test using the nested RT-PCR method targeting ORF1b (encoding replicase) as described initially by Bernhard Nocht Institute (BNI nested RT-PCR). Information relating to this test is available on the internet, at the address www.15.bni-hamburg.de/bni2/neu2/getfile.acgi?area_engl=diagnostics&pid=4112.

The various tests compared in this study are:

the quantitative RT-PCR method according to the invention, with the "series 2" N primers and probes described above (LightCycler N column),

the nested RT-PCR test targeting the RNA polymerase gene described above, developed by the CDC, BNI and Institut Pasteur (CDC/IP nested RT-PCR),

the ARTUS kit with the reference "HPA Corona LC RT-PCR Kit #5601-02", which is a real time RT-PCR test targeting the ORF1b gene,

the BNI nested RT-PCR test, also targeting the RNA polymerase gene mentioned above.

The inventors observed:

1) an inter-test variability for the same technique, linked to the degradation of the RNA preparation during repeated thawing, in particular for the samples containing the lowest quantities of RNA,

2) a reduced sensitivity of the CDC/IP nested RT-PCR compared with the BNI nested RT-PCR, and

3) a comparable sensitivity of the quantitative RT-PCR test according to the invention (LightCycler N) compared with the Artus LightCycler (LC) test.

These results, which are presented in table VII below, show that the quantitative RT-PCR test according to the invention constitutes an excellent addition—or an alternative—to the tests currently available. Indeed, the SARS-linked coronavirus is an emergent virus which is capable of changing rapidly. In particular, the gene for the RNA polymerase of the SARS-linked coronavirus, which is targeted in most of the tests currently available, can recombine with that of other coronaviruses not linked to SARS. The use of a test targeting this gene exclusively could then lead to the production of false-negatives.

The quantitative RT-PCR test according to the invention does not target the same genomic region as the ARTUS kit since it targets the gene encoding the N protein. By carrying out a diagnostic test targeting two different genes of the SARS-linked coronavirus, it can therefore be hoped to avoid false-negative type results which could be due to the genetic evolution of the virus.

Furthermore, it appears particularly advantageous to target the gene for the nucleocapsid protein because it is very stable because of the high selection pressure linked to the high structural constraints regarding this protein.

TABLE VII

Comparison of various methods of analysis by gene amplification, from 121 samples of probable cases of SARS at the French hospital in Hanoi, Vietnam (epidemic 2003)

NRC No.	Sample type (1)	Sample collection day	Patient	CDC/IP nested RT-PCR	BNI nested RT-PCR	Artus Light Cycler kit	Light Cycler N (IP)
107 samples	N and P			Negative	Negative	Negative	Negative
032529	P	10	NHB	Negative	Positive	Negative	Negative
032530	N	10	NHB	Positive	Positive	3.10E+01	4.20E+01
032531	P	7	LP	Positive	Positive	7.70E+00	3.10E+00
032534	N	15	BND	Positive	Positive	1.60E+00	Negative
032600	P	4	NHH	Negative	Positive	Negative	1.30E+02
032612	P	17	NTS	Negative	Positive	Negative	Negative
032688	P	9	BTX	Positive	Positive	Negative	Negative
032689	N	4	NVH	Positive	Positive	1.20E+01	2.30E+02
032690	P	4	NVH	Negative	Positive	1.60E+00	Negative
032727	P	8	NVH	Positive	Positive	2.30E+02	4.00E+02
032728	N	8	NVH	Positive	Positive	1.10E+03	1.60E+04
032729	P	14	NHB	Positive	Positive	5.90E+00	3.40E+01
032730	N	14	NHB	Positive	Positive	1.30E+02	4.80E+02
032741	P	8	NHH	Positive	Positive	2.10E+02	1.30E+02
positives				10	14	10	9
fraction detected from the 14 positives				71.4%	100.0%	71.4%	64.3%

(1) P = pharyngeal swab
N = nasal swab

EXAMPLE 9

Production and Characterization of Monoclonal Antibodies Directed Against the N Protein

Balb C mice were immunized with the purified recombinant N protein and their spleen cells fused with an appropriate murine myeloma according to the Köhler and Milstein techniques.

Nineteen anti-N antibody secreting hybridomas were pre-selected and their immunoreactivities determined. These

antibodies do indeed recognize the recombinant N protein (in ELISA) with variable intensities, and the natural viral N protein in ELISA and/or in Western blotting. FIGS. 18 to 20 show the results of these tests for 15 of these 19 monoclonal antibodies.

The highly reactive clones 12, 17, 28, 57, 72, 76, 86, 87, 98, 103, 146, 156, 166, 170, 199, 212, 218, 219 and 222 were subcloned. Specificity studies were carried out with the appropriate tools in order to determine the epitopes recognized and verify the absence of reactivity toward other human coronaviruses and certain respiratory viruses.

Epitope mapping studies (performed on spot membrane with the aid of overlapping peptides of 15 aa) and additional studies performed on the natural N protein in Western blotting revealed the existence of 4 groups of monoclonal antibodies:

1. Monoclonal antibodies specific for a major linear epitope at the N-ter position (75-81, sequence: INTNSVP).

The representative of this group is antibody 156. The hybridoma producing this antibody was deposited at the Collection Nationale de Cultures de Microorganismes (CNCM) of the Institut Pasteur (Paris, France) on Dec. 1, 2004, under the number I-3331. This same epitope is also recognized by a rabbit serum (anti-N polyclonal) obtained by conventional immunization with the aid of this same N protein.

2. Monoclonal antibodies specific for a major linear epitope located in a central position (position 217-224, sequence: ETALALL); the representatives of this group are

the monoclonal antibodies 87 and 166. The hybridoma producing antibody 87 was deposited at the CNCM on Dec. 1, 2004, under the number I-3328.

3. Monoclonal antibodies specific for a major linear epitope located at the C-terminal position (position 403-408, sequence: DFFRQL), the representatives of this group are the antibodies 28, 57 and 143. The hybridoma producing antibody 57 was deposited at the CNCM on Dec. 1, 2004, under the number I-3330.

4. Monoclonal antibodies specific for a discontinuous conformational epitope. This group of antibodies does not rec-

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ognize any of the peptides spanning the sequence of the N protein, but react strongly on the non-denatured natural protein. The representative of this final group is the antibody 86. The hybridoma producing this antibody was deposited at the CNCM on Dec. 1, 2004, under the number I-3329.

Table VIII below summarizes the epitope mapping results obtained:

TABLE VIII

Epitope mapping of the monoclonal antibodies			
Antibody	Epitope	Position	Region
28	DFSRQL Q	403 . . . 408	C - Ter.
143	DFSRQL Q		
76	DFSRQL Q		
57	DFSRQL Q		
	FFGMS RI	315 . . . 319	
146	LPQRQ	383 . . . 387	
166	ETALALLLL	217 . . . 224	central
87	ETALALL	217 . . . 224	
156	INTNSGP	75 . . . 81	N-Ter.
86	Conformational		
212	Conformational		
1170	Conformational		

In addition, as illustrated in particular in FIGS. 18 and 19, these antibodies exhibit no reactivity in ELISA and/or in WB toward the N protein of the human corona-virus 229 E.

EXAMPLE 10

Combinations of the Monoclonal Antibodies for the Development of a Sensitive Immunocapture Test Specific for the Viral N Antigen in the Serum or Biological Fluids of Patients Infected with the SARS-CoV Virus

The antibodies listed below were selected because of their very specific properties for an additional capture and detection study of the viral N protein, in the serum of the subjects or patients.

These antibodies were produced in ascites on mice, purified by affinity chromatography and used alone or in combination, as capture antibodies and as signal antibodies.

List of the antibodies selected:

Ab anti-C-ter region (No. 28, 57, 143)

Ab anti-central region (No. 87, 166)

Ab anti-N-ter region (No. 156)

Ab anti-discontinuous conformational epitope (86)

1) Preparation of the Reagents:

a) Immunocapture ELISA Plates

The plates are sensitized with the antibody solutions at 5 µg/ml in 0.1 M carbonate buffer, pH 9.6. The (monovalent or plurivalent) solutions are deposited in a volume of 100 µl in the wells and incubated overnight at room temperature. These plates are then washed with PBS buffer (10 mM pH 7.4 supplemented with 0.1% Tween 20) and then saturated with a PBS solution supplemented with 0.3% BSA and 5% sucrose).

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The plates are then dried and then packaged in a bag in the presence of a desiccant. They are ready to use.

b) Conjugates

The purified antibodies were coupled with peroxidase according to the Nakane protocol (Nakane et al.—1974, J. of Histo and cytochemistry, vol. 22, pp. 1084-1091) in a ratio of one molecule of IgG per 3 molecules of peroxidase. These conjugates were purified by exclusion chromatography and stored concentrated (concentration between 1 and 2 mg/ml) in the presence of 50% glycerol and at -20° C. They are diluted for their use in the assays at the final concentration of 1 or 2 µg/ml in PBS buffer (pH 7.4) supplemented with 1% BSA.

c) Other Reagents

Human sera negative for all the serum markers for the HIV, HBV, HCV and THLV viruses

Pool of negative human sera supplemented with 0.5% Triton X 100

Inactivated viral Ag: viral culture supernatant inactivated by irradiation and inactivation verified after placing in culture on sensitive cells—titer of the suspension before inactivation about 10⁷ infectious particles per ml or alternatively about 5×10⁹ physical viral particles per ml of antigen

The Ag samples diluted in negative human serum: these samples were prepared by diluting 1:100 and then by 5-fold serial dilution.

These noninfectious samples mimic human samples thought to contain low to very low concentrations of viral nucleoprotein N. Such samples are not available for routine work.

Washing solution R2, solution for visualization TMB R8, chromogen R9 and stop solution R10, are the generic reagents marketed by Bio-Rad in its ELISA kits (e.g.: Platelia *pylori* kit ref. 72778).

2) Procedure

The samples of human sera overloaded with inactivated viral Ag are distributed in an amount of 100 µl per well, directly in the ready-to-use sensitized plates, and then incubated for 1 hour at 37° C. (Bio-Rad IPS incubation).

The material not bound to the solid phase is removed by 3 washings (washing with dilute R2 solution, automatic LP 35 washer).

The appropriate conjugates, diluted to the final concentration of 1 or 2 µg/ml, are distributed in an amount of 100 µl per well and the plates are again incubated for one hour at 37° C. (IPS incubation).

The excess conjugate is removed by 4 successive washings (dilute R2 solution—LP 35 washer).

The presence of conjugate attached to the plates is visualized after adding 100 µl of visualization solution prepared before use (1 ml of R9 and 10 ml of R8) and after incubation for 30 minutes, at room temperature and protected from light.

The enzymatic reaction is finally blocked by adding 100 µl of R10 reagent (1 N H₂SO₄) to all the wells.

The reading is carried out with the aid of an appropriate microplate reader at double wavelength (450/620 nm).

The results can be interpreted by using, as provisional threshold value, the mean of at least two negative controls multiplied by a factor of 2 or alternatively the mean of 100 negative sera supplemented with an increment corresponding to 6 SD (standard deviation calculated on the 100 individual measurements).

3) Results

Various capture antibody and signal antibody combinations were tested based on the properties of the antibodies

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selected, and avoiding the combinations of antibodies specific for the same epitopes in solid phase and as conjugates.

The best results were obtained with the 4 combinations listed below. These results are reproduced in table IX below.

1. Combination F/28

Solid phase (Ab 166+87 central region): conjugate antibody 28 (C-ter)

2. Combination G/28

Solid phase (Ab 86—conformational epitope): conjugate antibody 28 (C-ter)

3. Combination H/28

Solid phase (Ab 86, 166 and 87 central region and conformational epitope): conjugate antibody 28 (C-ter)

4. Combination H/28+87

Solid phase (Ab 86, 166 and 87 central region and conformational epitope): mixed conjugate antibodies 28 (C-ter) and 87 (central)

5. Combination G/87

Solid phase (Ab 86—conformational epitope): conjugate antibody 87 (central region)

The first 4 combinations exhibit equivalent and reproduced performance levels, greater than the other combinations used such as for example the combination G/87). Of course, in these combinations, a monoclonal antibody may be replaced with another antibody recognizing the same epitope. Thus, the following variants may be mentioned:

6. Variant of the combination F/28

Solid phase (Ab 87 only): conjugate antibody 57 (C-ter)

7. Variant of the combination G/28

Solid phase (Ab 86—conformational epitope): conjugate antibody 57 (C-ter)

8. Variant of the combination H/28

Solid phase (Ab 86 and 87 central region and conformational epitope): conjugate antibody 57 (C-ter)

9. Variant of the combination H/28+87

Solid phase (Ab 86 and 87 central region and conformational epitope): mixed conjugate antibodies 57 (C-ter) and 87 (central)

TABLE IX

Test of immunoreactivity of the anti-SARS-CoV nucleoprotein Abs: optical densities measured with each combination of antibodies according to the dilutions of the inactivated viral antigen.						
No.	Dilution	F/28	G/28	G/87	H/28	H/28 + 87
0	1/100	5	5	3.495	3.900	5
1	1/500	3.795	3.814	1.379	3.702	3.804
2	1/2 500	2.815	2.950	0.275	3.268	2.680
3	1/12 500	0.987	1.038	0.135	1.374	0.865
4	1/62 500	0.404	0.348	0.125	0.480	0.328
5	1/312 500	0.285	0.211	0.123	0.240	0.215
6	Control	0.210	0.200	0.098	0.186	0.156
7	Control	0.269	0.153	0.104	0.193	0.202

The detection limit for these 4 experimental trials corresponds to the antigen dilution in negative serum 1:62 500. A rapid extrapolation suggests the detection of less than 10^3 infectious particles per ml of sera.

From this study, it is evident that the most appropriate antibodies for the capture of the native viral nucleoprotein are the antibodies specific for the central region and/or for a conformational epitope, both being antibodies also selected for their high affinity for the native antigen.

Having determined the best antibodies for the composition of the solid phase, the antibodies to be selected as a priority for the detection of the antigens attached to the solid phase are the complementary antibodies specific for a dominant epitope

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in the C-ter region. The use of any other complementary antibody specific for epitopes located in the N-ter region of the protein leads to average or poor results.

EXAMPLE 11

Eukaryotic Expression Systems for the SARS-Associated Coronavirus (SARS-CoV) Spicule (S) Protein

1) Optimization of the Conditions for Expression of the SARS-CoV S in Mammalian Cells

The conditions for transient expression of the SARS-CoV spicule (S) protein were optimized in mammalian cells (293T, VeroE6).

For that, a DNA fragment containing the cDNA for SARS-CoV S was amplified by PCR with the aid of the oligonucleotides 5'-ATAGGATCCA CCATGTTTAT TTTCT-TATTA TTTCTTACTC TCACT-3' and 5'-ATACTCGAGTT ATGTGTAATG TAATTTGACA CCCTTG-3' from the plasmid pSARS-S (C.N.C.M. No. I-3059) and then inserted between the BamH1 and Xho1 sites of the plasmid pTRIPAU3-CMV containing a lentiviral vector TRIP (Sirven, 2001, Mol. Ther., 3, 438-448) in order to obtain the plasmid pTRIP-S. The BamH1 and Xho1 fragment containing the cDNA for S was then subcloned between BamH1 and Xho1 of the eukaryotic expression plasmid pcDNA3.1(+) (Clontech) in order to obtain the plasmid pcDNA-S. The Nhe1 and Xho1 fragment containing the cDNA for S was then subcloned between the corresponding sites of the expression plasmid pCI (Promega) in order to obtain the plasmid pCI-S. The WPRE sequences of the woodchuck hepatitis virus ("Woodchuck Hepatitis Virus posttranscriptional regulatory element") and the CTE sequences ("constitutive transport element") of the simian retro-virus from Mason-Pfizer were inserted into each of the two plasmids pcDNA-S and pCI-S between the Xho1 and Xba1 sites in order to obtain respectively the plasmids pcDNA-S-CTE, pcDNA-S-WPRE, pCI-S-CTE and pCI-S-WPRE (FIG. 21). The plasmid pCI-S-WPRE was deposited at the CNCM, on Nov. 22, 2004, under the number I-3323. All the inserts were sequenced with the aid of a BigDye Terminator v1.1 kit (Applied Biosystems) and an automated sequencer ABI377.

The capacity of the plasmid constructs to direct the expression of SARS-CoV S in mammalian cells was assessed after transfection of VeroE6 cells (FIG. 22). In this experiment, monolayers of 5×10^5 VeroE6 cells in 35 mm Petri dishes were transfected with 2 μ g of plasmids pcDNA as control), pcDNA-S, pCI and pCI-S and 6 μ l of Fugene6 reagent according to the manufacturer's instructions (Roche). After 48 hours of incubation at 37° C. and under 5% CO₂, cellular extracts were prepared in loading buffer according to Laemmli, separated on 8% SDS polyacrylamide gel, and then transferred onto a PVDF membrane (BioRad). The detection of this immunoblot (Western blot) was carried out with the aid of an anti-S rabbit polyclonal serum (immune serum from the rabbit P11135; cf. example 4 above) and donkey polyclonal antibodies directed against rabbit IgGs and coupled with peroxidase (NA934V, Amersham). The bound antibodies were visualized by luminescence with the aid of the ECL+ kit (Amersham) and autoradiography films Hyperfilm MP (Amersham).

This experiment (FIG. 22) shows that the plasmid pcDNA-S does not make it possible to direct the expression of SARS-CoV S at detectable levels whereas the plasmid pCI-S allows a weak expression, close to the limit of detection, which may be detected when the film is overexposed. Similar

results were obtained when the expression of S was sought by immunofluorescence (data not shown). This impossibility to detect effective expression of S cannot be attributed to the detection techniques used since the S protein can be detected at the expected size (180 kDa) in an extract of cells infected with SARS-CoV or in an extract of VeroE6 cells infected with the recombinant vaccinia virus VV-TF7.3 and transfected with the plasmid pcDNA-S. In this latter experiment, the virus VV-TF7.3 expresses the RNA polymerase of the T7 phage and allows the cytoplasmic transcription of an uncapped RNA capable of being efficiently translated. This experiment suggests that the expression defects described above are due to an intrinsic inability of the cDNA for S to be efficiently expressed when the step for transcription to messenger RNA is carried out at the nuclear level.

In a second experiment, the effect of the CTE and WPRE signals on the expression of S was assessed after transfection of VeroE6 (FIG. 23A) and 293T (FIG. 23B) cells and according to a protocol similar to that described above. Whereas the expression of S cannot be detected after transfection of the plasmids pcDNA-S-CTE and pcDNA-S-WPRE derived from pcDNA-S, the insertion of the WPRE and CTE signals greatly improves the expression of S in the context of the expression plasmid pCI-S.

To specify this result, a second series of experiments were carried out where the immunoblot is quantitatively visualized by luminescence and acquisition on a digital imaging device (FluorS, BioRad). The analysis of the results obtained with the QuantityOne v4.2.3 software (BioRad) shows that the WPRE and CTE sequences increase respectively the expression of S by a factor of 20 to 42 and 10 to 26 in Vero E6 cells (table X). In 293T cells (table X), the effect of the CTE sequence is more moderate (4 to 5 times) whereas that of the WPRE sequence remains high (13 to 22 times).

TABLE X

Quantitative analysis of the effect of the CTE and WPRE signals on the expression of SARS-CoV S:			
Plasmid	cell	exp. 1	exp. 2
PCI	VeroE6	0.0	0.0
pCI-S	VeroE6	1.0 ± 0.1	1.0
pCI-S-CTE	VeroE6	9.8 ± 0.9	26.4
pCI-S-WPRE	VeroE6	20.1 ± 2.0	42.3
PCI	293T	0.0	0.0
pCI-S	293T	1.0	1.0
pCI-S-CTE	293T	4.6	4.0
pCI-S-WPRE	293T	27.6	12.8

Cellular extracts were prepared 48 hours after transfection of VeroE6 or 293T cells with the plasmid pCI, pCI-S, pCI-S-CTE and pCI-S-WPRE and analyzed by Western blotting as described in the legend to FIG. 22. The Western blot is visualized by luminescence (ECL+, Amersham) and acquisition on a digital imaging device (FluorS, BioRad). The expression levels are indicated according to an arbitrary scale where the value of 1 represents the level measured after transfection of the plasmid pCI-S. Two independent experiments were carried out for each of the two cell types. In experiment 1 on VeroE6 cells, the transfections were carried out in duplicate and the results are indicated in the form of the mean and standard deviation values for the expression levels measured.

In summary, all these results show that the expression, in mammalian cells, of the cDNA for the SARS-CoV S under the control of the RNA polymerase II promoter sequences requires, to be efficient, the expression of a splice signal and of either of the sequences WPRE and CTE.

2) Production of Stable Lines Allowing the Expression of SARS-CoV S

The cDNA for the SARS-CoV S protein was cloned in the form of a BamHI-XhoI fragment into the plasmid pTRIPAU3-CMV containing a defective lentiviral vector TRIP with central DNA flap (Sirven et al., 2001, Mol. Ther., 3: 438-448) in order to obtain the plasmid pTRIP-S (FIG. 24).

Transient cotransfection according to Zennou et al. (2000, Cell, 101: 173-185) of this plasmid, of an encapsidation plasmid (p8.2) and of a plasmid for expression of the VSV envelope glycoprotein, G (pHCMV-G) in 293T cells allowed the preparation of retroviral pseudoparticles containing the vector TRIP-S and pseudotyped with the envelope protein G. These pseudotyped TRIP-S vectors were used to translate 293T and FRhK-4 cells: no expression of the S protein could be detected by Western blotting and immunofluorescence in the transduced cells (data not presented).

The optimum expression cassettes consisting of the CMV virus immediate/early promoter, a splice signal, cDNA for S and either of the posttranscriptional signals WPRE or CTE described above were then substituted for the EF1 α -EGFP cassette of the defective lentiviral expression vector with central DNA flap TRIPAU3-EF1 α (Sirven et al., 2001, Mol. Ther., 3: 438-448) (FIG. 25). These substitutions were carried out by a series of successive subclonings of the S expression cassettes which were excised from the plasmids pCT-S-CTE (BglII-ApaI) or respectively pCI-S-WPRE (BglII-SalI) and then inserted between the MluI and KpnI sites or respectively MluI or XhoI sites of the plasmid TRIPAU3-EF1 α in order to obtain the plasmids pTRIP-SD/SA-S-CTE and pTRIP-SD/SA-S-WPRE, deposited at the CNCM, on Dec. 1, 2004, under the numbers I-3336 and I-3334, respectively. Pseudotyped vectors were produced according to Zennou et al. (2000, Cell, 101: 173-185) and used to transduce 293T cells (10 000 cells) and FRhK-4 cells (15 000 cells) according to a series of 5 successive transduction cycles with a quantity of vectors corresponding to 25 ng (TRIP-SD/SA-S-CTE) or 22 ng TRIP-SD/SA-S-WPRE) of p24 per cycle.

The transduced cells were cloned by limiting dilution and a series of clones were qualitatively analyzed for the expression of SARS-CoV S by immunofluorescence (data not shown), and then quantitatively by Western blotting (FIG. 25) with the aid of an anti-S rabbit polyclonal serum. The results presented in FIG. 25 show that clones 2 and 15 of FRhK4-S-CTE cells transduced with TRIP-SD/SA-S-CTE and clones 4, 9 and 12 of FRhK4-S-WPRE cells transduced with TRIP-SD/SA-S-WPRE allow the expression of the SARS-CoV S at respectively low, or moderate levels if they are compared to those which can be observed during infection with SARS-CoV.

In summary, the vectors TRIP-SD/SA-S-CTE and TRIP-SD/SA-S-WPRE allow the production of stable clones of FRhK-4 cells and similarly 293T cells expressing SARS-CoV S, whereas the assays carried out with the "parent" vector TRIP-S remained unsuccessful, which demonstrates the need for a splice signal and for either of the sequences CTE and WPRE for the production of stable cell clones expressing the S protein.

In addition, these modifications of the vector TRIP (insertion of a splice signal and of a post-transcriptional signal like CTE and WPRE) could prove advantageous for improving the expression of other cDNAs than that for S.

3) Production of Stable Lines Allowing the Expression of a Soluble Form of SARS-CoV S. Purification of this Recombinant Antigen.

A cDNA encoding a soluble form of the S protein (Ssol) was obtained by fusing the sequences encoding the ectodomain of the protein (amino acids 1 to 1193) with those of a tag (FLAG: DYKDDDDK) via a BspE1 linker encoding the SG dipeptide. Practically, in order to obtain the plasmid pcDNA-Ssol, a DNA fragment encoding the ectodomain of SARS-CoV S was amplified by PCR with the aid of the oligonucleotides 5'-ATAGGATCCA CCAATGTTTAT TTTCTTATTA TTCTTACTC TCACT-3' and 5'-ACCTC-

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CGGAT TTAATATATT GCTCATATTT TCCCAA-3' from the plasmid pcDNA-S, and then inserted between the unique BamHI and BspEI sites of a modified eukaryotic expression plasmid pcDNA3.1(+) (Clontech) containing the tag sequence FLAG between its BamHI and XhoI sites:

```
// GGATCC ...nnn... TCC GGA GAT TAT AAA GAT GAC
   BamHI           S  G  D  Y  K  D  D

GAC GAT AAA TAA CTCGAG //
  D  D  K  ter XhoI
```

The NheI-XhoI and BamHI-XhoI fragments, containing the cDNA for S, were then excised from the plasmid pcDNA-Ssol, and subcloned between the corresponding sites of the plasmid pTRIP-SD/SA-S-CTE and of the plasmid pTRIP-SD/SA-S-WPRE, respectively, in order to obtain the plasmids pTRIP-SD/SA-Ssol-CTE and pTRIP-SD/SA-Ssol-WPRE, deposited at the CNM, on Dec. 1, 2004, under the numbers I-3337 and I-3335, respectively.

Pseudotyped vectors were produced according to Zennou et al. (2000, Cell, 101:173-185) and used to transduce FRhK-4 cells (15 000 cells) according to a series of 5 successive transduction cycles (15 000 cells) with a quantity of vector corresponding to 24 ng (TRIP-SD/SA-Ssol-CTE) or 40 ng (TRIP-SD/SA-Ssol-WPRE) of p24 per cycle. The transduced cells were cloned by limiting dilution and a series of 16 clones transduced with TRIP-SD/SA-Ssol-CTE and of 15 clones with TRIP-SD/SA-Ssol-WPRE were analyzed for the expression of the Ssol polypeptide by Western blotting visualized with an anti-FLAG monoclonal antibody (FIG. 26 and data not presented), and by capture ELISA specific for the Ssol polypeptide which was developed for this purpose (table XI and data not presented). Part of the process for selecting the best secretory clones is shown in FIG. 26. Capture ELISA is based on the use of solid phases coated with polyclonal antibodies of rabbits immunized with purified and inactivated SARS-CoV. These solid phases allow the capture of the Ssol polypeptide secreted into the cellular supernatants, whose presence is then visualized with a series of steps successively involving the attachment of an anti-FLAG monoclonal antibody (M2, SIGMA), of anti-mouse IgG(H+L) biotinylated rabbit polyclonal antibodies (Jackson) and of a streptavidin-peroxidase conjugate (Amersham) and then the addition of chromogen and substrate (TMB+H₂O₂, KPL).

TABLE XI

Analysis of the expression of the Ssol polypeptide by cell lines transduced with the lentiviral vectors TRIP-SD/SA-Ssol-WPRE and TRIP-SD/SA-Ssol-CTE.		
Vector	Clone	OD (450 nm)
Control	—	0.031
TRIP-SD/SA-Ssol-CTE	CTE2	0.547
	CTE3	0.668
	CTE9	0.171
	CTE12	0.208
	CTE13	0.133
TRIP-SD/SA-Ssol-WPRE	WPRE1	0.061
	WPRE10	0.134

The secretion of the Ssol polypeptide was assessed in the supernatant of a series of cell clones isolated after transduction of FRhK-4 cells with the lentiviral vectors TRIP-SD/SA-Ssol-WPRE and TRIP-SD/SA-Ssol-CTE. The supernatants diluted 1/50 were analyzed by a capture ELISA test specific for SARS-CoV S.

The cell line secreting the highest quantities of Ssol polypeptide in the culture supernatant is the FRhK4-Ssol-CTE3 line. It was subjected to a second series of 5 cycles of

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transduction with the vector TRIP-SD/SA-Ssol-CTE under conditions similar to those described above and then cloned. The subclone secreting the highest quantities of Ssol was selected by a combination of Western blot and capture ELISA analysis: it is the subclone FRhK4-Ssol-30, which was deposited at the CNM, on Nov. 22, 2004, under the name I-3325.

The FRhK4-Ssol-30 line allows the quantitative production and purification of the recombinant Ssol polypeptide. In a typical experiment where the experimental conditions for growth, production and purification were optimized, the cells of the FRhK4-Ssol-30 line are inoculated in standard culture medium (pyruvate-free DMEM containing 4.5 g/l of glucose and supplemented with 5% FCS, 100 U/ml of penicillin and 100 µg/ml of streptomycin) in the form of a subconfluent monolayer (1 million cells per each 100 cm² in 20 ml of medium). At confluence, the standard medium is replaced with the secretion medium where the quantity of FCS is reduced to 0.5% and the quantity of medium reduced to 16 ml per each 100 cm². The culture supernatant is removed after 4 to 5 days of incubation at 35° C. and under 5% CO₂. The recombinant polypeptide Ssol is purified from the supernatant by the succession of steps of filtration on 0.1 µm polyethersulfone (PES) membrane, concentration by ultrafiltration on a PES membrane with a 50 kD cut-off, affinity chromatography on anti-FLAG matrix with elution with a solution of FLAG peptide (DYKDDDDK) at 100 µg/ml in TBS (50 mM tris, pH 7.4, 150 mM NaCl) and then gel filtration chromatography in TBS on sephadex G-75 beads (Pharmacia). The concentration of the purified recombinant Ssol polypeptide was determined by micro-BCA test (Pierce) and then its biochemical characteristics analyzed.

Analysis by 8% SDS acrylamide gel stained with silver nitrate demonstrates a predominant polypeptide whose molecular mass is about 180 kD and whose degree of purity may be evaluated at 98% (FIG. 27A). Two main peaks are detected by SELDI-TOF mass spectrometry (Cypherger): they correspond to single and double charged forms of a predominant polypeptide whose molecular mass is thus determined at 182.6±3.7 kD (FIGS. 27B and C). After transfer onto Prosorb membrane and rinsing in 0.1% TFA, the N-terminal end of the Ssol polypeptide was sequenced in liquid phase by Edman degradation on 5 residues (ABI494, Applied Biosystems) and determined as being SLDLDR (FIG. 27D). This demonstrates that the signal peptide located at the N-terminal end of the SARS-CoV S protein, composed of aa 1 to 13 (MFIFLLFLTLTSG) according to an analysis carried out with the software signalP v2.0 (Nielsen et al., 1997, Protein Engineering, 10:1-6), is cleaved from the mature Ssol polypeptide. The recombinant Ssol polypeptide therefore consists of amino acids 14 to 1193 of the SARS-CoV S protein fused at the C-terminals with a sequence SG DYKDDDDK containing the sequence of the FLAG tag (underlined). The difference between the theoretical molar mass of the naked Ssol polypeptide (132.0 kD) and the real molar mass of the mature polypeptide (182.6 kD) suggests that the Ssol polypeptide is glycosylated.

A preparation of purified Ssol polypeptide, whose protein concentration was determined by micro-BCA test, makes it possible to prepare a calibration series in order to measure, with the aid of the capture ELISA test described above, the concentrations of Ssol present in the culture supernatants and to review the characteristics of the secretory lines. According to this test, the FRhK4-Ssol-CT3 line secretes 4 to 6 µg/ml of polypeptide Ssol while the FRhK4-Ssol-30 line secretes 9 to 13 µg/ml of Ssol after 4 to 5 days of culture at confluence. In addition, the purification scheme presented above makes it

possible routinely to purify from 1 to 2 mg of Ssol polypeptide per liter of culture supernatant.

EXAMPLE 12

Gene Immunization Involving the SARS-Associated Corona Virus (SARS-CoV) Spicule (S) Protein

The effect of a splice signal and of the posttranscriptional signals WPRE and CTE was analyzed after gene immunization of BALB/c mice (FIG. 28).

For that, BALB/c mice were immunized at intervals of 4 weeks by injecting into the tibialis anterior a saline solution of 50 µg of plasmid DNA of pcDNA-S and pCI-S and, as a control, 50 µg of plasmid DNA of pcDNA-N (directing the expression of SARS-CoV N) or of pCI-HA (directing the expression of the HA of the influenza virus A/PR/8/34) and the immune sera collected 3 weeks after the 2nd injection. The presence of antibodies directed against the SARS-CoV S was assessed by indirect ELISA using as antigen a lysate of VeroE6 cells infected with SARS-CoV and, as a control, a lysate of noninfected VeroE6 cells. The anti-SARS-CoV antibody titers (TI) are calculated as the reciprocal of the dilution producing a specific OD of 0.5 (difference between OD measured on a lysate of infected cells and OD measured on a lysate of noninfected cells) after visualization with an anti-mouse IgG polyclonal antibody coupled with peroxidase (NA931V, Amersham) and TMB supplemented with H₂O₂ (KPL) (FIG. 28A).

Under these conditions, the expression plasmid pcDNA-S only allows the induction of low antibody titers directed against SARS-CoV S in 3 mice out of 6 ($\text{LOG}_{10}(\text{TI})=1.9\pm0.6$) whereas the plasmid pcDNA-N allows the induction of anti-N antibodies at high titers ($\text{LOG}_{10}(\text{TI})=3.9\pm0.3$) in all the animals, and the control plasmids (pCI, pCI-HA) do not result in any detectable antibody ($\text{LOG}_{10}(\text{TI})<1.7$). The plasmid pCI-S equipped with a splice signal allows the induction of antibodies at high titers ($\text{LOG}_{10}(\text{TI})=3.7\pm0.2$), which are approximately 60 times higher than those observed after injection of the plasmid pcDNA-S ($p<10^{-5}$).

The efficiency of the posttranscriptional signals was studied by carrying out a dose-response study of the anti-S antibody titers induced in the BALB/c mouse as a function of the quantity of plasmid DNA used as immunogen (2 µg, 10 µg and 50 µg). This study (FIG. 28B) demonstrates that the posttranscriptional signal WPRE greatly improves the efficiency of gene immunization when small doses of DNA are used ($p<10^{-5}$ for a dose of 2 µg of DNA and $p<10^{-2}$ for a dose of 10 µg), whereas the effect of the CTE signal remains marginal ($p=0.34$ for a dose of 2 µg of DNA).

Finally, the antibodies induced in mice after gene immunization neutralize the infectivity of SARS-CoV in vitro (FIGS. 29A and 29B) at titers which are consistent with the titers measured by ELISA.

In summary, the use of a splice signal and of the posttranscriptional signal WPRE of the woodchuck hepatitis virus considerably improves the induction of neutralizing antibodies directed against SARS-CoV after gene immunization with the aid of plasmid DNA directing the expression of the cDNA for SARS-CoV S.

EXAMPLE 13

Diagnostic Applications of the S Protein

The ELISA reactivity of the recombinant Ssol polypeptide was analyzed with respect to sera from patients suffering from SARS.

The sera from probable cases of SARS tested were chosen on the basis of the results (positive or negative) of analysis of their specific reactivity toward the native antigens of SARS-CoV by immunofluorescence test on VeroE6 cells infected with SARS-CoV and/or by indirect ELISA test using as antigen a lysate of VeroE6 cells infected with SARS-CoV. The sera of these patients are identified by a serial number of the National Reference Center for Influenza Viruses and by the initials of the patient and the number of days elapsed since the onset of the symptoms. All the sera of probable cases (cf. Table XII) recognize the native antigens of SARS-CoV, with the exception of the serum 032552 of the patient VTT for whom infection with SARS-CoV could not be confirmed by RT-PCR performed on respiratory samples of days 3, 8 and 12. A panel of control sera was used as control (TV sera): they are sera collected in France before the SARS epidemic that occurred in 2003.

TABLE XII

Sera of probable cases of SARS		
Serum	Patient	Sample collection day
031724	JYK	7
033168	JYK	38
033597	JYK	74
032632	NTM	17
032634	THA	15
032541	PHV	10
032542	NIH	17
032552	VTT	8
032633	PTU	16
032791	JLB	3
033258	JLB	27
032703	JCM	8
033153	JCM	29

Solid phases sensitized with the recombinant Ssol polypeptide were prepared by adsorption of a solution of purified Ssol polypeptide at 2 µg/ml in PBS in the wells of an ELISA plate, and then the plates are incubated overnight at 4° C. and washed with PBS-Tween buffer (PBS, 0.1% Tween 20). After saturating the ELISA plates with a solution of PBS-10% skimmed milk (weight/volume) and washing in PBS-Tween, the sera to be tested (100 µl) are diluted 1/400 in PBS skimmed milk-Tween buffer (PBS, 3% skimmed milk, 0.1% Tween) and then added to the wells of the sensitized ELISA plate. The plates are incubated for 1 h at 37° C. After 3 washings with PBS-Tween buffer, the anti-human IgG conjugate labeled with peroxidase (ref. NA933V, Amersham) diluted 1/4000 in PBS-skimmed milk-Tween buffer is added, and then the plates are incubated for 1 hour at 37° C. After 6 washings with PBS-Tween buffer, the chromogen (TMB) and the substrate (H₂O₂) are added and the plates are incubated for 10 minutes protected from light. The reaction is stopped by adding a 1 N H₃PO₄ solution, and then the absorbance is measured at 450 nm with a reference at 620 nm.

The ELISA tests (FIG. 30) demonstrate that the recombinant Ssol polypeptide is specifically recognized by the serum antibodies of patients suffering from SARS collected at the medium or late phase of infection (≥ 10 days after the onset of the symptoms) whereas it is not significantly recognized by the serum antibodies of 2 patients (JLB and JCM) collected in the early phase of infection (3 to 8 days after the onset of the symptoms) or by control sera of subjects not suffering from SARS. The serum antibodies of patients JLB and JCM show a seroconversion between days 3 and 27 for the first and 8 and

29 for the second after the onset of the symptoms, which confirms the specificity of the reactivity of these sera toward the Ssol polypeptide.

In conclusion, these results demonstrate that the recombinant Ssol polypeptide may be used as an antigen for the development of an ELISA test for serological diagnosis of infection with SARS-CoV.

EXAMPLE 14

Vaccine Applications of the Recombinant Soluble S Protein

The immunogenicity of the recombinant Ssol polypeptide was studied in mice.

For that, a group of 6 mice was immunized at 3 weeks' interval with 10 µg of recombinant Ssol polypeptide adjuvanted with 1 mg of aluminum hydroxide (Alu-gel-S, Serva) diluted in PBS. Three successive immunizations were performed and the immune sera were collected 3 weeks after each of the immunizations (IS1, IS2, IS3). As a control, a group of mice (mock group) received aluminum hydroxide alone according to the same protocol.

The immune sera were analyzed per pool for each of the 2 groups by indirect ELISA using a lysate of VeroE6 cells infected with SARS-CoV as antigen and as a control a lysate of noninfected VeroE6 cells. The anti-SARS-CoV antibody titers are calculated as the reciprocal of the dilution producing a specific OD of 0.5 after visualization with an anti-mouse IgG(H+L) polyclonal antibody coupled with peroxidase (NA931V, Amersham) and TMB supplemented with H₂O₂ (KPL). This analysis (FIG. 31) shows that the immunization with the Ssol polypeptide induces in mice, from the first immunization, antibodies directed against the native form of the SARS-CoV spicule protein present in the lysate of infected VeroE6 cells. After 2 then 3 immunizations, the anti-S antibody titers become very high.

The immune sera were analyzed per pool for each of the two groups for their capacity to seroneutralize the infectivity of SARS-CoV. 4 points of seroneutralization on FRhK-4 cells (100 TCID₅₀ of SARS-CoV) are produced for each of the 2-fold dilutions tested from 1/20. The seroneutralizing titer is calculated according to the Reed and Munsch method as the reciprocal of the dilution neutralizing the infectivity of 2 wells out of 4. This analysis shows that the antibodies induced in mice by the Ssol polypeptide are neutralizing: the titers observed are very high after 2 and then 3 immunizations (greater than 2560 and 5120 respectively, table XIII).

TABLE XIII

Induction of antibodies directed against SARS-CoV after immunization with the recombinant Ssol polypeptide.		
Group	Sera	Neutralizing Ab
Mock	pi	<20
	IS1	<20
	IS2	<20
	IS3	<20
Ssol	pi	<20
	IS1	57
	IS2	>2560
	IS3	>5120

The immune sera were analyzed per pool for each of the two groups for their capacity to seroneutralize the infectivity of 100 TCID₅₀ of SARS-CoV on FRhK-4 cells. 4 points are produced for each of the 2-fold dilutions tested from 1/20. The seroneutralizing titer is calculated according to the Reed and Munsch method as the reciprocal of the dilution neutralizing the infectivity of 2 wells out of 4.

The neutralizing titers observed in mice immunized with the Ssol polypeptide reach levels far greater than the titers observed by Yang et al. in mice (2004, Nature, 428:561-564) and those observed by Buchholz in the hamster (2004, PNAS 101:9804-9809) which protect respectively mice and hamsters from infection with SARS-CoV. It is therefore probable that the neutralizing antibodies induced in mice after immunization with the Ssol polypeptide protect these animals against infection with SARS-CoV.

EXAMPLE 15

Optimized Synthetic Gene for the Expression in Mammalian Cells of the SARS-Associated Coronavirus (SARS-CoV) Spicule (S) Protein

1) Design of the Synthetic Gene

A synthetic gene encoding the SARS-CoV spicule protein was designed from the gene of the isolate 031589 (plasmid pSARS-S, C.N.C.M. No. I-3059) so as to allow high levels of expression in mammalian cells and in particular in cells of human origin.

For that:

the use of codons of the wild-type gene of the isolate 031589 was modified so as to become close to the bias observed in humans and to improve the efficiency of translation of the corresponding mRNA

the overall GC content of the gene was increased so as to extend the half-life of the corresponding mRNA

the optionally cryptic motifs capable of interfering with an efficient expression of the gene were deleted (splice donor and acceptor sites, polyadenylation signals, sequences very rich (>80%) or very low (<30%) in GC, repeat sequences, sequences involved in the formation of secondary RNA structures, TATA boxes)

a second STOP codon was added to allow efficient termination of translation.

In addition, CpG motifs were introduced into the gene so as to increase its immunogenicity as DNA vaccine. In order to facilitate the manipulation of the synthetic gene, two BamH1 and Xho1 restriction sites were placed on either side of the open reading frame of the S protein, and the BamH1, Xho1, Nhe1, Kpn1, BspE1 and Sal1 restriction sites were avoided in the synthetic gene.

The sequence of the synthetic gene designed (gene 040530) is given in SEQ ID No: 140.

An alignment of the synthetic gene 040530 with the sequence of the wild-type gene of the isolate 031589 of SARS-CoV deposited at the C.N.C.M. under the number I-3059 (SEQ ID No: 4, plasmid pSRAS-S) is presented in FIG. 32.

2) Plasmid Constructs

The synthetic gene SEQ ID No: 140 was assembled from synthetic oligonucleotides and cloned between the Kpn1 and Sac1 sites of the plasmid pUC-Kana in order to give the plasmid 040530pUC-Kana. The nucleotide sequence of the insert of the plasmid 040530pUC-Kana was verified by automated sequencing (Applied).

A Kpn1-Xho1 fragment containing the synthetic gene 040530 was excised from the plasmid 040530pUC-Kana and subcloned between the Nhe1 and Xho1 sites of the expression plasmic pCI (Promega) in order to obtain the plasmid pCI-SSYNTH, deposited at the CNCM on Dec. 1, 2004, under the number I-3333.

A synthetic gene encoding the soluble form of the S protein was then obtained by fusing the synthetic sequences encoding the ectodomain of the S protein (amino acids 1 to 1193) with

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those of the tag (FLAG:DYKDDDDK) via a linker BspE1 encoding the dipeptide SG. Practically, a DNA fragment encoding the ectodomain of the SARS-CoV S was amplified by PCR with the aid of the oligonucleotides 5'-ACTA GCTAGC GGATCCACCATGTTTCATCTT CCTG-3' and 5'-AGTATCCGGAC TTG ATGTACT GCTCGTACTTGC-3' from the plasmid 040530pUC-Kana, digested with NheI and BspE1 and then inserted between the unique NheI and BspE1 sites of the plasmid pCI-Ssol, to give the plasmid pCI-SCUBE, deposited at the CNCM on Dec. 1, 2004, under the number I-3332. The plasmids pCI-Ssol, pCI-Ssol-CTE, and pCI-Ssol-WPRE (deposited at the CNCM, on Nov. 22, 2004, under the number I-3324) had been previously obtained by subcloning the KpnI-XhoI fragment excised from the plasmid pcDNA-Ssol (see technical note of DI 2004-106) between the NheI and XhoI sites of the plasmids pCI, pCI-S-CTE and pCI-S-WPRE respectively.)

The plasmids pCI-Scube and pCI-Ssol encode the same recombinant Ssol polypeptide.

3) Results

The capacity of the synthetic gene encoding the S protein to efficiently direct the expression of the SARS-CoV S in mammalian cells was compared with that of the wild-type gene after transient transfection of primate cells (VeroE6) and of human cells (293T).

In the experiment presented in FIG. 33 and in table XIV, monolayers of 5×10^5 VeroE6 cells or 7×10^5 293T cells in 35 mm Petri dishes were transfected with 2 μ g of plasmids pCI (as control), pCI-S, pCI-S-CTE, pCI-S-WPRE and pCI-S-Ssynth and 6 μ l of Fugene6 reagent according to the manufacturer's instructions (Roche). After 48 hours of incubation at 37° C. and under 5% CO₂, cell extracts were prepared in loading buffer according to Laemmli, separated on 8% SDS polyacrylamide gel and then transferred onto a PVDF membrane (BioRad). The detection of this immunoblot (Western blot) was carried out with the aid of an anti-S rabbit polyclonal serum (immune serum of the rabbit P11135: cf example 4 above) and of donkey polyclonal antibodies directed against rabbit IgGs and coupled with peroxidase (NA934V, Amersham). The immunoblot was quantitatively visualized by luminescence with the aid of the ECL+ kit (Amersham) and acquisition on a digital imaging device (FluorS, BioRad).

The analysis of the results obtained with the software QuantityOne v4.2.3 (BioRad) shows that in this experiment, the plasmid pCI-Synth allows the transient expression of the S protein at high levels in the VeroE6 and 293T cells, whereas the plasmid pCI-S does not make it possible to induce expression at sufficient levels to be detected. The expression. Levels observed are of the order of twice as high as those observed with the plasmid pCI-S-WPRE.

TABLE XIV

Use of a synthetic gene for the expression of the SARS-CoV S.		
Plasmid	VeroE6	293T
pCI	0.0	0.0
pCI-S	≤0.1	≤0.1
pCI-S-CTE	0.5	≤0.1

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TABLE XIV-continued

Use of a synthetic gene for the expression of the SARS-CoV S.		
Plasmid	VeroE6	293T
pCI-S-WPRE	1.0	1.0
pCI-Ssynth	1.8	1.9

Cell extracts prepared 48 hours after transfection of VeroE6 or 293T cells with the plasmids pCI, pCI-S, pCI-S-CTE, pCI-S-WPRE and pCI-S-Ssynth were separated on 8% SDS acrylamide gel and analyzed by Western blotting with the aid of an anti-S rabbit polyclonal antibody and an anti-rabbit IgG(H + L) polyclonal antibody coupled with peroxidase (NA934V, Amersham). The Western blot is visualized by luminescence (ECL+, Amersham) and acquisition on a digital imaging device (FluorS, BioRad). The expression levels of the S protein were measured by quantifying the two predominant bands identified on the image (see FIG. 33) and are indicated according to an arbitrary scale where the value 1 represents the level measured after transfection of the plasmid pCI-S-WPRE.

In a second instance, the capacity of the synthetic gene Scube to efficiently direct the synthesis and the secretion of the Ssol polypeptide by mammalian cells was compared with that of the wild-type gene after transient transfection of hamster cells (BHK-21) and of human cells (293T).

In the experiment presented in table XV, monolayers of 6×10^5 BHK-21 cells and 7×10^5 293T cells in 35 mm Petri dishes were transfected with 2 μ g of plasmids pCI (as control), pCI-Ssol, pCI-Ssol-CTE, pCI-Ssol-WPRE and pCI-Scube and 6 μ l of Fugene6 reagent according to the manufacturer's instructions (Roche). After 48 hours of incubation at 37° C. and under 5% CO₂, the cellular supernatants were collected and quantitatively analyzed for the secretion of the Ssol polypeptide by a capture ELISA test specific for the Ssol polypeptide.

Analysis of the results shows that, in this experiment, the plasmid pCI-Scube allows the expression of the Ssol polypeptide at levels 8 times (BHK-21 cells) to 20 times (293T cells) higher than the plasmid pCI-Ssol. The levels of expression observed are of the order of twice (293T cells) to 5 times (BHK-21 cells) as high as those observed with the plasmid pCI-Ssol-WPRE.

TABLE XV

Use of a synthetic gene for the expression of the Ssol polypeptide.		
Plasmid	BHK	293T
pCI	<20	<20
pCI-Ssol	<20	56 ± 10
pCI-Ssol-CTE	<20	63 ± 8
pCI-Ssol-WPRE	28 ± 1	531 ± 15
pCI-Scube	152 ± 6	1140 ± 20

The supernatants were harvested 48 hours after transfection of BHK or 293T cells with the plasmids pCI, pCI-Ssol, pCI-Ssol-CTE, pCI-Ssol-WPRE and pCI-Scube and quantitatively analyzed for the secretion of the Ssol polypeptide by an ELISA test specific for the Ssol polypeptide. The transfections were carried out in duplicate and the results are presented in the form of means and standard deviations of the concentrations of Ssol polypeptide (ng/ml) measured in the supernatants.

In summary, these results show that the expression, in mammalian cells, of the synthetic gene 040530 encoding SARS-CoV S under the control of RNA polymerase II promoter sequences is much more efficient than that of the wild-type gene of the 031589 isolate. This expression is even more efficient than that directed by the wild-type gene in the presence of the WPRE sequences of the woodchuck hepatitis virus.

4) Applications

The use of the synthetic gene 040530 encoding SARS-CoV S or its Scube variant encoding the polypeptide Ssol is capable of advantageously replacing the wild-type gene in numerous applications where the expression of S is necessary at high levels. In particular in order to:

improve the efficiency of gene immunization with plasmids of the pCI-Ssynth or even pCI-Ssynth-CTE or pCI-Ssynth-WPRE type
 establish novel cell lines expressing higher quantities of the S protein or of the Ssol polypeptide with the aid of recombinant lentiviral vectors carrying the Ssynth gene or the Scube gene respectively
 improve the immunogenicity of the recombinant lentiviral vectors allowing the expression of the S protein or of the Ssol polypeptide
 improve the immunogenicity of live vectors allowing the expression of the S protein or of the Ssol polypeptide like recombinant vaccinia viruses or recombinant measles viruses (see examples 16 and 17 below)

EXAMPLE 16

Expression of the SARS-Associated Coronavirus
 (SARS-CoV) Spicule (S) Protein with the Aid of
 Recombinant Vaccinia Viruses

Vaccine Application

Application to the Production of a Soluble Form of the Spicule (S) Protein and Design of a Serological Test for SARS

1) Introduction

The aim of this example is to evaluate the capacity of recombinant vaccinia viruses (VV) expressing various SARS-associated coronavirus (SARS-CoV) antigens to constitute novel vaccine candidates against SARS and a means of producing recombinant antigens in mammalian cells.

For that, the inventors focused on the SARS-CoV spicule (S) protein which makes it possible to induce, after gene immunization in animals, antibodies neutralizing the infectivity of SARS-CoV, and a soluble and secreted form of this protein, the Ssol polypeptide, which is composed of the ectodomain (aa 1-1193) of S fused at its C-ter end with a tag FLAG (DYKDDDDK) via a BspE1 linker encoding the SG dipeptide. This Ssol polypeptide exhibits an antigenicity similar to that of the S protein and allows, after injection into mice in the form of a purified protein adjuvanted with aluminum hydroxide, the induction of high neutralizing antibody titers against SARS-CoV.

The various forms of the S gene were placed under the control of the promoter of the 7.5K gene and then introduced into the thymidine kinase (TK) locus of the Copenhagen strain of the vaccinia virus by double homologous recombination in vivo. In order to improve the immunogenicity of the recombinant vaccinia viruses, a synthetic late promoter was chosen in place of the 7.5K promoter, in order to increase the production of S and Ssol during the late phases of the viral cycle.

After having isolated the recombinant vaccinia viruses and verified their capacity to express the SARS-CoV S antigen, their capacity to induce in mice an immune response against SARS was tested. After having purified the Ssol antigen from the supernatant of infected cells, an ELISA test for serodiagnosis of SARS was designed, and its efficiency was evaluated with the aid of sera from probable cases of SARS.

2) Construction of the Recombinant Viruses

Recombinant vaccinia viruses directing the expression of the S glycoprotein of the 031589 isolate of SARS-CoV and of a soluble and secreted form of this protein, the Ssol polypeptide, under the control of the 7.5K promoter were obtained.

With the aim of increasing the levels of expression of S and Ssol, recombinant viruses in which the cDNAs for S and for Ssol are placed under the control of a late synthetic promoter were also obtained.

The plasmid pTG186poly is a transfer plasmid for the construction of recombinant vaccinia viruses (Kieny, 1986, Biotechnology, 4:790-795). As such, it contains the VV thymidine kinase gene into which the promoter of the 7.5K gene has been inserted followed by a multiple cloning site allowing the insertion of heterologous genes (FIG. 34A). The promoter of the 7.5K gene in fact contains a tandem of two promoter sequences that are respectively active during the early (P_E) and late (P_L) phases of the vaccinia virus replication cycle. The BamHI-XhoI fragments were excised from the plasmids pTRIP-S and pcDNA-Ssol respectively and inserted between the BamHI and SmaI sites of the plasmid pTG186poly in order to give the plasmids pTG-S and pTG-Ssol (FIG. 34A). The plasmids pTG-S and pTG-Ssol were deposited at the CNCM, on Dec. 2, 2004, under the numbers I-3338 and I-3339, respectively.

The plasmids pTN480, pTN-S and pTN-Ssol were obtained from the plasmids pTG186poly, pTG-S and pTG-Ssol respectively, by substituting the NdeI-PstI fragment containing the 7.5K promoter by a DNA fragment containing the synthetic late promoter 480, which was obtained by hybridization of the oligonucleotides 5'-TATGAGCCTT TTTTTTTTTT TTTTTTTGGC ATATAAATAG ACTCG-GCGCG CCATCTGCA-3' and 5'-GATGGCGCGCG-CGAGTCTATT TATATGCCAA AAAAAAAAAA AAAAAAAAAAGC TCA-3' (FIG. 34B). The insert was sequenced with the aid of a BigDye Terminator v1.1 kit (Applied Biosystems) and an automated sequencer ABI377. The sequence of the late synthetic promoter 480 as cloned into the transfer plasmids of the pTN series is indicated in FIG. 34C. The plasmids pTN-S and pTN-Ssol were deposited at the CNCM, on Dec. 2, 2004, under the numbers I-3340 and I-3341, respectively.

The recombinant vaccinia viruses were obtained, by double homologous recombination in vivo between the TK cassette of the transfer plasmids of the series pTG and pTN and the TK gene of the Copenhagen strain of the vaccinia virus according to a procedure described by Kieny et al. (1984, Nature, 312:163-166). Briefly, CV-1 cells are transfected with the aid of DOTAP (Roche) with genomic DNA of the Copenhagen strain of the vaccinia virus and each of the transfer plasmids of the pTG and pTN series described above, and then superinfected with the helper vaccinia virus VV-ts7 for 24 hours at 33° C. The helper virus is counter-selected by incubation at 40° C. for 2 days and then the recombinant viruses (TK- phenotype) selected by two cloning cycles under agar medium on 143Btk- cells in the presence of BuDr (25 µg/ml). The 6 viruses VV-TG, VV-TG-S, VV-TG-Ssol, VV-TN, VV-TN-S, and VV-TN-Ssol are respectively obtained with the aid of the transfer plasmids pTG186poly, pTG-S, pTG-Ssol, pTN480, pTN-Ssol. The viruses VV-TG and VV-TN do not express any heterologous gene and were used as TK- control in the experiments. The preparations of recombinant viruses were performed on monolayers of CV-1 or BHK-21 cells and the titer in plaque forming units (p.f.u) determined on CV-1 cells according to Earl and Moss (1998, Current Protocols in Molecular Biology, 16.16.1-16.16.13).

3) Characterization of the Recombinant Viruses

The expression of the transgenes encoding the S protein and the Ssol polypeptide was assessed by Western blotting.

Monolayers of CV-1 cells were infected at a multiplicity of 2 with various recombinant vaccinia viruses VV-TG, VV-TG-S, VV-TG-Ssol, VV-TN, VV-TN-S and VV-TN-Ssol. After 18 hours of incubation at 37° C. and under 5% CO₂, cellular

extracts were prepared in loading buffer according to Laemmli, separated on 8% SDS polyacrylamide gel and then transferred onto a PVDF membrane (BioRad). The detection of this immunoblot (Western blot) was performed with the aid of an anti-S rabbit polyclonal serum (immune serum from the rabbit P11135: cf. example 4) and donkey polyclonal antibodies directed against rabbit IgGs and coupled with peroxidase (NA934V, Amersham). The bound antibodies were visualized by luminescence with the aid of the ECL+ kit (Amersham) and autoradiography films Hyperfilm MP (Amersham).

As shown in FIG. 35A, the recombinant virus VV-TN-S directs the expression of the S protein at levels which are comparable to those which can be observed 8 h after infection with SARS-CoV but which are much higher than those which can be observed after infection with VV-TG-S. In a second experiment (FIG. 35B), the analysis of variable quantities of cellular extracts shows that the levels of expression observed after infection with viruses of the TN series (VV-TN-S and VV-TN-Ssol) are about 10 times as high as those observed with the viruses of the TG series (VV-TG-S and VV-TG-Ssol, respectively). In addition, the Ssol polypeptide is secreted into the supernatant of CV-1 cells infected with the VV-TN-Ssol virus more efficiently than in the supernatant of cells infected with VV-TG-Ssol (FIG. 36A). In this experiment, the VV-TN-Sflag virus was used as a control because it expresses the membrane form of the S protein fused at its C-ter end with the FLAG tag. The Sflag protein is not detected in the supernatant of cells infected with VV-TN-Sflag, demonstrating that the Ssol polypeptide is indeed actively secreted after infection with VV-TN-Ssol.

These results demonstrate that the recombinant vaccinia viruses are indeed carriers of the transgenes and allow the expression of the SRAS glycoprotein in its membrane form (S) or in a soluble or secreted form (Ssol). The vaccinia viruses carrying the synthetic promoter 480 allow the expression of S and the secretion of Ssol at levels much higher than the viruses carrying the promoter of the 7.5K gene.

4) Application to the Production of a Soluble Form of SARS-CoV S. Purification of this Recombinant Antigen and Diagnostic Applications

The BHK-21 line is the cell line which secretes the highest quantities of Ssol polypeptide after infection with the VV-TN-Ssol virus among the lines tested (BHK-21, CV1, 293T and FrhK-4, FIG. 36B); it allows the quantitative production and purification of the recombinant Ssol polypeptide. In a typical experiment where the experimental conditions for infection, production and purification were optimized, the BHK-21 cells are inoculated in standard culture medium (pyruvate-free DMEM containing 4.5 g/l of glucose and supplemented with 5% TPB, 5% FCS, 100 U/ml of penicillin and 100 µg/ml of streptomycin) in the form of a subconfluent monolayer (10 million cells for each 100 cm² in 25 ml of medium). After 24 h of incubation at 37° C. under 5% CO₂, the cells are infected at an M.O.I. of 0.03 and the standard medium replaced with the secretion medium where the quantity of FCS is reduced to 0.5% and the TPB eliminated. The culture supernatant is removed after 2.5 days of incubation at 35° C. and under 5% CO₂ and the vaccinia virus inactivated by addition of Triton X-100 (0.1%). After filtration on 0.1 µm polyethersulfone (PES) membrane, the recombinant Ssol polypeptide is purified by affinity chromatography on an anti-FLAG matrix with elution with a solution of FLAG peptide (DYKDDDDK) at 100 µg/ml in TBS (50 mM Tris, pH 7.4, 150 mM NaCl).

The analysis by 8% SDS acrylamide gel stained with silver nitrate identified a predominant polypeptide whose molecu-

lar mass is about 180 kD and whose degree of purity is greater than 90% (FIG. 37). The concentration of the purified Ssol recombinant polypeptide was determined by comparison with molecular mass markers and estimated at 24 ng/µl.

This purified Ssol polypeptide preparation makes it possible to produce a calibration series in order to measure, with the aid of a capture ELISA test, the Ssol concentrations present in the culture supernatants. According to this test, the BHK-21 line secretes about 1 µg/ml of Ssol polypeptide under the production conditions described above. In addition, the purification scheme presented makes it possible to purify of the order of 160 µg of Ssol polypeptide per liter of culture supernatant.

The ELISA reactivity of the recombinant Ssol polypeptide was analyzed toward sera from patients suffering from SARS.

The sera of probable cases of SARS tested were chosen on the basis of the results (positive or negative) of analysis of their specific reactivity toward the native antigens of SARS-CoV by immunofluorescence test on VeroE6 cells infected with SARS-CoV and/or by indirect ELISA test using, as antigen, a lysate of VeroE6 cells infected with SARS-CoV. The sera of these patients are identified by a serial number of the National Reference Center for Influenza Viruses and by the patient's initials and the number of days elapsed since the onset of the symptoms. All the sera of probable cases (cf. table XVI) recognize the native antigens of SARS-CoV with the exception of the serum 032552 of the patient VTT, for which infection with SARS-CoV could not be confirmed by RT-PCR performed on respiratory samples of days 3, 8 and 12. A panel of control sera was used as control (TV sera): they are sera collected in France before the SARS epidemic which occurred in 2003.

TABLE XVI

Sera of probable cases of SARS			
Serum	Patient	Sample collection day	
033168	JYK	38	
033597	JYK	74	
032632	NTM	17	
032634	THA	15	
032541	PHV	10	
032542	NIH	17	
032552	VTT	8	
032633	PTU	16	

Solid phases sensitized with the recombinant Ssol polypeptide were prepared by adsorption of a solution of purified Ssol polypeptide at 4 µg/ml in PBS in the wells of an ELISA plate. The plates are incubated overnight at 4° C. and then washed with PES-Tween buffer (PBS, 0.1% Tween 20). After washing with PBS-Tween, the sera to be tested (100 µl) are diluted 1/100 and 1/400 in PBS-skimmed milk-Tween buffer (PBS, 3% skimmed milk, 0.1% Tween) and then added to the wells of the sensitized ELISA plate. The plates are then incubated for 1 h at 37° C. After 3 washings with PBS-Tween buffer, the anti-human IgG conjugate labeled with peroxidase (ref. NA933V, Amersham) diluted 1/4000 in PBS-skimmed milk-Tween buffer is added and then the plates are incubated for one hour at 37° C. After 6 washings with PBS-Tween buffer, the chromogen (TMB) and the substrate (H₂O₂) are added and the plates are incubated for 10 minutes protected from light. The reaction is stopped by adding a 1M solution of H₃PO₄ and then the absorbance is measured at 450 nm with a reference at 620 nm.

The ELISA tests (FIG. 38) demonstrate that the recombinant Ssol polypeptide is specifically recognized by the serum

antibodies of patients suffering from SARS, collected at the middle or late phase of infection (≥ 10 days after the onset of the symptoms), whereas it is not significantly recognized by the serum antibodies of the control sera of subjects not suffering from SARS.

In conclusion, these results demonstrate that the recombinant Ssol polypeptide can be purified from the supernatant of mammalian cells infected with the recombinant vaccinia virus VV-TN-Ssol and can be used as antigen for developing an ELISA test for serological diagnosis of infection with SARS-CoV.

5. Vaccine Applications

The immunogenicity of the recombinant vaccinia viruses was studied in mice.

For that, groups of 7 BALB/c mice were immunized by the i.v. route twice at 4 weeks' interval with 10^6 p.f.u. of recombinant vaccinia viruses VV-TG, VV-TG-S, VV-TG-Ssol, VV-TN, VV-TN-S and VV-TN-Ssol and, as a control, VV-TG-HA which directs the expression of hemagglutinin of the A/PR/8/34 strain of the influenza virus. The immune sera were collected 3 weeks after each of the immunizations (IS1, IS2).

The immune sera were analyzed per pool for each of the groups by indirect ELISA using a lysate of VeroE6 cells infected with SARS-CoV as antigen and, as control, a lysate of noninfected VeroE6 cells. The anti-SARS-CoV antibody titers (TI) are calculated as the reciprocal of the dilution producing a specific OD of 0.5 after visualization with an anti-mouse IgG(H+L) polyclonal antibody coupled with peroxidase (NA931V, Amersham) and TMB supplemented with H_2O_2 (KPL). This analysis (FIG. 39A) shows that immunization with the virus VV-TG-S and VV-TN-S induces in mice, from the first immunization, antibodies directed against the native form of the SARS-CoV spicule protein present in the lysate of infected VeroE6 cells. The responses induced by the VV-TN-S virus are higher than those induced by the VV-TG-S virus after the first (TI=740 and TI=270 respectively) and the second (TI=3230 and TI=600 respectively) immunization. The VV-TN-Ssol virus induces high anti-SARS-CoV antibody titers after two immunizations (TI=640), whereas the virus VV-TG-Ssol induces a response at the detection limit (TI=40).

The immune sera were analyzed per pool for each of the groups for their capacity to seroneutralize the infectivity of SARS-CoV. 4 seroneutralization points on FRhK-4 cells (100 TCID₅₀ of SARS-CoV) are produced for each of the 2-fold dilutions tested from 1/20. The seroneutralizing titer is calculated according to the Reed, and Munsch method as the reciprocal of the dilution neutralizing the infectivity of 2 wells out of 4. This analysis shows that the antibodies induced in mice by the vaccinia viruses expressing the S protein or the Ssol polypeptide are neutralizing and that the viruses with synthetic promoters are more efficient immunogens than the viruses carrying the 7.5K promoter: the highest titers (640) are observed after 2 immunizations with the virus VV-TN-S (FIG. 39B).

The protective power of the neutralizing antibodies induced in mice after immunization with the recombinant vaccinia viruses is evaluated with the aid of a challenge infection with SARS-CoV.

6) Other Applications

Third generation recombinant vaccinia viruses are constructed by substituting the wild-type sequences of the S and Ssol genes by synthetic genes optimized for the expression in mammalian cells, described above. These recombinant vaccinia viruses are capable of expressing larger quantities of S and Ssol antigens and therefore of exhibiting increased immunogenicity.

The recombinant vaccinia virus VV-TN-Ssol can be used for the quantitative production and purification of the Ssol antigen for diagnostic (serology by ELISA) and vaccine (sub-unit vaccine) applications.

EXAMPLE 17

Recombinant Measles Virus Expressing the SARS-Associated Coronavirus (SARS-CoV) Spicule (S) Protein. Vaccine Applications

1) Introduction

The measles vaccine (MV) induces a lasting protective immunity in humans after a single injection (Hilleman, 2002, Vaccine, 20: 651-665). The protection conferred is very robust and is based on the induction of an antibody response and of a CD4 and CD8 cell response. The MV genome is very stable and no reversion of the vaccine strains to virulence has ever been observed. The measles virus belongs to the genus *Morbillivirus* of the Paramyxoviridae family; it is an enveloped virus whose genome is a 16 kb single-stranded RNA of negative polarity (FIG. 40A) and whose exclusively cytoplasmic replication cycle excludes any possibility of integration into the genome of the host. The measles vaccine is thus one of the most effective and one of the safest live vaccines used in the human population. Frédéric Tangy's team recently developed an expression vector on the basis of the Schwarz strain of the measles virus, which is the safest attenuated strain and the most widely used in humans as vaccine against measles. This vaccine strain may be isolated from an infectious molecular clone while preserving its immunogenicity in primates and in mice that are sensitive to the infection. It constitutes, after insertion of additional transcription units, a vector for the expression of heterologous sequences (Combretet, 2003, J. Virol. 77: 11546-11554). In addition, a recombinant MV Schwarz expressing the envelope glycoprotein of the West Nile virus (WNV) induces an effective and lasting antibody response which protects mice from a lethal challenge infection with WNV (Despres et al., 2004, J. Infect. Dis., in press). All these characteristics make the attenuated Schwarz strain of the measles virus an extremely promising candidate vector for the construction of novel recombinant live vaccines.

The aim of this example is to evaluate the capacity of recombinant measles viruses (MV) expressing various SARS-associated coronavirus (SARS-CoV) antigens to constitute novel candidate vaccines against SARS.

The inventors focused on the SARS-CoV spicule (S) protein, which makes it possible to induce, after gene immunization in animals, antibodies neutralizing the infectivity of SARS-CoV, and on a soluble and secreted form of this protein, the Ssol polypeptide, which is composed of the ectodomain (aa 1-1193) of S fused at its C-ter end with a FLAG tag (DYKDDDDK) via a BspE1 linker encoding the SG dipeptide. This Ssol polypeptide exhibits a similar antigenicity to that of the S protein and allows, after injection into mice in the form of a purified protein adjuvanted with aluminum hydroxide, the induction of high neutralizing antibody titers against SARS-CoV.

The various forms of the S gene were introduced in the form of an additional transcription unit between the P (phosphoprotein) and M (matrix) genes into the cDNA of the Schwarz strain of MV previously described (Combretet, 2003, J. Virol. 77: 11546-11554; EP application No. 02291551.6 of Jun. 20, 2002, and EP application No. 02291550.8 of Jun. 20, 2002). After having isolated the recombinant viruses MVSchw2-SARS-S and MVSchw2-

SARS-Ssol and checked their capacity to express the SARS-CoV S antigen, their capacity to induce a protective immune response against SARS in mice and then in monkeys was tested.

2) Construction of the Recombinant Viruses

The plasmid pTM-MV Schw-ATU2 (FIG. 40B) contains an infectious cDNA corresponding to the antigenome of the Schwarz vaccine strain of the measles virus (MV) into which an additional transcription unit (ATU) has been introduced between the P (phosphoprotein) and M (matrix) genes (Combrete, 2003, *Journal of Virology*, 77: 11546-11554). Recombinant genomes MV Schw2-SARS-S and MV Schw2-SARS-Ssol of the measles virus were constructed by inserting ORFs of the S protein and of the Ssol polypeptide into the additional transcription unit of the MV Schw-ATU2 vector.

For that, a DNA fragment containing the SARS-CoV S cDNA was amplified by PCR with the aid of the oligonucleotides 5'-ATACGTACGA CCATGTTTAT TTTCTTATTA TTTCTTACTC TCACT-3' and 5'-ATAGCGCGCT CATTAT-GTGT AATGTAATTT GACACCCTTG-3' using the plasmid pCDNA-S as template and then inserted into the plasmid pCR®2.1-TOPO (Invitrogen) in order to obtain the plasmid pTOPO-S-MV. The two oligonucleotides used contain restriction sites BsiW1 and BssHII, so as to allow subsequent insertion into the measles vector, and were designed so as to generate a sequence of 3774 nt including the codons for initiation and termination, so as to observe the rule of 6 which stipulates that the length of the genome of a measles virus must be divisible by 6 (Calain & Roux, 1993, *J. Virol.*, 67: 4822-4830; Schneider et al., 1997, *Virology*, 227: 314-322). The insert was sequenced with the aid of a BigDye Terminator v1.1 kit (Applied Biosystems) and an automated sequencer ABI377.

To express a soluble and secreted form of SARS-CoV S, a plasmid containing the cDNA of the Ssol polypeptide corresponding to the ectodomain (aa 1-1193) of SARS-CoV S fused at its C-ter end with the sequence of a FLAG tag (DYKDDDDK) via a BspE1 linker encoding the SG dipeptide was then obtained. For that, a DNA fragment was amplified with the aid of the oligonucleotides 5'-CCATTCAAC AATTTGGCCG-3' and 5'-ATAGGATCCGCGCGCTCATT ATTTATCGTC GTCATCTTTA TAATC-3' from the plasmid pCDNA-Ssol and then inserted into the plasmid pTOPO-S-MV between the SalI and BamHI sites in order to obtain the plasmid pTOPO-S-MV-SF. The sequence generated is 3618 nt long between the BsiW1 and BssHII sites and observes the rule of 6. The insert was sequenced as indicated above.

The BsiW1-BssHII fragments containing the cDNAs for the S protein and the Ssol polypeptide were then excised by digestion of the plasmids pTOPO-S-MV and pTOPO-S-MV-SF and then subcloned between the corresponding sites of the plasmid pTM-MV Schw-ATU2 in order to give the plasmids pTM-MV Schw2-SARS-S and pTM-MV Schw2-SARS-Ssol (FIG. 40B). These two plasmids were deposited at the C.N.C.M. on Dec. 1, 2004, under the numbers I-3326 and I-3327, respectively.

The recombinant measles viruses corresponding to the plasmids pTM-MV Schw2-SARS-S and pTM-MV Schw2-SARS-Ssol were obtained by reverse genetics according to the system based on the use of a helper cell line, described by Radecke et al. (1995, *Embo J.*, 14: 5773-5784) and modified by Parks et al. (1999, *J. Virol.*, 73: 3560-3566). Briefly, the helper cells 293-3-46 are transfected according to the calcium phosphate method with 5 µg of the plasmids pTM-MV Schw2-SARS-S or pTM-MV Schw2-SARS-Ssol and 0.02 µg of the plasmid pEMC-La directing the expression of the MV L polymerase (gift from M. A. Billeter). After incu-

bating, overnight at 37° C., a heat shock is produced for 2 hours at 43° C. and the transfected cells are transferred onto a monolayer of Vero cells. For each of the two plasmids, syncytia appeared after 2 to 3 days of coculture and were transferred successively onto monolayers of Vero cells at 70% confluence in 35 mm Petri dishes and then in, 25 and 75 cm² flasks. When the syncytia have reached 80-90% confluence, the cells are recovered with the aid of a scraper and then frozen and thawed once. After low-speed centrifugation, the supernatant containing the virus is stored in aliquots at -80° C. The titers of the recombinant viruses MV Schw2-SARS-S and MV Schw2-SARS-Ssol were determined by limiting dilution on Vero cells and the titer as dose infecting 50% of the wells (TCID₅₀) calculated according to the Kärber method.

3) Characterization of the Recombinant Viruses

The expression of the transgenes encoding the S protein and the Ssol polypeptide was assessed by Western blotting and immunofluorescence.

Monolayers of Vero cells in T-25 flasks were infected at a multiplicity of 0.05 by various passages of the two viruses MV Schw2-SARS-S and MV Schw2-SARS-Ssol and the wild-type virus MWSchw as a control. When the syncytia had reached 80 to 90% confluence, cytoplasmic extracts were prepared in an extraction buffer (150 mM NaCl, 50 mM Tris-HCl, pH 7.2, 1% Triton X-100, 0.1% SDS, 1% DOC) and then diluted in loading buffer according to Laemmli, separated on 8% SDS polyacrylamide gel and transferred onto a PVDF membrane (BioRad). The detection of this immunoblot (Western blot) was carried out with the aid of an anti-S rabbit polyclonal serum (immune serum of the rabbit P11135: cf. example 4 above) and donkey polyclonal antibodies directed against rabbit IgGs and coupled with peroxidase (NA934V, Amersham). The bound antibodies were visualized by luminescence with the aid of the ECL+ kit (Amersham) and Hyperfilm MP autoradiography films (Amersham).

Vero cells in monolayers on glass slides were infected with the two viruses MV Schw2-SARS-S and MV Schw2-SARS-Ssol and the wild-type virus MWSchw as a control at multiplicities of infection of 0.05. When the syncytia had reached 90 to 100% (MV Schw2-SARS-Ssol virus) or 30 to 40% (MV Schw2-SARS-S, MWSchw) confluence, the cells were fixed in a 4% PBS-PFA solution, permeabilized with a PBS solution containing 0.2% Triton and then labeled with rabbit polyclonal antibodies hyperimmunized with purified and inactivated SARS-CoV virions and with an anti-rabbit IgG (H+L) goat antibody conjugate coupled with FITC (Jackson).

As shown in FIGS. 41 and 42, the recombinant viruses MV Schw2-SARS-S and MV Schw2-SARS-Ssol direct the expression of the S protein and the Ssol polypeptide respectively at levels comparable to those which can be observed 8 h after infection with SARS-CoV. The expression of these polypeptides is stable after 3 passages of the recombinant viruses in cell culture. These results demonstrate that the recombinant measles viruses are indeed carriers of the transgenes and allow the expression of the SARS glycoprotein in its membrane form (S) or in a soluble form (Ssol). The Ssol polypeptide is expected to be secreted by cells infected with the MV Schw2-SARS-Ssol virus as is the case when this same polypeptide is expressed in mammalian cells after transient transfection of the corresponding sequences (cf. example 11 above).

4) Applications

Having shown that the viruses, MV Schw2-SARS-S and MV Schw2-SARS-Ssol allow the expression of the SARS-CoV S, their capacity to induce a protective immune response against SARS-CoV in CD46^{+/+} IFN-αβR^{-/-} mice, which is

sensitive to infection by MV, is evaluated. The antibody response of the immunized mice is evaluated by ELISA test against the native antigens of SARS-CoV and for their capacity to neutralize the infectivity of SARS-CoV in vitro, using the methodologies described above. The protective power of the response will be evaluated by measuring the reduction in the pulmonary viral load 2 days after a nonlethal challenge infection with SARS-CoV.

Second generation recombinant measles viruses are constructed by substituting the wild-type sequences of the S and Sol genes by synthetic genes optimized for expression in mammalian cells, described in example 15 above. These recombinant measles viruses are capable of expressing larger quantities of the S and Ssol antigens and therefore of exhibiting increased immunogenicity.

Alternatively, the wild-type or synthetic genes encoding the S protein or the Ssol polypeptide may be inserted into the measles vector MVSchw-ATU3 in the form of an additional transcription unit located between the H and L genes, and then the recombinant viruses produced and characterized in a similar manner. This insertion is capable of generating recombinant viruses possessing different characteristics (multiplication of the virus, level of expression of the transgene) and possibly an improved immunogenicity compared with those obtained after insertion of the transgenes between the P and N genes.

The recombinant measles virus MVSchw2-SARS-Ssol may be used for the quantitative production and the purification of the Ssol antigen for diagnostic and vaccine applications.

EXAMPLE 18

Other Applications Linked to the S Protein

a) The lentiviral vectors allowing the expression of S or Ssol (or even of fragments of S) can constitute a recombinant vaccine against SARS-CoV, to be used in human or veterinary prophylaxis. In order to demonstrate the feasibility of such a vaccine, the immunogenicity of the recombinant lentiviral vectors TRIP-SD/SA-S-WPRE and TRIP-SD/SA-Ssol-WPRE is studied in mice.

b) Monoclonal antibodies are produced with the aid of the recombinant Ssol polypeptide. According to the results presented in example 14 above, these antibodies or at least the majority of them will recognize the native form of the SARS-CoV S and will be capable of diagnostic and/or prophylactic applications.

c) A serological test for SARS is developed with the Ssol polypeptide used as antigen and the double epitope methodology.

SEQUENCE LISTING

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Met	Ala	Lys	Thr	Ser	Val	Asp	Cys	Asn	Met	Tyr	Ile	Cys	Gly	Asp	Ser	725
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Leu	Asn	Arg	Ala	Leu	Ser	Gly	Ile	Ala	Ala	Glu	Gln	Asp	Arg	Asn	Thr	760
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Arg	Glu	Val	Phe	Ala	Gln	Val	Lys	Gln	Met	Tyr	Lys	Thr	Pro	Thr	Leu	775
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Lys	Tyr	Phe	Gly	Gly	Phe	Asn	Phe	Ser	Gln	Ile	Leu	Pro	Asp	Pro	Leu	790
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Thr	Val	Leu	Pro	Pro	Leu	Leu	Thr	Asp	Asp	Met	Ile	Ala	Ala	Tyr	Thr	855
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Gly	Ala	Ala	Leu	Gln	Ile	Pro	Phe	Ala	Met	Gln	Met	Ala	Tyr	Arg	Phe	885
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Thr Thr Thr Ser Thr	Ala Leu Gly Lys Leu	Gln Asp Val Val Asn Gln	
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aat gct caa gca tta aac	aca ctt gtt aaa caa	ctt agc tct aat ttt	2944
Asn Ala Gln Ala Leu	Asn Thr Leu Val Lys	Gln Leu Ser Ser Asn Phe	
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ggg gca att tca agt	gtg cta aat gat atc	ctt tgc cga ctt gat aaa	2992
Gly Ala Ile Ser Ser	Val Leu Asn Asp Ile	Leu Ser Arg Leu Asp Lys	
	955	960	965
gtc gag gcg gag gta	caa att gac agg tta	att aca ggc aga ctt caa	3040
Val Glu Ala Glu Val	Gln Ile Asp Arg Leu	Ile Thr Gly Arg Leu Gln	
	970	975	980
agc ctt caa acc tat	gta aca caa caa cta	atc agg gct gct gaa atc	3088
Ser Leu Gln Thr Tyr	Val Thr Gln Gln Leu	Ile Arg Ala Ala Glu Ile	
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agg gct tct gct aat	ctt gct gct act aaa	atg tct gag tgt gtt	3133
Arg Ala Ser Ala Asn	Leu Ala Ala Thr Lys	Met Ser Glu Cys Val	
	1005	1010	1015
ctt gga caa tca aaa	aga gtt gac ttt tgt	gga aag ggc tac cac	3178
Leu Gly Gln Ser Lys	Arg Val Asp Phe Cys	Gly Lys Gly Tyr His	
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ctt atg tcc ttc cca	caa gca gcc cgc cat	ggg gtt gtc ttc cta	3223
Leu Met Ser Phe Pro	Gln Ala Ala Pro His	Gly Val Val Phe Leu	
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cat gtc acg tat gtg	cca tcc cag gag agg	aac ttc acc aca gcg	3268
His Val Thr Tyr Val	Pro Ser Gln Glu Arg	Asn Phe Thr Thr Ala	
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cca gca att tgt cat	gaa ggc aaa gca tac	ttc cct cgt gaa ggt	3313
Pro Ala Ile Cys His	Glu Gly Lys Ala Tyr	Phe Pro Arg Glu Gly	
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Val Phe Val Phe Asn	Gly Thr Ser Trp Phe	Ile Thr Gln Arg Asn	
	1080	1085	1090
ttc ttt tct cca caa	ata att act aca gac	aat aca ttt gtc tca	3403
Phe Phe Ser Pro Gln	Ile Ile Thr Thr Asp	Asn Thr Phe Val Ser	
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gga aat tgt gat gtc	gtt att ggc atc att	aac aac aca gtt tat	3448
Gly Asn Cys Asp Val	Val Ile Gly Ile Ile	Asn Asn Thr Val Tyr	
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Asp Pro Leu Gln Pro	Glu Leu Asp Ser Phe	Lys Glu Glu Leu Asp	
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Lys Tyr Phe Lys Asn	His Thr Ser Pro Asp	Val Asp Leu Gly Asp	
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att tca ggc att aac	gct tct gtc gtc aac	att caa aaa gaa att	3583
Ile Ser Gly Ile Asn	Ala Ser Val Val Asn	Ile Gln Lys Glu Ile	
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gac cgc ctg aat gag	gtc gct aaa aat tta	aat gaa tca ctg att	3628
Asp Arg Leu Asn Glu	Val Ala Lys Asn Leu	Asn Glu Ser Leu Ile	
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gac ctt caa gaa ttg	gga aaa tat gag caa	tat att aaa tgg cct	3673
Asp Leu Gln Glu Leu	Gly Lys Tyr Glu Gln	Tyr Ile Lys Trp Pro	
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tgg tat gtt tgg ctg	ggc ttc att gct gga	cta att gcc atc gtc	3718
Trp Tyr Val Trp Leu	Gly Phe Ile Ala Gly	Leu Ile Ala Ile Val	
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atg gtt aca atc ttg	ctt tgt tgc atg act	agt tgt tgc agt tgc	3763

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<211> LENGTH: 1255
<212> TYPE: PRT
<213> ORGANISM: CORONAVIRUS
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His	Thr 35	Ser	Ser	Met	Arg	Gly	Val 40	Tyr	Tyr	Pro	Asp 45	Glu	Ile	Phe
Ser	Asp 50	Thr	Leu	Tyr	Leu	Thr 55	Gln	Asp	Leu	Phe 60	Leu	Pro	Phe	Tyr
Asn 65	Val	Thr	Gly	Phe	His 70	Thr	Ile	Asn	His	Thr 75	Phe	Gly	Asn	Pro
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Val	Val	Arg	Gly 100	Trp	Val	Phe	Gly	Ser 105	Thr	Met	Asn	Asn 110	Lys	Ser
Ser	Val 115	Ile	Ile	Ile	Asn	Asn	Ser 120	Thr	Asn	Val	Val 125	Ile	Arg	Ala
Asn	Phe 130	Glu	Leu	Cys	Asp 135	Asn	Pro	Phe	Phe	Ala 140	Val	Ser	Lys	Pro
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Phe	Glu	Tyr	Ile 165	Ser	Asp	Ala	Phe	Ser	Leu 170	Asp	Val	Ser	Glu	Lys
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Phe	Leu 195	Tyr	Val	Tyr	Lys	Gly	Tyr 200	Gln	Pro	Ile	Asp 205	Val	Val	Arg
Leu 210	Pro	Ser	Gly	Phe	Asn 215	Thr	Leu	Lys	Pro	Ile 220	Phe	Lys	Leu	Pro
Gly 225	Ile	Asn	Ile	Thr	Asn 230	Phe	Arg	Ala	Ile	Leu 235	Thr	Ala	Phe	Ser
Ala	Gln	Asp	Ile 245	Trp	Gly	Thr	Ser	Ala 250	Ala	Ala	Tyr	Phe	Val	Gly
Leu	Lys	Pro	Thr 260	Thr	Phe	Met	Leu	Lys 265	Tyr	Asp	Glu	Asn 270	Gly	Thr
Thr	Asp 275	Ala	Val	Asp	Cys	Ser	Gln 280	Asn	Pro	Leu	Ala 285	Glu	Leu	Lys
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Ser	Val	Leu	Tyr	Asn	Ser	Thr	Phe	Phe	Ser	Thr	Phe	Lys	Cys	Tyr	Gly	
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Val	Ser	Ala	Thr	Lys	Leu	Asn	Asp	Leu	Cys	Phe	Ser	Asn	Val	Tyr	Ala	
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Asp	Ser	Phe	Val	Val	Lys	Gly	Asp	Asp	Val	Arg	Gln	Ile	Ala	Pro	Gly	
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Lys	Pro	Cys	Thr	Pro	Pro	Ala	Leu	Asn	Cys	Tyr	Trp	Pro	Leu	Asn	Asp	
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<210> SEQ ID NO 4

<211> LENGTH: 3943

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

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gagttacca aaatgttctc tatgagaacc aaaaacaaat cgccaaccaa tttaacaagg	2820
cgattagtca aattcaagaa tcacttacaa caacatcaac tgcatgggc aagctgcaag	2880
acgttggtta ccagaatgct caagcattaa acacacttgt taaacaactt agctctaatt	2940
ttggtgcaat ttcaagtgtg ctaaatgata tcctttcgcg acttgataaa gtcgaggcgg	3000
aggatcaaat tgacaggcta attacaggca gacttcaaag ccttcaaacc tatgtaacac	3060
aaacaactat cagggtgct gaaatcaggg cttctgctaa tcttgctgct actaaaatgt	3120
ctgagtgtgt tcttgacaa tcaaaaagag ttgacttttg tggaaagggc taccacctta	3180
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cccaggagag gaacttcacc acagcgccag caatttgtca tgaaggcaaa gcatacttcc	3300
ctcgtgaagg tgtttttgtg tttaatggca cttcttggtt tattacacag aggaacttct	3360
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caggcattaa cgcttctgtc gtcaacatc aaaaagaaat tgaccgctc aatgaggctg	3600
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ttacaatctt gctttgttgc atgactagtt gttgcagttg cctcaagggt gcatgctctt	3780
gtggttcttg ctgcaagttt gatgaggatg actctgagcc agttctcaag ggtgtcaaat	3840
tacattacac ataaacgaac ttatggattt gtttatgaga ttttttactc ttggatcaat	3900

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tactgcacag ccagtaaaaa ttgacaatgc ttctcctgca agt 3943

<210> SEQ ID NO 5

<211> LENGTH: 2049

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 5

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atattcttgt taacaactaa acgaacatgt ttattttctt attatttctt actctcacta 120
gtggtagtga ccttgaccgg tgcaccactt ttgatgatgt tcaagctcct aattacactc 180
aacatacttc atctatgagg ggggtttact atcctgatga aattttttaga tcagacactc 240
tttatttaac tcaggattta ttcttccat ttatttctaa tgttacaggg ttccatacta 300
ttaatcatat gtttggaac cctgtcatat cttttaagga tggatttat ttgctgcca 360
cagagaaatc aaatgttgtc cgtggttggg tttttggttc taccatgaac aacaagtcac 420
agtcggtgat tattattaac aattctacta atgttggtat acgagcatgt aactttgaat 480
tgtgtgacaa ccctttcttt gctgtttcta aacccatggg tacacagaca catactatga 540
tattcgataa tgcatttaat tgcactttcg agtacatata tgatgccttt tcgcttgatg 600
tttcagaaaa gtcaggtaat tttaaacact tacgagagtt tgtgtttaa aataaagatg 660
ggtttctcta tgtttataag ggctatcaac ctatagatgt agttcgtgat ctacctctg 720
gttttaacac tttgaaacct atttttaagt tgcctcttgg tattaacatt acaaatttta 780
gagccattct tacagccttt tcacctgtct aagacatttg gggcacgtca gctgcagcct 840
attttgttgg ctattttaa ccaactacat ttatgctcaa gtatgatgaa aatggtacaa 900
tcacagatgc tgttgattgt tctcaaaatc cacttgctga actcaaatgc tctgttaaga 960
gctttgatag tgacaaagga atttaccaga cctctaattt cagggttgtt ccctcaggag 1020
atgttgtgag attccctaatt attacaaact tgtgtccttt tggagagggt tttaatgcta 1080
ctaaattccc ttctgtctat gcatgggaga gaaaaaaaaa ttctaattgt gttgctgatt 1140
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ctaagttaa tgatctttgc ttctccaatg tctatgcaga ttcttttcta gtcaggagg 1260
atgatgtaag acaaatagcg ccaggacaaa ctggtgttat tgctgattat aattataaat 1320
tgccagatga tttcatgggt tgtgtccttg cttggaatac taggaacatt gatgctactt 1380
caactggtaa ttataattat aaatataggt atcttagaca tggcaagcct aggccctttg 1440
agagagacat atctaattgt cctttctccc ctgatggcaa accttgacc ccacctgtc 1500
ttaattgtta ttggccatta aatgattatg gtttttacac cactactggc attggctacc 1560
aaccttacag agttgtagta ctttcttttg aactttttaa tgcaccggcc acgggttgtg 1620
gacccaaaatt atccactgac cttattaaga accagtgtgt caattttaat tttaatggac 1680
tcactggtag tgggtgtgta actccttctt caaagagatt tcaaccattt caacaatttg 1740
gccgtgatgt ctctgatttc actgattccg ttcgagatcc taaaacatct gaaatattag 1800
acatttcacc ttgctctttt gggggtgtaa gtgtaattac acctggaaca aatgcttcat 1860
ctgaagttgc tgttctatat caagatgta actgcactga tgtttctaca gcaatccatg 1920
cagatcaact cacaccagct tggcgcatat attctactgg aaacaatgta ttccagactc 1980
aagcaggctg tcttatagga gctgagcatg tcgacacttc ttatgagtgc gacattccta 2040
ttggagctg 2049

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<210> SEQ ID NO 6
 <211> LENGTH: 2027
 <212> TYPE: DNA
 <213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 6

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catgcagatc aactcacacc agcttggcgc atatatctta ctggaacaa tgtattccag    60
actcaagcag gctgtcttat aggagctgag catgtcgaca cttcttatga gtgcgacatt    120
cctattggag ctggcatttg tgctagttac catacagttt ctttattacg tagtactagc    180
caaaaatcta ttgtggctta tactatgtct ttaggtgctg atagttcaat tgcttactct    240
aataacacca ttgctatacc tactaacttt tcaattagca ttactacaga agtaatgcct    300
gtttctatgg ctaaaacctc cgtagattgt aatatgtaca tctgcggaga ttctactgaa    360
tgtgctaatt tgcttctcca atatggtagc ttttgcacac aactaaatcg tgcactctca    420
ggatttgctg ctgaacagga tcgcaacaca cgtgaagtgt tcgctcaagt caaacaatg    480
tacaaaaacc caactttgaa atattttggt gggtttaatt ttccacaaat attacctgac    540
cctctaaagc caactaagag gtcttttatt gaggacttgc tctttaataa ggtgacactc    600
gctgatgctg gcttcatgaa gcaatatggc gaatgcctag gtgatattaa tgctagagat    660
ctcatttgct cgcagaagtt caatgggctt acagtgttgc cacctctgct cactgatgat    720
atgattgctg cctacactgc tgctctagtt agtggtagct ccactgctgg atggacattt    780
gggtgctggc ctgctcttca aatacctttt gctatgcaaa tggcatatag gttcaatggc    840
attggagtta cccaaaatgt tctctatgag aaccaaaaac aaatcgccaa ccaatttaac    900
aaggcgatta gtcaaattca agaatcactt acaacaacat caactgcatt gggcaagctg    960
caagacgttg ttaaccagaa tgctcaagca ttaaacacac ttgttaacaa acttagctct    1020
aattttgggt caatttcaag tgtgctaaat gatatccttt cgcgacttga taaagtcgag    1080
gcggagggtac aaattgacag gttaattaca ggcagacttc aaagccttca aacctatgta    1140
acacaacaac taatcagggc tgctgaaatc agggcttctg ctaatcttgc tgctactaaa    1200
atgtctgagt gtgttcttgg acaatcaaaa agagttgact tttgtgaaa gggctaccac    1260
cttatgtcct tcccacaagc agccccgcgt ggtgttgtct tcctacatgt cactgatgtg    1320
ccatcccagg agaggaactt caccacagcg ccagcaattt gtcatgaagg caaagcatac    1380
ttccctcgct aagggtgttt tgtgtttaat ggcacttctt gggttattac acagaggaac    1440
ttcttttctc cacaataaat tactacagac aatacatttg tctcaggaaa ttgtgatgtc    1500
gttattggcg tcattaacaa cacagtttat gatcctctgc aacctgagct tgactcattc    1560
aaagaagagc tggacaagta cttcaaaaat catacatcac cagatgttga tcttggcgac    1620
atttcaggca ttaacgcttc tgtcgtcaac attcaaaaag aattggaccg cctcaatgag    1680
gtcgctaaaa atttaaatga atcactcatt gaccttcaag aattgggaaa atatgagcaa    1740
tatattaaat ggcttgggta tgtttggctc ggcttcattg ctggactaat tgccatcgtc    1800
atgggttaca tcttgctttg ttgcatgact agttgttgca gttgcctcaa gggtgcatgc    1860
tcttgtgggt cttgctgcaa gtttgatgag gatgactctg agccagttct caaggggtgc    1920
aaattacatt acacataaac gaacttatgg atttgtttat gagatttttt actcttggat    1980
caattactgc acagccagta aaaattgaca atgcttctcc tgcaagt                    2027

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<210> SEQ ID NO 7
 <211> LENGTH: 1096

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<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 7

tcttgctttg ttgcatgact agttgttgca gttgectcaa ggggtgcatgc tcttggtggtt	60
cttgtgcaaa gtttgatgag gatgactctg agccagttct caagggtgtc aaattacatt	120
acacataaac gaacttatgg atttgtttat gagatTTTTT actcttgat caattactgc	180
acagccagta aaaattgaca atgcttctcc tgcaagtact gttcatgcta cagcaacgat	240
accgctacaa gcctcactcc ctttcggatg gcttgttatt ggcgttgcat ttcttgctgt	300
ttttcagagc gctacaaaa taattgcgct caataaaaaga tggcagctag ccctttataa	360
gggcttcag ttcatttgca atttactgct gctatttggt accatctatt cacatctttt	420
gcttgctgct gcaggtatgg aggcgcaatt tttgtacctc tatgcctga tatattttct	480
acaatgcatc aacgcatgta gaattattat gagatgttgg ctttggtgga agtgcaaatc	540
caagaaccca ttactttatg atgccaaacta ctttgtttgc tggcacacac ataactatga	600
ctactgtata ccatataaca gtgtcacaga tacaattgtc gttactgaag gtgacggcat	660
ttcaacacca aaactcaaag aagactacca aattgggtgg tattctgagg ataggcactc	720
agggtttaaa gactatgtcg ttgtacatgg ctatttcacc gaagtttact accagcttga	780
gtctacacaa attactacag aacttggtat tgaaaatgct acattcttca tctttaacaa	840
gcttggttaa gacccaccga atgtgcaaat acacacaatc gacggctctt caggagtgtc	900
taatccagca atggatccaa tttatgatga gccgacgacg actactagcg tgcctttgta	960
agcacaagaa agtgagtacg aacttatgta ctcattcgtt tcggaagaaa caggtagctt	1020
aatagttaat agcgtacttc tttttcttgc tttcgtggta ttcttgctag tcacactagc	1080
catecttact gcgctt	1096

<210> SEQ ID NO 8

<211> LENGTH: 1135

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 8

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aagggtgcat gctcttggtg ttcttgctgc aagtttgatg aggatgactc tgagccagtt	120
ctcaagggtg tcaaattaca ttacacataa acgaacttat ggatttgttt atgagatttt	180
ttactcttgg atcaattact gcacagccag taaaaattga caatgcttct cctgcaagta	240
ctgttcacgc tacagcaacg ataccgctac aagcctcact ccctttcgga tggcttggtta	300
ttggcggtgc atttcttgct gtttttcaga gcgctaccaa aataattgcg ctcaataaaa	360
gatggcagct agccctttat aagggtctcc agttcatttg caatttactg ctgctatttg	420
ttaccatcta ttcacatctt ttgcttgctg ctgcaggatg ggaggcgcaa tttttgtacc	480
tctatgcctt gatataTTTT ctacaatgca tcaacgcacg tagaattatt atgagatggt	540
ggctttgttg gaagtgcacaa tccaagaacc cttacttcta tgatgccaac tactttgttt	600
gctggcacac acataactat gactactgta taccatataa cagtgtcaca gatacaattg	660
tcgttactga aggtgacggc atttcaacac caaaactcaa agaagactac caaattgggtg	720
gttattctga ggataggcac tcagggtgta aagactatgt cgttgtagat ggctatttca	780
ccgaagttaa ctaccagctt gagtctacac aaattactac agacactggg attgaaaatg	840
ctacattctt catctttaac aagcttggtta aagaccacc gaatgtgcaa atacacacaa	900

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tcgacggctc ttcaggagtt gctaataccag caatggatcc aatttatgat gagccgacga 960
cgactactag cgtgcctttg taagcacaag aaagtgagta cgaacttatg tactcattcg 1020
tttcggaaga aacaggtacg ttaatagtta atagcgtact tctttttcctt gctttcgtgg 1080
tattcttgct agtcacacta gccatcctta ctgcgcttcg attgtgtgcg tactg 1135

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<210> SEQ ID NO 9
<211> LENGTH: 1096
<212> TYPE: DNA
<213> ORGANISM: CORONAVIRUS
<220> FEATURE:
<221> NAME/KEY: CDS
<222> LOCATION: (137) .. (958)
<223> OTHER INFORMATION:

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<400> SEQUENCE: 9

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tcttgctttg ttgcatgact agttgttgca gttgcctcaa gggtgcatgc tcttggtggtt 60
cttgctgcaa gtttgatgag gatgactctg agccagttct caaggggtgc aaattacatt 120
acacataaac gaactt atg gat ttg ttt atg aga ttt ttt act ctt gga tca 172
      Met Asp Leu Phe Met Arg Phe Phe Thr Leu Gly Ser
      1             5             10

att act gca cag cca gta aaa att gac aat gct tct cct gca agt act 220
Ile Thr Ala Gln Pro Val Lys Ile Asp Asn Ala Ser Pro Ala Ser Thr
      15             20             25

gtt cat gct aca gca acg ata ccg cta caa gcc tca ctc cct ttc gga 268
Val His Ala Thr Ala Thr Ile Pro Leu Gln Ala Ser Leu Pro Phe Gly
      30             35             40

tgg ctt gtt att ggc gtt gca ttt ctt gct gtt ttt cag agc gct acc 316
Trp Leu Val Ile Gly Val Ala Phe Leu Ala Val Phe Gln Ser Ala Thr
      45             50             55             60

aaa ata att gcg ctc aat aaa aga tgg cag cta gcc ctt tat aag ggc 364
Lys Ile Ile Ala Leu Asn Lys Arg Trp Gln Leu Ala Leu Tyr Lys Gly
      65             70             75

ttc cag ttc att tgc aat tta ctg ctg cta ttt gtt acc atc tat tca 412
Phe Gln Phe Ile Cys Asn Leu Leu Leu Leu Phe Val Thr Ile Tyr Ser
      80             85             90

cat ctt ttg ctt gtc gct gca ggt atg gag gcg caa ttt ttg tac ctc 460
His Leu Leu Leu Val Ala Ala Gly Met Glu Ala Gln Phe Leu Tyr Leu
      95             100             105

tat gcc ttg ata tat ttt cta tgc atc aac gca tgt aga att att 508
Tyr Ala Leu Ile Tyr Phe Leu Gln Cys Ile Asn Ala Cys Arg Ile Ile
      110             115             120

atg aga tgt tgg ctt tgt tgg aag tgc aaa tcc aag aac cca tta ctt 556
Met Arg Cys Trp Leu Cys Trp Lys Cys Lys Ser Lys Asn Pro Leu Leu
      125             130             135             140

tat gat gcc aac tac ttt gtt tgc tgg cac aca cat aac tat gac tac 604
Tyr Asp Ala Asn Tyr Phe Val Cys Trp His Thr His Asn Tyr Asp Tyr
      145             150             155

tgt ata cca tat aac agt gtc aca gat aca att gtc gtt act gaa ggt 652
Cys Ile Pro Tyr Asn Ser Val Thr Asp Thr Ile Val Val Thr Glu Gly
      160             165             170

gac gcc att tca aca cca aaa ctc aaa gaa gac tac caa att ggt ggt 700
Asp Gly Ile Ser Thr Pro Lys Leu Lys Glu Asp Tyr Gln Ile Gly Gly
      175             180             185

tat tct gag gat agg cac tca ggt gtt aaa gac tat gtc gtt gta cat 748
Tyr Ser Glu Asp Arg His Ser Gly Val Lys Asp Tyr Val Val Val His
      190             195             200

ggc tat ttc acc gaa gtt tac tac cag ctt gag tct aca caa att act 796
Gly Tyr Phe Thr Glu Val Tyr Tyr Gln Leu Glu Ser Thr Gln Ile Thr
      205             210             215             220

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aca gac act ggt att gaa aat gct aca ttc ttc atc ttt aac aag ctt      844
Thr Asp Thr Gly Ile Glu Asn Ala Thr Phe Phe Ile Phe Asn Lys Leu
                225                230                235

gtt aaa gac cca ccg aat gtg caa ata cac aca atc gac ggc tct tca      892
Val Lys Asp Pro Pro Asn Val Gln Ile His Thr Ile Asp Gly Ser Ser
                240                245                250

gga gtt gct aat cca gca atg gat cca att tat gat gag ccg acg acg      940
Gly Val Ala Asn Pro Ala Met Asp Pro Ile Tyr Asp Glu Pro Thr Thr
                255                260                265

act act agc gtg cct ttg taagcacaag aaagtgagta cgaacttatg      988
Thr Thr Ser Val Pro Leu
                270

tactcattcg ttccggaaga aacagggtacg ttaatagtta atagcgtact tctttttcctt 1048

gctttcgtgg tattcttgcct agtcacacta gccatcctta ctgcgctt      1096

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<210> SEQ ID NO 10
<211> LENGTH: 274
<212> TYPE: PRT
<213> ORGANISM: CORONAVIRUS

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<400> SEQUENCE: 10

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Met Asp Leu Phe Met Arg Phe Phe Thr Leu Gly Ser Ile Thr Ala Gln
1          5          10          15

Pro Val Lys Ile Asp Asn Ala Ser Pro Ala Ser Thr Val His Ala Thr
20          25          30

Ala Thr Ile Pro Leu Gln Ala Ser Leu Pro Phe Gly Trp Leu Val Ile
35          40          45

Gly Val Ala Phe Leu Ala Val Phe Gln Ser Ala Thr Lys Ile Ile Ala
50          55          60

Leu Asn Lys Arg Trp Gln Leu Ala Leu Tyr Lys Gly Phe Gln Phe Ile
65          70          75          80

Cys Asn Leu Leu Leu Leu Phe Val Thr Ile Tyr Ser His Leu Leu Leu
85          90          95

Val Ala Ala Gly Met Glu Ala Gln Phe Leu Tyr Leu Tyr Ala Leu Ile
100         105         110

Tyr Phe Leu Gln Cys Ile Asn Ala Cys Arg Ile Ile Met Arg Cys Trp
115         120         125

Leu Cys Trp Lys Cys Lys Ser Lys Asn Pro Leu Leu Tyr Asp Ala Asn
130         135         140

Tyr Phe Val Cys Trp His Thr His Asn Tyr Asp Tyr Cys Ile Pro Tyr
145         150         155         160

Asn Ser Val Thr Asp Thr Ile Val Val Thr Glu Gly Asp Gly Ile Ser
165         170         175

Thr Pro Lys Leu Lys Glu Asp Tyr Gln Ile Gly Gly Tyr Ser Glu Asp
180         185         190

Arg His Ser Gly Val Lys Asp Tyr Val Val Val His Gly Tyr Phe Thr
195         200         205

Glu Val Tyr Tyr Gln Leu Glu Ser Thr Gln Ile Thr Thr Asp Thr Gly
210         215         220

Ile Glu Asn Ala Thr Phe Phe Ile Phe Asn Lys Leu Val Lys Asp Pro
225         230         235         240

Pro Asn Val Gln Ile His Thr Ile Asp Gly Ser Ser Gly Val Ala Asn
245         250         255

Pro Ala Met Asp Pro Ile Tyr Asp Glu Pro Thr Thr Thr Thr Ser Val
260         265         270

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Pro Leu

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<210> SEQ ID NO 11
<211> LENGTH: 1096
<212> TYPE: DNA
<213> ORGANISM: CORONAVIRUS
<220> FEATURE:
<221> NAME/KEY: CDS
<222> LOCATION: (558) .. (1019)
<223> OTHER INFORMATION:

<400> SEQUENCE: 11

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acacataaac gaacttatgg atttgtttat gagatttttt actcttggat caattactgc    180
acagccagta aaaattgaca atgcttctcc tgcaagtact gttcatgcta cagcaacgat    240
accgctacaa gcctcactcc ctttcggatg gcttgttatt ggcgttgcat ttcttgctgt    300
ttttcagagc gctacaaaaa taattgcgct caataaaaga tggcagctag ccttttataa    360
gggcttccag ttcatttgca atttactgct gctatttggt accatctatt cacatctttt    420
gcttgctgct gcaggtatgg aggcgcaatt tttgtacctc tatgcctga tatattttct    480
acaatgcatc aacgcatgta gaattattat gagatgttgg ctttgttgga agtgcaaatc    540
caagaaccca ttactttt atg atg cca act act ttg ttt gct ggc aca cac    590
          Met Met Pro Thr Thr Leu Phe Ala Gly Thr His
          1             5             10

ata act atg act act gta tac cat ata aca gtg tca cag ata caa ttg    638
Ile Thr Met Thr Thr Val Tyr His Ile Thr Val Ser Gln Ile Gln Leu
          15             20             25

tcg tta ctg aag gtg acg gca ttt caa cac caa aac tca aag aag act    686
Ser Leu Leu Lys Val Thr Ala Phe Gln His Gln Asn Ser Lys Lys Thr
          30             35             40

acc aaa ttg gtg gtt att ctg agg ata ggc act cag gtg tta aag act    734
Thr Lys Leu Val Val Ile Leu Arg Ile Gly Thr Gln Val Leu Lys Thr
          45             50             55

atg tcg ttg tac atg gct att tca ccg aag ttt act acc agc ttg agt    782
Met Ser Leu Tyr Met Ala Ile Ser Pro Lys Phe Thr Thr Ser Leu Ser
          60             65             70             75

cta cac aaa tta cta cag aca ctg gta ttg aaa atg cta cat tct tca    830
Leu His Lys Leu Leu Gln Thr Leu Val Leu Lys Met Leu His Ser Ser
          80             85             90

tct tta aca agc ttg tta aag acc cac cga atg tgc aaa tac aca caa    878
Ser Leu Thr Ser Leu Leu Lys Thr His Arg Met Cys Lys Tyr Thr Gln
          95             100             105

tcg acg gct ctt cag gag ttg cta atc cag caa tgg atc caa ttt atg    926
Ser Thr Ala Leu Gln Glu Leu Leu Ile Gln Gln Trp Ile Gln Phe Met
          110             115             120

atg agc cga cga cga cta cta gcg tgc ctt tgt aag cac aag aaa gtg    974
Met Ser Arg Arg Arg Leu Leu Ala Cys Leu Cys Lys His Lys Lys Val
          125             130             135

agt acg aac tta tgt act cat tcg ttt cgg aag aaa cag gta cgt    1019
Ser Thr Asn Leu Cys Thr His Ser Phe Arg Lys Lys Gln Val Arg
          140             145             150

taatagttaa tagcgtactt ctttttcttg ctttcgtggt attcttgcta gtcacactag    1079
ccatccttac tgcgctt    1096

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<210> SEQ ID NO 12
<211> LENGTH: 154
<212> TYPE: PRT

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<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 12

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Met Met Pro Thr Thr Leu Phe Ala Gly Thr His Ile Thr Met Thr Thr
 1           5           10           15

Val Tyr His Ile Thr Val Ser Gln Ile Gln Leu Ser Leu Leu Lys Val
      20           25           30

Thr Ala Phe Gln His Gln Asn Ser Lys Lys Thr Thr Lys Leu Val Val
      35           40           45

Ile Leu Arg Ile Gly Thr Gln Val Leu Lys Thr Met Ser Leu Tyr Met
 50           55           60

Ala Ile Ser Pro Lys Phe Thr Thr Ser Leu Ser Leu His Lys Leu Leu
65           70           75           80

Gln Thr Leu Val Leu Lys Met Leu His Ser Ser Ser Leu Thr Ser Leu
      85           90           95

Leu Lys Thr His Arg Met Cys Lys Tyr Thr Gln Ser Thr Ala Leu Gln
      100          105          110

Glu Leu Leu Ile Gln Gln Trp Ile Gln Phe Met Met Ser Arg Arg Arg
      115          120          125

Leu Leu Ala Cys Leu Cys Lys His Lys Lys Val Ser Thr Asn Leu Cys
      130          135          140

Thr His Ser Phe Arg Lys Lys Gln Val Arg
145           150

```

<210> SEQ ID NO 13

<211> LENGTH: 332

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<220> FEATURE:

<221> NAME/KEY: CDS

<222> LOCATION: (36)..(263)

<223> OTHER INFORMATION:

<400> SEQUENCE: 13

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tgcctttgta agcacaagaa agtgagtacg aactt atg tac tca ttc gtt tcg      53
              Met Tyr Ser Phe Val Ser
              1           5

gaa gaa aca ggt acg tta ata gtt aat agc gta ctt ctt ttt ctt gct      101
Glu Glu Thr Gly Thr Leu Ile Val Asn Ser Val Leu Leu Phe Leu Ala
      10           15           20

ttc gtg gta ttc ttg cta gtc aca cta gcc atc ctt act gcg ctt cga      149
Phe Val Val Phe Leu Leu Val Thr Leu Ala Ile Leu Thr Ala Leu Arg
      25           30           35

ttg tgt gcg tac tgc tgc aat att gtt aac gtg agt tta gta aaa cca      197
Leu Cys Ala Tyr Cys Cys Asn Ile Val Asn Val Ser Leu Val Lys Pro
      40           45           50

acg gtt tac gtc tac tcg cgt gtt aaa aat ctg aac tct tct gaa gga      245
Thr Val Tyr Val Tyr Ser Arg Val Lys Asn Leu Asn Ser Ser Glu Gly
      55           60           65           70

gtt cct gat ctt ctg gtc taaacgaact aactattatt attattctgt      293
Val Pro Asp Leu Leu Val
      75

ttggaacttt aacattgctt atcatggcag acaacggta      332

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<210> SEQ ID NO 14

<211> LENGTH: 76

<212> TYPE: PRT

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 14

-continued

Met	Tyr	Ser	Phe	Val	Ser	Glu	Glu	Thr	Gly	Thr	Leu	Ile	Val	Asn	Ser
1				5					10					15	
Val	Leu	Leu	Phe	Leu	Ala	Phe	Val	Val	Phe	Leu	Leu	Val	Thr	Leu	Ala
			20					25					30		
Ile	Leu	Thr	Ala	Leu	Arg	Leu	Cys	Ala	Tyr	Cys	Cys	Asn	Ile	Val	Asn
		35					40					45			
Val	Ser	Leu	Val	Lys	Pro	Thr	Val	Tyr	Val	Tyr	Ser	Arg	Val	Lys	Asn
	50					55					60				
Leu	Asn	Ser	Ser	Glu	Gly	Val	Pro	Asp	Leu	Leu	Val				
65					70					75					

<210> SEQ ID NO 15
 <211> LENGTH: 332
 <212> TYPE: DNA
 <213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 15

tgcccttgta agcacaagaa agtgagtacg aacttatgta ctcattcggt tcggaagaaa	60
caggtagcgtt aatagttaat agcgtacttc tttttcttgc tttcgtggta ttcttgctag	120
tcacactagc catccttact gcgcttcgat tgtgtgcgta ctgctgcaat attgttaacg	180
tgagtttagt aaaaccaacg gtttacgtct actcgcggtg taaaaatctg aactcttctg	240
aaggagtcc tgatcttctg gtctaaacga actaactatt attattattc tgtttggaac	300
tttaacattg cttatcatgg cagacaacgg ta	332

<210> SEQ ID NO 16
 <211> LENGTH: 708
 <212> TYPE: DNA
 <213> ORGANISM: CORONAVIRUS
 <220> FEATURE:
 <221> NAME/KEY: CDS
 <222> LOCATION: (41)..(703)
 <223> OTHER INFORMATION:

<400> SEQUENCE: 16

tattattatt attctgtttg gaactttaac attgcttacc	atg gca gac aac ggt	55
	Met Ala Asp Asn Gly	
	1 5	
act att acc gtt gag gag ctt aaa caa ctc ctg gaa caa tgg aac cta	103	
Thr Ile Thr Val Glu Glu Leu Lys Gln Leu Leu Glu Gln Trp Asn Leu		
	10 15 20	
gta ata ggt ttc cta ttc cta gcc tgg att atg tta cta caa ttt gcc	151	
Val Ile Gly Phe Leu Phe Leu Ala Trp Ile Met Leu Leu Gln Phe Ala		
	25 30 35	
tat tct aat cgg aac agg ttt ttg tac ata ata aag ctt gtt ttc ctc	199	
Tyr Ser Asn Arg Asn Arg Phe Leu Tyr Ile Ile Lys Leu Val Phe Leu		
	40 45 50	
tgg ctc ttg tgg cca gta aca ctt gct tgt ttt gtg ctt gct gct gtc	247	
Trp Leu Leu Trp Pro Val Thr Leu Ala Cys Phe Val Leu Ala Ala Val		
	55 60 65	
tac aga att aat tgg gtg act ggc ggg att gcg att gca atg gct tgt	295	
Tyr Arg Ile Asn Trp Val Thr Gly Gly Ile Ala Ile Ala Met Ala Cys		
	70 75 80 85	
att gta ggc ttg atg tgg ctt agc tac ttc gtt gct tcc ttc agg ctg	343	
Ile Val Gly Leu Met Trp Leu Ser Tyr Phe Val Ala Ser Phe Arg Leu		
	90 95 100	
ttt gct cgt acc cgc tca atg tgg tca ttc aac cca gaa aca aac att	391	
Phe Ala Arg Thr Arg Ser Met Trp Ser Phe Asn Pro Glu Thr Asn Ile		
	105 110 115	
ctt ctc aat gtg cct ctc cgg ggg aca att gtg acc aga ccg ctc atg	439	

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Leu	Leu	Asn	Val	Pro	Leu	Arg	Gly	Thr	Ile	Val	Thr	Arg	Pro	Leu	Met	
		120					125					130				
gaa	agt	gaa	ctt	gtc	att	ggg	gct	gtg	atc	att	cgt	ggg	cac	ttg	cga	487
Glu	Ser	Glu	Leu	Val	Ile	Gly	Ala	Val	Ile	Ile	Arg	Gly	His	Leu	Arg	
		135				140					145					
atg	gcc	gga	cac	tcc	cta	ggg	cgc	tgt	gac	att	aag	gac	ctg	cca	aaa	535
Met	Ala	Gly	His	Ser	Leu	Gly	Arg	Cys	Asp	Ile	Lys	Asp	Leu	Pro	Lys	
		150				155					160				165	
gag	atc	act	gtg	gct	aca	tca	cga	acg	ctt	tct	tat	tac	aaa	tta	gga	583
Glu	Ile	Thr	Val	Ala	Thr	Ser	Arg	Thr	Leu	Ser	Tyr	Tyr	Lys	Leu	Gly	
				170					175					180		
gcg	tcg	cag	cgt	gta	ggc	act	gat	tca	ggg	ttt	gct	gca	tac	aac	cgc	631
Ala	Ser	Gln	Arg	Val	Gly	Thr	Asp	Ser	Gly	Phe	Ala	Ala	Tyr	Asn	Arg	
			185				190						195			
tac	cgt	att	gga	aac	tat	aaa	tta	aat	aca	gac	cac	gcc	ggg	agc	aac	679
Tyr	Arg	Ile	Gly	Asn	Tyr	Lys	Leu	Asn	Thr	Asp	His	Ala	Gly	Ser	Asn	
		200					205					210				
gac	aat	att	gct	ttg	cta	gta	cag	taagt								708
Asp	Asn	Ile	Ala	Leu	Leu	Val	Gln									
		215				220										

<210> SEQ ID NO 17

<211> LENGTH: 221

<212> TYPE: PRT

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 17

Met	Ala	Asp	Asn	Gly	Thr	Ile	Thr	Val	Glu	Glu	Leu	Lys	Gln	Leu	Leu	
1			5						10					15		
Glu	Gln	Trp	Asn	Leu	Val	Ile	Gly	Phe	Leu	Phe	Leu	Ala	Trp	Ile	Met	
		20					25						30			
Leu	Leu	Gln	Phe	Ala	Tyr	Ser	Asn	Arg	Asn	Arg	Phe	Leu	Tyr	Ile	Ile	
		35					40					45				
Lys	Leu	Val	Phe	Leu	Trp	Leu	Leu	Trp	Pro	Val	Thr	Leu	Ala	Cys	Phe	
		50					55				60					
Val	Leu	Ala	Ala	Val	Tyr	Arg	Ile	Asn	Trp	Val	Thr	Gly	Gly	Ile	Ala	
		65			70				75					80		
Ile	Ala	Met	Ala	Cys	Ile	Val	Gly	Leu	Met	Trp	Leu	Ser	Tyr	Phe	Val	
			85					90						95		
Ala	Ser	Phe	Arg	Leu	Phe	Ala	Arg	Thr	Arg	Ser	Met	Trp	Ser	Phe	Asn	
		100					105						110			
Pro	Glu	Thr	Asn	Ile	Leu	Leu	Asn	Val	Pro	Leu	Arg	Gly	Thr	Ile	Val	
		115					120					125				
Thr	Arg	Pro	Leu	Met	Glu	Ser	Glu	Leu	Val	Ile	Gly	Ala	Val	Ile	Ile	
		130				135					140					
Arg	Gly	His	Leu	Arg	Met	Ala	Gly	His	Ser	Leu	Gly	Arg	Cys	Asp	Ile	
		145			150				155					160		
Lys	Asp	Leu	Pro	Lys	Glu	Ile	Thr	Val	Ala	Thr	Ser	Arg	Thr	Leu	Ser	
			165					170						175		
Tyr	Tyr	Lys	Leu	Gly	Ala	Ser	Gln	Arg	Val	Gly	Thr	Asp	Ser	Gly	Phe	
		180					185					190				
Ala	Ala	Tyr	Asn	Arg	Tyr	Arg	Ile	Gly	Asn	Tyr	Lys	Leu	Asn	Thr	Asp	
		195					200					205				
His	Ala	Gly	Ser	Asn	Asp	Asn	Ile	Ala	Leu	Leu	Val	Gln				
		210				215					220					

<210> SEQ ID NO 18

<211> LENGTH: 769

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<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 18

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cctgatcttc tggctctaac gaactaacta ttattattat tctgtttgga actttaacat    60
tgcttatcat ggcagacaac ggtactatta cgttgagga gcttaacaa ctcttggaac    120
aatggaacct agtaataggt ttcctattcc tagcctggat tatgttacta caatttgacct    180
attctaactcg gaacagggtt ttgtacataa taaagcttgt tttcctctgg ctcttggtggc    240
cagtaaacact tgcttgtttt gtgcttgctg ctgtctacag aattaattgg gtgactggcg    300
ggattgcatg tgcaatggct tgtattgtag gcttgatgtg gcttagctac ttcgttgctt    360
ccttcaggct gtttgctcgt acccgctcaa tgtggtcatt caaccagaa acaaacattc    420
ttctcaatgt gcctctccgg gggacaattg tgaccagacc gctcatggaa agtgaacttg    480
tcattggtgc tgtgatcatt cgtggtcact tgcgaatggc cggacactcc ctaggcgct    540
gtgacattaa ggacctgcca aaagagatca ctgtggctac atcacgaacg ctttcttatt    600
acaaattagg agcgtcgcag cgtgtaggca ctgattcagg ttttgctgca tacaaccgct    660
accgtatttg aaactataaa ttaaatacag accacgccgg tagcaacgac aatattgctt    720
tgctagtaca gtaagtgaca acagatgttt catcttggtg acttccagg    769

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<210> SEQ ID NO 19

<211> LENGTH: 1231

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 19

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taccgtattg gaaactataa attaaataca gaccacgccg gtagcaacga caatattgct    60
ttgctagtac agtaagtgc aacagatggt tcatcttggt gacttccagg ttacaatagc    120
agagatattg attatcatta tgaggacttt caggattgct atttggaatc ttgacgttat    180
aataagttca atagtgcagc aattatttaa gcctctaact aagaagaatt attcggagtt    240
agatgatgaa gaacctatgg agttagatta tccataaaac gaacatgaaa attattctct    300
tcctgacatt gattgtatct acatcttgct agctatatca ctatcaggag tgtgtagtag    360
gtacgactgt actactaaaa gaaccttgcc catcaggaa atacgagggc aattcaccat    420
ttcaccctct tgctgacaat aaatttgcac taacttgcac tagcacacac tttgcttttg    480
cttggtctga cggctactga catacctatc agctgcgtgc aagatcagtt tcacaaaaac    540
ttttcatcag acaagaggag gttcaacaag agctctactc gccacttttt ctcatgtgtg    600
ctgctctagt atttttaata ctttgcttca ccattaagag aaagacagaa tgaatgagct    660
cactttaatt gacttctatt tgtgcttttt agcctttctg ctattccttg ttttaataat    720
gcttattata ttttggtttt cactcgaaat ccaggatcta gaagaacctt gtaccaaagt    780
ctaaacgaac atgaaacttc tcattgtttt gacttgattt tctctatgca gttgcatatg    840
cactgtagta cagcgcgtgt catctaataa acctcatgtg cttgaagatc cttgtaaggt    900
acaacactag gggtaatact tatagcactg cttggccttg tgctctagga aagggttttac    960
cttttcatag atggcacact atggttcaaa catgcacacc taatgttact atcaactgtc    1020
aagatccagc tgggtggtgc cttatagcta ggtgttggtg ccttcatgaa ggtcaccaaa    1080
ctgctgcatt tagagacgta cttgtgtttt taaataaacg aacaaattaa aatgtctgat    1140
aatggacccc aatcaaacca acgtagtgcc ccccgcatca catttggttg acccacagat    1200
tcaactgaca ataaccagaa tggaggacgc a                                1231

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<210> SEQ ID NO 20
 <211> LENGTH: 1242
 <212> TYPE: DNA
 <213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 20

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gcatacaacc gctaccgtat tggaaactat aaattaaata cagaccacgc cggtagcaac    60
gacaatattg ctttgctagt acagtaagtg acaacagatg tttcatcttg ttgacttcca    120
ggttacaata gcagagatat tgattatcat tatgaggact ttcaggattg ctatttggaa    180
tcttgacggt ataataagtt caatagttag acagttattt aagcctctaa ctaagaagaa    240
ttattcggag ttagatgatg aagaacctat ggagttagat tatccataaa acgaacatga    300
aaattattct cttcctgaca ttgattgtat ttacatcttg cgagctatat cactatcagg    360
agtggtgttag aggtacgact gtactactaa aagaaccttg cccatcagga acatacgagg    420
gcaattcacc atttcacctt cttgtcgaca ataaatttgc actaacttgc actagcacac    480
actttgcttt tgcttgctgt gacggtactc gacataccta tcagctgcgt gcaagatcag    540
tttcacaaaa acttttctac agacaagagg aggttcaaca agagctctac tcgccacttt    600
ttctcattgt tgctgctcta gtatttttaa tactttgctt caccattaag agaaagacag    660
aatgaatgag ctacttttaa ttgacttcta tttgtgcttt ttagcctttc tgctattcct    720
tgttttaata atgcttatta tattttggtt ttcactcgaa atccaggatc tagaagaacc    780
ttgtacaaaa gtctaaacga acatgaaact tctcattggt ttgacttgta tttctctatg    840
cagttgcata tgcactgtag tacagcgtg tgcatctaataaacctcatg tgcttgaaga    900
tccttgtaag gtacaacact aggggtaata cttatagcac tgcttggtt tgtgctctag    960
gaaagggttt accttttcat agatggcaca ctatgggttca aacatgcaca cctaattgta   1020
ctatcaactg tcaagatcca gctggtggtg cgcttatagc taggtgttgg taccttcatg   1080
aagggtcacca aactgctgca tttagagacg tacttggtgt tttaaataaa cgaacgaatt   1140
aaaaatgtctg ataatggacc ccaatcaaac caacgtagtg ccccccgcac tacatttggt   1200
ggacccacag attcaactga caataaccag aatggaggac gc                        1242
```

<210> SEQ ID NO 21
 <211> LENGTH: 1231
 <212> TYPE: DNA
 <213> ORGANISM: CORONAVIRUS
 <220> FEATURE:
 <221> NAME/KEY: CDS
 <222> LOCATION: (86)..(274)
 <223> OTHER INFORMATION:

<400> SEQUENCE: 21

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taccgtattg gaaactataa attaaatata gaccacgccg gtagcaacga caatattgct    60
ttgctagtac agtaagtgc aacag atg ttt cat ctt gtt gac ttc cag gtt    112
                               Met Phe His Leu Val Asp Phe Gln Val
                               1                     5
aca ata gca gag ata ttg att atc att atg agg act ttc agg att gct    160
Thr Ile Ala Glu Ile Leu Ile Ile Ile Met Arg Thr Phe Arg Ile Ala
10                     15                     20                     25
att tgg aat ctt gac gtt ata ata agt tca ata gtg aga caa tta ttt    208
Ile Trp Asn Leu Asp Val Ile Ile Ser Ser Ile Val Arg Gln Leu Phe
30                     35                     40
aag cct cta act aag aag aat tat tgc gag tta gat gat gaa gaa cct    256
Lys Pro Leu Thr Lys Lys Asn Tyr Ser Glu Leu Asp Asp Glu Glu Pro
45                     50                     55
```


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atg gag tta gat tat cca taaaacgaac atgaaaatta ttctcttcct      304
Met Glu Leu Asp Tyr Pro
60

gacattgatt gtatttacct ctgtcgagct atatcactat caggagtgtg ttagaggtag      364
gactgtacta ctaaaagaac ctgtcccatc aggaacatac gagggcaatt caccatttca      424
ccctcttgct gacaataaat ttgcactaac ttgcactagc acacactttg cttttgcttg      484
tgctgacggt actcgacata cctatcagct gcgtgcaaga tcagtttcac caaaactttt      544
catcagacaa gaggagggtc aacaagagct ctactcgcca ctttttctca ttgttgctgc      604
tctagtattt ttaatacttt gcttcacat taagagaaag acagaatgaa tgagctcact      664
ttaattgact tctatttggt ctttttagcc tttctgctat tccttgtttt aataatgctt      724
attatatttt ggttttcact cgaaatccag gatctagaag aaccttgtag caaagtctaa      784
acgaacatga aacttctcat tgttttgact tgtattttct tatgcagttg catatgcact      844
gtagtacagc gctgtgcac taataaacct catgtgcttg aagatccttg taaggtagaa      904
cactaggggt aatacttata gcaactgctg gctttgtgct ctaggaaaag ttttaccttt      964
tcatagatgg cacactatgg ttcaaacatg cacaccta gttactatca actgtcaaga     1024
tccagctggt ggtgcgctta tagctagggt ttggtacett catgaagggt accaaactgc     1084
tgcatttaga gacgtacttg ttgttttaaa taaacgaaca aattaaaatg tctgataatg     1144
gaccccaatc aaaccaacgt agtgccccc gcattacatt tgggtggacc acagattcaa     1204
ctgacaataa ccagaatgga ggacgca                                     1231

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<210> SEQ ID NO 22
<211> LENGTH: 63
<212> TYPE: PRT
<213> ORGANISM: CORONAVIRUS

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<400> SEQUENCE: 22

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```

Met Phe His Leu Val Asp Phe Gln Val Thr Ile Ala Glu Ile Leu Ile
1           5           10          15

Ile Ile Met Arg Thr Phe Arg Ile Ala Ile Trp Asn Leu Asp Val Ile
20          25          30

Ile Ser Ser Ile Val Arg Gln Leu Phe Lys Pro Leu Thr Lys Lys Asn
35          40          45

Tyr Ser Glu Leu Asp Asp Glu Glu Pro Met Glu Leu Asp Tyr Pro
50          55          60

```

```

<210> SEQ ID NO 23
<211> LENGTH: 1231
<212> TYPE: DNA
<213> ORGANISM: CORONAVIRUS
<220> FEATURE:
<221> NAME/KEY: CDS
<222> LOCATION: (285) .. (650)
<223> OTHER INFORMATION:

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<400> SEQUENCE: 23

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```

taccgtattg gaaactataa attaaatata gaccacgccg gtagcaacga caatattgct      60
ttgctagtac agtaagttag aacagatggt tcatcttggt gacttccagg ttacaatagc     120
agagatattg attatcatta tgaggacttt caggattgct atttggaaac ttgacgttat     180
aataagttca atagttagac aattatttaa gcctctaact aagaagaatt attcgaggtt     240
agatgatgaa gaacctatgg agtttagatta tccataaaac gaac atg aaa att att     296
Met Lys Ile Ile
1

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ctc ttc ctg aca ttg att gta ttt aca tct tgc gag cta tat cac tat    344
Leu Phe Leu Thr Leu Ile Val Phe Thr Ser Cys Glu Leu Tyr His Tyr
5          10          15          20

cag gag tgt gtt aga ggt acg act gta cta cta aaa gaa cct tgc cca    392
Gln Glu Cys Val Arg Gly Thr Thr Val Leu Leu Lys Glu Pro Cys Pro
          25          30          35

tca gga aca tac gag ggc aat tca cca ttt cac cct ctt gct gac aat    440
Ser Gly Thr Tyr Glu Gly Asn Ser Pro Phe His Pro Leu Ala Asp Asn
          40          45          50

aaa ttt gca cta act tgc act agc aca cac ttt gct ttt gct tgt gct    488
Lys Phe Ala Leu Thr Cys Thr Ser Thr His Phe Ala Phe Ala Cys Ala
          55          60          65

gac ggt act cga cat acc tat cag ctg cgt gca aga tca gtt tca cca    536
Asp Gly Thr Arg His Thr Tyr Gln Leu Arg Ala Arg Ser Val Ser Pro
          70          75          80

aaa ctt ttc atc aga caa gag gag gtt caa caa gag ctc tac tcg cca    584
Lys Leu Phe Ile Arg Gln Glu Glu Val Gln Gln Glu Leu Tyr Ser Pro
          85          90          95          100

ctt ttt ctc att gtt gct gct cta gta ttt tta ata ctt tgc ttc acc    632
Leu Phe Leu Ile Val Ala Ala Leu Val Phe Leu Ile Leu Cys Phe Thr
          105          110          115

att aag aga aag aca gaa tgaatgagct cactttaatt gacttctatt    680
Ile Lys Arg Lys Thr Glu
          120

tgtgtctttt agcctttctg ctattccttg ttttaataat gcttattata ttttggtttt    740

cactcgaaat ccaggatcta gaagaacctt gtaccaaagt ctaaactgaac atgaaacttc    800

tcattgtttt gacttgattt tctctatgca gttgcatatg cactgtagta cagcgtgtg    860

catctaataa acctcatgtg cttgaagatc cttgtaaggt acaacactag gggtaatact    920

tatagcactg cttggttttg tgctctagga aagggttttac cttttcatag atggcacact    980

atggttcaaa catgcacacc taatgttact atcaactgtc aagatccagc tgggtgtgctg    1040

cttatagcta ggtgttggtta ccttcatgaa ggtcaccaaa ctgctgcatt tagagacgta    1100

cttggtgttt taaataaacg aacaaattaa aatgtctgat aatggacccc aatcaaacca    1160

acgtagtgcc ccccgcatcattttggtgg acccacagat tcaactgaca ataaccagaa    1220

tggaggacgc a    1231

```

<210> SEQ ID NO 24

<211> LENGTH: 122

<212> TYPE: PRT

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 24

```

Met Lys Ile Ile Leu Phe Leu Thr Leu Ile Val Phe Thr Ser Cys Glu
1          5          10          15

Leu Tyr His Tyr Gln Glu Cys Val Arg Gly Thr Thr Val Leu Leu Lys
          20          25          30

Glu Pro Cys Pro Ser Gly Thr Tyr Glu Gly Asn Ser Pro Phe His Pro
          35          40          45

Leu Ala Asp Asn Lys Phe Ala Leu Thr Cys Thr Ser Thr His Phe Ala
          50          55          60

Phe Ala Cys Ala Asp Gly Thr Arg His Thr Tyr Gln Leu Arg Ala Arg
          65          70          75          80

Ser Val Ser Pro Lys Leu Phe Ile Arg Gln Glu Glu Val Gln Gln Glu
          85          90          95

Leu Tyr Ser Pro Leu Phe Leu Ile Val Ala Ala Leu Val Phe Leu Ile

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100	105	110	
Leu Cys Phe Thr Ile Lys Arg Lys Thr Glu			
115	120		
<210> SEQ ID NO 25 <211> LENGTH: 1231 <212> TYPE: DNA <213> ORGANISM: CORONAVIRUS <220> FEATURE: <221> NAME/KEY: CDS <222> LOCATION: (650)..(781) <223> OTHER INFORMATION: <400> SEQUENCE: 25			
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ctgctctagt atttttaata ctttgcttca ccattaagag aaagacaga atg aat gag		658	
		Met Asn Glu	
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ctc act tta att gac ttc tat ttg tgc ttt tta gcc ttt ctg cta ttc		706	
Leu Thr Leu Ile Asp Phe Tyr Leu Cys Phe Leu Ala Phe Leu Leu Phe			
5 10 15			
ctt gtt tta ata atg ctt att ata ttt tgg ttt tca ctc gaa atc cag		754	
Leu Val Leu Ile Met Leu Ile Ile Phe Trp Phe Ser Leu Glu Ile Gln			
20 25 30 35			
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Asp Leu Glu Glu Pro Cys Thr Lys Val			
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cattgttttg acttgtattt ctctatgcag ttgcatatgc actgtagtac agcgtgtgc		861	
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<210> SEQ ID NO 27

<211> LENGTH: 1231

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<220> FEATURE:

<221> NAME/KEY: CDS

<222> LOCATION: (791) .. (907)

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 ctaaacgaac atg aaa ctt ctc att gtt ttg act tgt att tct cta tgc 829
 Met Lys Leu Leu Ile Val Leu Thr Cys Ile Ser Leu Cys
 1 5 10
 agt tgc ata tgc act gta gta cag cgc tgt gca tct aat aaa cct cat 877
 Ser Cys Ile Cys Thr Val Val Gln Arg Cys Ala Ser Asn Lys Pro His
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 Val Leu Glu Asp Pro Cys Lys Val Gln His
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<211> LENGTH: 39

<212> TYPE: PRT

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 28

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Cys Thr Val Val Gln Arg Cys Ala Ser Asn Lys Pro His Val Leu Glu

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cactgtagta cagcgtgtg catctaataa acctc atg tgc ttg aag atc ctt			893
	Met Cys Leu Lys Ile Leu		
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Val Arg Tyr Asn Thr Arg Gly Asn Thr Tyr Ser Thr Ala Trp Leu Cys			
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Ala Leu Gly Lys Val Leu Pro Phe His Arg Trp His Thr Met Val Gln			
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Thr Cys Thr Pro Asn Val Thr Ile Asn Cys Gln Asp Pro Ala Gly Gly			
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gcg ctt ata gct agg tgt tgg tac ctt cat gaa ggt cac caa act gct			1085
Ala Leu Ile Ala Arg Cys Trp Tyr Leu His Glu Gly His Gln Thr Ala			
55 60 65 70			
gca ttt aga gac gta ctt gtt gtt tta aat aaa cga aca aat			1127
Ala Phe Arg Asp Val Leu Val Val Leu Asn Lys Arg Thr Asn			
75 80			
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 35 40 45
 Gln Asp Pro Ala Gly Gly Ala Leu Ile Ala Arg Cys Trp Tyr Leu His
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actttctcga	gctcgtatg	gatgaattca	tacagcgata	taagctcgag	ggctatgcct	19980
tcgaacacat	cgtttatgga	gatttcagtc	atggacaact	tggcggctct	catttaatga	20040
taggcttagc	caagcgtca	caagattcac	cacttaaatt	agaggatttt	atccctatgg	20100
acagcacagt	gaaaaattac	ttcataacag	atgcgcaaac	aggttcatca	aatgtgtgt	20160
gttctgtgat	tgatctttta	cttgatgact	ttgtcgagat	aataaagtca	caagatttgt	20220
cagtgatctc	aaaagtgggc	aagggttaca	ttgactatgc	tgaatttca	ttcatgcttt	20280
ggtgtaagga	tggacatgtt	gaaaccttct	acccaaaact	acaagcaagt	caagcgtggc	20340
aaccaggtgt	tgcatgacct	aacttgatca	agatgcaaag	aatgcttctt	gaaaagtgtg	20400
accttcagaa	ttatggtgaa	aatgctgtta	tacaaaaagg	aataatgatg	aatgtcgcaa	20460
agtatactca	actgtgtcaa	tacttaaata	cacttacttt	agctgtaccc	tacaacatga	20520
gagtatttca	ctttggtgct	ggctctgata	aaggagttgc	accaggtaca	gctgtgctca	20580
gacaatgggt	gccaaactgc	acactacttg	tcgattcaga	tcttaatgac	ttcgtctccg	20640
acgcagattc	tactttaatt	ggagactgtg	caacagtaca	tacggcta	aatgggacc	20700
ttattattag	cgatatgtat	gaccctagga	ccaaacatgt	gacaaaagag	aatgactcta	20760
aagaaggggt	tttcaacttat	ctgtgtggat	ttataaagca	aaaactagcc	ctgggtgggt	20820

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ctatagctgt aaagataaca gagcattctt ggaatgctga cctttacaag cttatgggcc 20880
atcttctcatg gtggacagct tttgttacaa atgtaaatgc atcatcatcg gaagcatttt 20940
taattggggc taactatctt ggcaagccga aggaacaaat tgatggctat accatgcatg 21000
ctaactacat tttctggagg aacacaaatc ctatccagtt gtcttcctat tcactctttg 21060
acatgagcaa atttctctct aaattaagag gaactgctgt aatgtctctt aaggagaatc 21120
aaatcaatga tatgatttat tctcttctgg aaaaaggtag gcttatcatt agagaaaaca 21180
acagagtgtt ggtttcaagt gatattcttg ttaacaacta a 21221

```

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<210> SEQ ID NO 32
<211> LENGTH: 297
<212> TYPE: DNA
<213> ORGANISM: CORONAVIRUS

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<400> SEQUENCE: 32

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```

atggacccca atcaaaccaa cgtagtgtccc cccgcattac atttgggtgga cccacagatt 60
caactgacaa taaccagaat ggaggacgca atgggggcaag gccaaaacag cgccgacccc 120
aagggtttacc caataatact gcgtcttggt tcacagctct cactcagcat ggcaaggagg 180
aacttagatt ccctcgaggc cagggcgctc caatcaacac caatagtggg ccagatgacc 240
aaattggcta ctaccgaaga gctaccgcac gagttcgtgg tggtgacggc aaaatga 297

```

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<210> SEQ ID NO 33
<211> LENGTH: 98
<212> TYPE: PRT
<213> ORGANISM: CORONAVIRUS

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<400> SEQUENCE: 33

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```

Met Asp Pro Asn Gln Thr Asn Val Val Pro Pro Ala Leu His Leu Val
1          5          10          15
Asp Pro Gln Ile Gln Leu Thr Ile Thr Arg Met Glu Asp Ala Met Gly
20          25          30
Gln Gly Gln Asn Ser Ala Asp Pro Lys Val Tyr Pro Ile Ile Leu Arg
35          40          45
Leu Gly Ser Gln Leu Ser Leu Ser Met Ala Arg Arg Asn Leu Asp Ser
50          55          60
Leu Glu Ala Arg Ala Phe Gln Ser Thr Pro Ile Val Val Gln Met Thr
65          70          75          80
Lys Leu Ala Thr Thr Glu Glu Leu Pro Asp Glu Phe Val Val Val Thr
85          90          95

```

```

Ala Lys

```

```

<210> SEQ ID NO 34
<211> LENGTH: 213
<212> TYPE: DNA
<213> ORGANISM: CORONAVIRUS

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<400> SEQUENCE: 34

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```

atgctgccac cgtgctacaa cttcctcaag gaacaacatt gccaaaaggc ttctacgcag 60
agggaagcag aggcggcagt caagcctctt ctcgctcttc atcacgtagt cgcggtaatt 120
caagaaattc aactcctggc agcagtaggg gaaattctcc tgctcgaatg gctagcggag 180
gtggtgaaac tgccctcgcg ctattgctgc tag 213

```

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<210> SEQ ID NO 35
<211> LENGTH: 70
<212> TYPE: PRT

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<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 35

```

Met Leu Pro Pro Cys Tyr Asn Phe Leu Lys Glu Gln His Cys Gln Lys
1           5           10           15

Ala Ser Thr Gln Arg Glu Ala Glu Ala Ala Val Lys Pro Leu Leu Ala
20           25           30

Pro His His Val Val Ala Val Ile Gln Glu Ile Gln Leu Leu Ala Ala
35           40           45

Val Gly Glu Ile Leu Leu Leu Glu Trp Leu Ala Glu Val Val Lys Leu
50           55           60

Pro Ser Arg Tyr Cys Cys
65           70

```

<210> SEQ ID NO 36

<211> LENGTH: 1377

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<220> FEATURE:

<221> NAME/KEY: CDS

<222> LOCATION: (67)..(1335)

<223> OTHER INFORMATION:

<400> SEQUENCE: 36

```

atgaagggtca ccaaactgct gcatttagag acgtacttgt tgttttaaat aaacgaacaa      60

attaaa atg tct gat aat gga ccc caa tca aac caa cgt agt gcc ccc      108
Met Ser Asp Asn Gly Pro Gln Ser Asn Gln Arg Ser Ala Pro
1           5           10

cgc att aca ttt ggt gga ccc aca gat tca act gac aat aac cag aat      156
Arg Ile Thr Phe Gly Gly Pro Thr Asp Ser Thr Asp Asn Asn Gln Asn
15          20          25          30

gga gga cgc aat ggg gca agg cca aaa cag cgc cga ccc caa ggt tta      204
Gly Gly Arg Asn Gly Ala Arg Pro Lys Gln Arg Arg Pro Gln Gly Leu
35          40          45

ccc aat aat act gcg tct tgg ttc aca gct ctc act cag cat ggc aag      252
Pro Asn Asn Thr Ala Ser Trp Phe Thr Ala Leu Thr Gln His Gly Lys
50          55          60

gag gaa ctt aga ttc cct cga ggc cag ggc gtt cca atc aac acc aat      300
Glu Glu Leu Arg Phe Pro Arg Gly Gln Gly Val Pro Ile Asn Thr Asn
65          70          75

agt ggt cca gat gac caa att ggc tac tac cga aga gct acc cga cga      348
Ser Gly Pro Asp Asp Gln Ile Gly Tyr Tyr Arg Arg Ala Thr Arg Arg
80          85          90

gtt cgt ggt ggt gac ggc aaa atg aaa gag ctc agc ccc aga tgg tac      396
Val Arg Gly Gly Asp Gly Lys Met Lys Glu Leu Ser Pro Arg Trp Tyr
95          100         105         110

ttc tat tac cta gga act ggc cca gaa gct tca ctt ccc tac ggc gct      444
Phe Tyr Tyr Leu Gly Thr Gly Pro Glu Ala Ser Leu Pro Tyr Gly Ala
115         120         125

aac aaa gaa ggc atc gta tgg gtt gca act gag gga gcc ttg aat aca      492
Asn Lys Glu Gly Ile Val Trp Val Ala Thr Glu Gly Ala Leu Asn Thr
130         135         140

ccc aaa gac cac att ggc acc cgc aat cct aat aac aat gct gcc acc      540
Pro Lys Asp His Ile Gly Thr Arg Asn Pro Asn Asn Asn Ala Ala Thr
145         150         155

gtg cta caa ctt cct caa gga aca aca ttg cca aaa ggc ttc tac gca      588
Val Leu Gln Leu Pro Gln Gly Thr Thr Leu Pro Lys Gly Phe Tyr Ala
160         165         170

gag gga agc aga ggc ggc agt caa gcc tct tct cgc tcc tca tca cgt      636
Glu Gly Ser Arg Gly Gly Ser Gln Ala Ser Ser Arg Ser Ser Ser Arg
175         180         185         190

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agt cgc ggt aat tca aga aat tca act cct ggc agc agt agg gga aat      684
Ser Arg Gly Asn Ser Arg Asn Ser Thr Pro Gly Ser Ser Arg Gly Asn
      195                200                205

tct cct gct cga atg gct agc gga ggt ggt gaa act gcc ctc gcg cta      732
Ser Pro Ala Arg Met Ala Ser Gly Gly Gly Glu Thr Ala Leu Ala Leu
      210                215                220

ttg ctg cta gac aga ttg aac cag ctt gag agc aaa gtt tct ggt aaa      780
Leu Leu Leu Asp Arg Leu Asn Gln Leu Glu Ser Lys Val Ser Gly Lys
      225                230                235

ggc caa caa caa caa ggc caa act gtc act aag aaa tct gct gct gag      828
Gly Gln Gln Gln Gln Gly Gln Thr Val Thr Lys Lys Ser Ala Ala Glu
      240                245                250

gca tct aaa aag cct cgc caa aaa cgt act gcc aca aaa cag tac aac      876
Ala Ser Lys Lys Pro Arg Gln Lys Arg Thr Ala Thr Lys Gln Tyr Asn
      255                260                265                270

gtc act caa gca ttt ggg aga cgt ggt cca gaa caa acc caa gga aat      924
Val Thr Gln Ala Phe Gly Arg Arg Gly Pro Glu Gln Thr Gln Gly Asn
      275                280                285

ttc ggg gac caa gac cta atc aga caa gga act gat tac aaa cat tgg      972
Phe Gly Asp Gln Asp Leu Ile Arg Gln Gly Thr Asp Tyr Lys His Trp
      290                295                300

ccg caa att gca caa ttt gct cca agt gcc tct gca ttc ttt gga atg     1020
Pro Gln Ile Ala Gln Phe Ala Pro Ser Ala Ser Ala Phe Phe Gly Met
      305                310                315

tca cgc att ggc atg gaa gtc aca cct tcg gga aca tgg ctg act tat     1068
Ser Arg Ile Gly Met Glu Val Thr Pro Ser Gly Thr Trp Leu Thr Tyr
      320                325                330

cat gga gcc att aaa ttg gat gac aaa gat cca caa ttc aaa gac aac     1116
His Gly Ala Ile Lys Leu Asp Asp Lys Asp Pro Gln Phe Lys Asp Asn
      335                340                345                350

gtc ata ctg ctg aac aag cac att gac gca tac aaa aca ttc cca cca     1164
Val Ile Leu Leu Asn Lys His Ile Asp Ala Tyr Lys Thr Phe Pro Pro
      355                360                365

aca gag cct aaa aag gac aaa aag aaa aag act gat gaa gct cag cct     1212
Thr Glu Pro Lys Lys Asp Lys Lys Lys Lys Thr Asp Glu Ala Gln Pro
      370                375                380

ttg ccg cag aga caa aag aag cag ccc act gtg act ctt ctt cct gcg     1260
Leu Pro Gln Arg Gln Lys Lys Gln Pro Thr Val Thr Leu Leu Pro Ala
      385                390                395

gct gac atg gat gat ttc tcc aga caa ctt caa aat tcc atg agt gga     1308
Ala Asp Met Asp Asp Phe Ser Arg Gln Leu Gln Asn Ser Met Ser Gly
      400                405                410

gct tct gct gat tca act cag gca taa acactcatga tgaccacaca           1355
Ala Ser Ala Asp Ser Thr Gln Ala
      415                420

aggcagatgg gctatgtaaa cg                                           1377

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<210> SEQ ID NO 37
<211> LENGTH: 422
<212> TYPE: PRT
<213> ORGANISM: CORONAVIRUS

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<400> SEQUENCE: 37

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```

Met Ser Asp Asn Gly Pro Gln Ser Asn Gln Arg Ser Ala Pro Arg Ile
1      5      10      15

Thr Phe Gly Gly Pro Thr Asp Ser Thr Asp Asn Asn Gln Asn Gly Gly
20     25     30

Arg Asn Gly Ala Arg Pro Lys Gln Arg Arg Pro Gln Gly Leu Pro Asn
35     40     45

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Asn	Thr	Ala	Ser	Trp	Phe	Thr	Ala	Leu	Thr	Gln	His	Gly	Lys	Glu	Glu
50					55					60					
Leu	Arg	Phe	Pro	Arg	Gly	Gln	Gly	Val	Pro	Ile	Asn	Thr	Asn	Ser	Gly
65					70				75					80	
Pro	Asp	Asp	Gln	Ile	Gly	Tyr	Tyr	Arg	Arg	Ala	Thr	Arg	Arg	Val	Arg
			85					90						95	
Gly	Gly	Asp	Gly	Lys	Met	Lys	Glu	Leu	Ser	Pro	Arg	Trp	Tyr	Phe	Tyr
			100					105					110		
Tyr	Leu	Gly	Thr	Gly	Pro	Glu	Ala	Ser	Leu	Pro	Tyr	Gly	Ala	Asn	Lys
		115					120					125			
Glu	Gly	Ile	Val	Trp	Val	Ala	Thr	Glu	Gly	Ala	Leu	Asn	Thr	Pro	Lys
		130					135				140				
Asp	His	Ile	Gly	Thr	Arg	Asn	Pro	Asn	Asn	Asn	Ala	Ala	Thr	Val	Leu
145					150					155					160
Gln	Leu	Pro	Gln	Gly	Thr	Thr	Leu	Pro	Lys	Gly	Phe	Tyr	Ala	Glu	Gly
			165					170						175	
Ser	Arg	Gly	Gly	Ser	Gln	Ala	Ser	Ser	Arg	Ser	Ser	Ser	Arg	Ser	Arg
		180						185					190		
Gly	Asn	Ser	Arg	Asn	Ser	Thr	Pro	Gly	Ser	Ser	Arg	Gly	Asn	Ser	Pro
		195					200					205			
Ala	Arg	Met	Ala	Ser	Gly	Gly	Gly	Glu	Thr	Ala	Leu	Ala	Leu	Leu	Leu
		210					215				220				
Leu	Asp	Arg	Leu	Asn	Gln	Leu	Glu	Ser	Lys	Val	Ser	Gly	Lys	Gly	Gln
225					230					235					240
Gln	Gln	Gln	Gly	Gln	Thr	Val	Thr	Lys	Lys	Ser	Ala	Ala	Glu	Ala	Ser
			245					250					255		
Lys	Lys	Pro	Arg	Gln	Lys	Arg	Thr	Ala	Thr	Lys	Gln	Tyr	Asn	Val	Thr
		260					265						270		
Gln	Ala	Phe	Gly	Arg	Arg	Gly	Pro	Glu	Gln	Thr	Gln	Gly	Asn	Phe	Gly
		275					280				285				
Asp	Gln	Asp	Leu	Ile	Arg	Gln	Gly	Thr	Asp	Tyr	Lys	His	Trp	Pro	Gln
		290				295					300				
Ile	Ala	Gln	Phe	Ala	Pro	Ser	Ala	Ser	Ala	Phe	Phe	Gly	Met	Ser	Arg
305					310					315					320
Ile	Gly	Met	Glu	Val	Thr	Pro	Ser	Gly	Thr	Trp	Leu	Thr	Tyr	His	Gly
			325						330					335	
Ala	Ile	Lys	Leu	Asp	Asp	Lys	Asp	Pro	Gln	Phe	Lys	Asp	Asn	Val	Ile
		340						345					350		
Leu	Leu	Asn	Lys	His	Ile	Asp	Ala	Tyr	Lys	Thr	Phe	Pro	Pro	Thr	Glu
		355					360					365			
Pro	Lys	Lys	Asp	Lys	Lys	Lys	Lys	Thr	Asp	Glu	Ala	Gln	Pro	Leu	Pro
		370				375					380				
Gln	Arg	Gln	Lys	Lys	Gln	Pro	Thr	Val	Thr	Leu	Leu	Pro	Ala	Ala	Asp
385					390					395					400
Met	Asp	Asp	Phe	Ser	Arg	Gln	Leu	Gln	Asn	Ser	Met	Ser	Gly	Ala	Ser
			405					410						415	
Ala	Asp	Ser	Thr	Gln	Ala										
			420												

<210> SEQ ID NO 38

<211> LENGTH: 1377

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 38

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atgaaggtca ccaaactgct gcatttagag acgtacttgt tgttttaaat aaacgaacaa	60
attaaaaatgt ctgataatgg accccaatca aaccaacgta gtgcccccg cattacattt	120
ggtggaccca cagattcaac tgacaataac cagaatggag gacgcaatgg ggcaaggcca	180
aaacagcgcc gacccaagg ttacccaat aatactgcgt cttgggtcac agctctcact	240
cagcatggca aggaggaact tagattccct cgaggccagg gcgttccaat caacaccaat	300
agtggccag atgaccaaat tggctactac cgaagagcta cccgacgagt tcgtggtggt	360
gacggcaaaa tgaaagagct cagccccaga tggctacttct attacctagg aactggccca	420
gaagcttcac ttcctacagg cgctaacaaa gaaggcatcg tatgggttgc aactgaggga	480
gccttgaata caccacaaaga ccacattggc acccgcaatc ctaataacaa tgtgcccacc	540
gtgctacaac ttcctcaagg aacaacattg ccaaaaggct tctacgcaga gggaagcaga	600
ggcggcagtc aagcctcttc tcgctcctca tcacgtagtc gcgtaattc aagaaattca	660
actcctggca gcagtagggg aaattctcct gctcgaatgg ctacggagg tggtgaaact	720
gccctcgcgc tattgctgct agacagattg aaccagcttg agagcaaagt ttctggtaaa	780
ggccaacaac aacaaggcca aactgtcact aagaaatctg ctgctgaggc atctaaaaag	840
cctcgccaaa aacgtactgc cacaaaacag tacaacgtca ctcaagcatt tgggagacgt	900
ggtccagaac aaaccaagg aaatttcggg gaccaagacc taatcagaca aggaactgat	960
tacaaaacatt ggccgaaaat tgcacaattt gctccaagtg cctctgcatt ctttggaatg	1020
tcacgcattg gcatggaagt cacaccttcg ggaacatggc tgacttatca tggagccatt	1080
aaattggatg acaaagatcc acaattcaaa gacaacgtca tactgctgaa caagcacatt	1140
gacgcataca aaacattccc accaacagag cctaaaaagg acaaaaagaa aaagactgat	1200
gaagctcagc ctttgccgca gagacaaaag aagcagccca ctgtgactct tcttctcg	1260
gctgacatgg atgattttct cagacaactt caaaattcca tgagtggagc ttctgctgat	1320
tcaactcagg cataaacact catgatgacc acacaaggca gatgggctat gtaaacg	1377

<210> SEQ ID NO 39
 <211> LENGTH: 204
 <212> TYPE: DNA
 <213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 39

atattaggtt ttacctacc caggaaaagc caaccaacct cgatctcttg tagatctgtt	60
ctctaaacga actttaaaat ctgtgtagct gtgctcggc tgcattgcta gtgcacctac	120
gcagtataaa caataataaa ttttactgtc gttgacaaga aacgagtaac tcgtccctct	180
tctgcagact gcttacgggt tcgt	204

<210> SEQ ID NO 40
 <211> LENGTH: 809
 <212> TYPE: DNA
 <213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 40

actcaagcat tggggagacg tgggtccagaa caaacccaag gaaatttcgg ggaccaagac	60
ctaatacagac aaggaaactga ttacaaacat tggccgcaaa ttgcacaatt tgcctcaagt	120
gcctctgcat tctttggaat gtcacgcatt ggcatggaag tcacaccttc gggaacatgg	180
ctgacttatc atggagccat taaattggat gacaaagatc cacaattcaa agacaacgtc	240
atactgctga acaagcacat tgacgcatac aaaacattcc caccaacaga gcctaaaaag	300

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gacaaaaaga aaaagactga tgaagctcag cctttgccgc agagacaaaa gaagcagccc	360
actgtgactc ttcttctctgc ggctgacatg gatgatttct ccagacaact tcaaaattcc	420
atgagtggag cttctgctga ttcaactcag gcataaacac tcatgatgac cacacaaggc	480
agatgggcta tgtaaactgt ttcgcaattc cgtttacgat acatagtcta ctcttgtgca	540
gaatgaattc tcgtaactaa acagcacaag taggttttagt taactttaat ctccatagc	600
aatctttaat caatgtgtaa cattagggag gacttgaaag agccaccaca ttttcatcga	660
ggccacgcgg agtacgatcg agggtagagt gaataatgct agggagagct gcctatatgg	720
aagagcccta atgtgtaaaa ttaatttttag tagtgctatc cccatgtgat tttaatagct	780
tcttaggaga atgacaaaaa aaaaaaaaa	809

<210> SEQ ID NO 41
 <211> LENGTH: 448
 <212> TYPE: DNA
 <213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 41

aatgaacaca tagggctgtt caagctgggg cagtacgcct tttccagct ctactagacc	60
acaagtgcc a tttttgaggt gttcaactgc ctccgatagg gcctcttcca cagagtcccc	120
gaagccacgc actagcacgt ctctaactcg aaggacaggg aaactgagtt ggacgtgtgt	180
tttctcgttg acaccaagaa caaggctctc catcttacct ttccgtcaca cccggacgaa	240
acctaggtat gctgatgatc gactgcaaca cggacgaaac cgtaagcagt ctgcagaaga	300
gggacgagtt actcgtttct tgtcaacgac agtaaaattt attattgttt atactgcgta	360
gggtgcactag gcatgcagcc gagcgacagc tacacagatt ttaaagttcg tttagagaac	420
agatctacaa gagatcgagg ttggttgg	448

<210> SEQ ID NO 42
 <211> LENGTH: 2033
 <212> TYPE: DNA
 <213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 42

atacctaggt ttccgtccggg tgtgaccgaa aggtaagatg gagagccttg ttcttggtgt	60
caacgagaaa acacacgtcc aactcagttt gcctgtcctt caggtagag acgtgctagt	120
gcgtggcttc ggggactctg tggaagaggg cctatcgagg gcacgtgaac acctcaaaaa	180
tggcacttgt ggtctagtag agctggaaaa aggcgtactg cccagccttg aacagcccta	240
tgtgttcatt aaacgttctg atgccttaag caccaatcac ggccacaagg tcgttgagct	300
ggttgcagaa atggacggca ttcagtacgg tcgtagcggg ataacactgg gagtactcgt	360
gccacatgtg ggcgaaaccc caattgcata ccgcaatgtt cttcttcgta agaacggtaa	420
taagggagcc ggtggtcata gctatggcat cgatctaaag tcttatgact taggtgacga	480
gcttggcact gatcccatgg aagattatga acaaaactgg aacactaagc atggcagtggt	540
tgcactccgt gaactcactc gtgagctcaa tggaggtgca gtcactcgtc atgtcgacaa	600
caattttctgt ggccagatg ggtaccctct tgattgcac aaagattttc tcgcacgcgc	660
gggcaagtca atgtgcactc tttccgaaca acttgattac atcgagtcga agagaggtgt	720
ctactgctgc cgtgaccatg agcatgaaat tgcttggttc actgagcgtc ctgataagag	780
ctacgagcac cagacaccct tcgaaattaa gagtgccaag aaatttgaca ctttcaaagg	840
ggaatgcccc aagtttgtgt ttctctttaa ctcaaaagtc aaagtcattc aaccacgtgt	900

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tgaaaagaaa aagactgagg gtttcatggg gcgtatacgc tctgtgtacc ctgttgcatc	960
tcacacaggag tgtaacaata tgcacttgtc taccttgatg aaatgtaatc attgcgatga	1020
agtttcatgg cagacgtgcg actttctgaa agccacttgt gaacattgtg gcaactgaaaa	1080
tttagttatt gaaggaccta ctacatgtgg gtacctacct actaatgctg tagtgaaaat	1140
gccatgtcct gcctgtcaag acccagagat tggacctgag catagtgttg cagattatca	1200
caaccactca aacattgaaa ctgcactccg caagggaggt aggactagat gttttggagg	1260
ctgtgtgttt gcctatgttg gctgtctataa taagcgtgcc tactgggttc ctgctgctag	1320
tgctgatatt ggctcaggcc atactggcat tactgggtgac aatgtggaga ccttgaatga	1380
ggatctcctt gagatactga gtcgtgaacg tgttaacatt aacattgttg gcgattttca	1440
tttgaatgaa gaggttgcca tcattttggc atctttctct gcttctacaa gtgcctttat	1500
tgacactata aagagtcttg attacaagtc tttcaaaacc attgttgagt cctgcggtaa	1560
ctataaagtt accaagggaa agcccgtaaa aggtgcttgg aacattggac aacagagatc	1620
agttttaaca ccactgtgtg gttttccctc acaggctgct ggtgttatca gatcaatttt	1680
tgcgcgacac ctgtatgcag caaaccactc aattcctgat ttgcaaagag cagctgtcac	1740
catacttgat ggtatttctg aacagtcatt acgtcttgtc gacgccatgg tttatacttc	1800
agacctgctc accaacagtg tcattattat ggcatatgta actgggtggc ttgtacaaca	1860
gacttctcag tggttgtcta atcttttggg cactactggt gaaaaactca ggcctatctt	1920
tgaatggatt gaggggaaac ttagtgcagg agttgaattt ctcaaggatg cttgggagat	1980
tctcaaatct ctcattacag gtgtttttga catcgtcaag ggtcaaatac agg	2033

<210> SEQ ID NO 43

<211> LENGTH: 2018

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 43

ggattgaggc gaaacttagt gcaggagttg aatttctcaa ggatgcttgg gagattctca	60
aatttctcat tacagggtgt tttgacatcg tcaagggtca aatacagggt gcttcagata	120
acatcaagga ttgtgtaaaa tgcttcattg atgttggtta caaggcactc gaaatgtgca	180
ttgatcaagt cactatcgct ggcgcaaagt tgcgatcact caacttaggt gaagtcttca	240
tcgctcaaag caagggaact taccgctcagt gtatacgtgg caaggagcag ctgcaactac	300
tcatgcctct taaggcacca aaagaagtaa cctttcttga aggtgattca catgacacag	360
tacttacctc tgaggaggtt gttctcaaga acggtgaact cgaagcactc gagacgcccg	420
ttgatagctt cacaatgga gctatcgttg gcacaccagt ctgtgtaaat ggcctcatgc	480
tcttagagat taaggacaaa gaacaatact gcgcattgtc tcctgggttta ctggctacaa	540
acaatgtctt tcgcttaaaa gggggtgcac caattaaagg tgtaaccttt ggagaagata	600
ctgtttggga agttcaagggt tacaagaatg tgagaatcac atttgagctt gatgaacgtg	660
ttgacaaagt gcttaatgaa aagtgcctctg tctacactgt tgaatccggt accgaagtta	720
ctgagtttgc atgtgttgta gcagaggctg ttgtgaagac tttacaacca gtttctgac	780
tccttaccaa catgggtatt gatcttgatg agtggagtgt agctacattc tacttatttg	840
atgatgctgg tgaagaaaaa ttttcatcac gtatgtattg ttcccttttac cctccagatg	900
aggaagaaga ggacgatgca gagtgtgagg aagaagaaat tgatgaaacc tgtgaacatg	960
agtacggtac agaggatgat tatcaaggtc tccctctgga atttgggtgcc tcagctgaaa	1020

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cagttcaggt	tgaggaagaa	gaagaggaag	actggctgga	tgatactact	gagcaatcag	1080
agattgagcc	agaaccagaa	cctacacctg	aagaaccagt	taatcagttt	actgggttatt	1140
taaaacttac	tgacaatgtt	gccattaaat	gtgttgacat	cgttaaggag	gcacaaagtg	1200
ctaatacctat	gggtattgta	aatgctgcta	acatacacct	gaaacatggg	gggtgggtag	1260
cagggtgcaat	caacaaggca	accaatgggt	ccatgcacaa	ggagagtgat	gattacatta	1320
agctaaatgg	ccctcttaca	gtaggagggt	cttggttctt	ttctggacat	aatcttgcta	1380
agaagtgtct	gcatgttgtt	ggacctaacc	taaatgcagg	tgaggacatc	cagcttctta	1440
aggcagcata	tgaaaatttc	aattcacagg	acatcttact	tgcaccattg	ttgtcagcag	1500
gcatatttgg	tgctaaacca	cttcagttct	tacaagtgtg	cgtgcagacg	gttcgtacac	1560
agggtttatat	tgcatgcaat	gacaaagctc	tttatgagca	ggttgtcatg	gattatcttg	1620
ataacctgaa	gcctagagtg	gaagcaccta	aacaagagga	gccacacaa	acagaagatt	1680
ccaaaactga	ggagaaatct	gtcgtacaga	agcctgtcga	tgtgaagcca	aaaattaagg	1740
cctgcattga	tgagggttacc	acaacactgg	aagaaactaa	gtttcttacc	aataagttac	1800
tcttgtttgc	tgatatcaat	ggtaagcttt	accatgatct	tcagaacatg	cttagagggtg	1860
aagatatgtc	tttccttgag	aaggatgcac	cttacatggg	agggtgatgt	atcactagtg	1920
gtgatatac	ttgtgttgta	ataccctcca	aaaaggctgg	tggcactact	gagatgctct	1980
caagagcttt	gaagaaagtg	ccagttgatg	agtatata			2018

<210> SEQ ID NO 44

<211> LENGTH: 1442

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 44

ttgatgaggt	taccacaaca	ctggaagaaa	ctaagtttct	taccaataag	ttactcttgt	60
ttgctgatat	caatggttaag	ctttaccatg	attctcagaa	catgcttaga	ggatgaagata	120
tgcttttct	tgagaaggat	gcaccttaca	tggtagggtg	tggtatcact	agtgggtgata	180
tcacttgtgt	tgtaataccc	tccaaaaagg	ctggtggcac	tactgagatg	ctctcaagag	240
ctttgaagaa	agtgccagtt	gatgagtata	taaccacgta	ccctggacaa	ggatgtgctg	300
gttatatact	tgaggaagct	aagactgtct	ttaagaaatg	caaactctga	ttttatgtac	360
taccttcaga	agcaccta	gctaagggaag	agattctagg	aactgtatcc	tggaatttga	420
gagaaatgct	tgctcatgct	gaagagacaa	gaaaattaat	gcctatatgc	atggatgtta	480
gagccataat	ggcaaccatc	caacgtaagt	ataaaggaat	taaaattcaa	gagggcatcg	540
ttgactatgg	tgctccgatc	ttcttttata	ctagtaaaga	gcctgtagct	tctattatta	600
cgaagctgaa	ctctctaaat	gagccgcttg	tcacaatgcc	aattgggttat	gtgacacatg	660
gttttaaatct	tgaagaggct	gcgcgctgta	tgcgttctct	taaagctcct	gccgtagtgt	720
cagtatcatc	accagatgct	gttactacat	ataatggata	cctcacttcg	tcacaaaga	780
catctgagga	gcacttttga	gaaacagttt	ctttggctgg	ctcttacaga	gattggctct	840
attcaggaca	gcgtacagag	ttaggtgttg	aatttcttaa	gcgtgggtgac	aaaattgtgt	900
accacactct	ggagagcccc	gtcaggttct	atcttgacgg	tgagggtctt	tcacttgaca	960
aactaaagag	tctcttatcc	ctgcgggagg	ttaagactat	aaaagtgttc	acaactgtgg	1020
acaacactaa	tctccacaca	cagcttgttg	atatgtctat	gacatagga	cagcagtttg	1080
gtccaacata	cttgatgggt	gctgatgtta	caaaaattaa	acctcatgta	aatcatgagg	1140

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gtaagacttt ctttgtacta cctagtgtatg acacactacg tagtgaagct ttcgagtact	1200
accatactct tcatgagagt tttcttggtta ggtacatgtc tgctttaaac cacacaaaga	1260
aatggaaatt tcctcaagtt ggtggtttta cttcaattaa atgggctgat aacaattggt	1320
atttgtctag tgttttatta gcacttcaac agcttgaagt caaattcaat gcaccagcac	1380
ttcaagaggc ttattataga gcccggtgctg gtgatgtctg taacttttgt gcaactcatac	1440
tc	1442

<210> SEQ ID NO 45
 <211> LENGTH: 1050
 <212> TYPE: DNA
 <213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 45

atatgtctat gacatatgga cagcagtttg gtccaacata cttggatggg gctgatgta	60
caaaaattaa acctcatgta aatcatgagg gtaagacttt ctttgtacta cctagtgtatg	120
acacactacg tagtgaagct ttcgagtact accatactct tcatgagagt tttcttggtta	180
ggtacatgtc tgctttaaac cacacaaaga aatggaaatt tcctcaagtt ggtggtttta	240
cttcaattaa atgggctgat aacaattggt atttgtctag tgttttatta gcacttcaac	300
agcttgaagt caaattcaat gcaccagcac ttcaagagc ttattataga gcccggtgctg	360
gtgatgtctg taacttttgt gcaactcatac tcgcttacag taataaaaact gttggcgagc	420
ttggtgatgt cagagaaact atgacctatc ttctacagca tgctaatttg gaatctgcaa	480
agcgagttct taatgtggtg tgtaaacatt gtggtcagaa aactactacc ttaacgggtg	540
tagaagctgt gatgtatatg ggtactctat cttatgataa tcttaagaca ggtgtttcca	600
ttccatgtgt gtgtggtcgt gatgtacac aatatctagt acaacaagag tcttcttttg	660
ttatgatgtc tgcaccacct gctgagtata aattacagca aggtacattc ttatgtgcga	720
atgagtacac tggtaactat cagtgtggtc attacactca tataactgct aaggagacct	780
tctatcgtat tgacggagct caccttaca agatgtcaga gtacaaagga ccagtgtctg	840
atgttttcta caaggaaaca tcttactata caaccatcaa gccgtgtctg tataaactcg	900
atggagttac ttacacagag attgaacaa aattggatgg gtattataaa aaggataatg	960
cttactatac agagcagcct atagacctg taccaactca accattacca atgcgaggt	1020
ttgataattt caaactcaca tgttctaaca	1050

<210> SEQ ID NO 46
 <211> LENGTH: 1995
 <212> TYPE: DNA
 <213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 46

tttgtgcact cactactcgt tacagtaata aaactgttgg cgagcttggg gatgtcagag	60
aaactatgac ccatcttcta cagcatgcta atttggaaac tgcaaagcga gttcttaatg	120
tggtgtgtaa acattgtggt cagaaaacta ctaccttaac ggggtgtaga gctgtgatgt	180
atatgggtac tctatcttat gataatctta agacaggtgt ttccattcca tgtgtgtgtg	240
gtcgtgatgc tacacaatat ctagtacaac aagagtcttc tttgttatg atgtctgcac	300
cacctgctga gtataaatta cagcaaggta cattcttatg tgcgaatgag tacactggta	360
actatcagtg tggtcattac actcatataa ctgctaagga gacctctat cgtattgacg	420
gagctcacct tacaaagatg tcagagtaca aaggaccagt gactgatgtt ttctacaagg	480

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aaacatctta cactacaacc atcaagcctg tgtcgtataa actcgatgga gttacttaca	540
cagagattga accaaaaattg gatgggtatt ataaaaagga taatgcttac tatacagagc	600
agcctataga ccttgtacca actcaaccat taccaaatgc gagttttgat aatttcaaac	660
tcacatgttc taacacaaaa ttgtctgatg atttaaatca aatgacaggc ttcacaaagc	720
cagcttcacg agagctatct gtcacattct tcccagactt gaatggcgat gtagtggcta	780
ttgactatag acactattca gcgagtttca agaaagggtgc taaattactg cataagccaa	840
ttgtttggca cattaaccag gctacaacca agacaacggt caaaccaaac acttgggtgtt	900
tacgttgtct ttggagtaca aagccagtag atacttcaaa ttcatttgaa gttctggcag	960
tagaagacac acaaggaatg gacaatcttg cttgtgaaag tcaacaaccc acctctgaag	1020
aagtagtgga aaatcctacc atacagaagg aagtcataga gtgtgacgtg aaaactaccg	1080
aagttgtagg caatgtcata cttaaaccat cagatgaagg tgttaaagta acacaagagt	1140
taggtcatga ggatcttatg gctgcttatg tggaaaacac aagcattacc attaagaaac	1200
ctaagttagct ttcactagcc ttaggtttta aaacaattgc cactcatggt attgctgcaa	1260
ttaatagtgt tccttgaggat aaaattttgg cttatgtcaa accattctta ggacaagcag	1320
caattacaac atcaaattgc gctaagagat tagcacaacg tgtgtttaac aattatatgc	1380
cttatgtgtt tacattattg ttccaattgt gtacttttac taaaagtacc aattctagaa	1440
ttagagcttc actacctaca actattgcta aaaatagtgt taagagtgtt gctaaattat	1500
gtttggatgc cggcattaat tatgtgaagt caccctaaatt ttctaaattg ttcacaatcg	1560
ctatgtggct attgttgta agtatgtgt taggttctct aatctgtgta actgctgctt	1620
ttggtgtact cttatctaatt ttgtgtgtc cttcttattg taatggcggt agagaattgt	1680
atcttaattc gtctaacgtt actactatgg atttctgtga aggttctttt ccttcagca	1740
tttgtttaag tggattagac tcccttgatt cttatccagc tctgaaacc attcaggtga	1800
cgatttcacg gtacaagcta gacttgacaa ttttaggtct ggccgctgag tgggttttgg	1860
catatatgtt gttcacaaaa ttcttttatt tattaggtct ttcagctata atgcaggtgt	1920
tctttggcta ttttgctagt catttcatca gcaattcttg gctcatgtgg tttatcatta	1980
gtattgtaca aatgg	1995

<210> SEQ ID NO 47

<211> LENGTH: 1884

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 47

aattcttggc tcatgtggtt tatcattagt attgtacaaa tggcaccggt ttctgcaatg	60
gtaggatgt acatcttctt tgcttcttct tactacatat ggaagagcta tgttcatatc	120
atggatggtt gcacctcttc gacttgcatg atgtgctata agcgcaatcg tgccacacgc	180
gttgagtgt caactattgt taatggcatg aagagatctt tctatgtcta tgcaaatgga	240
ggccgtggct tctgcaagac tcacaattgg aattgtctca attgtgacac attttgcact	300
ggtagtacat tcattagtga tgaagttgct cgtgatttgt cactccagtt taaaagacca	360
atcaacccta ctgaccagtc atcgatatatt gttgatagtg ttgctgtgaa aaatggcgcg	420
cttcacctct actttgacaa ggctgggtcaa aagacctatg agagacatcc gctctcccat	480
tttgtcaatt tagacaattt gagagctaac aacactaaag gttcactgcc tattaatgtc	540
atagtttttg atggcaagtc caaatgcgac gagtctgctt ctaagtctgc ttctgtgtac	600

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tacagtcagc	tgatgtgcc	acctattctg	ttgcttgacc	aagctcttgt	atcagacgtt	660
ggagatagta	ctgaagtttc	cgtaagatg	tttgatgctt	atgtcgacac	cttttcagca	720
acttttagtg	ttctatgga	aaaacttaag	gcacttggtg	ctacagctca	cagcgagtta	780
gcaaaggggtg	tagctttaga	tggtgtcctt	tctacattcg	tgtcagctgc	ccgacaaggt	840
gttggtgata	ccgatgttga	cacaaaggat	gttattgaat	gtctcaaaact	ttcacatcac	900
tctgacttag	aagtgcagg	tgacagttgt	aacaatttca	tgctcaccta	taataaggtt	960
gaaaacatga	cgcccagaga	tcttggcgca	tgtattgact	gtaatgcaag	gcatatcaat	1020
gcccagtag	caaaaagtca	caatgtttca	ctcatctgga	atgtaaaaga	ctacatgtct	1080
ttatctgaac	agctgcgtaa	acaaattcgt	agtgtgcc	agaagaacaa	catacctttt	1140
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aagggtggta	agattgttag	tacttgtttt	aaacttatgc	ttaaggccac	attattgtgc	1260
gttcttgctg	cattggtttg	ttatatcgtt	atgccagtac	atacattgtc	aatccatgat	1320
ggttacacaa	atgaaatcat	tggttacaaa	gccattcagg	atggtgtcac	tcgtgacatc	1380
atttctactg	atgattgttt	tgcaaaaaa	catgctggtt	ttgacgcatg	gtttagccag	1440
cggtgtggtt	catacaaaaa	tgacaaaagc	tgccctgtag	tagctgctat	cattacaaga	1500
gagattgggt	tcatagtgcc	tggttacccg	ggtagctgtc	tgagagcaat	caatgggtgac	1560
ttcttgcaat	ttctacctcg	tggttttagt	gctgttgcca	acattttgcta	cacaccttcc	1620
aaactcattg	agtatagtga	ttttgttacc	tctgcttgcg	ttcttgctgc	tgagtgtaca	1680
atttttaagg	atgctatggg	caaacctgtg	ccatattggt	atgacactaa	tttgctagag	1740
ggttctatct	cttatagtga	gcttcgtcca	gacactcgtt	atgtgcttat	ggatgggtcc	1800
atcatacagt	ttctaacac	ttacctggag	ggttctgtta	gagtagtaac	aacttttgat	1860
gctgagtact	gtagacatgg	taca				1884

<210> SEQ ID NO 48

<211> LENGTH: 2020

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 48

cactcgttat	gtgcttatgg	atggttccat	catacagttt	cctaacactt	acctggaggg	60
ttctgttaga	gtagtaacaa	cttttgatgc	tgagtactgt	agacatggta	catgcgaaag	120
gtcagaagta	ggtatttgcc	tatctaccag	tggtagatgg	gttcttaata	atgagcatta	180
cagagctcta	tcaggagttt	tctgtgggtg	tgatgcgatg	aatctcatag	ctaacatctt	240
tactcctctt	gtgcaacctg	tggtgtgctt	agatgtgtct	gcttcagtag	tggtgtgtgg	300
tattattgcc	atattgggtga	cttgtgtgc	ctactacttt	atgaaattca	gacgtgtttt	360
tggtgagtag	aacctgttg	ttgctgctaa	tgcaactttg	tttttgatgt	ctttcactat	420
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gttttctcct	attgtgcctt	tttgataac	agcaatctat	gtattctgta	tttctctgaa	600
gcactgccat	tggtttctta	acaactatct	taggaaaaga	gtcatgttta	atggagttac	660
atttagtacc	ttcgaggagg	ctgctttgtg	tacctttttg	ctcaacaagg	aatgtacct	720
aaaaattgct	agcgagacac	tggtgccact	tacacagtat	aacaggtatc	ttgctctata	780
taacaagtag	aagtatttca	gtggagcctt	agatactacc	agctatcgtg	aagcagcttg	840

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ctgccactta gcaaaggctc taaatgactt tagcaactca ggtgctgatg ttctctacca	900
accaccacag acatcaatca cttctgctgt tctgcagagt ggttttagga aaatggcatt	960
cccgtcaggc aaagtgaag ggtgcatggt acaagtaacc tgtggaacta caactcttaa	1020
tggattgtgg ttggatgaca cagtatactg tocaagacat gtcatttgca cagcagaaga	1080
catgcttaat cctaactatg aagatctgct cattcgcaaa tccaaccata gctttcttgt	1140
tcaggctggc aatgttcaac ttcgtgttat tggccattct atgcaaaatt gctgcttag	1200
gcttaaagtt gatacttcta accctaagac acccaagtat aaatttgtcc gtatccaacc	1260
tggtaaaca ttttcagttc tagcatgcta caatgggtca ccactcgttg tttatcagtg	1320
tgccatgaga cctaatacata ccattaaagg ttctttcctt aatggatcat gtggtagtgt	1380
tggttttaac attgattatg attgcgtgtc tttctgctat atgcatcata tggagcttcc	1440
aacaggagta cacgctggta ctgacttaga aggtaaattc tatggtccat ttgttgacag	1500
acaaactgca caggctgcag gtacagacac aaccataaca ttaaatgttt tggcatggct	1560
gtatgctgct gttatcaatg gtgatagggt gtttcttaac agattcacca ctactttgaa	1620
tgactttaac cttgtggcaa tgaagtacaa ctatgaacct ttgacacaag atcatgttga	1680
catattggga cctctttctg ctcaaacagg aattgccgtc ttagatatgt gtgctgcttt	1740
gaaagagctg ctgcagaatg gtatgaatgg tcgtactatc cttggtagca ctattttaga	1800
agatgagttt acaccatttg atgttgtagt acaatgctct ggtgttacct tccaaggtaa	1860
gttcaagaaa attgttaagg gcaactcatc ttggatgctt ttaactttct tgacatcact	1920
attgattcct gttcaaagta cacagtggtc actgttttct tttgtttacg agaatgcttt	1980
cttgccattt actcttggtt ttatggcaat tgctgcatgt	2020

<210> SEQ ID NO 49

<211> LENGTH: 2040

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 49

agcatttcca gcctgaagac gtactgtagc agctaaactg ccagcacca tacctctatt	60
taggttggtt aagcctttga tgaagtacaa gtatttcact ttaggccctt ttggtgtgtc	120
tgtacaaaac ctacaagggt gttccagttc tgtgtaaatt gtacctgtac catcactctt	180
agggaaatcta gcccatttga gatcttggtg gtctgatagt aatgccagca caaacctacc	240
tcccttcgaa ttgttatagt aggcaagtgc attgtcatca gtacaagctg tttgtgtggt	300
accagccgca caggacatct gtcgtagtgc tactggactc agttcattat tctgtagtgt	360
aacagctgag ttggctctta gagctgtaac aataagaggc caagccaaat ttggtgaatt	420
gtccatgtta atttcactaa gttgaacaat cttgctatcc gcatcaacaa cttgctggat	480
ttcccagagt gcagatgcat atgtaaaggt gttaccatca caagtgttct tgtaggtagc	540
ataatcaggg acaacaacca tgagtttggc tgctgtagtc aatggtaga tgttgagtgg	600
aacacaacca tcacgcgcgt tgttgataat gttgttaagt gcatcattat caagcttcct	660
aagcatagtg aagagcattg tttgcatagc actagttact tttgccctct tgcctcaga	720
tcttgctctg ttgtacattt gggctcatagc ctgatctgcc atcttttcca acttgctgtg	780
catggcagca tcacgggtcaa actcagattt agccacattc aaagatttct ttaacttttt	840
gagaacgact tcagaatcac cattagctac agcctgctca taggcctcct gggcagtggt	900
ataagcgga tatgatggta aagaactaaa ttctgaagca atagcctgaa gagtagcacg	960

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gttatcgagc atttcctcgc acaacctatt aatgtctaca gcacctgca tggatagcaa	1020
aacagacaaa agagaaacca tcttctcgaa agcttcagtt gtgtcttttg caagaagaat	1080
atcattgtgg agttgtacac atttgcccc caatttagaa gatgactcta ctctaagttg	1140
ttgaagaacc gagagcagta ccacagatgt gcactttacg tcagacattt tagactgtac	1200
agtagcaacc ttgatacatg gtttacctcc aatacccaac aacttaatgt taagcttgaa	1260
agcatcaata ctactcttag gaggcataag ccctgggag ttcataatac taaattcttg	1320
tgtagagacc aagtagtcat aaacaccaag agtaagcctg aagtaacggg tgagtaaaaa	1380
gaaaaggcca aagtagcagc agcaacaata gcctaagaaa caataaaca gcatgataga	1440
ctgtaagggt ttgccagtaa taaataacaa tgggtaatac tcaacacaca caaacactat	1500
agctctagct aaaaacatga tagtcgtaac gacaccagaa tagttagagg ttacagaaat	1560
aactaaggcc cacatggaaa tagcttgatc taaagcatta ccataagtaga ctttgtaaac	1620
aagtgtaatg acattcatca gtgtccaaac acgtctagca gcatcatcat aaacagtgcg	1680
agctgtcatg agaataagca aaactaaagc tgaagcatac ataacacaat ccttaagcct	1740
ataaccagac aagctagtgt cagccaattc aagccatgtc atgatacga tccccagct	1800
agcaggcatg tagaccatat taaagtaagc aactgttgca agagaaggta acagaaacaa	1860
gcacaagaat gcgtgcttat gcttaacaag cagcatagca catgcagcaa ttgccataat	1920
accaagagta aatggcaaga aagcattctc gtaaacaaag aaaaacagtg accactgtgt	1980
actttgaaca agaatacaata gtgatgtcaa gaaagttaaa agcatccaat gatgagtga	2040

<210> SEQ ID NO 50

<211> LENGTH: 2012

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 50

cttgtagggt tggtacagac acacaaaaag ggctaaagt gaaatacttg tacttcatca	60
aaggcttaaa caacctaat agaggatgg tgctgggagc tttagctgct acagtacgtc	120
ttcaggctgg aaatgtaca gaagtacctg ccaattcaac tgtgctttcc ttctgtgctt	180
ttgcagtaga ccctgctaaa gcatataagg attacctagc aagtggagga caaccaatca	240
ccaactgtgt gaagatgttg tgtacacaca ctggtacagg acaggcaatt actgtaacac	300
cagaagctaa catggaccaa gagtcctttg gtggtgcttc atgttgtctg tattgtagat	360
gccacattga ccatccaaat cctaaaggat tctgtgactt gaaaggtaag tacgtccaaa	420
tacctaccac ttgtgctaata gacctagtg gttttactac tagaaacaca gtctgtaccg	480
tctgcggaat gtggaagggt tatggctgta gttgtgacca actccgagaa cccttgatgc	540
agtctgcgga tgcatacaag tttttaaacg ggtttgcggg gtaagtgcag ccgctcttac	600
accgtgcggc acaggcacta gtactgatgt cgtctacagg gcttttgata tttacaacga	660
aaaagtgtgt ggttttgcaa agttcctaaa aactaattgc tgtcgcttcc aggagaagga	720
tgaggaaggc aatttattag actcttactt tgtagttaag aggcatacta tgtctaacta	780
ccaacatgaa gagactattt ataacttggt taaagattgt ccagcggttg ctgtccatga	840
ctttttcaag tttagagtag atgggtgacat ggtaccacat atatcacgtc agcgtctaac	900
taaatacaca atggctgatt tagtctatgc tctacgtcat tttgatgagg gtaattgtga	960
tacattaaaa gaaatactcg tcacatacaa ttgctgtgat gatgattatt tcaataagaa	1020
ggattgggtat gacttcgtag agaatacctga catcttacgc gtatatgcta acttaggtga	1080

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gcgtgtacgc caatcattat taaagactgt acaattctgc gatgctatgc gtgatgcagg	1140
cattgttaggc gtactgacat tagataatca ggatcttaat gggaactggg acgatttcgg	1200
tgatttcgta caagtagcac caggctgcgg agttcctatt gtggattcat attactcatt	1260
gctgatgccc atcctcactt tgactagggc attggctgct gagtcccata tggatgctga	1320
tctcgcaaaa ccacttatta agtgggattt gctgaaatat gattttacgg aagagagact	1380
ttgtctcttc gaccgttatt ttaaatattg ggaccagaca taccatccca attgtattaa	1440
ctgtttggat gatagggtga tccttcattg tgcaaaacttt aatgtgttat tttctactgt	1500
gtttccacct acaagttttg gaccactagt aagaaaaata tttgtagatg gtgttccttt	1560
tggtgtttca actggatacc attttcgtga gttaggagtc gtacataatc aggatgtaaa	1620
cttcatagc tcgcgtctca gtttcaagga acttttagtg tatgctgctg atccagctat	1680
gcatgcagct tctggcaatt tattgctaga taaacgcact acatgctttt cagtagctgc	1740
actaacaac aatgttgctt ttcaaaactgt caaacccggg aattttaata aagactttta	1800
tgactttgct gtgtctaaag gtttctttaa ggaagggaagt tctgttgaac taaaacactt	1860
cttctttgct caggatggca acgctgctat cagtgattat gactattatc gttataatct	1920
gccacaatg tgtgatatca gacaactcct attcgtagtt gaagttgttg ataaatactt	1980
tgattgttac gatggtggct gtattaatgc ca	2012

<210> SEQ ID NO 51

<211> LENGTH: 1877

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 51

gtacttcgcg tacagtggca ataccatag acagcttaaa tgtttcctca gtggctttga	60
gcgtttctgc tgcgaaaagc ttgagtctct cagtacaagt gttggcaagt atgtaatcgc	120
cagcattagt ccaatcacat gttgctatcg cattgaagtc agtgacattg tcaactgccta	180
cacatgtggt tttgtataaa ccaaaaacct gaccattagc acataatgga aaactaatgg	240
gaggcttatg tgacttgcaa taatagctca tacctcctag atacagttgt gtcacatcag	300
tgacatcaca acctggggca ttgcaaacat agggattaac agacaacact aatttgtgtg	360
atgttgaaat gacatggtca tagcagcact tgcaacatag gaatggtctc ctaatacagg	420
caccgcaacg aagtgaagtc tgtgaattgc acaatacaca agcacctaca gctgcaaga	480
ctgtatgtgg tgtgtacata gcctcataaa actcaggttc ccagtaccgt gaggtgttat	540
cattagttag cattacggaa tacatgtcca acatgtggcc agtaagctca tcatgtaact	600
ttctaataa ttgtaaatc aagtgaaga catcagcata ctctgatta ggatgttttg	660
taagtgggta agcatcaata gccagtgaca cgaacctttc aatcataagt gtaccatctg	720
ttttgacaat atcatcgaca aaacagcctg cgcctaatat tcttgatgga tctgggtaag	780
gcaggtaac gtaatcatct ccttgtttaa ctagcattgt atgctgtgag caaaattcgt	840
gaggtccttt agtaagggtc gtctcagtc aacattttgc ctcagacatg aacacattat	900
tttgataata aagaactgcc ttaaagtctt taatgctagc tactaaacct tgagccgcat	960
agttactggt atagcacaca acggcatcat cagaaagaat catcatggag aaatgtttac	1020
gcaggtaagc gtaaaactca tccacgaatt catgatcaac atccctatct ctatagagac	1080
actcatagag cctgtgttgt agattgcgga catacttgtc agctatctta ttaccatcag	1140
ttgaaagaag tgcatttaca ttggctgtaa cagcttgaca aatgttaaag acactattag	1200

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cataagcagt tgtagcatca ccg gatgatg ttcacacctgg tttaacatat agtgagccgc	1260
cacacatgac catctcactt aatacttgcg cacactcggt agctaacctg tagaaacgggt	1320
gtgataagtt acagcaagtg ttatgtttgc gagcaagaac aagagaggcc attatcctaa	1380
gcatgttagg catggctctg tcacattttg gataatccca acccataaagg tgtggagttt	1440
ctacatcact gtaaacagtt tttaacatat tatgccagcc accgtaaaac ttgcttggtc	1500
caattaccac agtagctcct ctagtggcgg ctattgactt caataatttc tgatgaaact	1560
gtctatttgt catagtacta cagatagaga caccagctac ggtgcgagct ctattctttg	1620
cactaatggc atacttaaga ttcatttgag ttatagtagg gatgacatta cgcttagtat	1680
acgcgaaaag tgcattctga tcctcataac tcattgagtc ataataaagt ctagccttac	1740
cccatattatt aaatgggaaa ccagctgatt tatccagatt gttaacgatt acttggttgg	1800
cattaataca gccaccatcg taacaatcaa agtatatttc aacaacttca actacgaata	1860
ggagttgtct gatatca	1877

<210> SEQ ID NO 52

<211> LENGTH: 2051

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 52

tcaggccaa tcttgacaaa gtacttcatt gatgtaagct caaagccatg cgcacaaagg	60
acgaacacga ctctgtctga caatccttc agtgtatcac tgagcatttg tactatctta	120
atacgacta cattccaggg caagccttta tacatgagtg gtataagatg tttaaactgg	180
tcacctgggtg gaggttttgc attaaactctg gtgaattctg tgttattttc agtgtcaaca	240
taaccagtcg gtacagctac taagttaaca cctgtagaaa atcctagctg gagaggtagg	300
ttagtaccga cagcatctct agttgcatga cagccctcta catcaaagcc aatccacgca	360
cgaacgtgac gaatagcttc ttcgcggtg ataaacatat tagggtaacc attgacttgg	420
taattcattt tgaacccat catagagatg agtctacggt aggtcatgtc ctttggtatg	480
cctggtagt caacacataa tccttcagtc ttgaacttta tatcaacgct gaggtgtgta	540
ggtgcctgtg taggatgaag accagtaatg atcttactac agtccttaaa aagtcagtt	600
acattttctg ctgtaatgt agccacattg cgacgtggta tttctagact tgtaaattgc	660
agtttgtcat aaagatctct atcagacatt atgcacaaaa tgccaatttt tgccttctg	720
atagccacat tgaagcggt gacattacaa gagtgtgtctg tttcagtagt ttgtgtgaat	780
atgacatagt catattcaga accctgtgat gaatcaacag tctgcgtagg caatcctaag	840
atttttgaag ctacacggt ctgtgaatta taaggtaga taaaaacagc ttttctccaa	900
gcaggattgc gtgtaagaaa ttctcttaca acgcctatct gaggtctgtt gattgcagat	960
gaaacatcat gtgtaataac acctttgtag aacattttga agcattgagc tgacttatcc	1020
ttgtgtgctt ttagcttatt gtcataaact aaagcactca cagtgtcaac aatttcagca	1080
ggacaacggc gacaagttcc aaggaacatg tctggacctt ttgttttcat aagtctgcac	1140
actgaattaa aatattctgg ttctagtgtg ccttagtca gcaatgtgctg gggggtggt	1200
aattgagcag gatcgccaat atagacgtag tgttttgac gaagtctagc attgacaaca	1260
ctcaagtcat aattagtagc catagagatt tcatcaaaga ctacaatgtc agcagttggt	1320
tctggcaatg catttacagt gcagaaaaca tactgttcta gtgttgatt cactttgaat	1380
ttatcaaaac actctacgcg cgcacgcgca ggtatgatcc tactacattt atctatgggc	1440

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aaatatttta atgccttttc acatagggca tcaacagctg catgagagca tgcggtatac	1500
actatgcgag cagatgggta atagagagca agtccgatgg caaaatgact cttaccagta	1560
ccagggtggc cttggagtgt agagtacttt tgcagccga ccttttgata atttgcaaca	1620
ttgctagaaa actcatctga gatgttgagt gttgggtaca agccagtaat tctcacatag	1680
tgctcttggt gactagagt aggtgcacta agtggcatta cagtgtgaga tgtcaacaca	1740
aagtaatcac caacattcaa cttgtatgtc gtagtacctc tgtacacaac agcatcacca	1800
tagtcacctt tttcaaaggt gtactctcca atctgtactt tactattttt agttacacgg	1860
taaccagtaa agacatagtt tctgttcaat ggtggtctag gttttccaac ctcccatgaa	1920
agatgcaatt ctctgtcaga gagtacttcg cgtacagtgg caataccata tgacagctta	1980
aatgtttcct cagtggcttt gagcgtttct gctgcgaaaa gcttgagtct ctacagtaca	2040
gtgttggtgaa g	2051

<210> SEQ ID NO 53

<211> LENGTH: 2075

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 53

tgcttgtagt tttgggtaga aggtttcaac atgtccatcc ttacacccaa gcatgaatga	60
aatttcagca tagtcaattg taaccttgac cacttttgaa atcactgaca aatcttggtga	120
ctttattatc tcgacaaagt catcaagtaa aagatcaatc acagaacaca cacattttga	180
tgaacctgtt tgcgcatctg ttatgaagta atttttact gtgctgtcca tagggataaa	240
atcctctaata ttaagtgggt aatcttggtg gcgcttggtt aagcctatca ttaaatgaag	300
accgccaagt tgtccatgac tgaaatctcc ataaacgatg tgttcgaagg catagccctc	360
gagccttatat cgctgtatga attcatccat agcgagctcg agaaagtcag tttccatttg	420
tgatctgggc ttaaaatcct ctaagtctct gctctgagta aagtaggttt caggcaactg	480
ttgaataatg cgtctactt tcttaaagta gttaaactgt gtttttactg attctccaat	540
taatgtgact ccattgacgc tagcttggtc tgggtccctt gaagggtgta gacctttgac	600
tgaaccttct gttattaaaa caccattacg ggcgtttcta aaaagggtcta cctgtccttc	660
cactctacca tcaaacaga cagtaagtga agaacaagca ctctcagtag gtttcttggc	720
aatgtcagtc attgtgcaga cacctattgt agatacatgt gctggggctt ctcttttgta	780
gtcccagatt acagtattag cagcgatata aacacccaaa ttattgagta tcttaatctc	840
tggcactggt ttaatgttac gcttagccca aagctcaaat gcaacattaa caggaagtgt	900
tgtcttattt tcaaagatct ccacatcaat accatctacc tttgtgtaaa cagcattatt	960
aatgatggaa acaggtgctt cgccggcggtg tccatcaaag tgccttttat taacaacatt	1020
ataagccaca ttttctaacc tctgtaacct ggtaaatgta ttccacaggt tataagtatc	1080
aaattgtttg taaatccata ggctaaatcc agcagaaatc atcatattat atgcatccaa	1140
gtactgtcgg tactcatttg catggtgtct gcaaacagca ccacctaaat tgcacgtgtg	1200
aatacacgta gcagatttga gtggaacata atcaatatcc gacactactt gtttgccatg	1260
agactcacia ggactatcag aatagtaaaa gaaaggcaat tgctttaaata tagtaaatgc	1320
acttttatcg aaagctggag tgtggaatgc atgcttatcc acatacaaac taccaccatc	1380
acagcctggt aagttcaagt ttgacaagac tcttgtgtca aacctacaca caattgcatt	1440
ggctgggtaa cgatcaacgt tacaattcca aaacaaacaa acaccatcag tgaatttatc	1500

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gtgatgtgta	gcataagaat	agaagagttc	ctctattttg	taagctttgt	cactacatgg	1560
ctgagcatcg	tagaacttcc	attctacttc	agcctgaggc	acacacttga	tagccttttg	1620
atttccaatg	tcataagaa	ctggaaactt	atcagcaagc	aatgcagact	tcacaaccat	1680
gtgtgtgtact	tttctgcaag	cagaattaac	cctcagttca	tctcctataa	taggggtattc	1740
aacagaccaa	tcaacgcgct	taacaaagca	ctcatggact	gctaaacatc	tagtcatgat	1800
agcatcacia	ctagccacat	gtgcatttcc	atgtacctgg	caatgttggt	catgggttact	1860
ctgaagggtta	cccgtaaagc	cccactgctg	aacatcaatc	ataaatgggt	tatagacata	1920
gtcaaaaccc	acagaatgat	tccagcaggc	ataagtatct	gatgaagtag	aaaagcaagt	1980
tgcacgtttg	tcacacagac	aacacgttct	ttcaggtcca	atcttgacaa	agtacttcat	2040
tgatgtaagc	tcaaagccat	gcgcccacaa	gacga			2075

<210> SEQ ID NO 54

<211> LENGTH: 1891

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 54

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tcataacaga	tgcgcaaa	ggttcatcaa	aatgtgtgtg	ttctgtgatt	gatcttttac	120
ttgatgactt	tgctgagata	ataaagtcac	aagattttgc	agtgatttca	aaagtgggtca	180
aggttacaat	tgactatgct	gaaatttcat	tcctgctttg	gtgtaaggat	ggacatgttg	240
aaaccttcta	cccaaaacta	caagcaagtc	aagcgtggca	accaggtggt	gcgatgccta	300
acttgtacaa	gatgcaaaga	atgcttcttg	aaaagtgtga	ccttcagaat	tatgggtgaaa	360
atgctgttat	acaaaagga	ataatgatga	atgtcgcaaa	gtatactcaa	ctgtgtcaat	420
acttaaatca	acttacttta	gctgtacct	acaacatgag	agttattcac	tttgggtctg	480
gctctgataa	aggagtgtga	ccaggtacag	ctgtgctcag	acaatgggtg	ccaactggca	540
cactacttgt	cgattcagat	cttaatgact	tcgtctccga	cgcagattct	actttaattg	600
gagactgtgc	aacagtacat	acggctaata	aatgggacct	tattattagc	gatatgtatg	660
accctaggac	caaacatgtg	acaaaagaga	atgactctaa	agaaggggtt	ttcacttacc	720
tgtgtggatt	tataaagcaa	aaactagccc	tgggtgggtc	tatagctgta	aagataacag	780
agcattcttg	gaatgctgac	ctttacaagc	ttatgggcca	tttctcatgg	tggacagctt	840
ttgttacaaa	tgtaaatgca	tcctcatcgg	aagcattttt	aattggggct	aactatcttg	900
gcaagccgaa	ggaacaaatt	gatggctata	ccatgcatgc	taactacatt	ttctggagga	960
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aattaagagg	aactgctgta	atgtctctta	aggagaatca	aatcaatgat	atgatttatt	1080
ctcttctgga	aaaaggtagg	cttatcatta	gagaaaacaa	cagagtgttg	gtttcaagtg	1140
atattcttgt	taacaactaa	acgaacatgt	ttattttctt	attatttctt	actctcacta	1200
gtggtagtga	ccttgaccgg	tgcaccactt	ttgatgatgt	tcaagctcct	aattacactc	1260
aacatacttc	atctatgagg	ggggtttact	atcctgatga	aatttttaga	tcagacactc	1320
ttttattaac	tcaggattta	tttcttccat	tttattctaa	tggtacaggg	tttcatacta	1380
ttaatcatac	gtttggcaac	cctgtcatac	cttttaagga	tggatattat	tttctgcca	1440
cagagaaatc	aaatgttgtc	cgtgggtggg	tttttggttc	taccatgaac	aacaagtcac	1500
agtcggtgat	tattattaac	aattctacta	atgttggtat	acgagcatgt	aactttgaat	1560

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tgtgtgacaa cccctttcttt gctgtttcta aaccatggg tacacagaca catactatga	1620
tattcgataa tgcatttaat tgcactttcg agtacatata tgatgccttt tcgcttgatg	1680
tttcagaaaa gtcaggtaat tttaaacact tacgagagtt tgtgtttaaa aataaagatg	1740
ggttttctcta tgtttataag ggctatcaac ctatagatgt agttcgtgat ctaccttctg	1800
gttttaacac tttgaaacct atttttaagt tgcctcttgg tattaacatt acaaatttta	1860
gagccattct tacagccttt tcacctgctc a	1891

<210> SEQ ID NO 55
 <211> LENGTH: 32
 <212> TYPE: DNA
 <213> ORGANISM: artificial sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: N sens primer

<400> SEQUENCE: 55

cccatatgtc tgataatgga ccccaatcaa ac	32
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<210> SEQ ID NO 56
 <211> LENGTH: 32
 <212> TYPE: DNA
 <213> ORGANISM: artificial sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: N antisens primer

<400> SEQUENCE: 56

cccccggtg cctgagttga atcagcagaa gc	32
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<210> SEQ ID NO 57
 <211> LENGTH: 31
 <212> TYPE: DNA
 <213> ORGANISM: artificial sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Sc sens primer

<400> SEQUENCE: 57

cccatatgag tgaccttgac cgggtcacca c	31
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<210> SEQ ID NO 58
 <211> LENGTH: 30
 <212> TYPE: DNA
 <213> ORGANISM: artificial sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: SL sens primer

<400> SEQUENCE: 58

cccatatgaa accttgacac ccacctgctc	30
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<210> SEQ ID NO 59
 <211> LENGTH: 33
 <212> TYPE: DNA
 <213> ORGANISM: Sc and SL antisens primer

<400> SEQUENCE: 59

cccccggtt taatatattg ctcattttt ccc	33
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<210> SEQ ID NO 60
 <211> LENGTH: 16
 <212> TYPE: DNA
 <213> ORGANISM: Sens set 1 primer

<400> SEQUENCE: 60

ggcatcgat ggggtg	16
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<210> SEQ ID NO 61
<211> LENGTH: 16
<212> TYPE: DNA
<213> ORGANISM: Antisens set 2 (28774-28759) primer

<400> SEQUENCE: 61

cagtttcacc acctcc 16

<210> SEQ ID NO 62
<211> LENGTH: 16
<212> TYPE: DNA
<213> ORGANISM: Sens set 2 (28375-28390) primer

<400> SEQUENCE: 62

ggctactacc gaagag 16

<210> SEQ ID NO 63
<211> LENGTH: 16
<212> TYPE: DNA
<213> ORGANISM: Antisens set 2 (28702-28687)primer

<400> SEQUENCE: 63

aattaccgcg actacg 16

<210> SEQ ID NO 64
<211> LENGTH: 26
<212> TYPE: DNA
<213> ORGANISM: Probe 1/set 1 (28561-28586)

<400> SEQUENCE: 64

ggcaccgca atcctaataa caatgc 26

<210> SEQ ID NO 65
<211> LENGTH: 21
<212> TYPE: DNA
<213> ORGANISM: Probe 2/set 1 (28588-28608)

<400> SEQUENCE: 65

gccaccgtgc tacaacttcc t 21

<210> SEQ ID NO 66
<211> LENGTH: 23
<212> TYPE: DNA
<213> ORGANISM: Probe 1/set 2 /probe N/FL (28541-28563)

<400> SEQUENCE: 66

atacacccaa agaccacatt ggc 23

<210> SEQ ID NO 67
<211> LENGTH: 25
<212> TYPE: DNA
<213> ORGANISM: Probe 2/set 2/probe SARS/N/LC705 (28565-28589)

<400> SEQUENCE: 67

cccgcaatcc taataacaat gctgc 25

<210> SEQ ID NO 68
<211> LENGTH: 30
<212> TYPE: DNA
<213> ORGANISM: artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: Anchor primer 14T

<400> SEQUENCE: 68

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agatgaattc ggtacctttt tttttttttt

30

<210> SEQ ID NO 69
 <211> LENGTH: 13
 <212> TYPE: PRT
 <213> ORGANISM: artificial sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: M2-14 peptide

<400> SEQUENCE: 69

Ala Asp Asn Gly Thr Ile Thr Val Glu Glu Leu Lys Gln
 1 5 10

<210> SEQ ID NO 70
 <211> LENGTH: 12
 <212> TYPE: PRT
 <213> ORGANISM: artificial sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: E1-12 peptide

<400> SEQUENCE: 70

Met Tyr Ser Phe Val Ser Glu Glu Thr Gly Thr Leu
 1 5 10

<210> SEQ ID NO 71
 <211> LENGTH: 24
 <212> TYPE: PRT
 <213> ORGANISM: artificial sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: E53-72 peptide

<400> SEQUENCE: 71

Lys Pro Thr Val Tyr Val Tyr Ser Arg Val Lys Asn Leu Asn Ser Ser
 1 5 10 15

Glu Gly Val Pro Asp Leu Leu Val
 20

<210> SEQ ID NO 72
 <211> LENGTH: 153
 <212> TYPE: DNA
 <213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 72

gatattaggt ttttacctac ccaggaaaag ccaaccaacc tcgatctctt gtagatctgt 60

tctctaaacg aactttaaaa tctgtgtagc tgctgctcgg ctgcatgcct agtgcaccta 120

cgcagtataa acaataataa attttactgt cgt 153

<210> SEQ ID NO 73
 <211> LENGTH: 410
 <212> TYPE: DNA
 <213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 73

ttctccagac aactttcaaaa ttccatgagt ggagcttctg ctgattcaac tcaggcataa 60

acactcatga tgaccacaca aggcagatgg gctatgtaaa cgttttcgca attccgttta 120

cgatacatag tctactcttg tgcagaatga attctcgtaa ctaaacagca caagtagggt 180

tagttaactt taatctcaca tagcaatctt taatcaatgt gtaacattag ggaggacttg 240

aaagagccac cacattttca tcgaggccac gccgagtagc atcgagggta cagtgaataa 300

tgctaggggag agctgcctat atggaagagc cctaattgtgt aaaattaatt ttagtagtgc 360

tatcccatg tgattttaat agcttcttag gagaatgaca aaaaaaaaaa 410

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<210> SEQ ID NO 74
 <211> LENGTH: 4382
 <212> TYPE: PRT
 <213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 74

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Met Glu Ser Leu Val Leu Gly Val Asn Glu Lys Thr His Val Gln Leu
1          5          10          15

Ser Leu Pro Val Leu Gln Val Arg Asp Val Leu Val Arg Gly Phe Gly
          20          25          30

Asp Ser Val Glu Glu Ala Leu Ser Glu Ala Arg Glu His Leu Lys Asn
          35          40          45

Gly Thr Cys Gly Leu Val Glu Leu Lys Gly Val Leu Pro Gln Leu
          50          55          60

Glu Gln Pro Tyr Val Phe Ile Lys Arg Ser Asp Ala Leu Ser Thr Asn
          65          70          75          80

His Gly His Lys Val Val Glu Leu Val Ala Glu Met Asp Gly Ile Gln
          85          90          95

Tyr Gly Arg Ser Gly Ile Thr Leu Gly Val Leu Val Pro His Val Gly
          100          105          110

Glu Thr Pro Ile Ala Tyr Arg Asn Val Leu Leu Arg Lys Asn Gly Asn
          115          120          125

Lys Gly Ala Gly Gly His Ser Tyr Gly Ile Asp Leu Lys Ser Tyr Asp
          130          135          140

Leu Gly Asp Glu Leu Gly Thr Asp Pro Ile Glu Asp Tyr Glu Gln Asn
          145          150          155          160

Trp Asn Thr Lys His Gly Ser Gly Ala Leu Arg Glu Leu Thr Arg Glu
          165          170          175

Leu Asn Gly Gly Ala Val Thr Arg Tyr Val Asp Asn Asn Phe Cys Gly
          180          185          190

Pro Asp Gly Tyr Pro Leu Asp Cys Ile Lys Asp Phe Leu Ala Arg Ala
          195          200          205

Gly Lys Ser Met Cys Thr Leu Ser Glu Gln Leu Asp Tyr Ile Glu Ser
          210          215          220

Lys Arg Gly Val Tyr Cys Cys Arg Asp His Glu His Glu Ile Ala Trp
          225          230          235          240

Phe Thr Glu Arg Ser Asp Lys Ser Tyr Glu His Gln Thr Pro Phe Glu
          245          250          255

Ile Lys Ser Ala Lys Lys Phe Asp Thr Phe Lys Gly Glu Cys Pro Lys
          260          265          270

Phe Val Phe Pro Leu Asn Ser Lys Val Lys Val Ile Gln Pro Arg Val
          275          280          285

Glu Lys Lys Lys Thr Glu Gly Phe Met Gly Arg Ile Arg Ser Val Tyr
          290          295          300

Pro Val Ala Ser Pro Gln Glu Cys Asn Asn Met His Leu Ser Thr Leu
          305          310          315          320

Met Lys Cys Asn His Cys Asp Glu Val Ser Trp Gln Thr Cys Asp Phe
          325          330          335

Leu Lys Ala Thr Cys Glu His Cys Gly Thr Glu Asn Leu Val Ile Glu
          340          345          350

Gly Pro Thr Thr Cys Gly Tyr Leu Pro Thr Asn Ala Val Val Lys Met
          355          360          365

Pro Cys Pro Ala Cys Gln Asp Pro Glu Ile Gly Pro Glu His Ser Val
          370          375          380

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Ala	Asp	Tyr	His	Asn	His	Ser	Asn	Ile	Glu	Thr	Arg	Leu	Arg	Lys	Gly	385	390	395	400
Gly	Arg	Thr	Arg	Cys	Phe	Gly	Gly	Cys	Val	Phe	Ala	Tyr	Val	Gly	Cys	405	410	415	
Tyr	Asn	Lys	Arg	Ala	Tyr	Trp	Val	Pro	Arg	Ala	Ser	Ala	Asp	Ile	Gly	420	425	430	
Ser	Gly	His	Thr	Gly	Ile	Thr	Gly	Asp	Asn	Val	Glu	Thr	Leu	Asn	Glu	435	440	445	
Asp	Leu	Leu	Glu	Ile	Leu	Ser	Arg	Glu	Arg	Val	Asn	Ile	Asn	Ile	Val	450	455	460	
Gly	Asp	Phe	His	Leu	Asn	Glu	Glu	Val	Ala	Ile	Ile	Leu	Ala	Ser	Phe	465	470	475	480
Ser	Ala	Ser	Thr	Ser	Ala	Phe	Ile	Asp	Thr	Ile	Lys	Ser	Leu	Asp	Tyr	485	490	495	
Lys	Ser	Phe	Lys	Thr	Ile	Val	Glu	Ser	Cys	Gly	Asn	Tyr	Lys	Val	Thr	500	505	510	
Lys	Gly	Lys	Pro	Val	Lys	Gly	Ala	Trp	Asn	Ile	Gly	Gln	Gln	Arg	Ser	515	520	525	
Val	Leu	Thr	Pro	Leu	Cys	Gly	Phe	Pro	Ser	Gln	Ala	Ala	Gly	Val	Ile	530	535	540	
Arg	Ser	Ile	Phe	Ala	Arg	Thr	Leu	Asp	Ala	Ala	Asn	His	Ser	Ile	Pro	545	550	555	560
Asp	Leu	Gln	Arg	Ala	Ala	Val	Thr	Ile	Leu	Asp	Gly	Ile	Ser	Glu	Gln	565	570	575	
Ser	Leu	Arg	Leu	Val	Asp	Ala	Met	Val	Tyr	Thr	Ser	Asp	Leu	Leu	Thr	580	585	590	
Asn	Ser	Val	Ile	Ile	Met	Ala	Tyr	Val	Thr	Gly	Gly	Leu	Val	Gln	Gln	595	600	605	
Thr	Ser	Gln	Trp	Leu	Ser	Asn	Leu	Leu	Gly	Thr	Thr	Val	Glu	Lys	Leu	610	615	620	
Arg	Pro	Ile	Phe	Glu	Trp	Ile	Glu	Ala	Lys	Leu	Ser	Ala	Gly	Val	Glu	625	630	635	640
Phe	Leu	Lys	Asp	Ala	Trp	Glu	Ile	Leu	Lys	Phe	Leu	Ile	Thr	Gly	Val	645	650	655	
Phe	Asp	Ile	Val	Lys	Gly	Gln	Ile	Gln	Val	Ala	Ser	Asp	Asn	Ile	Lys	660	665	670	
Asp	Cys	Val	Lys	Cys	Phe	Ile	Asp	Val	Val	Asn	Lys	Ala	Leu	Glu	Met	675	680	685	
Cys	Ile	Asp	Gln	Val	Thr	Ile	Ala	Gly	Ala	Lys	Leu	Arg	Ser	Leu	Asn	690	695	700	
Leu	Gly	Glu	Val	Phe	Ile	Ala	Gln	Ser	Lys	Gly	Leu	Tyr	Arg	Gln	Cys	705	710	715	720
Ile	Arg	Gly	Lys	Glu	Gln	Leu	Gln	Leu	Leu	Met	Pro	Leu	Lys	Ala	Pro	725	730	735	
Lys	Glu	Val	Thr	Phe	Leu	Glu	Gly	Asp	Ser	His	Asp	Thr	Val	Leu	Thr	740	745	750	
Ser	Glu	Glu	Val	Val	Leu	Lys	Asn	Gly	Glu	Leu	Glu	Ala	Leu	Glu	Thr	755	760	765	
Pro	Val	Asp	Ser	Phe	Thr	Asn	Gly	Ala	Ile	Val	Gly	Thr	Pro	Val	Cys	770	775	780	
Val	Asn	Gly	Leu	Met	Leu	Leu	Glu	Ile	Lys	Asp	Lys	Glu	Gln	Tyr	Cys	785	790	795	800
Ala	Leu	Ser	Pro	Gly	Leu	Leu	Ala	Thr	Asn	Asn	Val	Phe	Arg	Leu	Lys	805	810	815	

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Gly Gly Ala Pro Ile Lys Gly Val Thr Phe Gly Glu Asp Thr Val Trp
 820 825 830

Glu Val Gln Gly Tyr Lys Asn Val Arg Ile Thr Phe Glu Leu Asp Glu
 835 840 845

Arg Val Asp Lys Val Leu Asn Glu Lys Cys Ser Val Tyr Thr Val Glu
 850 855 860

Ser Gly Thr Glu Val Thr Glu Phe Ala Cys Val Val Ala Glu Ala Val
 865 870 875 880

Val Lys Thr Leu Gln Pro Val Ser Asp Leu Leu Thr Asn Met Gly Ile
 885 890 895

Asp Leu Asp Glu Trp Ser Val Ala Thr Phe Tyr Leu Phe Asp Asp Ala
 900 905 910

Gly Glu Glu Asn Phe Ser Ser Arg Met Tyr Cys Ser Phe Tyr Pro Pro
 915 920 925

Asp Glu Glu Glu Glu Asp Asp Ala Glu Cys Glu Glu Glu Glu Ile Asp
 930 935 940

Glu Thr Cys Glu His Glu Tyr Gly Thr Glu Asp Asp Tyr Gln Gly Leu
 945 950 955 960

Pro Leu Glu Phe Gly Ala Ser Ala Glu Thr Val Arg Val Glu Glu Glu
 965 970 975

Glu Glu Glu Asp Trp Leu Asp Asp Thr Thr Glu Gln Ser Glu Ile Glu
 980 985 990

Pro Glu Pro Glu Pro Thr Pro Glu Glu Pro Val Asn Gln Phe Thr Gly
 995 1000 1005

Tyr Leu Lys Leu Thr Asp Asn Val Ala Ile Lys Cys Val Asp Ile
 1010 1015 1020

Val Lys Glu Ala Gln Ser Ala Asn Pro Met Val Ile Val Asn Ala
 1025 1030 1035

Ala Asn Ile His Leu Lys His Gly Gly Gly Val Ala Gly Ala Leu
 1040 1045 1050

Asn Lys Ala Thr Asn Gly Ala Met Gln Lys Glu Ser Asp Asp Tyr
 1055 1060 1065

Ile Lys Leu Asn Gly Pro Leu Thr Val Gly Gly Ser Cys Leu Leu
 1070 1075 1080

Ser Gly His Asn Leu Ala Lys Lys Cys Leu His Val Val Gly Pro
 1085 1090 1095

Asn Leu Asn Ala Gly Glu Asp Ile Gln Leu Leu Lys Ala Ala Tyr
 1100 1105 1110

Glu Asn Phe Asn Ser Gln Asp Ile Leu Leu Ala Pro Leu Leu Ser
 1115 1120 1125

Ala Gly Ile Phe Gly Ala Lys Pro Leu Gln Ser Leu Gln Val Cys
 1130 1135 1140

Val Gln Thr Val Arg Thr Gln Val Tyr Ile Ala Val Asn Asp Lys
 1145 1150 1155

Ala Leu Tyr Glu Gln Val Val Met Asp Tyr Leu Asp Asn Leu Lys
 1160 1165 1170

Pro Arg Val Glu Ala Pro Lys Gln Glu Glu Pro Pro Asn Thr Glu
 1175 1180 1185

Asp Ser Lys Thr Glu Glu Lys Ser Val Val Gln Lys Pro Val Asp
 1190 1195 1200

Val Lys Pro Lys Ile Lys Ala Cys Ile Asp Glu Val Thr Thr Thr
 1205 1210 1215

Leu Glu Glu Thr Lys Phe Leu Thr Asn Lys Leu Leu Leu Phe Ala

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1220	1225	1230
Asp Ile Asn Gly Lys Leu Tyr	His Asp Ser Gln Asn Met Leu Arg	
1235	1240	1245
Gly Glu Asp Met Ser Phe Leu	Glu Lys Asp Ala Pro Tyr Met Val	
1250	1255	1260
Gly Asp Val Ile Thr Ser Gly	Asp Ile Thr Cys Val Val Ile Pro	
1265	1270	1275
Ser Lys Lys Ala Gly Gly Thr	Thr Glu Met Leu Ser Arg Ala Leu	
1280	1285	1290
Lys Lys Val Pro Val Asp Glu	Tyr Ile Thr Thr Tyr Pro Gly Gln	
1295	1300	1305
Gly Cys Ala Gly Tyr Thr Leu	Glu Glu Ala Lys Thr Ala Leu Lys	
1310	1315	1320
Lys Cys Lys Ser Ala Phe Tyr	Val Leu Pro Ser Glu Ala Pro Asn	
1325	1330	1335
Ala Lys Glu Glu Ile Leu Gly	Thr Val Ser Trp Asn Leu Arg Glu	
1340	1345	1350
Met Leu Ala His Ala Glu Glu	Thr Arg Lys Leu Met Pro Ile Cys	
1355	1360	1365
Met Asp Val Arg Ala Ile Met	Ala Thr Ile Gln Arg Lys Tyr Lys	
1370	1375	1380
Gly Ile Lys Ile Gln Glu Gly	Ile Val Asp Tyr Gly Val Arg Phe	
1385	1390	1395
Phe Phe Tyr Thr Ser Lys Glu	Pro Val Ala Ser Ile Ile Thr Lys	
1400	1405	1410
Leu Asn Ser Leu Asn Glu Pro	Leu Val Thr Met Pro Ile Gly Tyr	
1415	1420	1425
Val Thr His Gly Phe Asn Leu	Glu Glu Ala Ala Arg Cys Met Arg	
1430	1435	1440
Ser Leu Lys Ala Pro Ala Val	Val Ser Val Ser Ser Pro Asp Ala	
1445	1450	1455
Val Thr Thr Tyr Asn Gly Tyr	Leu Thr Ser Ser Ser Lys Thr Ser	
1460	1465	1470
Glu Glu His Phe Val Glu Thr	Val Ser Leu Ala Gly Ser Tyr Arg	
1475	1480	1485
Asp Trp Ser Tyr Ser Gly Gln	Arg Thr Glu Leu Gly Val Glu Phe	
1490	1495	1500
Leu Lys Arg Gly Asp Lys Ile	Val Tyr His Thr Leu Glu Ser Pro	
1505	1510	1515
Val Glu Phe His Leu Asp Gly	Glu Val Leu Ser Leu Asp Lys Leu	
1520	1525	1530
Lys Ser Leu Leu Ser Leu Arg	Glu Val Lys Thr Ile Lys Val Phe	
1535	1540	1545
Thr Thr Val Asp Asn Thr Asn	Leu His Thr Gln Leu Val Asp Met	
1550	1555	1560
Ser Met Thr Tyr Gly Gln Gln	Phe Gly Pro Thr Tyr Leu Asp Gly	
1565	1570	1575
Ala Asp Val Thr Lys Ile Lys	Pro His Val Asn His Glu Gly Lys	
1580	1585	1590
Thr Phe Phe Val Leu Pro Ser	Asp Asp Thr Leu Arg Ser Glu Ala	
1595	1600	1605
Phe Glu Tyr Tyr His Thr Leu	Asp Glu Ser Phe Leu Gly Arg Tyr	
1610	1615	1620

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Met	Ser	Ala	Leu	Asn	His	Thr	Lys	Lys	Trp	Lys	Phe	Pro	Gln	Val
1625						1630					1635			
Gly	Gly	Leu	Thr	Ser	Ile	Lys	Trp	Ala	Asp	Asn	Asn	Cys	Tyr	Leu
1640						1645					1650			
Ser	Ser	Val	Leu	Leu	Ala	Leu	Gln	Gln	Leu	Glu	Val	Lys	Phe	Asn
1655						1660					1665			
Ala	Pro	Ala	Leu	Gln	Glu	Ala	Tyr	Tyr	Arg	Ala	Arg	Ala	Gly	Asp
1670						1675					1680			
Ala	Ala	Asn	Phe	Cys	Ala	Leu	Ile	Leu	Ala	Tyr	Ser	Asn	Lys	Thr
1685						1690					1695			
Val	Gly	Glu	Leu	Gly	Asp	Val	Arg	Glu	Thr	Met	Thr	His	Leu	Leu
1700						1705					1710			
Gln	His	Ala	Asn	Leu	Glu	Ser	Ala	Lys	Arg	Val	Leu	Asn	Val	Val
1715						1720					1725			
Cys	Lys	His	Cys	Gly	Gln	Lys	Thr	Thr	Thr	Leu	Thr	Gly	Val	Glu
1730						1735					1740			
Ala	Val	Met	Tyr	Met	Gly	Thr	Leu	Ser	Tyr	Asp	Asn	Leu	Lys	Thr
1745						1750					1755			
Gly	Val	Ser	Ile	Pro	Cys	Val	Cys	Gly	Arg	Asp	Ala	Thr	Gln	Tyr
1760						1765					1770			
Leu	Val	Gln	Gln	Glu	Ser	Ser	Phe	Val	Met	Met	Ser	Ala	Pro	Pro
1775						1780					1785			
Ala	Glu	Tyr	Lys	Leu	Gln	Gln	Gly	Thr	Phe	Leu	Cys	Ala	Asn	Glu
1790						1795					1800			
Tyr	Thr	Gly	Asn	Tyr	Gln	Cys	Gly	His	Tyr	Thr	His	Ile	Thr	Ala
1805						1810					1815			
Lys	Glu	Thr	Leu	Tyr	Arg	Ile	Asp	Gly	Ala	His	Leu	Thr	Lys	Met
1820						1825					1830			
Ser	Glu	Tyr	Lys	Gly	Pro	Val	Thr	Asp	Val	Phe	Tyr	Lys	Glu	Thr
1835						1840					1845			
Ser	Tyr	Thr	Thr	Thr	Ile	Lys	Pro	Val	Ser	Tyr	Lys	Leu	Asp	Gly
1850						1855					1860			
Val	Thr	Tyr	Thr	Glu	Ile	Glu	Pro	Lys	Leu	Asp	Gly	Tyr	Tyr	Lys
1865						1870					1875			
Lys	Asp	Asn	Ala	Tyr	Tyr	Thr	Glu	Gln	Pro	Ile	Asp	Leu	Val	Pro
1880						1885					1890			
Thr	Gln	Pro	Leu	Pro	Asn	Ala	Ser	Phe	Asp	Asn	Phe	Lys	Leu	Thr
1895						1900					1905			
Cys	Ser	Asn	Thr	Lys	Phe	Ala	Asp	Asp	Leu	Asn	Gln	Met	Thr	Gly
1910						1915					1920			
Phe	Thr	Lys	Pro	Ala	Ser	Arg	Glu	Leu	Ser	Val	Thr	Phe	Phe	Pro
1925						1930					1935			
Asp	Leu	Asn	Gly	Asp	Val	Val	Ala	Ile	Asp	Tyr	Arg	His	Tyr	Ser
1940						1945					1950			
Ala	Ser	Phe	Lys	Lys	Gly	Ala	Lys	Leu	Leu	His	Lys	Pro	Ile	Val
1955						1960					1965			
Trp	His	Ile	Asn	Gln	Ala	Thr	Thr	Lys	Thr	Thr	Phe	Lys	Pro	Asn
1970						1975					1980			
Thr	Trp	Cys	Leu	Arg	Cys	Leu	Trp	Ser	Thr	Lys	Pro	Val	Asp	Thr
1985						1990					1995			
Ser	Asn	Ser	Phe	Glu	Val	Leu	Ala	Val	Glu	Asp	Thr	Gln	Gly	Met
2000						2005					2010			
Asp	Asn	Leu	Ala	Cys	Glu	Ser	Gln	Gln	Pro	Thr	Ser	Glu	Glu	Val
2015						2020					2025			

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Val Glu	Asn Pro Thr Ile Gln	Lys Glu Val Ile Glu	Cys Asp Val
2030	2035	2040	
Lys Thr	Thr Glu Val Val Gly	Asn Val Ile Leu Lys	Pro Ser Asp
2045	2050	2055	
Glu Gly	Val Lys Val Thr Gln	Glu Leu Gly His Glu	Asp Leu Met
2060	2065	2070	
Ala Ala	Tyr Val Glu Asn Thr	Ser Ile Thr Ile Lys	Lys Pro Asn
2075	2080	2085	
Glu Leu	Ser Leu Ala Leu Gly	Leu Lys Thr Ile Ala	Thr His Gly
2090	2095	2100	
Ile Ala	Ala Ile Asn Ser Val	Pro Trp Ser Lys Ile	Leu Ala Tyr
2105	2110	2115	
Val Lys	Pro Phe Leu Gly Gln	Ala Ala Ile Thr Thr	Ser Asn Cys
2120	2125	2130	
Ala Lys	Arg Leu Ala Gln Arg	Val Phe Asn Asn Tyr	Met Pro Tyr
2135	2140	2145	
Val Phe	Thr Leu Leu Phe Gln	Leu Cys Thr Phe Thr	Lys Ser Thr
2150	2155	2160	
Asn Ser	Arg Ile Arg Ala Ser	Leu Pro Thr Thr Ile	Ala Lys Asn
2165	2170	2175	
Ser Val	Lys Ser Val Ala Lys	Leu Cys Leu Asp Ala	Gly Ile Asn
2180	2185	2190	
Tyr Val	Lys Ser Pro Lys Phe	Ser Lys Leu Phe Thr	Ile Ala Met
2195	2200	2205	
Trp Leu	Leu Leu Leu Ser Ile	Cys Leu Gly Ser Leu	Ile Cys Val
2210	2215	2220	
Thr Ala	Ala Phe Gly Val Leu	Leu Ser Asn Phe Gly	Ala Pro Ser
2225	2230	2235	
Tyr Cys	Asn Gly Val Arg Glu	Leu Tyr Leu Asn Ser	Ser Asn Val
2240	2245	2250	
Thr Thr	Met Asp Phe Cys Glu	Gly Ser Phe Pro Cys	Ser Ile Cys
2255	2260	2265	
Leu Ser	Gly Leu Asp Ser Leu	Asp Ser Tyr Pro Ala	Leu Glu Thr
2270	2275	2280	
Ile Gln	Val Thr Ile Ser Ser	Tyr Lys Leu Asp Leu	Thr Ile Leu
2285	2290	2295	
Gly Leu	Ala Ala Glu Trp Val	Leu Ala Tyr Met Leu	Phe Thr Lys
2300	2305	2310	
Phe Phe	Tyr Leu Leu Gly Leu	Ser Ala Ile Met Gln	Val Phe Phe
2315	2320	2325	
Gly Tyr	Phe Ala Ser His Phe	Ile Ser Asn Ser Trp	Leu Met Trp
2330	2335	2340	
Phe Ile	Ile Ser Ile Val Gln	Met Ala Pro Val Ser	Ala Met Val
2345	2350	2355	
Arg Met	Tyr Ile Phe Phe Ala	Ser Phe Tyr Tyr Ile	Trp Lys Ser
2360	2365	2370	
Tyr Val	His Ile Met Asp Gly	Cys Thr Ser Ser Thr	Cys Met Met
2375	2380	2385	
Cys Tyr	Lys Arg Asn Arg Ala	Thr Arg Val Glu Cys	Thr Thr Ile
2390	2395	2400	
Val Asn	Gly Met Lys Arg Ser	Phe Tyr Val Tyr Ala	Asn Gly Gly
2405	2410	2415	
Arg Gly	Phe Cys Lys Thr His	Asn Trp Asn Cys Leu	Asn Cys Asp

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2420	2425	2430
Thr Phe Cys Thr Gly Ser 2435	Thr Phe Ile Ser Asp 2440	Glu Val Ala Arg 2445
Asp Leu Ser Leu Gln Phe 2450	Lys Arg Pro Ile Asn 2455	Pro Thr Asp Gln 2460
Ser Ser Tyr Ile Val Asp 2465	Ser Val Ala Val Lys 2470	Asn Gly Ala Leu 2475
His Leu Tyr Phe Asp Lys 2480	Ala Gly Gln Lys Thr 2485	Tyr Glu Arg His 2490
Pro Leu Ser His Phe Val 2495	Asn Leu Asp Asn Leu 2500	Arg Ala Asn Asn 2505
Thr Lys Gly Ser Leu Pro 2510	Ile Asn Val Ile Val 2515	Phe Asp Gly Lys 2520
Ser Lys Cys Asp Glu Ser 2525	Ala Ser Lys Ser Ala 2530	Ser Val Tyr Tyr 2535
Ser Gln Leu Met Cys Gln 2540	Pro Ile Leu Leu Leu 2545	Asp Gln Ala Leu 2550
Val Ser Asp Val Gly Asp 2555	Ser Thr Glu Val Ser 2560	Val Lys Met Phe 2565
Asp Ala Tyr Val Asp Thr 2570	Phe Ser Ala Thr Phe 2575	Ser Val Pro Met 2580
Glu Lys Leu Lys Ala Leu 2585	Val Ala Thr Ala His 2590	Ser Glu Leu Ala 2595
Lys Gly Val Ala Leu Asp 2600	Gly Val Leu Ser Thr 2605	Phe Val Ser Ala 2610
Ala Arg Gln Gly Val Val 2615	Asp Thr Asp Val Asp 2620	Thr Lys Asp Val 2625
Ile Glu Cys Leu Lys Leu 2630	Ser His His Ser Asp 2635	Leu Glu Val Thr 2640
Gly Asp Ser Cys Asn Asn 2645	Phe Met Leu Thr Tyr 2650	Asn Lys Val Glu 2655
Asn Met Thr Pro Arg Asp 2660	Leu Gly Ala Cys Ile 2665	Asp Cys Asn Ala 2670
Arg His Ile Asn Ala Gln 2675	Val Ala Lys Ser His 2680	Asn Val Ser Leu 2685
Ile Trp Asn Val Lys Asp 2690	Tyr Met Ser Leu Ser 2695	Glu Gln Leu Arg 2700
Lys Gln Ile Arg Ser Ala 2705	Ala Lys Lys Asn Asn 2710	Ile Pro Phe Arg 2715
Leu Thr Cys Ala Thr Thr 2720	Arg Gln Val Val Asn 2725	Val Ile Thr Thr 2730
Lys Ile Ser Leu Lys Gly 2735	Gly Lys Ile Val Ser 2740	Thr Cys Phe Lys 2745
Leu Met Leu Lys Ala Thr 2750	Leu Leu Cys Val Leu 2755	Ala Ala Leu Val 2760
Cys Tyr Ile Val Met Pro 2765	Val His Thr Leu Ser 2770	Ile His Asp Gly 2775
Tyr Thr Asn Glu Ile Ile 2780	Gly Tyr Lys Ala Ile 2785	Gln Asp Gly Val 2790
Thr Arg Asp Ile Ile Ser 2795	Thr Asp Asp Cys Phe 2800	Ala Asn Lys His 2805
Ala Gly Phe Asp Ala Trp 2810	Phe Ser Gln Arg Gly 2815	Gly Ser Tyr Lys 2820

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Asn Asp 2825	Lys Ser Cys Pro	Val 2830	Val Ala Ala Ile	Ile Thr Arg Glu 2835
Ile Gly 2840	Phe Ile Val Pro	Gly 2845	Leu Pro Gly Thr	Val Leu Arg Ala 2850
Ile Asn 2855	Gly Asp Phe Leu	His 2860	Phe Leu Pro Arg	Val Phe Ser Ala 2865
Val Gly 2870	Asn Ile Cys Tyr	Thr 2875	Pro Ser Lys Leu	Ile Glu Tyr Ser 2880
Asp Phe 2885	Ala Thr Ser Ala	Cys 2890	Val Leu Ala Ala	Glu Cys Thr Ile 2895
Phe Lys 2900	Asp Ala Met Gly	Lys 2905	Pro Val Pro Tyr	Cys Tyr Asp Thr 2910
Asn Leu 2915	Leu Glu Gly Ser	Ile 2920	Ser Tyr Ser Glu	Leu Arg Pro Asp 2925
Thr Arg 2930	Tyr Val Leu Met	Asp 2935	Gly Ser Ile Ile	Gln Phe Pro Asn 2940
Thr Tyr 2945	Leu Glu Gly Ser	Val 2950	Arg Val Val Thr	Thr Phe Asp Ala 2955
Glu Tyr 2960	Cys Arg His Gly	Thr 2965	Cys Glu Arg Ser	Glu Val Gly Ile 2970
Cys Leu 2975	Ser Thr Ser Gly	Arg 2980	Trp Val Leu Asn	Asn Glu His Tyr 2985
Arg Ala 2990	Leu Ser Gly Val	Phe 2995	Cys Gly Val Asp	Ala Met Asn Leu 3000
Ile Ala 3005	Asn Ile Phe Thr	Pro 3010	Leu Val Gln Pro	Val Gly Ala Leu 3015
Asp Val 3020	Ser Ala Ser Val	Val 3025	Ala Gly Gly Ile	Ile Ala Ile Leu 3030
Val Thr 3035	Cys Ala Ala Tyr	Tyr 3040	Phe Met Lys Phe	Arg Arg Val Phe 3045
Gly Glu 3050	Tyr Asn His Val	Val 3055	Ala Ala Asn Ala	Leu Leu Phe Leu 3060
Met Ser 3065	Phe Thr Ile Leu	Cys 3070	Leu Val Pro Ala	Tyr Ser Phe Leu 3075
Pro Gly 3080	Val Tyr Ser Val	Phe 3085	Tyr Leu Tyr Leu	Thr Phe Tyr Phe 3090
Thr Asn 3095	Asp Val Ser Phe	Leu 3100	Ala His Leu Gln	Trp Phe Ala Met 3105
Phe Ser 3110	Pro Ile Val Pro	Phe 3115	Trp Ile Thr Ala	Ile Tyr Val Phe 3120
Cys Ile 3125	Ser Leu Lys His	Cys 3130	His Trp Phe Phe	Asn Asn Tyr Leu 3135
Arg Lys 3140	Arg Val Met Phe	Asn 3145	Gly Val Thr Phe	Ser Thr Phe Glu 3150
Glu Ala 3155	Ala Leu Cys Thr	Phe 3160	Leu Leu Asn Lys	Glu Met Tyr Leu 3165
Lys Leu 3170	Arg Ser Glu Thr	Leu 3175	Leu Pro Leu Thr	Gln Tyr Asn Arg 3180
Tyr Leu 3185	Ala Leu Tyr Asn	Lys 3190	Tyr Lys Tyr Phe	Ser Gly Ala Leu 3195
Asp Thr 3200	Thr Ser Tyr Arg	Glu 3205	Ala Ala Cys Cys	His Leu Ala Lys 3210
Ala Leu 3215	Asn Asp Phe Ser	Asn 3220	Ser Gly Ala Asp	Val Leu Tyr Gln 3225

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Pro	Pro	Gln	Thr	Ser	Ile	Thr	Ser	Ala	Val	Leu	Gln	Ser	Gly	Phe
3230						3235					3240			
Arg	Lys	Met	Ala	Phe	Pro	Ser	Gly	Lys	Val	Glu	Gly	Cys	Met	Val
3245						3250					3255			
Gln	Val	Thr	Cys	Gly	Thr	Thr	Thr	Leu	Asn	Gly	Leu	Trp	Leu	Asp
3260						3265					3270			
Asp	Thr	Val	Tyr	Cys	Pro	Arg	His	Val	Ile	Cys	Thr	Ala	Glu	Asp
3275						3280					3285			
Met	Leu	Asn	Pro	Asn	Tyr	Glu	Asp	Leu	Leu	Ile	Arg	Lys	Ser	Asn
3290						3295					3300			
His	Ser	Phe	Leu	Val	Gln	Ala	Gly	Asn	Val	Gln	Leu	Arg	Val	Ile
3305						3310					3315			
Gly	His	Ser	Met	Gln	Asn	Cys	Leu	Leu	Arg	Leu	Lys	Val	Asp	Thr
3320						3325					3330			
Ser	Asn	Pro	Lys	Thr	Pro	Lys	Tyr	Lys	Phe	Val	Arg	Ile	Gln	Pro
3335						3340					3345			
Gly	Gln	Thr	Phe	Ser	Val	Leu	Ala	Cys	Tyr	Asn	Gly	Ser	Pro	Ser
3350						3355					3360			
Gly	Val	Tyr	Gln	Cys	Ala	Met	Arg	Pro	Asn	His	Thr	Ile	Lys	Gly
3365						3370					3375			
Ser	Phe	Leu	Asn	Gly	Ser	Cys	Gly	Ser	Val	Gly	Phe	Asn	Ile	Asp
3380						3385					3390			
Tyr	Asp	Cys	Val	Ser	Phe	Cys	Tyr	Met	His	His	Met	Glu	Leu	Pro
3395						3400					3405			
Thr	Gly	Val	His	Ala	Gly	Thr	Asp	Leu	Glu	Gly	Lys	Phe	Tyr	Gly
3410						3415					3420			
Pro	Phe	Val	Asp	Arg	Gln	Thr	Ala	Gln	Ala	Ala	Gly	Thr	Asp	Thr
3425						3430					3435			
Thr	Ile	Thr	Leu	Asn	Val	Leu	Ala	Trp	Leu	Tyr	Ala	Ala	Val	Ile
3440						3445					3450			
Asn	Gly	Asp	Arg	Trp	Phe	Leu	Asn	Arg	Phe	Thr	Thr	Thr	Leu	Asn
3455						3460					3465			
Asp	Phe	Asn	Leu	Val	Ala	Met	Lys	Tyr	Asn	Tyr	Glu	Pro	Leu	Thr
3470						3475					3480			
Gln	Asp	His	Val	Asp	Ile	Leu	Gly	Pro	Leu	Ser	Ala	Gln	Thr	Gly
3485						3490					3495			
Ile	Ala	Val	Leu	Asp	Met	Cys	Ala	Ala	Leu	Lys	Glu	Leu	Leu	Gln
3500						3505					3510			
Asn	Gly	Met	Asn	Gly	Arg	Thr	Ile	Leu	Gly	Ser	Thr	Ile	Leu	Glu
3515						3520					3525			
Asp	Glu	Phe	Thr	Pro	Phe	Asp	Val	Val	Arg	Gln	Cys	Ser	Gly	Val
3530						3535					3540			
Thr	Phe	Gln	Gly	Lys	Phe	Lys	Lys	Ile	Val	Lys	Gly	Thr	His	His
3545						3550					3555			
Trp	Met	Leu	Leu	Thr	Phe	Leu	Thr	Ser	Leu	Leu	Ile	Leu	Val	Gln
3560						3565					3570			
Ser	Thr	Gln	Trp	Ser	Leu	Phe	Phe	Phe	Val	Tyr	Glu	Asn	Ala	Phe
3575						3580					3585			
Leu	Pro	Phe	Thr	Leu	Gly	Ile	Met	Ala	Ile	Ala	Ala	Cys	Ala	Met
3590						3595					3600			
Leu	Leu	Val	Lys	His	Lys	His	Ala	Phe	Leu	Cys	Leu	Phe	Leu	Leu
3605						3610					3615			
Pro	Ser	Leu	Ala	Thr	Val	Ala	Tyr	Phe	Asn	Met	Val	Tyr	Met	Pro

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3620	3625	3630
Ala Ser Trp Val Met Arg Ile Met Thr Trp Leu Glu Leu Ala Asp		
3635	3640	3645
Thr Ser Leu Ser Gly Tyr Arg Leu Lys Asp Cys Val Met Tyr Ala		
3650	3655	3660
Ser Ala Leu Val Leu Leu Ile Leu Met Thr Ala Arg Thr Val Tyr		
3665	3670	3675
Asp Asp Ala Ala Arg Arg Val Trp Thr Leu Met Asn Val Ile Thr		
3680	3685	3690
Leu Val Tyr Lys Val Tyr Tyr Gly Asn Ala Leu Asp Gln Ala Ile		
3695	3700	3705
Ser Met Trp Ala Leu Val Ile Ser Val Thr Ser Asn Tyr Ser Gly		
3710	3715	3720
Val Val Thr Thr Ile Met Phe Leu Ala Arg Ala Ile Val Phe Val		
3725	3730	3735
Cys Val Glu Tyr Tyr Pro Leu Leu Phe Ile Thr Gly Asn Thr Leu		
3740	3745	3750
Gln Cys Ile Met Leu Val Tyr Cys Phe Leu Gly Tyr Cys Cys Cys		
3755	3760	3765
Cys Tyr Phe Gly Leu Phe Cys Leu Leu Asn Arg Tyr Phe Arg Leu		
3770	3775	3780
Thr Leu Gly Val Tyr Asp Tyr Leu Val Ser Thr Gln Glu Phe Arg		
3785	3790	3795
Tyr Met Asn Ser Gln Gly Leu Leu Pro Pro Lys Ser Ser Ile Asp		
3800	3805	3810
Ala Phe Lys Leu Asn Ile Lys Leu Leu Gly Ile Gly Gly Lys Pro		
3815	3820	3825
Cys Ile Lys Val Ala Thr Val Gln Ser Lys Met Ser Asp Val Lys		
3830	3835	3840
Cys Thr Ser Val Val Leu Leu Ser Val Leu Gln Gln Leu Arg Val		
3845	3850	3855
Glu Ser Ser Ser Lys Leu Trp Ala Gln Cys Val Gln Leu His Asn		
3860	3865	3870
Asp Ile Leu Leu Ala Lys Asp Thr Thr Glu Ala Phe Glu Lys Met		
3875	3880	3885
Val Ser Leu Leu Ser Val Leu Leu Ser Met Gln Gly Ala Val Asp		
3890	3895	3900
Ile Asn Arg Leu Cys Glu Glu Met Leu Asp Asn Arg Ala Thr Leu		
3905	3910	3915
Gln Ala Ile Ala Ser Glu Phe Ser Ser Leu Pro Ser Tyr Ala Ala		
3920	3925	3930
Tyr Ala Thr Ala Gln Glu Ala Tyr Glu Gln Ala Val Ala Asn Gly		
3935	3940	3945
Asp Ser Glu Val Val Leu Lys Lys Leu Lys Lys Ser Leu Asn Val		
3950	3955	3960
Ala Lys Ser Glu Phe Asp Arg Asp Ala Ala Met Gln Arg Lys Leu		
3965	3970	3975
Glu Lys Met Ala Asp Gln Ala Met Thr Gln Met Tyr Lys Gln Ala		
3980	3985	3990
Arg Ser Glu Asp Lys Arg Ala Lys Val Thr Ser Ala Met Gln Thr		
3995	4000	4005
Met Leu Phe Thr Met Leu Arg Lys Leu Asp Asn Asp Ala Leu Asn		
4010	4015	4020

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Asn Ile	Ile Asn Asn Ala Arg	Asp Gly Cys Val Pro	Leu Asn Ile
4025	4030	4035	
Ile Pro	Leu Thr Thr Ala Ala	Lys Leu Met Val Val	Val Pro Asp
4040	4045	4050	
Tyr Gly	Thr Tyr Lys Asn Thr	Cys Asp Gly Asn Thr	Phe Thr Tyr
4055	4060	4065	
Ala Ser	Ala Leu Trp Glu Ile	Gln Gln Val Val Asp	Ala Asp Ser
4070	4075	4080	
Lys Ile	Val Gln Leu Ser Glu	Ile Asn Met Asp Asn	Ser Pro Asn
4085	4090	4095	
Leu Ala	Trp Pro Leu Ile Val	Thr Ala Leu Arg Ala	Asn Ser Ala
4100	4105	4110	
Val Lys	Leu Gln Asn Asn Glu	Leu Ser Pro Val Ala	Leu Arg Gln
4115	4120	4125	
Met Ser	Cys Ala Ala Gly Thr	Thr Gln Thr Ala Cys	Thr Asp Asp
4130	4135	4140	
Asn Ala	Leu Ala Tyr Tyr Asn	Asn Ser Lys Gly Gly	Arg Phe Val
4145	4150	4155	
Leu Ala	Leu Leu Ser Asp His	Gln Asp Leu Lys Trp	Ala Arg Phe
4160	4165	4170	
Pro Lys	Ser Asp Gly Thr Gly	Thr Ile Tyr Thr Glu	Leu Glu Pro
4175	4180	4185	
Pro Cys	Arg Phe Val Thr Asp	Thr Pro Lys Gly Pro	Lys Val Lys
4190	4195	4200	
Tyr Leu	Tyr Phe Ile Lys Gly	Leu Asn Asn Leu Asn	Arg Gly Met
4205	4210	4215	
Val Leu	Gly Ser Leu Ala Ala	Thr Val Arg Leu Gln	Ala Gly Asn
4220	4225	4230	
Ala Thr	Glu Val Pro Ala Asn	Ser Thr Val Leu Ser	Phe Cys Ala
4235	4240	4245	
Phe Ala	Val Asp Pro Ala Lys	Ala Tyr Lys Asp Tyr	Leu Ala Ser
4250	4255	4260	
Gly Gly	Gln Pro Ile Thr Asn	Cys Val Lys Met Leu	Cys Thr His
4265	4270	4275	
Thr Gly	Thr Gly Gln Ala Ile	Thr Val Thr Pro Glu	Ala Asn Met
4280	4285	4290	
Asp Gln	Glu Ser Phe Gly Gly	Ala Ser Cys Cys Leu	Tyr Cys Arg
4295	4300	4305	
Cys His	Ile Asp His Pro Asn	Pro Lys Gly Phe Cys	Asp Leu Lys
4310	4315	4320	
Gly Lys	Tyr Val Gln Ile Pro	Thr Thr Cys Ala Asn	Asp Pro Val
4325	4330	4335	
Gly Phe	Thr Leu Arg Asn Thr	Val Cys Thr Val Cys	Gly Met Trp
4340	4345	4350	
Lys Gly	Tyr Gly Cys Ser Cys	Asp Gln Leu Arg Glu	Pro Leu Met
4355	4360	4365	
Gln Ser	Ala Asp Ala Ser Thr	Phe Leu Asn Gly Phe	Ala Val
4370	4375	4380	

<210> SEQ ID NO 75

<211> LENGTH: 2695

<212> TYPE: PRT

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 75

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Arg	Val	Cys	Gly	Val	Ser	Ala	Ala	Arg	Leu	Thr	Pro	Cys	Gly	Thr	Gly
1				5					10					15	
Thr	Ser	Thr	Asp	Val	Val	Tyr	Arg	Ala	Phe	Asp	Ile	Tyr	Asn	Glu	Lys
			20					25					30		
Val	Ala	Gly	Phe	Ala	Lys	Phe	Leu	Lys	Thr	Asn	Cys	Cys	Arg	Phe	Gln
		35					40					45			
Glu	Lys	Asp	Glu	Glu	Gly	Asn	Leu	Leu	Asp	Ser	Tyr	Phe	Val	Val	Lys
	50					55				60					
Arg	His	Thr	Met	Ser	Asn	Tyr	Gln	His	Glu	Glu	Thr	Ile	Tyr	Asn	Leu
65					70					75					80
Val	Lys	Asp	Cys	Pro	Ala	Val	Ala	Val	His	Asp	Phe	Phe	Lys	Phe	Arg
				85					90					95	
Val	Asp	Gly	Asp	Met	Val	Pro	His	Ile	Ser	Arg	Gln	Arg	Leu	Thr	Lys
			100					105					110		
Tyr	Thr	Met	Ala	Asp	Leu	Val	Tyr	Ala	Leu	Arg	His	Phe	Asp	Glu	Gly
		115					120					125			
Asn	Cys	Asp	Thr	Leu	Lys	Glu	Ile	Leu	Val	Thr	Tyr	Asn	Cys	Cys	Asp
	130					135					140				
Asp	Asp	Tyr	Phe	Asn	Lys	Lys	Asp	Trp	Tyr	Asp	Phe	Val	Glu	Asn	Pro
145					150					155					160
Asp	Ile	Leu	Arg	Val	Tyr	Ala	Asn	Leu	Gly	Glu	Arg	Val	Arg	Gln	Ser
				165					170					175	
Leu	Leu	Lys	Thr	Val	Gln	Phe	Cys	Asp	Ala	Met	Arg	Asp	Ala	Gly	Ile
		180						185					190		
Val	Gly	Val	Leu	Thr	Leu	Asp	Asn	Gln	Asp	Leu	Asn	Gly	Asn	Trp	Tyr
		195					200					205			
Asp	Phe	Gly	Asp	Phe	Val	Gln	Val	Ala	Pro	Gly	Cys	Gly	Val	Pro	Ile
	210					215					220				
Val	Asp	Ser	Tyr	Tyr	Ser	Leu	Leu	Met	Pro	Ile	Leu	Thr	Leu	Thr	Arg
225					230					235					240
Ala	Leu	Ala	Ala	Glu	Ser	His	Met	Asp	Ala	Asp	Leu	Ala	Lys	Pro	Leu
				245					250					255	
Ile	Lys	Trp	Asp	Leu	Leu	Lys	Tyr	Asp	Phe	Thr	Glu	Glu	Arg	Leu	Cys
			260					265					270		
Leu	Phe	Asp	Arg	Tyr	Phe	Lys	Tyr	Trp	Asp	Gln	Thr	Tyr	His	Pro	Asn
		275					280					285			
Cys	Ile	Asn	Cys	Leu	Asp	Asp	Arg	Cys	Ile	Leu	His	Cys	Ala	Asn	Phe
	290					295					300				
Asn	Val	Leu	Phe	Ser	Thr	Val	Phe	Pro	Pro	Thr	Ser	Phe	Gly	Pro	Leu
305					310					315					320
Val	Arg	Lys	Ile	Phe	Val	Asp	Gly	Val	Pro	Phe	Val	Val	Ser	Thr	Gly
				325					330					335	
Tyr	His	Phe	Arg	Glu	Leu	Gly	Val	Val	His	Asn	Gln	Asp	Val	Asn	Leu
			340				345						350		
His	Ser	Ser	Arg	Leu	Ser	Phe	Lys	Glu	Leu	Leu	Val	Tyr	Ala	Ala	Asp
		355					360					365			
Pro	Ala	Met	His	Ala	Ala	Ser	Gly	Asn	Leu	Leu	Leu	Asp	Lys	Arg	Thr
	370					375					380				
Thr	Cys	Phe	Ser	Val	Ala	Ala	Leu	Thr	Asn	Asn	Val	Ala	Phe	Gln	Thr
385					390					395					400
Val	Lys	Pro	Gly	Asn	Phe	Asn	Lys	Asp	Phe	Tyr	Asp	Phe	Ala	Val	Ser
				405					410					415	
Lys	Gly	Phe	Phe	Lys	Glu	Gly	Ser	Ser	Val	Glu	Leu	Lys	His	Phe	Phe
			420					425					430		

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Phe Ala Gln Asp Gly Asn Ala Ala Ile Ser Asp Tyr Asp Tyr Tyr Arg
 435 440 445
 Tyr Asn Leu Pro Thr Met Cys Asp Ile Arg Gln Leu Leu Phe Val Val
 450 455 460
 Glu Val Val Asp Lys Tyr Phe Asp Cys Tyr Asp Gly Gly Cys Ile Asn
 465 470 475 480
 Ala Asn Gln Val Ile Val Asn Asn Leu Asp Lys Ser Ala Gly Phe Pro
 485 490 495
 Phe Asn Lys Trp Gly Lys Ala Arg Leu Tyr Tyr Asp Ser Met Ser Tyr
 500 505 510
 Glu Asp Gln Asp Ala Leu Phe Ala Tyr Thr Lys Arg Asn Val Ile Pro
 515 520 525
 Thr Ile Thr Gln Met Asn Leu Lys Tyr Ala Ile Ser Ala Lys Asn Arg
 530 535 540
 Ala Arg Thr Val Ala Gly Val Ser Ile Cys Ser Thr Met Thr Asn Arg
 545 550 555 560
 Gln Phe His Gln Lys Leu Leu Lys Ser Ile Ala Ala Thr Arg Gly Ala
 565 570 575
 Thr Val Val Ile Gly Thr Ser Lys Phe Tyr Gly Gly Trp His Asn Met
 580 585 590
 Leu Lys Thr Val Tyr Ser Asp Val Glu Thr Pro His Leu Met Gly Trp
 595 600 605
 Asp Tyr Pro Lys Cys Asp Arg Ala Met Pro Asn Met Leu Arg Ile Met
 610 615 620
 Ala Ser Leu Val Leu Ala Arg Lys His Asn Thr Cys Cys Asn Leu Ser
 625 630 635 640
 His Arg Phe Tyr Arg Leu Ala Asn Glu Cys Ala Gln Val Leu Ser Glu
 645 650 655
 Met Val Met Cys Gly Gly Ser Leu Tyr Val Lys Pro Gly Gly Thr Ser
 660 665 670
 Ser Gly Asp Ala Thr Thr Ala Tyr Ala Asn Ser Val Phe Asn Ile Cys
 675 680 685
 Gln Ala Val Thr Ala Asn Val Asn Ala Leu Leu Ser Thr Asp Gly Asn
 690 695 700
 Lys Ile Ala Asp Lys Tyr Val Arg Asn Leu Gln His Arg Leu Tyr Glu
 705 710 715 720
 Cys Leu Tyr Arg Asn Arg Asp Val Asp His Glu Phe Val Asp Glu Phe
 725 730 735
 Tyr Ala Tyr Leu Arg Lys His Phe Ser Met Met Ile Leu Ser Asp Asp
 740 745 750
 Ala Val Val Cys Tyr Asn Ser Asn Tyr Ala Ala Gln Gly Leu Val Ala
 755 760 765
 Ser Ile Lys Asn Phe Lys Ala Val Leu Tyr Tyr Gln Asn Asn Val Phe
 770 775 780
 Met Ser Glu Ala Lys Cys Trp Thr Glu Thr Asp Leu Thr Lys Gly Pro
 785 790 795 800
 His Glu Phe Cys Ser Gln His Thr Met Leu Val Lys Gln Gly Asp Asp
 805 810 815
 Tyr Val Tyr Leu Pro Tyr Pro Asp Pro Ser Arg Ile Leu Gly Ala Gly
 820 825 830
 Cys Phe Val Asp Asp Ile Val Lys Thr Asp Gly Thr Leu Met Ile Glu
 835 840 845
 Arg Phe Val Ser Leu Ala Ile Asp Ala Tyr Pro Leu Thr Lys His Pro

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850	855	860
Asn Gln Glu Tyr Ala Asp Val Phe His Leu Tyr Leu Gln Tyr Ile Arg		
865	870	875 880
Lys Leu His Asp Glu Leu Thr Gly His Met Leu Asp Met Tyr Ser Val		
	885	890 895
Met Leu Thr Asn Asp Asn Thr Ser Arg Tyr Trp Glu Pro Glu Phe Tyr		
	900	905 910
Glu Ala Met Tyr Thr Pro His Thr Val Leu Gln Ala Val Gly Ala Cys		
	915	920 925
Val Leu Cys Asn Ser Gln Thr Ser Leu Arg Cys Gly Ala Cys Ile Arg		
	930	935 940
Arg Pro Phe Leu Cys Cys Lys Cys Cys Tyr Asp His Val Ile Ser Thr		
	945	950 955 960
Ser His Lys Leu Val Leu Ser Val Asn Pro Tyr Val Cys Asn Ala Pro		
	965	970 975
Gly Cys Asp Val Thr Asp Val Thr Gln Leu Tyr Leu Gly Gly Met Ser		
	980	985 990
Tyr Tyr Cys Lys Ser His Lys Pro Pro Ile Ser Phe Pro Leu Cys Ala		
	995	1000 1005
Asn Gly Gln Val Phe Gly Leu Tyr Lys Asn Thr Cys Val Gly Ser		
	1010	1015 1020
Asp Asn Val Thr Asp Phe Asn Ala Ile Ala Thr Cys Asp Trp Thr		
	1025	1030 1035
Asn Ala Gly Asp Tyr Ile Leu Ala Asn Thr Cys Thr Glu Arg Leu		
	1040	1045 1050
Lys Leu Phe Ala Ala Glu Thr Leu Lys Ala Thr Glu Glu Thr Phe		
	1055	1060 1065
Lys Leu Ser Tyr Gly Ile Ala Thr Val Arg Glu Val Leu Ser Asp		
	1070	1075 1080
Arg Glu Leu His Leu Ser Trp Glu Val Gly Lys Pro Arg Pro Pro		
	1085	1090 1095
Leu Asn Arg Asn Tyr Val Phe Thr Gly Tyr Arg Val Thr Lys Asn		
	1100	1105 1110
Ser Lys Val Gln Ile Gly Glu Tyr Thr Phe Glu Lys Gly Asp Tyr		
	1115	1120 1125
Gly Asp Ala Val Val Tyr Arg Gly Thr Thr Thr Tyr Lys Leu Asn		
	1130	1135 1140
Val Gly Asp Tyr Phe Val Leu Thr Ser His Thr Val Met Pro Leu		
	1145	1150 1155
Ser Ala Pro Thr Leu Val Pro Gln Glu His Tyr Val Arg Ile Thr		
	1160	1165 1170
Gly Leu Tyr Pro Thr Leu Asn Ile Ser Asp Glu Phe Ser Ser Asn		
	1175	1180 1185
Val Ala Asn Tyr Gln Lys Val Gly Met Gln Lys Tyr Ser Thr Leu		
	1190	1195 1200
Gln Gly Pro Pro Gly Thr Gly Lys Ser His Phe Ala Ile Gly Leu		
	1205	1210 1215
Ala Leu Tyr Tyr Pro Ser Ala Arg Ile Val Tyr Thr Ala Cys Ser		
	1220	1225 1230
His Ala Ala Val Asp Ala Leu Cys Glu Lys Ala Leu Lys Tyr Leu		
	1235	1240 1245
Pro Ile Asp Lys Cys Ser Arg Ile Ile Pro Ala Arg Ala Arg Val		
	1250	1255 1260

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Glu Cys Phe Asp Lys Phe Lys Val Asn Ser Thr Leu Glu Gln Tyr	1265	1270	1275
Val Phe Cys Thr Val Asn Ala Leu Pro Glu Thr Thr Ala Asp Ile	1280	1285	1290
Val Val Phe Asp Glu Ile Ser Met Ala Thr Asn Tyr Asp Leu Ser	1295	1300	1305
Val Val Asn Ala Arg Leu Arg Ala Lys His Tyr Val Tyr Ile Gly	1310	1315	1320
Asp Pro Ala Gln Leu Pro Ala Pro Arg Thr Leu Leu Thr Lys Gly	1325	1330	1335
Thr Leu Glu Pro Glu Tyr Phe Asn Ser Val Cys Arg Leu Met Lys	1340	1345	1350
Thr Ile Gly Pro Asp Met Phe Leu Gly Thr Cys Arg Arg Cys Pro	1355	1360	1365
Ala Glu Ile Val Asp Thr Val Ser Ala Leu Val Tyr Asp Asn Lys	1370	1375	1380
Leu Lys Ala His Lys Asp Lys Ser Ala Gln Cys Phe Lys Met Phe	1385	1390	1395
Tyr Lys Gly Val Ile Thr His Asp Val Ser Ser Ala Ile Asn Arg	1400	1405	1410
Pro Gln Ile Gly Val Val Arg Glu Phe Leu Thr Arg Asn Pro Ala	1415	1420	1425
Trp Arg Lys Ala Val Phe Ile Ser Pro Tyr Asn Ser Gln Asn Ala	1430	1435	1440
Val Ala Ser Lys Ile Leu Gly Leu Pro Thr Gln Thr Val Asp Ser	1445	1450	1455
Ser Gln Gly Ser Glu Tyr Asp Tyr Val Ile Phe Thr Gln Thr Thr	1460	1465	1470
Glu Thr Ala His Ser Cys Asn Val Asn Arg Phe Asn Val Ala Ile	1475	1480	1485
Thr Arg Ala Lys Ile Gly Ile Leu Cys Ile Met Ser Asp Arg Asp	1490	1495	1500
Leu Tyr Asp Lys Leu Gln Phe Thr Ser Leu Glu Ile Pro Arg Arg	1505	1510	1515
Asn Val Ala Thr Leu Gln Ala Glu Asn Val Thr Gly Leu Phe Lys	1520	1525	1530
Asp Cys Ser Lys Ile Ile Thr Gly Leu His Pro Thr Gln Ala Pro	1535	1540	1545
Thr His Leu Ser Val Asp Ile Lys Phe Lys Thr Glu Gly Leu Cys	1550	1555	1560
Val Asp Ile Pro Gly Ile Pro Lys Asp Met Thr Tyr Arg Arg Leu	1565	1570	1575
Ile Ser Met Met Gly Phe Lys Met Asn Tyr Gln Val Asn Gly Tyr	1580	1585	1590
Pro Asn Met Phe Ile Thr Arg Glu Glu Ala Ile Arg His Val Arg	1595	1600	1605
Ala Trp Ile Gly Phe Asp Val Glu Gly Cys His Ala Thr Arg Asp	1610	1615	1620
Ala Val Gly Thr Asn Leu Pro Leu Gln Leu Gly Phe Ser Thr Gly	1625	1630	1635
Val Asn Leu Val Ala Val Pro Thr Gly Tyr Val Asp Thr Glu Asn	1640	1645	1650
Asn Thr Glu Phe Thr Arg Val Asn Ala Lys Pro Pro Pro Gly Asp	1655	1660	1665

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Gln Phe 1670	Lys His Leu Ile Pro 1675	Leu Met Tyr Lys Gly 1680	Leu Pro Trp
Asn Val 1685	Val Arg Ile Lys Ile 1690	Val Gln Met Leu Ser 1695	Asp Thr Leu
Lys Gly 1700	Leu Ser Asp Arg Val 1705	Val Phe Val Leu Trp 1710	Ala His Gly
Phe Glu 1715	Leu Thr Ser Met Lys 1720	Tyr Phe Val Lys Ile 1725	Gly Pro Glu
Arg Thr 1730	Cys Cys Leu Cys Asp 1735	Lys Arg Ala Thr Cys 1740	Phe Ser Thr
Ser Ser 1745	Asp Thr Tyr Ala Cys 1750	Trp Asn His Ser Val 1755	Gly Phe Asp
Tyr Val 1760	Tyr Asn Pro Phe Met 1765	Ile Asp Val Gln Gln 1770	Trp Gly Phe
Thr Gly 1775	Asn Leu Gln Ser Asn 1780	His Asp Gln His Cys 1785	Gln Val His
Gly Asn 1790	Ala His Val Ala Ser 1795	Cys Asp Ala Ile Met 1800	Thr Arg Cys
Leu Ala 1805	Val His Glu Cys Phe 1810	Val Lys Arg Val Asp 1815	Trp Ser Val
Glu Tyr 1820	Pro Ile Ile Gly Asp 1825	Glu Leu Arg Val Asn 1830	Ser Ala Cys
Arg Lys 1835	Val Gln His Met Val 1840	Val Lys Ser Ala Leu 1845	Leu Ala Asp
Lys Phe 1850	Pro Val Leu His Asp 1855	Ile Gly Asn Pro Lys 1860	Ala Ile Lys
Cys Val 1865	Pro Gln Ala Glu Val 1870	Glu Trp Lys Phe Tyr 1875	Asp Ala Gln
Pro Cys 1880	Ser Asp Lys Ala Tyr 1885	Lys Ile Glu Glu Leu 1890	Phe Tyr Ser
Tyr Ala 1895	Thr His His Asp Lys 1900	Phe Thr Asp Gly Val 1905	Cys Leu Phe
Trp Asn 1910	Cys Asn Val Asp Arg 1915	Tyr Pro Ala Asn Ala 1920	Ile Val Cys
Arg Phe 1925	Asp Thr Arg Val Leu 1930	Ser Asn Leu Asn Leu 1935	Pro Gly Cys
Asp Gly 1940	Gly Ser Leu Tyr Val 1945	Asn Lys His Ala Phe 1950	His Thr Pro
Ala Phe 1955	Asp Lys Ser Ala Phe 1960	Thr Asn Leu Lys Gln 1965	Leu Pro Phe
Phe Tyr 1970	Tyr Ser Asp Ser Pro 1975	Cys Glu Ser His Gly 1980	Lys Gln Val
Val Ser 1985	Asp Ile Asp Tyr Val 1990	Pro Leu Lys Ser Ala 1995	Thr Cys Ile
Thr Arg 2000	Cys Asn Leu Gly Gly 2005	Ala Val Cys Arg His 2010	His Ala Asn
Glu Tyr 2015	Arg Gln Tyr Leu Asp 2020	Ala Tyr Asn Met Met 2025	Ile Ser Ala
Gly Phe 2030	Ser Leu Trp Ile Tyr 2035	Lys Gln Phe Asp Thr 2040	Tyr Asn Leu
Trp Asn 2045	Thr Phe Thr Arg Leu 2050	Gln Ser Leu Glu Asn 2055	Val Ala Tyr
Asn Val	Val Asn Lys Gly His	Phe Asp Gly His Ala	Gly Glu Ala

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2060	2065	2070
Pro Val Ser Ile Ile Asn 2075	Asn Ala Val Tyr Thr 2080	Lys Val Asp Gly 2085
Ile Asp Val Glu Ile Phe 2090	Glu Asn Lys Thr Thr 2095	Leu Pro Val Asn 2100
Val Ala Phe Glu Leu Trp 2105	Ala Lys Arg Asn Ile 2110	Lys Pro Val Pro 2115
Glu Ile Lys Ile Leu Asn 2120	Asn Leu Gly Val Asp 2125	Ile Ala Ala Asn 2130
Thr Val Ile Trp Asp Tyr 2135	Lys Arg Glu Ala Pro 2140	Ala His Val Ser 2145
Thr Ile Gly Val Cys Thr 2150	Met Thr Asp Ile Ala 2155	Lys Lys Pro Thr 2160
Glu Ser Ala Cys Ser Ser 2165	Leu Thr Val Leu Phe 2170	Asp Gly Arg Val 2175
Glu Gly Gln Val Asp Leu 2180	Phe Arg Asn Ala Arg 2185	Asn Gly Val Leu 2190
Ile Thr Glu Gly Ser Val 2195	Lys Gly Leu Thr Pro 2200	Ser Lys Gly Pro 2205
Ala Gln Ala Ser Val Asn 2210	Gly Val Thr Leu Ile 2215	Gly Glu Ser Val 2220
Lys Thr Gln Phe Asn Tyr 2225	Phe Lys Lys Val Asp 2230	Gly Ile Ile Gln 2235
Gln Leu Pro Glu Thr Tyr 2240	Phe Thr Gln Ser Arg 2245	Asp Leu Glu Asp 2250
Phe Lys Pro Arg Ser Gln 2255	Met Glu Thr Asp Phe 2260	Leu Glu Leu Ala 2265
Met Asp Glu Phe Ile Gln 2270	Arg Tyr Lys Leu Glu 2275	Gly Tyr Ala Phe 2280
Glu His Ile Val Tyr Gly 2285	Asp Phe Ser His Gly 2290	Gln Leu Gly Gly 2295
Leu His Leu Met Ile Gly 2300	Leu Ala Lys Arg Ser 2305	Gln Asp Ser Pro 2310
Leu Lys Leu Glu Asp Phe 2315	Ile Pro Met Asp Ser 2320	Thr Val Lys Asn 2325
Tyr Phe Ile Thr Asp Ala 2330	Gln Thr Gly Ser Ser 2335	Lys Cys Val Cys 2340
Ser Val Ile Asp Leu Leu 2345	Leu Asp Asp Phe Val 2350	Glu Ile Ile Lys 2355
Ser Gln Asp Leu Ser Val 2360	Ile Ser Lys Val Val 2365	Lys Val Thr Ile 2370
Asp Tyr Ala Glu Ile Ser 2375	Phe Met Leu Trp Cys 2380	Lys Asp Gly His 2385
Val Glu Thr Phe Tyr Pro 2390	Lys Leu Gln Ala Ser 2395	Gln Ala Trp Gln 2400
Pro Gly Val Ala Met Pro 2405	Asn Leu Tyr Lys Met 2410	Gln Arg Met Leu 2415
Leu Glu Lys Cys Asp Leu 2420	Gln Asn Tyr Gly Glu 2425	Asn Ala Val Ile 2430
Pro Lys Gly Ile Met Met 2435	Asn Val Ala Lys Tyr 2440	Thr Gln Leu Cys 2445
Gln Tyr Leu Asn Thr Leu 2450	Thr Leu Ala Val Pro 2455	Tyr Asn Met Arg 2460

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Val Ile	His Phe Gly Ala Gly	Ser Asp Lys Gly	Val Ala Pro Gly
2465	2470	2475	
Thr Ala	Val Leu Arg Gln Trp	Leu Pro Thr Gly Thr	Leu Leu Val
2480	2485	2490	
Asp Ser	Asp Leu Asn Asp Phe	Val Ser Asp Ala Asp	Ser Thr Leu
2495	2500	2505	
Ile Gly	Asp Cys Ala Thr Val	His Thr Ala Asn Lys	Trp Asp Leu
2510	2515	2520	
Ile Ile	Ser Asp Met Tyr Asp	Pro Arg Thr Lys His	Val Thr Lys
2525	2530	2535	
Glu Asn	Asp Ser Lys Glu Gly	Phe Phe Thr Tyr Leu	Cys Gly Phe
2540	2545	2550	
Ile Lys	Gln Lys Leu Ala Leu	Gly Gly Ser Ile Ala	Val Lys Ile
2555	2560	2565	
Thr Glu	His Ser Trp Asn Ala	Asp Leu Tyr Lys Leu	Met Gly His
2570	2575	2580	
Phe Ser	Trp Trp Thr Ala Phe	Val Thr Asn Val Asn	Ala Ser Ser
2585	2590	2595	
Ser Glu	Ala Phe Leu Ile Gly	Ala Asn Tyr Leu Gly	Lys Pro Lys
2600	2605	2610	
Glu Gln	Ile Asp Gly Tyr Thr	Met His Ala Asn Tyr	Ile Phe Trp
2615	2620	2625	
Arg Asn	Thr Asn Pro Ile Gln	Leu Ser Ser Tyr Ser	Leu Phe Asp
2630	2635	2640	
Met Ser	Lys Phe Pro Leu Lys	Leu Arg Gly Thr Ala	Val Met Ser
2645	2650	2655	
Leu Lys	Glu Asn Gln Ile Asn	Asp Met Ile Tyr Ser	Leu Leu Glu
2660	2665	2670	
Lys Gly	Arg Leu Ile Ile Arg	Glu Asn Asn Arg Val	Val Val Ser
2675	2680	2685	
Ser Asp	Ile Leu Val Asn Asn		
2690	2695		

<210> SEQ ID NO 76
 <211> LENGTH: 20
 <212> TYPE: DNA
 <213> ORGANISM: Artificial sequence
 <220> FEATURE:
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<400> SEQUENCE: 76

ccacacacag cttgtggata

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<210> SEQ ID NO 77
 <211> LENGTH: 20
 <212> TYPE: DNA
 <213> ORGANISM: Artificial sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: S/L4/+/6401 primer

<400> SEQUENCE: 77

ccgaagttgt aggcaatgtc

20

<210> SEQ ID NO 78
 <211> LENGTH: 20
 <212> TYPE: DNA
 <213> ORGANISM: Artificial sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: S/L4/+/6964 primer

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<400> SEQUENCE: 78

tttgggtgctc cttcttattg

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<210> SEQ ID NO 79

<211> LENGTH: 20

<212> TYPE: DNA

<213> ORGANISM: Artificial sequence

<220> FEATURE:

<223> OTHER INFORMATION: S/L4/-/6817 primer

<400> SEQUENCE: 79

ccggcatcca aacataattt

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<210> SEQ ID NO 80

<211> LENGTH: 20

<212> TYPE: DNA

<213> ORGANISM: Artificial sequence

<220> FEATURE:

<223> OTHER INFORMATION: S/L5/-/7633 primer

<400> SEQUENCE: 80

tggtcagtag ggttgattgg

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<210> SEQ ID NO 81

<211> LENGTH: 20

<212> TYPE: DNA

<213> ORGANISM: Artificial sequence

<220> FEATURE:

<223> OTHER INFORMATION: S/L5/-/8127 primer

<400> SEQUENCE: 81

catcctttgt gtcaacatcg

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<210> SEQ ID NO 82

<211> LENGTH: 20

<212> TYPE: DNA

<213> ORGANISM: Artificial sequence

<220> FEATURE:

<223> OTHER INFORMATION: S/L5/-/8633 primer

<400> SEQUENCE: 82

gtcacgagtg acaccatcct

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<210> SEQ ID NO 83

<211> LENGTH: 20

<212> TYPE: DNA

<213> ORGANISM: Artificial sequence

<220> FEATURE:

<223> OTHER INFORMATION: S/L5/+ /7839 primer

<400> SEQUENCE: 83

atgcgacgag tctgcttcta

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<210> SEQ ID NO 84

<211> LENGTH: 20

<212> TYPE: DNA

<213> ORGANISM: Artificial sequence

<220> FEATURE:

<223> OTHER INFORMATION: S/L5/+ /8785 primer

<400> SEQUENCE: 84

ttcatagtag ctggcttacc

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<210> SEQ ID NO 85

<211> LENGTH: 20

<212> TYPE: DNA

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<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L5/+ /8255 primer

<400> SEQUENCE: 85

atcttggcgc atgtattgac

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<210> SEQ ID NO 86
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L6/- /9422 primer

<400> SEQUENCE: 86

tgcattagca gcaacaacat

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<210> SEQ ID NO 87
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L6/- /9966 primer

<400> SEQUENCE: 87

tctgcagaac agcagaagtg

20

<210> SEQ ID NO 88
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L6/- /10542 primer

<400> SEQUENCE: 88

cctgtgcagt ttgtctgtca

20

<210> SEQ ID NO 89
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L6/+ /10677 primer

<400> SEQUENCE: 89

ccttgtggca atgaagtaca

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<210> SEQ ID NO 90
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L6/+ /10106 primer

<400> SEQUENCE: 90

atgtcatttg cacagcagaa

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<210> SEQ ID NO 91
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L6/+ /9571 primer

<400> SEQUENCE: 91

cttcaatggt ttgccatggt

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<210> SEQ ID NO 92
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L7/-/11271 primer

<400> SEQUENCE: 92

tgcgagctgt catgagaata

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<210> SEQ ID NO 93
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L7/-/11801 primer

<400> SEQUENCE: 93

aaccgagagc agtaccacag

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<210> SEQ ID NO 94
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L7/-/12383 primer

<400> SEQUENCE: 94

tttggctgct gtagtcaatg

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<210> SEQ ID NO 95
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L7/+/12640 primer

<400> SEQUENCE: 95

ctacgacaga tgtcctgtgc

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<210> SEQ ID NO 96
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
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<223> OTHER INFORMATION: S/L7/+/12088 primer

<400> SEQUENCE: 96

gagcaggctg tagctaattg

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<210> SEQ ID NO 97
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L7/+/11551 primer

<400> SEQUENCE: 97

ttaggctatt gttgctgctg

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<210> SEQ ID NO 98
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L8/-/13160 primer

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<400> SEQUENCE: 98

cagacaacat gaagcaccac

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<210> SEQ ID NO 99

<211> LENGTH: 20

<212> TYPE: DNA

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<220> FEATURE:

<223> OTHER INFORMATION: S/L8/-/13704 primer

<400> SEQUENCE: 99

cgctgacgtg atatatgtgg

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<210> SEQ ID NO 100

<211> LENGTH: 20

<212> TYPE: DNA

<213> ORGANISM: Artificial sequence

<220> FEATURE:

<223> OTHER INFORMATION: S/L8/-/14284 primer

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<210> SEQ ID NO 101

<211> LENGTH: 20

<212> TYPE: DNA

<213> ORGANISM: Artificial sequence

<220> FEATURE:

<223> OTHER INFORMATION: S/L8/+14453 primer

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<210> SEQ ID NO 102

<211> LENGTH: 20

<212> TYPE: DNA

<213> ORGANISM: Artificial sequence

<220> FEATURE:

<223> OTHER INFORMATION: S/L8/+13968 primer

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ggcattgtag gcgtactgac

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<210> SEQ ID NO 103

<211> LENGTH: 19

<212> TYPE: DNA

<213> ORGANISM: Artificial sequence

<220> FEATURE:

<223> OTHER INFORMATION: S/L8/+13401 primer

<400> SEQUENCE: 103

gtttgcggtg taagtgcag

19

<210> SEQ ID NO 104

<211> LENGTH: 20

<212> TYPE: DNA

<213> ORGANISM: Artificial sequence

<220> FEATURE:

<223> OTHER INFORMATION: S/L9/-/15098 primer

<400> SEQUENCE: 104

tagtggcggc tattgacttc

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<210> SEQ ID NO 105

<211> LENGTH: 20

<212> TYPE: DNA

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<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L9/-/15677 primer

<400> SEQUENCE: 105

ctaaaccttg agccgcatag

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<210> SEQ ID NO 106
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L9/-/16247 primer

<400> SEQUENCE: 106

catggtcata gcagcacttg

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<210> SEQ ID NO 107
<211> LENGTH: 21
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L9/+16323 primer

<400> SEQUENCE: 107

ccagggtgtg atgtcactga t

21

<210> SEQ ID NO 108
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L9/+15858 primer

<400> SEQUENCE: 108

ccttaccag atccatcaag

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<210> SEQ ID NO 109
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L9/+15288 primer

<400> SEQUENCE: 109

cgcaaacata acacttgctg

20

<210> SEQ ID NO 110
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L10/-/16914 primer

<400> SEQUENCE: 110

agtggtgggt acaagccagt

20

<210> SEQ ID NO 111
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L10/-/17466 primer

<400> SEQUENCE: 111

gttccaagga acatgtctgg

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<210> SEQ ID NO 112
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L10/-/18022 primer

<400> SEQUENCE: 112

aggtgcctgt gtaggatgaa 20

<210> SEQ ID NO 113
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L10/+18245 primer

<400> SEQUENCE: 113

gggctgtcat gcaactagag 20

<210> SEQ ID NO 114
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L10/+17663 primer

<400> SEQUENCE: 114

tcttacacgc aatcctgctt 20

<210> SEQ ID NO 115
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L10/+17061 primer

<400> SEQUENCE: 115

taccatctg ctgcatagt 20

<210> SEQ ID NO 116
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L11/-/18877 primer

<400> SEQUENCE: 116

gcaagcagaa ttaaccctca 20

<210> SEQ ID NO 117
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L11/-/19396 primer

<400> SEQUENCE: 117

agcaccacct aaattgcatc 20

<210> SEQ ID NO 118
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L11/-/20002 primer

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<400> SEQUENCE: 118

tggtcccttt gaagtggtta

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<210> SEQ ID NO 119

<211> LENGTH: 20

<212> TYPE: DNA

<213> ORGANISM: Artificial sequence

<220> FEATURE:

<223> OTHER INFORMATION: S/L11/+20245 primer

<400> SEQUENCE: 119

tcgaacacat cgtttatgga

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<210> SEQ ID NO 120

<211> LENGTH: 20

<212> TYPE: DNA

<213> ORGANISM: Artificial sequence

<220> FEATURE:

<223> OTHER INFORMATION: S/L11/+19611 primer

<400> SEQUENCE: 120

gaagcacctg tttccatcat

20

<210> SEQ ID NO 121

<211> LENGTH: 20

<212> TYPE: DNA

<213> ORGANISM: Artificial sequence

<220> FEATURE:

<223> OTHER INFORMATION: S/L11/+19021 primer

<400> SEQUENCE: 121

acgatgctca gccatgtagt

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<210> SEQ ID NO 122

<211> LENGTH: 20

<212> TYPE: DNA

<213> ORGANISM: Artificial sequence

<220> FEATURE:

<223> OTHER INFORMATION: SARS/L1/F3/+800 primer

<400> SEQUENCE: 122

gaggtgcagt cactcgctat

20

<210> SEQ ID NO 123

<211> LENGTH: 20

<212> TYPE: DNA

<213> ORGANISM: Artificial sequence

<220> FEATURE:

<223> OTHER INFORMATION: SARS/L1/F4/+1391 primer

<400> SEQUENCE: 123

cagagattgg acctgagcat

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<210> SEQ ID NO 124

<211> LENGTH: 20

<212> TYPE: DNA

<213> ORGANISM: Artificial sequence

<220> FEATURE:

<223> OTHER INFORMATION: SARS/L1/F5/+1925 primer

<400> SEQUENCE: 124

cagcaaacca ctcaattcct

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<210> SEQ ID NO 125

<211> LENGTH: 20

<212> TYPE: DNA

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<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: SARS/L1/R3/-/1674 primer

<400> SEQUENCE: 125

aaatgatggc aacctcttca 20

<210> SEQ ID NO 126
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: SARS/L1/R4/-/1107 primer

<400> SEQUENCE: 126

cacgtgggtg aatgactttg 20

<210> SEQ ID NO 127
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: SARS/L1/R5/-/520 primer

<400> SEQUENCE: 127

atttctgcaa ccagctcaac 20

<210> SEQ ID NO 128
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: SARS/L2/F3/+2664 primer

<400> SEQUENCE: 128

cgcatgtgtc cctggtttac 20

<210> SEQ ID NO 129
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: SARS/L2/F4/+3232 primer

<400> SEQUENCE: 129

gagattgagc cagaaccaga 20

<210> SEQ ID NO 130
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: SARS/L2/F5/+3746 primer

<400> SEQUENCE: 130

atgagcaggt tgtcatggat 20

<210> SEQ ID NO 131
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<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: SARS/L2/R3/-/3579 primer

<400> SEQUENCE: 131

ctgccttaag aagctggatg 20

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<210> SEQ ID NO 132
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: SARS/L2/R4/-/2991 primer

<400> SEQUENCE: 132

tttcttcacc agcatcatca

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<210> SEQ ID NO 133
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<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: SARS/L2/R5/-/2529 primer

<400> SEQUENCE: 133

caccgttctt gagaacaacc

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<210> SEQ ID NO 134
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: SARS/L3/F3+/4708 primer

<400> SEQUENCE: 134

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<210> SEQ ID NO 135
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: SRAS/L3/F4+/5305 primer

<400> SEQUENCE: 135

gctggtgatg ctgctaactt

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<210> SEQ ID NO 136
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: SARS/L3/F5+/5822 primer

<400> SEQUENCE: 136

ccatcaagcc tgtgtcgat

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<210> SEQ ID NO 137
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: SARS/L3/R3/-/5610 primer

<400> SEQUENCE: 137

caggtggtgc agacatcata

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<210> SEQ ID NO 138
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: SARS/L3/R4/-/4988 primer

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<400> SEQUENCE: 138

aacatcagca ccatccaagt

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<210> SEQ ID NO 139

<211> LENGTH: 20

<212> TYPE: DNA

<213> ORGANISM: Artificial sequence

<220> FEATURE:

<223> OTHER INFORMATION: SARS/L3/R5/-/4437 primer

<400> SEQUENCE: 139

atcggacacc atagtcaacg

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<210> SEQ ID NO 140

<211> LENGTH: 7788

<212> TYPE: DNA

<213> ORGANISM: Artificial sequence

<220> FEATURE:

<223> OTHER INFORMATION: synthetic S gene

<400> SEQUENCE: 140

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gcagtacatc tacgtattag tcatcgctat taccatgggtg atgcgggtttt ggcagtacac	540
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caatgggagt ttgttttgge accaaaatca acgggacttt ccaaaatgtc gtaataaccc	660
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tgacatccac ttgaccttc tctccacagg tgtccactcc cagttcaatt acagctctta	1020
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caccagaccc cacacatga tcttcgacaa cgccttcaac tgcaccttcg agtacatcag	1560
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cgtgttcaag aacaaggacg gcttcctgta cgtgtacaag ggctaccagc ccatcgacgt	1680
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catcaacatc accaacttcc gggccatcct gaccgccttt agccctgccc aggacatctg	1800
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45

The invention claimed is:

1. A method for the detection of a SARS-associated coronavirus infection, from a biological sample, by indirect IgG ELISA using the SARS-associated coronavirus N protein, which comprises providing ELISA plates that have been sensitized with a solution consisting of N protein at a concentration of between 0.5 and 4 µg/ml in a 10 mM PBS buffer, pH 7.2, phenol red at 0.25 ml/l.

2. A method for the detection of a SARS-associated coronavirus infection, from a biological sample, by double epitope ELISA, comprising mixing a serum to be tested with the antigen attached to a solid support, wherein said antigen is a SARS-associated coronavirus N protein and wherein said solid support is sensitized with a solution consisting of N protein at a concentration of between 0.5 and 4 µg/ml in a 10 mM PBS buffer, pH 7.2, phenol red at 0.25 ml/l.

3. The method as claimed in claim 2, wherein said N protein is at a concentration of 1 µg/ml.

4. The method as claimed in claim 1, wherein said biological sample is collected 12 days or more after said infection.

5. The method as claimed in claim 2, wherein said biological sample is collected 12 days or more after said infection.

6. The method as claimed in claim 1, wherein said N protein is at a concentration of 2 µg/ml.

7. The method as claimed in claim 2, wherein said visualizing antigen consists of said SARS-associated coronavirus N protein conjugated to a visualizing molecule selected from the group consisting of a radioactive atom, a dye, a fluorescent molecule, a fluorophore, and an enzyme.

8. The method as claimed in claim 7, wherein said enzyme is a peroxidase.

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